



# Enfermedades Infecciosas y Microbiología Clínica

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Review article

## Management of infection and febrile neutropenia in patients with solid cancer<sup>☆, □, ▲</sup>



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### ABSTRACT

A group of experts from the Spanish Society of Infectious Diseases and Clinical Microbiology (SEIMC) and the Spanish Society of Medical Oncology (SEOM) have reviewed in this paper the main aspects to be considered in the evaluation of patients with solid cancer and infectious diseases. They have established a series of recommendations on the prevention of the most prevalent infections in these patients, the use of vaccines, the control measures of vascular catheter infection and prevention of infections before certain surgical procedures. Also the criteria for management of febrile neutropenia and the use of colony-stimulating factors were revised. Finally they provide a series of recommendations for the treatment of cancer patients with severe infection. The document is completed with a series of measures for the control of hospital infection.

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## Manejo de la infección y la neutropenia febril en el paciente con cáncer sólido

### RESUMEN

#### Palabras clave:

Cáncer

Neutropenia febril

Infección

Profilaxis

Factores de riesgo

Un grupo de expertos de la Sociedad Española de Enfermedades Infecciosas y Microbiología Clínica (SEIMC) y de la Sociedad Española de Oncología Médica (SEOM) han revisado en este documento los principales aspectos que deben considerarse en la evaluación de los pacientes con cáncer sólido y complicaciones infecciosas. Para ello se han establecido unas recomendaciones sobre la profilaxis de las infecciones más prevalentes en estos pacientes, el uso de vacunas, las medidas de control de la infección por catéteres vasculares y la prevención de la infección ante determinadas maniobras quirúrgicas.

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A continuación, se han revisado los criterios de manejo de la neutropenia febril y del uso de factores estimulantes de colonias, para terminar dando una serie de pautas sobre el tratamiento del paciente oncológico con infección grave. El documento se completa con una serie de medidas para el control de la infección hospitalaria.

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## Introduction

Over the last two decades, notable advances have been made as regards the treatment of cancer patients. Undoubtedly, one of the most noteworthy has been the reduction in morbidity and mortality due to infectious complications, as a result of the progress achieved in preventing and treating these infections, as well as a decrease in the duration of neutropenia thanks to the use of haematopoietic growth factors.

Despite these advances, infectious complications continue to be one of the main causes of death in cancer patients. These patients are at a greater risk of certain infections being reactivated and also have an increased risk of suffering nosocomial infections as a result of surgery, the use of venous or urinary catheters and other devices, as well as the procedures that they undergo. The emergence of multidrug-resistant microorganisms in recent years has made an antibiotic approach difficult in these patients. Moreover, the increasingly frequent use of new monoclonal antibodies and biological therapies has increased the risk of these patients suffering a number of severe infections.

Although there are various clinical guidelines intended for haematological patients, there are few that specifically address those with a solid tumour. As such, experts from the Spanish Society of Infectious Diseases (SEIMC) and the Spanish Society of Medical Oncology (SEOM) have decided to produce this document, which reviews existing information on the topic and makes a series of recommendations based on the best available evidence, for use by oncologists and infectious diseases specialists in daily clinical practice.

## Initial cancer patient assessment

The initial assessment of the cancer patient seeks to detect active or latent infections that are at risk of reactivation in those with a solid cancer who are due to receive a potentially immunosuppressive treatment.

The clinical assessment should include: (1) previous history of infectious diseases that may have remained latent and be reactivated in the event of immunosuppression; (2) complete epidemiological history, including contact with patients suffering from an infectious disease, as well as with other immunocompromised patients; (3) patient origin and stays in or trips to foreign countries with endemic diseases that could be potentially reactivated; and (4) history of any drug reactions to antibiotics. In women, a gynaecological examination should also be advised, along with human papillomavirus (HPV) screening.

The initial microbiological assessment is intended to screen for the main chronic or latent infections that may be reactivated in the event of the patient's immunosuppression, and will depend on the type of chemotherapy treatment received as well as the specific immunosuppression risk in each cancer patient. In indicated cases, according to the treatment and immunosuppression risk, serological testing for the following viruses would be useful: (1) hepatitis A, B and C (HAV, HBV and HCV); (2) varicella zoster virus (VZV); and (3) human immunodeficiency virus (HIV). Moreover, the existence of latent tuberculosis (TB) should be ruled out by means of a Mantoux test and/or an interferon gamma release assay (IGRA) in

all patients with a suspected history of the disease, those in contact with affected patients or in high-risk populations such as institutionalised patients. In patients from certain geographical areas, a number of regional diseases should be taken into account ([Table 1](#)).

## Preventing infection

### Vaccination

The vaccines indicated in solid cancer patients are described in [Table 2](#) and should be inactivated vaccines.<sup>1</sup> Vaccines with live attenuated microorganisms such as rotavirus, MMR (measles, mumps and rubella) and varicella are contraindicated during chemotherapy.<sup>2</sup>

Patients with active solid tumours and those undergoing treatment with chemotherapy should receive an annual flu vaccine.<sup>2</sup> It is recommended that patients be vaccinated against pneumococcus according to the guidelines established for immunocompromised patients.

Depending on the aforementioned characteristics (type and duration of chemotherapy, clinical situation of the patient), a tetanus-diphtheria booster dose is advisable. Patients who have not been vaccinated against whooping cough are recommended to have the tetanus-diphtheria-pertussis vaccine (Tdap). Likewise, vaccination against HPV, meningococcus and HAV should be considered provided there is a specific indication. The administration of the HBV vaccine should be considered in unimmunised patients after evaluating their serological and clinical situation.

Patients should receive the indicated vaccines before beginning chemotherapy. As for inactivated vaccines, their administration is recommended at least two weeks before the start of treatment (except the flu vaccine, which will be administered annually, even during the chemotherapy regimen), while live attenuated vaccines should be given at least four weeks prior to commencing treatment.<sup>3</sup>

### Preventing hepatitis B

HBV screening is especially important in patients who are deemed high-risk (e.g. those treated with everolimus, temozolamide, rituximab, etc.) and should be considered, according to medical judgement, in other patients. Screening will be carried out by detecting the surface antigen (HBsAg), the hepatitis B core antibody (anti-HBc) and the hepatitis B surface antibody (anti-HBs). If they all come back negative, there is no infection and the patient should be vaccinated before beginning immunosuppressive therapy. If a patient tests positive for HbsAg, the study should be completed with the determination of the viral load, e antigen (HBeAg), liver function tests and a liver biopsy, if applicable. Based on the results, it can be determined whether the patient has chronic hepatitis, is in the immunotolerant phase or is an inactive carrier of HBV. In case of chronic hepatitis, the patient should receive antiviral treatment with entecavir or tenofovir. In the other two cases, the patient should receive antiviral prophylaxis.

If HBsAg is negative and anti-HBc positive, this indicates resolved hepatitis B. In this case, regardless of the anti-HBs result, viral deoxyribonucleic acid (DNA) levels should be determined. If

**Table 1**

Regional or imported diseases according to geographical area of origin.

Countries of origin	Possible microorganisms	Screening technique
Mexico, Panama, Venezuela, Guatemala or southern USA	<i>Histoplasma capsulatum</i>	Serology
Southern USA, Mexico, Guatemala, Honduras, Nicaragua, Argentina, Paraguay, Venezuela and Colombia	<i>Coccidioides immitis</i>	Serology
Caribbean, South west Japan, Central and South America, Sub-Saharan Africa	<i>HTLV-I-II</i>	Serology
Mexico, Central America or Southern Cone (Chile, Argentina, Bolivia, Brazil, Paraguay)	<i>Trypanosoma cruzi</i>	Two serological techniques
Tropical and subtropical areas, including southern USA	<i>Strongyloides stercoralis</i>	Agar technique-Stool culture
Malaria-endemic areas in the past 2–5 years: rule out asymptomatic parasitaemia	<i>Plasmodium spp.</i>	Serology PCR Coarse droplet

HTLV-I-II, human T-lymphotropic virus, types I and II; PCR, polymerase chain reaction.

**Table 2**

Recommended vaccines in adults with solid tumours.

Vaccine	Recommendation	Regimen
Pneumococcus	Recommended	1st dose (VNC13) on diagnosis before starting treatment; subsequent doses: one dose of VNP23 after 8 weeks
Flu	Recommended	Annually
Hepatitis A	Only in case of risk factors	1st dose on diagnosis; 2nd dose after 6–12 months
Hepatitis B	Recommended in unimmunised patients	1st dose: month 0; 2nd dose: 1st month; 3rd dose: 6th month
Tetanus-diphtheria-acellular pertussis (Tdap)	Booster TD or Tdap dose if no previous vaccine against whooping cough	–
Human papillomavirus (HPV)	If due on immunisation schedule	1st dose: month 0; 2nd dose: 1st or 2nd month; 3rd dose: 6th month
Meningococcus	Only in case of risk factors	–

the viral load is positive, the infection is occult and the patient should therefore receive prophylaxis. If the viral load is negative, the possibility of reactivation should be checked periodically throughout the immunosuppressive treatment for early detection and in order to initiate treatment as soon as possible. This monitoring will be performed by determining the liver biochemistry, HbsAg and/or viral load. In high-risk patients, most authors feel a prophylaxis regimen should be instituted immediately.<sup>4,5</sup> For patients with no HBV risk factors, cancer treatment is not expected to activate the risk of disease; current evidence does not support HBV detection before starting anti-cancer treatment.<sup>6</sup>

#### Preventing tuberculosis

Once active disease has been ruled out, all patients who meet one or more of the following criteria should receive prophylaxis against TB<sup>4–10</sup>: (1) positive purified protein derivative (PPD) skin test ( $\geq 5$  mm); (2) positive IGRA test; (3) history of incorrectly-treated TB; (4) radiological findings suggestive of residual TB lesions, such as apical fibronodular lesions, pleural thickening, etc.; or (5) contact with a patient with active TB. The routine regimens are to be used, with the known precautions.

#### Preventing central venous catheter infection

At present, there is not enough evidence to recommend a specific type of long-term central venous catheter (CVC)—either a tunneled CVC (Hickman), “port-a-cath” (PAC) or a peripherally inserted CVC (PICC)—and there is no specific insertion site, although femoral access is generally discouraged due to a greater risk of infection.<sup>8</sup>

The most important measures in the prevention of CVC infections are: (i) education and training of healthcare professionals; (ii) strict hand hygiene; and (iii) the use of aseptic techniques when setting and changing dressings.<sup>9</sup> Routine CVC changes and the application of topical antimicrobial agents at the insertion site

are not recommended, as they may encourage fungal infections and the emergence of resistances. The use of CVCs impregnated or coated with antimicrobial/antiseptic agents such as chlorhexidine and silver sulfadiazine or minocycline/rifampicin and/or heparin-impregnated catheters may reduce the risk of infections, although their benefit is relative and the cost high.<sup>10</sup> The prophylactic administration of antibiotics before the placement of a CVC has not been shown to reduce the incidence of infections.<sup>11</sup>

#### Preventing infection after endoscopic procedures

The administration of prophylactic antibiotics is not generally recommended before an endoscopic procedure in order to avoid the development of bacterial endocarditis as cases are rare and there are insufficient data supporting their relationship and the utility of antibiotics in this context.<sup>12</sup>

In case of an endoscopic retrograde cholangiopancreatography (ERCP), the administration of prophylactic antibiotic therapy should be considered to cover Gram-negative enteric bacilli and enterococci in patients with a blockage in whom full biliary drainage may not be possible. If the procedure does not resolve the blockage, continued antibiotic therapy is advised.<sup>12</sup> In percutaneous endoscopic gastrostomies (PEG), the administration of antibiotics (cefazolin, 1 g iv; 30 min before the procedure) has been proven to significantly reduce the risk of infection.<sup>13</sup>

#### Preventing infection by *Pneumocystis jiroveci*

Prophylaxis should be considered in the face of *Pneumocystis jiroveci* (*P. jiroveci*) in patients who are due to receive: (1) temozolamide with radiotherapy; (2) drugs that cause profound T-cell lymphocytopenia; and (3) steroids at a dose equivalent to  $\geq 20$  mg/day of prednisone for four or more weeks.<sup>14</sup>

The regimen of choice is co-trimoxazole (800/160 mg, one tablet, three times per week). In case of an allergy to co-trimoxazole,

the possibility of desensitisation should be considered.<sup>15,16</sup> Alternatively, atovaquone (1.5 g/day)<sup>17</sup> or dapsone (100 mg/day) may be used, although glucose-6-phosphate dehydrogenase deficiency (G6PD)<sup>18</sup> should be ruled out, or pentamidine (300 mg, four times per week or monthly iv).<sup>19,20</sup> Prophylaxis should be maintained at least throughout the chemotherapy treatment, and prolonging it for at least two months—or until the CD4 lymphocyte count is above 200 U/mm<sup>3</sup>—is recommended.

#### Special situations

Given the current characteristics of the resident Spanish population and frequent links between different geographical areas, the prevention of *Strongyloides stercoralis* hyperinfestation<sup>20</sup> and Chagas disease (*Trypanosoma cruzi*)<sup>21</sup> should be taken into account.

#### Prevention with granulocyte-colony stimulating factors

The prophylactic administration of granulocyte-colony stimulating factors (G-CSF) reduces the incidence, duration and severity of neutropenia and prevents associated infections.<sup>22</sup> As such, the risk of febrile neutropenia (FN) should be estimated before initiating chemotherapy, taking into account different factors, such as the type of tumour, the chemotherapy regimen applied, the patient's characteristics and the treatment intention. The prophylactic use of G-CSFs is recommended in patients with a estimated FN risk of over 20%.<sup>23,24</sup> If the estimated risk lies between 10% and 20%, an individual assessment is advised, with G-CSF administration primarily being considered if the treatment intention is curative, in order to avoid delays and dosage reductions, or in high-risk patients, such as those over 65 years of age with previous episodes of FN, extensive bone marrow involvement or who have recently undergone extensive surgery, especially if this includes an intestinal resection. Prophylactic use is more controversial in patients with very advanced tumours, a fragile general or nutritional status, significant comorbidities or in those in whom the benefit of chemotherapy and maintaining a dose intensity is dubious. Routine G-CSF use is not indicated in patients with a risk below 10%, except in specific circumstances that may entail serious consequences in the event of FN.

In FN, treatment with G-CSFs reduces the length of the patient's hospital stay and the neutrophil recovery time. However, it is not associated with a patient survival benefit.<sup>25,26</sup> As such, administration should be considered in cases associated with a high complication risk, as occurs with severe neutropenia (neutrophils <100/mm<sup>3</sup>) or when a long duration is expected (>10 days). Moreover, the use of G-CSFs should be considered in patients aged over 65 years, in cases of sepsis, pneumonia, invasive fungal infections, hospitalisation at the time of fever onset or previous episodes of FN.<sup>27</sup>

#### Antibiotic prophylaxis

Patients with solid tumours receiving conventional chemotherapy are thought to have a low risk of suffering infectious complications.<sup>25,28</sup> In these patients, fluoroquinolones have a protective effect<sup>29,30</sup> but do not reduce mortality. In high-risk patients, fluoroquinolones have demonstrated their efficacy in preventing infections in the neutropenic phase,<sup>30</sup> particularly in the first chemotherapy cycle.<sup>31</sup> Given the high number of patients requiring treatment to prevent an infection, the cost, the adverse effects, the onset of superinfections and the range of resistances,<sup>32–37</sup> antibacterial prophylaxis is contraindicated in low-risk patients receiving conventional chemotherapy with or without biological agents.<sup>34,36</sup> In specific situations, such as during the first chemotherapy cycle, when profound and prolonged neutropenia is expected, with very

aggressive cytostatic regimens, when there is a high base morbidity or in elderly patients, its administration will be considered on an individual basis.<sup>38,39</sup>

#### Febrile neutropenia

##### Assessing infection risk in a patient with febrile neutropenia

The rate of infectious complications in patients with FN is 25–30%, with mortality reaching 11% in some groups.<sup>27</sup> However, this risk is not homogeneous, so the overtreatment of low-risk episodes is common.<sup>39</sup> The objective of assessing the infection risk in these patients is to predict the risk of serious complications and thus the need for hospital admission and parenteral therapy. The initial assessment should include the evaluation of: (1) systemic inflammatory response data, by means of checking vital signs such as temperature, heart and respiratory rate; (2) severe sepsis data such as hypotension, signs of tissue hypoperfusion or acute organ dysfunction; and (3) existence of primary or secondary infection foci, taking into account the clinical and epidemiological context.

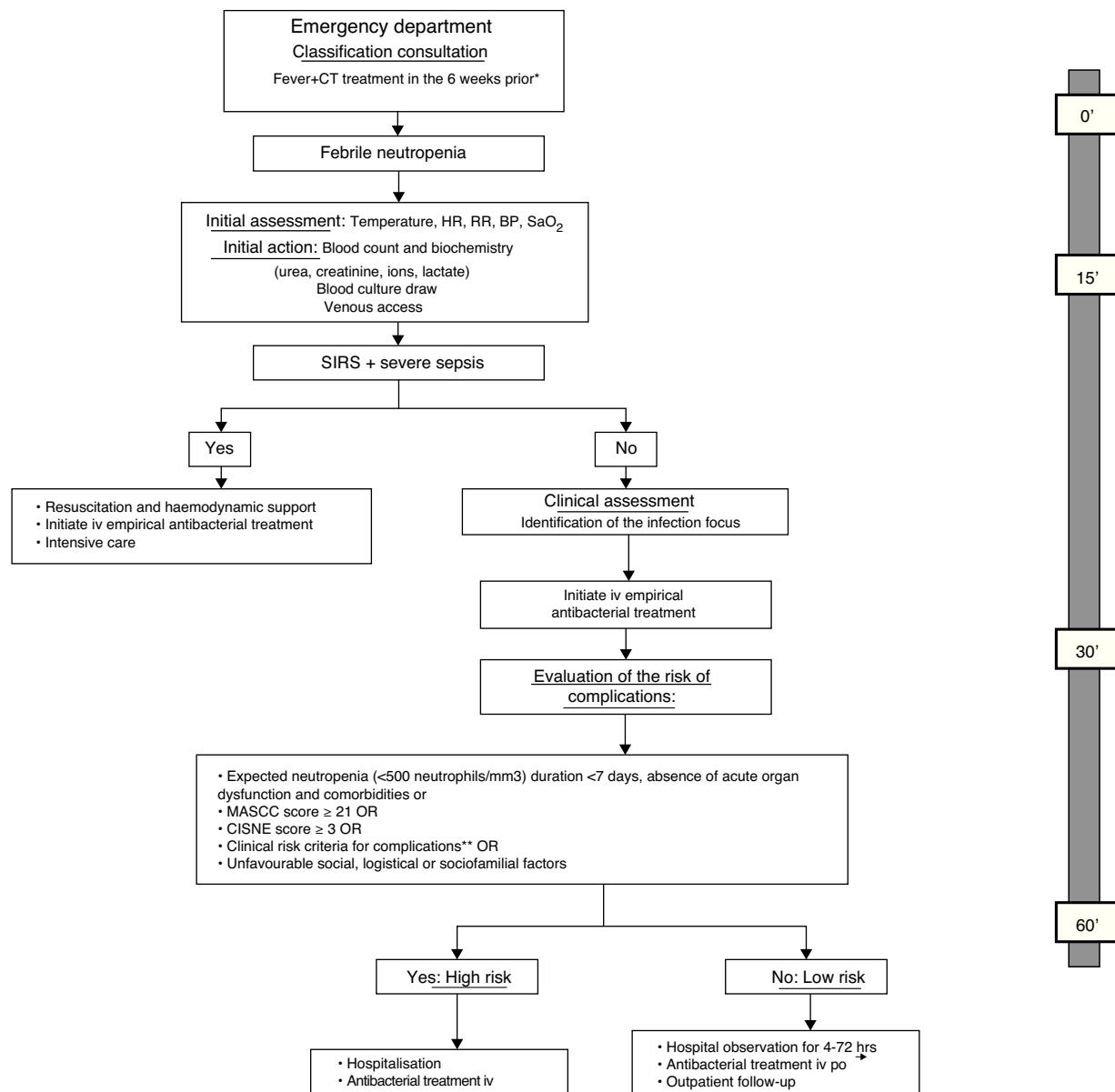
The most widely validated prognostic tool is the Multinational Association of Supportive Care in Cancer (MASCC)<sup>40</sup> score, although this is not specific to patients with solid tumours, and infectious complications may develop in 9–15% of episodes classified as low-risk.<sup>40–42</sup> The selection of patients in clinical trials on oral/outpatient treatments has been based on exclusion criteria, with the results deemed to have been satisfactory.<sup>43</sup> Patients defined as empirically “low-risk” are those with neutropenia (<500 neutrophils/mm<sup>3</sup>) lasting less than seven days, with no complications at the initial assessment and no acute organ dysfunction<sup>43,44</sup> (Table 3).

The American Society of Clinical Oncology (ASCO) recommends avoiding the outpatient management of patients who meet any of the clinical risk criteria summarised in Table 3, regardless of the patient's classification on one risk scale or another.<sup>33</sup> Moreover, the first prognostic index was published recently in order to predict the incidence of severe complications in patients with solid tumours and apparently stable FN episodes.<sup>45</sup> The Clinical Index of Stable Febrile Neutropenia (CISNE) includes six predictors that are independently associated with the incidence of severe complications (ECOG PS ≥ 2 [2 points], chronic bronchitis [1 point], cardiovascular disease [1 point], NCI grade ≥ 2 mucositis [1 point], monocytes <200/mm<sup>3</sup> [1 point] and stress-induced hyperglycaemia [2 points]). These factors are added to a scale ranging from 0 to 8, allowing patients to be classified into three prognostic groups: low risk (0 points), moderate risk (1–2 points) and high risk (≥3 points). The ultimate goal of this index is to prevent the early discharge of patients who, despite their apparent clinical stability, have a high risk of complications (≥3 points). Other social, psychological and logistical factors must be taken into account in order to decide upon the method of treatment. Fig. 1 proposes an action algorithm for treating patients with FN at the Emergency Department that helps the clinician to choose the method of treatment.

#### Febrile neutropenia treatment

Although hospitalisation and the intravenous treatment of FN patients has achieved a significant reduction in mortality, hospitalisation may in itself cause multiple problems, such as toxicity due to intravenous treatments, increased costs, exposure to nosocomial pathogens and a loss of patient quality of life. For this reason, hospital and outpatient treatment strategies have been developed, according to the individual risk stratification.

Empirical antibiotic therapy should be started as soon as possible, as a delay may compromise the patient's prognosis. Before



**Fig. 1.** Action algorithm for the initial care of febrile neutropenia patients at the Emergency Department and evaluation of the risk of complications and treatment method, including the maximum desirable time for each of the actions. Adapted from: Bell MS, Scullen P, McParlan D, et al. Neutropenic sepsis guideline. In edition Northern Ireland Cancer Network 2010; 1–11. \*It is not necessary to wait for an analytical confirmation of neutropenia in order to begin the evaluation; \*\*clinical risk criteria: onset or deterioration of organ dysfunction, comorbidity, altered vital signs, clinical signs or symptoms, documented focal infection or analytical/imaging data. BP, blood pressure; CT, chemotherapy; HR, heart rate; iv, intravenous; MASCC, Multinational Association for Supportive Care in Cancer; po, per os; RR, respiratory rate; SaO<sub>2</sub>, oxygen saturation; SIRS, systemic inflammatory response syndrome.

initiating antibiotic therapy, blood cultures should be collected (if the patient has a CVC, take one of the draws through the catheter) as well as samples of possible infection foci based on clinical data (urine, sputum, exudate, mucous or skin lesions, faeces, cerebrospinal fluid, urinary antigens for pneumococcus and/or *Legionella*, nasal smears for flu in the seasonal period, etc.).

#### Oral outpatient treatment

Low-risk patients are candidates for outpatient treatment, provided the patient tolerates the oral route and has good social and familial support. The most frequent combination is ciprofloxacin with amoxicillin/clavulanic acid and, in patients allergic to β-lactam antibiotics, ciprofloxacin with clindamycin.<sup>46</sup> In a recently-published randomised, double-blind, multicentre clinical trial, moxifloxacin was proven to be as effective as the

combination of amoxicillin/clavulanic acid and ciprofloxacin, with fewer gastrointestinal adverse effects.<sup>47</sup> However, moxifloxacin has a lower antipseudomonal activity and a greater risk of hepatotoxicity. It is also necessary to take rates of local quinolone resistance in Gram-negative bacilli into consideration. Patients who are receiving prophylaxis with fluoroquinolones should not receive empirical treatment with these antibiotics due to the risk of an infection being caused by bacteria that have become resistant to this treatment.

Patients who are discharged under oral outpatient treatment should undergo a check-up after 48 h in order to verify their positive clinical evolution and microbiology results, and in order to adjust the antibiotic treatment and determine its duration. In the event of a clinical deterioration, further diagnostic tests should be considered, as well as hospital admission with intravenous antibiotic therapy.

**Table 3**

Risk criteria for complications that exclude the patient from oral/outpatient management.

Category	Severity criteria
Haematological	Severe thrombocytopenia ( $\leq 10,000$ cells/mm $^3$ ) Anaemia ( $\leq 8$ g/dl)
Cardiovascular	Thromboembolic disease Hypotension (systolic blood pressure $\leq 90$ mmHg) Clinically significant arrhythmia Acute heart failure Chronic cardiovascular disease Severe haemorrhage
Digestive/hepatic	Oral intolerance Nausea or vomiting Diarrhoea Acute abdominal pain Elevated transaminases ( $5 \times$ upper limit of normal) Bilirubin ( $\geq 2$ mg/dl)
Central nervous system	Acute confusional state Meningitis Focal neurological deficit
Infectious	Serious infection (pneumonia, intra-abdominal infection, catheter infection, cellulitis $\geq 5$ cm, pyelonephritis) Signs of sepsis Previous antibiotic use ( $\leq 72$ h prior) Allergy to oral antibiotics Tachycardia, tachypnoea, hypotension Hypoxaemia, hypercapnia, any clinically significant alteration of an analytical value in comparison to the previous value
Vital signs Other laboratory data	
Renal	Dehydration Oliguria Acute kidney failure Hydroelectrolytic disorders
Another relevant comorbidity	Any serious complication or organ dysfunction seen at the outset, pregnancy

### Intravenous treatment

High-risk FN patients require hospital admission and intravenous antibiotics. Treatment options include antipseudomonal  $\beta$ -lactam antibiotics such as piperacillin in combination with tazobactam, cefepime, meropenem or imipenem with cilastatin. At many centres, ceftazidime is no longer considered adequate in monotherapy due to its low activity against many Gram-positive microorganisms, such as streptococcus. In case of allergy to  $\beta$ -lactam antibiotics, the alternative is vancomycin in combination with aztreonam (and metronidazole if there is an abdominal focus). In patients who present complications or those in whom an infection caused by resistant pathogens is suspected, the use of other drugs should be considered, such as aminoglycosides, quinolones and glycopeptides, and more occasionally daptomycin, linezolid, fosfomycin, tigecycline and rifampicin. The empirical addition of a glycopeptide to the initial antibiotic regimen does not generally improve the prognosis of these patients,<sup>48</sup> although it may be considered in specific cases (mucositis, catheter infection, colonisation by methicillin-resistant *Staphylococcus aureus* [*S. aureus*], positive blood cultures for Gram-positive cocci pending an antibiogram, etc.). Table 4 depicts routine doses of oral and intravenous antibiotics.

### Empirical treatment strategies in a patient with febrile neutropenia

The recently-published guidelines of the Infectious Diseases Society of America (IDSA) recommend the use of an antipseudomonal  $\beta$ -lactam drug in monotherapy as initial antibiotic

**Table 4**

Dose of routine oral and intravenous antibiotics.

	Dose
<i>Oral antibiotics</i>	
Amoxicillin/clavulanic acid	875 mg/8 h
Ciprofloxacin	750 mg/12 h
Moxifloxacin	400 mg/24 h
Levofloxacin	500 mg/24 h
Clindamycin	600 mg/6 h
<i>Intravenous antibiotics</i>	
Cefepime	2 g/8 h
Ceftazidime	2 g/8 h
Piperacillin-tazobactam	4 g/8 h
Imipenem	500 mg/6 h
Meropenem	1 g/8 h
Amikacin	1 g/24 h
Tobramycin	3 mg/kg/24 h
Gentamicin	3 mg/kg/24 h
Ciprofloxacin	200–400 mg/8–12 h
Colistin	4.5 MU/12 h (loading dose 9 MU)
Tigecycline	100 mg/12 h (loading dose 150 mg)
Fosfomycin	2 g/6 h
Vancomycin <sup>a</sup>	1 g/12 h
Teicoplanin	400 mg/12 h $\times$ 3 doses, 400 mg/24 h
Daptomycin <sup>b</sup>	10 mg/kg/24 h
Linezolid	600 mg/12 h

MU, million units.

<sup>a</sup> Adjust dose according to the type of infection and microorganism and according to plasma levels.

<sup>b</sup> The dose may vary according to the type of infection and microorganism.

therapy in FN.<sup>25</sup> A meta-analysis found monotherapy to be significantly more beneficial than the combination of a  $\beta$ -lactam antibiotic and aminoglycoside, with fewer adverse effects, lower morbidity and similar survival rates.<sup>49</sup> However, it cannot be completely ruled out that certain patient subgroups may benefit from the initial use of antibiotic combinations, and two types of strategies have thus been used pragmatically: escalation and de-escalation. Specifically, in recent decades, we are seeing an increase in Gram-negative bacterial infections in cancer patients, whilst we also observe a multidrug resistance emergency among these microorganisms.<sup>50,51</sup> In this context, it is doubtful as to whether initial empirical treatment with a  $\beta$ -lactam antibiotic in monotherapy is sufficiently safe in patients with FN,<sup>52</sup> particularly when there are associated severity criteria.

The escalation strategy involves beginning an intravenous treatment in monotherapy and, if the patient deteriorates or a resistant pathogen is isolated, the treatment will be escalated to an antibiotic or a combination of broader-spectrum antibiotics. The advantages of this strategy are that it avoids the early use of some broad-spectrum antibiotics, generates less toxicity, has a lower financial cost and a lower risk of resistance selection, fundamentally to carbapenem antibiotics. In contrast, the patients' prognosis may be compromised if the resistant microorganisms are not adequately covered from the outset.

This escalation strategy should be used in high-risk patients in the following situations: (1) uncomplicated clinical presentation; (2) no risk factors for resistant bacterial infections; and (3) at centres with a low prevalence of resistant microorganisms.

The initial therapeutic options include a non-carbapenem antipseudomonal  $\beta$ -lactam antibiotic such as cefepime, ceftazidime or piperacillin in combination with tazobactam. Carbapenems should be avoided in patients with no complications who lack risk factors for resistant bacteria. Nevertheless, they may be the most suitable option in patients who have had a recent hospital admission (<1 month), previous antibiotic use or previous invasive procedures, who might have a greater risk of Gram-negative bacilli infections with extended-spectrum beta-lactamases.

In the de-escalation strategy, on the other hand, the initial antibiotic treatment administered covers even the most resistant pathogens. Subsequently, therapy is de-escalated to a narrower-spectrum treatment, once the presence of resistant pathogens is ruled out or if a pathogen is identified and its antibiotic sensitivity profile is defined. The main advantage of de-escalation is that it is more likely to achieve an adequate initial antibiotic coverage. However, this strategy often leads to an unnecessary use of broad-spectrum antibiotics, with physicians generally not tending to de-escalate when they have the opportunity to do so, and there is a higher risk of resistance selection.

The de-escalation strategy should be used: (1) in complicated clinical presentations; (2) when there are risk factors for resistant bacterial infections; and (3) at centres with a high prevalence of resistant microorganisms.

The initial therapeutic options include: (1) monotherapy with meropenem or imipenem in seriously ill patients or when there is a previous history of colonisation/infection by extended-spectrum  $\beta$ -lactamase producing enterobacteriaceae; (2) an antipseudomonal  $\beta$ -lactam in combination with aminoglycoside or quinolone in seriously ill patients or if the presence of resistant nonfermenting Gram-negative bacilli (*Pseudomonas aeruginosa* or *Acinetobacter spp.*) is suspected; (3) a  $\beta$ -lactam alongside colistin with or without aminoglycoside, fosfomycin or tigecycline if an infection caused by carbapenemase-producing Gram-negative bacilli or multidrug-resistant nonfermenting Gram-negative bacilli is suspected; (4) a  $\beta$ -lactam in combination with co-trimoxazole if an infection caused by *Stenotrophomonas maltophilia* is suspected. In any case, if there are risk factors for an infection caused by a

resistant Gram-positive microorganism or there is a serious infection related to a venous catheter, the skin and soft tissues, a glycopeptide, daptomycin or linezolid may be added to the initial therapy.

#### Clinical follow-up once empirical treatment has begun

48–72 h after starting the empirical treatment, the patient's clinical evolution and results should be assessed from a microbiological point of view. In case an aetiological agent or clinical focus is isolated, treatment should be simplified and adjusted to the antibiotic sensitivity profile of each microorganism or type of infection, as reflected in Table 5.

In situations where no clinical focus or aetiological agent has been documented and the patient is stable, the antibiotic treatment should be de-escalated to a narrower-spectrum agent and/or the drugs administered in combination should be withdrawn (aminoglycoside, quinolone, colistin, etc.). If the initial presentation was not serious, and the patient has been afebrile for over 72 h and is asymptomatic, the possibility of suspending the treatment may be considered. However, if the patient was in a serious or unstable clinical situation, modifying the initial antibiotic treatment is not advisable.

In most documented infections, 10–14 days of antibiotic treatment is usually sufficient. In some cases, treatment may be extended beyond the resolution of the fever and neutropenia, if necessary. If a catheter infection is documented, its withdrawal or sealing with antimicrobial agents should be considered, depending on the patient's characteristics and the microorganism isolated. In

**Table 5**  
Recommendations for treating febrile neutropenia when there is an evident clinical focus.

Location	Microorganisms	Therapy
Mucositis	Viridans group streptococci, <i>S. aureus</i> , Gram positive and Gram negative anaerobes, herpes simplex virus, <i>Candida spp.</i>	Ensure anaerobicidal coverage In case highly penicillin-resistant viridans group streptococci are present at the centre, add a glycopeptide, daptomycin or linezolid Consider anti-herpes treatment (aciclovir) Consider antifungal treatment (fluconazole, echinocandins, other azoles)
Oesophagitis Neutropenic colitis	<i>Candida</i> , herpes simplex virus <i>Aerobic and anaerobic Gram-negative bacilli</i> , <i>Clostridium spp.</i> (typhlitis), <i>Clostridium difficile</i>	Add fluconazole or echinocandins and aciclovir Ensure anaerobicidal coverage Metronidazole or oral vancomycin in case of suspected <i>C. difficile</i> or dysbiosis In case of a previous history of extended-spectrum $\beta$ -lactamases (ESBL) or a very high incidence at the centre, add amikacin Add metronidazole or oral vancomycin in case of suspected <i>C. difficile</i> or dysbiosis
Diarrhoea	<i>Clostridium difficile</i> , Gram-negative bacteria ( <i>Campylobacter spp.</i> , <i>Salmonella spp.</i> ), virus <i>S. pneumoniae</i> , GNB, viridans group streptococci, anaerobes, respiratory viruses, <i>P. jiroveci</i> , <i>M. pneumoniae</i> , <i>C. pneumoniae</i> , <i>L. pneumoniae</i> , <i>Aspergillus</i> , Nocardia, mycobacteria, etc.	In case of clinical suspicion of atypical pneumonia, add levofloxacin or azithromycin Add oseeltamivir during flu season in case of clinical suspicion TMP-SMX if there is a possibility of <i>P. jiroveci</i> (maintained lymphocytopenia, interstitial pattern, in a patient on high-dose corticosteroids, temozolamide, immunomodulators, etc.) In case of suspected MRSA due to previous colonisation, add vancomycin or linezolid Add a glycopeptide (vancomycin, teicoplanin) or daptomycin or linezolid
Lung infiltrates		Echinocandin or fluconazole if Candidiasis is suspected In case of a high incidence of MRSA or previous colonisation, add a glycopeptide (vancomycin, teicoplanin) or daptomycin or linezolid In case of ectyma gangrenosum or a high incidence of ESBLs at the centre or known patient colonisation, add amikacin In case of suspected ESBLs, consider a beta-lactam with a beta-lactamase inhibitor, carbapenem and fosfomycin Consider adding linezolid, aciclovir and ampicillin with/without amphotericin B
PICC infection	<i>CoNS</i> , <i>S. aureus</i> , <i>C. jeikeium</i> , <i>Bacillus spp.</i> , Gram-negative ( <i>Pseudomonas spp.</i> , <i>S. maltophilia</i> ), <i>Candida spp.</i>	
Cellulitis	<i>CoNS</i> , <i>S. aureus</i> , <i>Streptococcus spp.</i> , <i>C. jeikeium</i> , <i>Bacillus spp.</i> , Gram-negative ( <i>Pseudomonas spp.</i> , <i>E. coli</i> , <i>K. pneumoniae</i> )	
Urinary infection	Enterobacteriaceae, <i>Enterococcus spp.</i> , <i>P. aeruginosa</i> , <i>Candida</i>	
CNS and neurosurgical infections	<i>CoNS</i> , <i>S. aureus</i> , <i>Streptococcus pneumoniae</i> , <i>Cryptococcus neoformans</i> , herpes simplex virus, <i>Listeria monocytogenes</i>	

CNS, central nervous system; CoNS, coagulase-negative staphylococci; MRSA, methicillin-resistant *Staphylococcus aureus*; PICC, peripherally inserted central catheter; TMP-SMX, trimethoprim and sulfamethoxazole.

patients with persistent fever, a full reassessment should be performed, with an active search for possible infection foci or other causes of fever such as drug toxicity, tumour fever, etc.

Biomarkers are analytical parameters that may complement other clinical and microbiological variables in the assessment of a FN episode and its severity. Likewise, normalised values support the treatment response. Of the biochemical parameters, those of greatest interest are stress hyperglycaemia as an acute phase reactant and hypoalbuminaemia as a marker of malnutrition and fragility.<sup>42</sup> Of the serum markers specific to the inflammatory/infectious process, those most frequently used are lactate,

procalcitonin and C-reactive protein. Their utility is not well defined due to the heterogeneity of the populations studied and the few patients included in published clinical trials.<sup>53</sup> Procalcitonin (value > 0.5 ng/ml) is a more useful and earlier marker than C-reactive protein (value ≥ 90 mg/dl), especially in the diagnosis of bacteraemia, as it is not elevated in viral infections, and for predicting the severity of FN complications. The addition of procalcitonin to clinical risk scores could increase the sensitivity and negative predictive value for detecting bacteraemia and the failure of the antibiotic treatment.<sup>54</sup> Interleukin 6, 8 and 10 could be better predictors of severity and complications but are less used due to their

**Table 6**

Application of standard precautions and types of specific precautions to be adopted according to the infectious disease or microorganism, and their transmissibility period.

Procedure	Examples	Hand hygiene <sup>a</sup>	Gloves <sup>b</sup>	Additional gown <sup>c</sup>	Mask <sup>d</sup>
No contact	Talk to the patient	No	No	No	No
Contact with intact skin or uncontaminated clothing	Physical examination, vital signs	Before and after	No	No	No
Contact with (or the possibility thereof) non-intact skin, mucous membranes, fluids, secretions, excretions	Draws, dressings, handling of catheters, probe, drains, etc.	Before and after	Yes <sup>b</sup>	No <sup>d</sup>	No <sup>d</sup>
Respiratory secretions	Aspiration, respiratory therapy Tracheotomy dressing	Before and after	Yes <sup>b</sup>	Yes	Yes
Disease or microorganisms	Types of precautions or isolation measures		Possible transmission period		
Multidrug-resistant bacterial pathogens (MRSA, VRE, ESBL+ Enterobacteriaceae, <i>Acinetobacter baumannii</i> , MDR <i>Pseudomonas aeruginosa</i> )	Contact		Cross-species transmission during colonisation or infection by the corresponding microorganism. In case of a prolonged hospital stay, carry out weekly epidemiological surveillance cultures for three consecutive weeks; if they are negative, lift precautions. Short hospital stays: throughout the stay.		
Adenovirus Flu (Influenza)	Droplets and contact Droplets		Adenovirus infections may be transmitted for up to 14 days after their onset. From 3 to 5 days after the onset of symptoms in adults. In children, transmissibility may extend to 7 days.		
Respiratory syncytial virus	Contact		The period immediately before the active disease and the entire duration of the active disease.		
Human parainfluenza virus	Droplets-fomites		Prior to the onset of symptoms up until their resolution (it may be transmitted by asymptomatic carriers).		
Measles	Airborne		From 4 days prior to the exanthema to 4 days after (minimum contagion after the 2nd day of the exanthema).		
Rubella (congenital) Rubella Mumps	Contact Droplets Droplets		Virus can be transmitted for months in infants. From 1-week before the exanthema to 7 days after. The virus is isolated in the saliva from 7 days before to 9 days after the onset of clear symptoms. The maximum contagion risk covers the period from 2 days before the onset of the disease to 4 days afterwards.		
Hepatitis A	Contact (Faecal – oral)		Infectivity period: From 2 to 3 weeks prior to the onset of symptoms and one week after symptom onset.		
Rotavirus Parvovirus B19	Contact (Faecal – oral) Droplets		During the acute phase and while virus excretion persists. If the patient only presents an exanthema, transmissibility is at its highest before the rash and unlikely thereafter. In case of an aplastic crisis, the transmissibility period is up to one week post-onset.		
Varicella-Zoster	Airborne and contact		4–5 days before the rash and up until the lesions have scabbed over (around 7 days)		
Salmonella	Contact (Faecal – oral)		From the first week up to the end of convalescence (1–2 weeks). In <i>S. Typhi</i> , assess chronic carriers.		
Tuberculosis	Airborne		Lasts for as long as tuberculosis bacilli are expelled in sputum. Effective antimicrobial chemotherapy eliminates transmissibility within 2–4 weeks		
Impetigo Mycoplasma (Primary Atypical Pneumonia)	Contact Droplets-(recently contaminated fomites or respiratory secretions)		Until the lesions have completely healed (usually 1–2 weeks). Duration of under 20 days. Treatment does not eradicate the microorganism from the airways, where it may persist for up to 13 weeks.		
Whooping cough Type B <i>H. influenzae</i>	Droplets Droplets		Persists for up to 5 days after effective treatment. Stops being transmissible 24–48 h after the onset of effective antibiotic treatment.		
<i>Neisseria meningitidis</i>	Droplets		Persists until the live meningococci disappear from nose and mouth secretions, that is, 24 h after starting an appropriate treatment.		
Scarlet fever <i>Clostridium difficile</i> Scabies	Droplets Contact Contact		Persists for up to 24 h after effective treatment. May persist for weeks and months in its non-vegetative or spore forms. Very persistent up until the mites and eggs have been destroyed. Not transmitted 24 h after effective treatment (Permethrin 5%).		

ESBL, extended-spectrum beta-lactamases; MDR, multidrug-resistant; MRSA, methicillin-resistant *Staphylococcus aureus*; VRE, vancomycin-resistant enterococci.

<sup>a</sup> Soap and water or a water-alcohol solution (provided there is no visible dirt).

<sup>b</sup> Change them between patients and between different contaminated and uncontaminated areas.

<sup>c</sup> Refers to a cotton or disposable gown for specific use in procedures, not to normal scrubs.

<sup>d</sup> Except wound dressings.

high cost, lack of availability and low specificity. The lipopolysaccharide binding protein, interleukin 2 and tumour necrosis factor, among others, have no current application in the context of cancer patients with FN.

### Specific precaution measures

The objective of these measures is to avoid the transmission of certain pathogens from a colonised patient or one with an active infection to other patients or healthcare personnel. One key aspect is that the application of such measures should not affect the quality of the healthcare received by the patient, and that these measures should be added to the so-called standard precautions, such as hand hygiene and decontamination, the use of gloves, gowns and/or a mask, according to cases, situations and indications, which are shown in [Table 6](#).

Specific precaution measures are classified based on the microorganisms' modes of transmission as follows: (1) respiratory precautions, the objective of which is to avoid the airborne dissemination of particles under 5 µ, which may remain suspended in the air for prolonged periods of time as in the case of respiratory TB, disseminated VZV, measles, etc.; (2) precautions against droplets, aimed at avoiding the transmission of pathogenic microorganisms through larger droplets and which requires close contact between the source of exposure and susceptible host, as in meningococcal infection, flu, etc.; and (3) contact precautions, which seek to avoid transmission through direct or indirect contact by means of objects or contaminated surfaces. [Table 6](#) summarises the recommendations and specific measures to be adopted according to the infectious disease or microorganism in question.

Contact precautions are those needed most frequently in cancer patients, and are indicated in the following situations: (1) respiratory, gastrointestinal or skin infections and/or wounds colonised or infected by multidrug-resistant pathogens; (2) diarrhoeal infections, including infections caused by *Clostridium difficile*; (3) infections caused by respiratory viruses; and (4) skin or mucous membrane infections.

As regards multidrug-resistant microorganisms, the Infection Control Committee or Team of each centre should decide which are the most important and susceptible to be subject to the implementation of contact precautions, based on the existing recommendations, and always considering the local epidemiology and capacity for transmission between patients of each of the multidrug-resistant pathogens assessed. To that effect, it may be necessary to perform epidemiological surveillance cultures.

In most hospitals, the application of contact precautions is recommended in the following scenarios: (1) all cases of methicillin-resistant *S. aureus*; (2) vancomycin-resistant *Enterococci*; (3) extended-spectrum beta-lactamase producing enterobacteriaceae; (4) carbapenemase-producing enterobacteriaceae; (5) nonfermenting Gram-negative bacilli, such as *Pseudomonas aeruginosa* or *Acinetobacter baumannii* with multidrug or pandrug resistance patterns.

Reverse isolation measures would only be indicated in those solid cancer patients who are receiving chemotherapy regimens that lead to profound and prolonged neutropenia. Reverse isolation rooms must fulfil a series of special characteristics that reduce environmental contamination by circulating microorganism-stripped air and preventing the penetration of microorganisms into the room by means of positive pressure.

### Conflicts of interest

The authors declare that they have no conflicts of interest.

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### References

1. Prevention and Treatment of Cancer-Related Infections. Version 2.2014; 2014. Vol. (National Comprehensive Cancer Network. (NCCN).
2. Rubin LG, Levin MJ, Ljungman P, Davies EG, Avery R, Tomblyn M, et al. 2013 IDSA clinical practice guideline for vaccination of the immunocompromised host. *Clin Infect Dis.* 2014;58:309–18.
3. General recommendations on immunization. Recommendations of the Advisory Committee on Immunization Practices (ACIP). *MMWR Recomm Rep.* 2011;60:1–64.
4. EASL Clinical Practice Guidelines: management of chronic hepatitis B. *J Hepatol.* 2009;50:227–42.
5. Castellano G, Manzano ML. Tratamiento y profilaxis de la hepatitis B en pacientes imunosuprimidos. *Gastroenterol Hepatol.* 2012;35:1–19.
6. Hwang JP, Somerter MR, Alston-Johnson DE, Cryer DR, Feld JJ, Kramer BS, et al. Hepatitis B virus screening for patients with cancer before therapy: American Society of Clinical Oncology Provisional Clinical Opinion Update. *J Clin Oncol.* 2015;33:2212–20.
7. Lorente L, Henry C, Martin MM, Jimenez A, Mora ML. Central venous catheter-related infection in a prospective and observational study of 2,595 catheters. *Crit Care.* 2005;9:R631–5.
8. O'Grady NP, Alexander M, Burns LA, Dellinger EP, Garland J, Heard SO, et al. Guidelines for the prevention of intravascular catheter-related infections. *Clin Infect Dis.* 2011;52:e162–93.
9. Schiffer CA, Mangi PB, Wade JC, Camp-Sorrell D, Cope DG, El-Rayes BF, et al. Central venous catheter care for the patient with cancer: American Society of Clinical Oncology clinical practice guideline. *J Clin Oncol.* 2013;31:1357–70.
10. Ranson MR, Oppenheim BA, Jackson A, Kamthan AG, Scarffe JH. Double-blind placebo-controlled study of vancomycin prophylaxis for central venous catheter insertion in cancer patients. *J Hosp Infect.* 1990;15:95–102.
11. Banerjee S, Shen B, Baron TH, Nelson DB, Anderson MA, Cash BD, et al. Antibiotic prophylaxis for GI endoscopy. *Gastrointest Endosc.* 2008;67:791–8.
12. Jain NK, Larson DE, Schroeder KW, Burton DD, Cannon KP, Thompson RL, et al. Antibiotic prophylaxis for percutaneous endoscopic gastrostomy. A prospective, randomized, double-blind clinical trial. *Ann Intern Med.* 1987;107:824–8.
13. Sepkowitz KA. *Pneumocystis carinii* pneumonia in patients without AIDS. *Clin Infect Dis.* 1993;17 Suppl. 2:S416–22.
14. Green H, Paul M, Vidal L, Leibovici L. Prophylaxis of *Pneumocystis pneumonia* in immunocompromised non-HIV-infected patients: systematic review and meta-analysis of randomized controlled trials. *Mayo Clin Proc.* 2007;82:1052–9.
15. Hughes WT, Rivera GK, Schell MJ, Thornton D, Lott L. Successful intermittent chemoprophylaxis for *Pneumocystis carinii* pneumonitis. *N Engl J Med.* 1987;316:1627–32.
16. Colby C, McAfee S, Sackstein R, Finkelstein D, Fishman J, Spitzer T. A prospective randomized trial comparing the toxicity and safety of atovaquone with trimethoprim/sulfamethoxazole as *Pneumocystis carinii* pneumonia prophylaxis following autologous peripheral blood stem cell transplantation. *Bone Marrow Transpl.* 1999;24:897–902.
17. Sangiolo D, Storer B, Nash R, Corey L, Davis C, Flowers M, et al. Toxicity and efficacy of daily dapsone as *Pneumocystis jiroveci* prophylaxis after hematopoietic stem cell transplantation: a case-control study. *Biol Blood Marrow Transpl.* 2005;11:521–9.
18. De Masi JM, Cox JA, Leonard D, Koh AY, Aquino VM. Intravenous pentamidine is safe and effective as primary *Pneumocystis pneumonia* prophylaxis in children and adolescents undergoing hematopoietic stem cell transplantation. *Pediatr Infect Dis J.* 2013;32:933–6.
19. Marras TK, Sanders K, Lipton JH, Messner HA, Conly J, Chan CK. Aerosolized pentamidine prophylaxis for *Pneumocystis carinii* pneumonia after allogeneic marrow transplantation. *Transpl Infect Dis.* 2002;4:66–74.
20. Keiser PB, Nutman TB. *Strongyloides stercoralis* in the immunocompromised population. *Clin Microbiol Rev.* 2004;17:208–17.
21. Martinez-Perez A, Norman FF, Monge-Maillo B, Perez-Molina JA, Lopez-Velez R. An approach to the management of *Trypanosoma cruzi* infection (Chagas' disease) in immunocompromised patients. *Expert Rev Anti Infect Ther.* 2014;12:357–73.
22. Kuderer NM, Dale DC, Crawford J, Lyman GH. Impact of primary prophylaxis with granulocyte colony-stimulating factor on febrile neutropenia and mortality in adult cancer patients receiving chemotherapy: a systematic review. *J Clin Oncol.* 2007;25:3158–67.
23. Crawford J, Caserta C, Roila F. Hematopoietic growth factors: ESMO Clinical Practice Guidelines for the applications. *Ann Oncol.* 2010;21 Suppl. 5:v248–51.
24. Freifeld AG, Bow EJ, Sepkowitz KA, Boeckh MJ, Ito JI, Mullen CA, et al. Clinical practice guideline for the use of antimicrobial agents in neutropenic patients with cancer: 2010 update by the infectious diseases society of America. *Clin Infect Dis.* 2011;52:e56–93.
25. Berghmans T, Paesmans M, Lafitte JJ, Mascaux C, Meert AP, Jacqy C, et al. Therapeutic use of granulocyte and granulocyte-macrophage colony-stimulating factors in febrile neutropenic cancer patients. A systematic review of the literature with meta-analysis. *Support Care Cancer.* 2002;10:181–8.

26. Clark OA, Lyman GH, Castro AA, Clark LG, Djulbegovic B. Colony-stimulating factors for chemotherapy-induced febrile neutropenia: a meta-analysis of randomized controlled trials. *J Clin Oncol.* 2005;23:4198–214.
27. Kuderer NM, Dale DC, Crawford J, Cosler LE, Lyman GH. Mortality, morbidity, and costs associated with febrile neutropenia in adult cancer patients. *Cancer.* 2006;106:2258–66.
28. Cullen M, Steven N, Billingham L, Gaunt C, Hastings M, Simmonds P, et al. Antibacterial prophylaxis after chemotherapy for solid tumors and lymphomas. *N Engl J Med.* 2005;353:988–98.
29. Gafter-Gvili A, Fraser A, Paul MLeibovici L. Meta-analysis: antibiotic prophylaxis reduces mortality in neutropenic patients. *Ann Intern Med.* 2005;142:979–95.
30. Bucaneve G, Micozzi A, Menichetti F, Martino P, Dionisi MS, Martinelli G, et al. Levofloxacin to prevent bacterial infection in patients with cancer and neutropenia. *N Engl J Med.* 2005;353:977–87.
31. Bow EJ. Fluoroquinolones, antimicrobial resistance and neutropenic cancer patients. *Curr Opin Infect Dis.* 2011;24:545–53.
32. Carratala J, Fernandez-Sevilla A, Tubau F, Callis MGudiol F. Emergence of quinolone-resistant *Escherichia coli* bacteremia in neutropenic patients with cancer who have received prophylactic norfloxacin. *Clin Infect Dis.* 1995;20:557–60, discussion 561–53.
33. Flowers CR, Seidenfeld J, Bow EJ, Kartem C, Gleason C, Hawley DK, et al. Antimicrobial prophylaxis and outpatient management of fever and neutropenia in adults treated for malignancy: American Society of Clinical Oncology clinical practice guideline. *J Clin Oncol.* 2013;31:794–810.
34. Kern WV, Andriof E, Oethinger M, Kern P, Hacker J, Marre R. Emergence of fluoroquinolone-resistant *Escherichia coli* at a cancer center. *Antimicrob Agents Chemother.* 1994;38:681–7.
35. Neumann S, Krause SW, Maschmeyer G, Schiel X, von Lilienfeld-Toal M. Primary prophylaxis of bacterial infections and *Pneumocystis jirovecii* pneumonia in patients with hematological malignancies and solid tumors: guidelines of the Infectious Diseases Working Party (AGIHO) of the German Society of Hematology and Oncology (DGHO). *Ann Hematol.* 2013;92:433–42.
36. Pepin J, Saheb N, Coulombe MA, Alary ME, Corriveau MP, Authier S, et al. Emergence of fluoroquinolones as the predominant risk factor for *Clostridium difficile*-associated diarrhea: a cohort study during an epidemic in Quebec. *Clin Infect Dis.* 2005;41:1254–60.
37. Cullen M, Baijal S. Prevention of febrile neutropenia: use of prophylactic antibiotics. *Br J Cancer.* 2009;101 Suppl. 1:S11–4.
38. Tjan-Heijnen VC, Postmus PE, Ardizzone A, Manegold CH, Burghouts J, van Meerbeeck J, et al. Reduction of chemotherapy-induced febrile leucopenia by prophylactic use of ciprofloxacin and roxithromycin in small-cell lung cancer patients: an EORTC double-blind placebo-controlled phase III study. *Ann Oncol.* 2001;12:1359–68.
39. Mayordomo JL, Lopez A, Vinolas N, Castellanos J, Pernas S, Domingo Alonso J, et al. Retrospective cost analysis of management of febrile neutropenia in cancer patients in Spain. *Curr Med Res Opin.* 2009;25:2533–42.
40. Klastersky J, Paesmans M, Georgala A, Muanza F, Plehiers B, Dubreucq L, et al. Outpatient oral antibiotics for febrile neutropenic cancer patients using a score predictive for complications. *J Clin Oncol.* 2006;24:4129–34.
41. Carmona-Bayonas A, Gomez J, Gonzalez-Billalabertia E, Canteras M, Navarrete A, Gonzalez ML, et al. Prognostic evaluation of febrile neutropenia in apparently stable adult cancer patients. *Br J Cancer.* 2011;105:612–7.
42. Talcott JA, Finberg R, Mayer RJ, Goldman L. The medical course of cancer patients with fever and neutropenia. Clinical identification of a low-risk subgroup at presentation. *Arch Intern Med.* 1988;148:2561–8.
43. Teuffel O, Ethier MC, Alibhai SM, Beyene J, Sung L. Outpatient management of cancer patients with febrile neutropenia: a systematic review and meta-analysis. *Ann Oncol.* 2011;22:2358–65.
44. Dellinger RP, Levy MM, Rhodes A, Annane D, Gerlach H, Opal SM, et al. Surviving sepsis campaign: international guidelines for management of severe sepsis and septic shock: 2012. *Crit Care Med.* 2013;41:580–637.
45. Carmona-Bayonas A, Jimenez-Fonseca P, Virizuela Echaburu J, Antonio M, Font C, Biosca M, et al. Prediction of serious complications in patients with seemingly stable febrile neutropenia: validation of the Clinical Index of Stable Febrile Neutropenia in prospective cohort of patients from the FINITE study. *J Clin Oncol.* 2015;33:465–71.
46. Kern WV, Marchetti O, Drgona L, Akan H, Aoun M, Akova M, et al. Oral antibiotics for fever in low-risk neutropenic patients with cancer: a double-blind, randomized, multicenter trial comparing single daily moxifloxacin with twice daily ciprofloxacin plus amoxicillin/clavulanic acid combination therapy – EORTC infectious diseases group trial XV. *J Clin Oncol.* 2013;31:1149–56.
47. Paul M, Dickstein Y, Borok S, Vidal I, Leibovici L. Empirical antibiotics targeting Gram-positive bacteria for the treatment of febrile neutropenic patients with cancer. *Cochrane Database Syst Rev.* 2014;1:CD003914.
48. Paul M, Soares-Weiser K, Grozinsky S, Leibovici L. Beta-lactam versus beta-lactam-aminoglycoside combination therapy in cancer patients with neutropenia. *Cochrane Database Syst Rev.* 2003;CD003038.
49. Gudiol C, Tubau F, Calatayud L, Garcia-Vidal C, Cisnal M, Sanchez-Ortega I, et al. Bacteremia due to multidrug-resistant Gram-negative bacilli in cancer patients: risk factors, antibiotic therapy and outcomes. *J Antimicrob Chemother.* 2011;66:657–63.
50. Marin M, Gudiol C, Garcia-Vidal C, Ardanuy C, Carratala J. Bloodstream infections in patients with solid tumors: epidemiology, antibiotic therapy, and outcomes in 528 episodes in a single cancer center. *Medicine (Baltimore).* 2014;93:143–9.
51. Averbuch D, Orasch C, Cordonnier C, Livermore DM, Mikulska M, Viscoli C, et al. European guidelines for empirical antibacterial therapy for febrile neutropenic patients in the era of growing resistance: summary of the 2011 4th European Conference on Infections in Leukemia. *Haematologica.* 2013;98:1826–35.
52. Meidani M, Khorvash F, Abolghasem H, Jamali B. Procalcitonin and quantitative C-reactive protein role in the early diagnosis of sepsis in patients with febrile neutropenia. *South Asian J Cancer.* 2013;2:216–9.
53. Jimeno A, Garcia-Velasco A, del Val O, Gonzalez-Billalabertia E, Hernando S, Hernandez R, et al. Assessment of procalcitonin as a diagnostic and prognostic marker in patients with solid tumors and febrile neutropenia. *Cancer.* 2004;100:2462–9.
54. Sahr Y, Sponholz C, Tuche F, Brunkhorst F, Reinhart K. The role of procalcitonin in febrile neutropenic patients: review of the literature. *Infection.* 2008;36:396–407.