



Enfermedades Infecciosas y Microbiología Clínica

www.elsevier.es/eimc



Letter to the Editor

Antigen-detecting rapid tests or real-time PCR, what test to use and why?[☆]



Test rápidos antigénicos o PCR en tiempo real para SARS-CoV-2, ¿qué test usar y por qué?

Dear Editor,

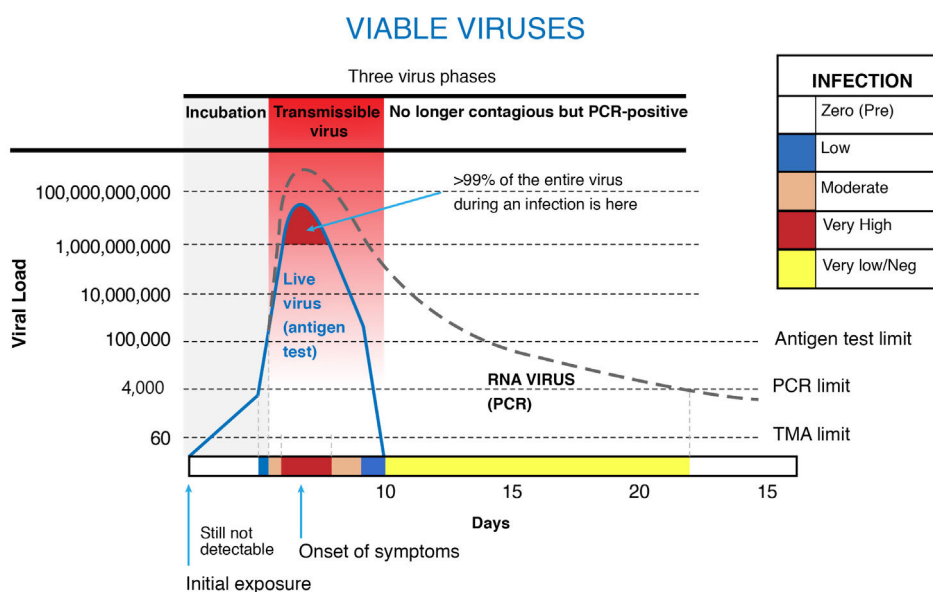
We read with interest the letter by Marco et al.¹ concerning the low sensitivity of rapid antigen tests (RAT) to screen for SARS-CoV-2 infection.

The authors reported an outbreak of SARS-CoV-2 in a prison. Having diagnosed three cases by RAT, they screened a total of 81 inmates, with a SARS-CoV-2 infection incidence of 11% (9/81) by RAT. Between three and five days later, the 72 negative cases were screened again by real-time PCR (rt-PCR), 37% of which (27/72) tested positive. The authors labelled the previous RAT results as false negatives, concluding that rt-PCR should be the SARS-CoV-2 screening technique of choice given the low sensitivity of RATs.

It is vital to understand certain important points concerning RAT screening strategies, such as how to correctly interpret the results, when their use is indicated and their advantages over rt-PCR.

RATs have a high sensitivity for detecting individuals with a high viral load who could potentially transmit the virus (symptomatic and asymptomatic)². This sensitivity is dependent on the viral load of the patient, which could be related to the 'cycle threshold' (Ct) of rt-PCR, an indirect marker of viral load in a infected subject. RATs are effective tools for diagnosing infected subjects with Cts <25, which are correlated with viruses that grow in cell cultures and are transmissible^{3,4}, having shown sensitivity rates approaching 100%². As a result, RATs may often be negative from the 5th day of symptoms onset (or 10th day since exposure) and universally in subjects with a low viral load.

The rt-PCR testing conducted between three and five days after the initial screening yielded a positivity rate of 37% in subjects with a prior negative RAT. SARS-CoV-2 nucleic acid amplification testing (NAAT) with rt-PCR or TMA (transcription-mediated amplification) detects positive results several days (in the case of rt-PCR)



Adapted from Michael Mina, MD, PhD. Harvard T.H. Chan School of Public Health/Medical School.

Fig. 1. SARS-CoV-2 phases of infection.

DOI of original article: <https://doi.org/10.1016/j.eimc.2021.06.001>

[☆] Please cite this article as: Revollo Barriga B, Llibre Codina JM. Test rápidos antigénicos o PCR en tiempo real para SARS-CoV-2, ¿qué test usar y por qué? *Enferm Infecc Microbiol Clin.* 2021;39:531–532.

or weeks (in the case of TMA) after the abatement of symptoms, when subjects are no longer infectious.

As such, RAT and rt-PCR detect different stages of the disease. RATs yield positive results over a shorter period of time during the acute phase of infection.

An analysis of the SARS-CoV-2 viral kinetics (Fig. 1) reveals that high-frequency rapid antigen test screening strategies are just as effective in detecting infectious individuals as a low-frequency rt-PCR testing regimen⁵. The difference lies in the characteristics of each test: unlike rt-PCR, RATs are “versatile” tests that can be used anywhere, are inexpensive and return results in 15 min. Applying a RAT screening strategy could identify the same number of infectious individuals as rt-PCR.

Returning to the report by Marco et al., it would be useful to know the Cts of the rt-PCR-positive samples three and five days after the initial test, primarily to ascertain the patients' actual risk of virus transmission. Cts greater than 25–30 represent a low risk of virus transmission and a high probability of a negative RAT result, regardless of whether the subject is symptomatic or asymptomatic. A negative RAT followed by a positive rt-PCR in already isolated subjects essentially suggests that the Ct is elevated and the final phase of infection or a resolved infection is being detected.

This is particularly true of RAT, an ultrasensitive technique that detects up to 60 copies of SARS-CoV-2 (as opposed to 3,000–5,000 copies for rt-PCR)⁶. RAT is currently the most commonly used technique in mass screening strategies because samples can be pooled in the laboratory and because positive results can be returned for up to eight weeks after infection.

In conclusion, we believe that RAT and NAAT (rt-PCR or TMA) detect different phenomena. If you want a simple way to identify subjects with the potential to transmit SARS-CoV-2, RATs are the ideal tool. However, if you would like to screen a cross-sectional cohort that identifies the largest possible number of subjects infected (current and recent), NAAT would be the technique that detects the most number of cases.

Reply to «Antigen-detecting rapid tests or real-time PCR, what test to use and why?»^{*}



Respuesta a «Test rápidos antigénicos o PCR en tiempo real para SARS-CoV-2, ¿qué test usar y por qué?»

Dear Editor,

We appreciate Revollo and Llibre's comments¹ on the letter recently published by our group on an outbreak of SARS-CoV-2 infection in Figueras prison (Girona)². As a reminder, infection was detected by rapid antigen test (RAT) in three mildly symptomatic inmates between 23 and 25 December. As a result, in the afternoon of 25 December, the 81 remaining inmates of that prison block were screened using RAT and nine positive results were identified. On 28 December, the 72 inmates who tested negative by RAT underwent rt-PCR testing, 27 (37.5%) of which were positive. The sensitivity of the RAT in this scenario was very low at just 25%, which is why we reported it.

DOI of original article: <https://doi.org/10.1016/j.eimc.2021.06.015>.

^{*} Please cite this article as: Marco A, Solé C, Abdo JJ, Turu E. Respuesta a «Test rápidos antigénicos o PCR en tiempo real para SARS-CoV-2, ¿qué test usar y por qué?». *Enferm Infecc Microbiol Clin.* 2021;39:532–533.

Conflicts of interest

The authors declare that they have no conflicts of interest.

References

1. Marco A, Solé C, Abdo JJ, Turu E. Low sensitivity of rapid antigenic tests as a screening method in an outbreak of SARS-CoV-2 infection in prison. *Enferm Infecc Microbiol Clin.* 2021. <http://dx.doi.org/10.1016/j.eimc.2021.01.016>.
2. Pray IW, Ford L, Cole D, Lee C, Bigouette JP, Abedi GR, et al. Performance of an antigen-based test for asymptomatic and symptomatic SARS-CoV-2 testing at two University Campuses – Wisconsin, September–October 2020. *MMWR Morb Mortal Wkly Rep.* 2021;69:1642–7.
3. Porte L, Legarraga P, Vollrath V, Aguilera X, Munita JM, Araos R, et al. Evaluation of a novel antigen-based rapid detection test for the diagnosis of SARS-CoV-2 in respiratory samples. *Int J Infect Dis.* 2020;99:328–33.
4. Alemany A, Baro B, Ouchi D, Ubals M, Corbacho-Monné M, Rodon J, et al. Analytical and Clinical Performance of the Panbio COVID-19 antigen-detecting rapid diagnostic test. *J Infect.* 2021 [Accessed 8 January 2021]. Available from: <https://doi.org/10.1101/2020.10.30.20223198>
5. Mina MJ, Parker R, Larremore DB. Rethinking Covid-19 test sensitivity – a strategy for containment. *N Engl J Med.* 2020;383:e120.
6. Chan JF-W, Yip CC-Y, To KK-W, Tang TH-C, Wong SC-Y, Leung K-H, et al. Improved molecular diagnosis of COVID-19 by the novel, highly sensitive and specific COVID-19-RdRp/Hex real-time reverse transcription-PCR assay validated in vitro and with clinical specimens. *J Clin Microbiol.* 2020;58(5).

Boris Revollo Barriga*, Josep M. Llibre Codina

Servicio de Enfermedades Infecciosas y Fundación Lucha Contra el Sida y Enfermedades Infecciosas, Hospital Universitario Germans Trias i Pujol, Badalona, Spain

* Corresponding author.

E-mail address: brevollo@flsida.org (B. Revollo Barriga).

<https://doi.org/10.1016/j.eimc.2021.09.008>

2529-993X/ © 2021 Sociedad Española de Enfermedades Infecciosas y Microbiología Clínica. Published by Elsevier España, S.L.U. All rights reserved.

For reasons of brevity, we did not include any information about the population studied in our original letter, which, according to Revollo and Llibre's comments, could be relevant. Since 1 July 2020, new prisoners in Catalonia have been screened by rt-PCR. In total, 46.2% of those infected by the outbreak had been incarcerated after that date and had a prior negative rt-PCR test. The rest of the infected inmates had been in prison for many months and had not been diagnosed with SARS-CoV-2 infection nor monitored due to close contact with an infected individual. As such, the risk of there being a persistently positive or residual rt-PCR result in an infected inmate, as raised by Revollo and Llibre, we consider to be extremely small. Regarding the use of rt-cycle thresholds (Ct) that Revollo and Llibre also discuss, their use in initial phases of infection is low as the values vary over time³. In fact, we only use them very rarely, almost exclusively to assess infection risk in cases with persistently positive PCR results that require prolonged isolation, as discharge without knowing whether or not the subject is infectious is a risk in a confined environment.

We agree with Revollo and Llibre's assessment of RATs' high sensitivity for detecting symptomatic cases with a high viral load and transmission potential, typically in the first five days. However, current data are not as conclusive when it comes to their use in pre-symptomatic or asymptomatic patients. The Centers for