



## Case Report

# Monitoring the extracorporeal photopheresis by immunophenotyping in a Sézary syndrome patient



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## ABSTRACT

The extracorporeal photopheresis is indicated as immunomodulatory treatment in primary cutaneous T-cell lymphoma. There are no guidelines about the most suitable immunophenotypic panel to monitor and correlate clinical evolution and laboratory parameters.

In this study we characterize a patient with Sézary syndrome stage III, who performed 98 sessions of photopheresis; in the last 30, the expression of T-cells in peripheral blood was evaluated.

The patient had exfoliative dermatitis (mainly in the lower limbs) and several analytical disorders: anemia, neutropenia and lymphocytosis; abnormal number of effector T-cells and a CD4/CD8 ratio higher than 10; atypical cells (CD7, CD26) with aberrant phenotype (characteristic of malignant T-cell clones); and regulatory T-cells indicating an immunotolerance state. The clinical evolution verified can be related with the therapeutic scheme adopted.

We propose a multiparameter flow cytometry approach to monitor patients with Sézary syndrome that realize photopheresis, including the aberrant cases.

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## Introduction

Sézary syndrome (SS) is the most aggressive form of Mycosis fungoides, a cutaneous T-cell lymphoma (CTCL), classified as non-Hodgkin lymphoma. This disease derives from skin-homing memory T-cells and is characterized by exfoliative erythroderma, generalized lymphadenopathy, Sézary cells with cerebriform nuclei circulating in peripheral blood (PB) ( $\geq 1000/\text{mm}^3$ ) and poor quality of life. The diagnosis represents a challenge due to the similarity with other dermatologic diseases and should be based on criteria defined by the International Society for Cutaneous Lymphomas (ISCL) and the European Organization of Research and Treatment of Cancer (EORTC).<sup>1–3</sup> After the diagnosis and the beginning of the treatment, it is important to monitorize the patient; however, there are no immunophenotyping standards to characterize the therapy's effectiveness. Extracorporeal photopheresis (ECP) is recommended as an immunomodulator treatment, offering better life quality for the patient. It is a personalized therapy, only

available in specialized healthcare centers; normally performed as an adjuvant treatment, associated with protein inhibitors, steroids and chemotherapy. The ECP procedure is based on 8-methoxysoralen effect (a photosensitizing agent), combined with UVA light, which is added to mononuclear cells collected by apheresis, and finally reinfused to the patient.<sup>4,5</sup>

We propose a multiparameter flow cytometry approach that can be used to monitorize Sézary syndrome patients performing ECP, including aberrant cases.

## Clinical case

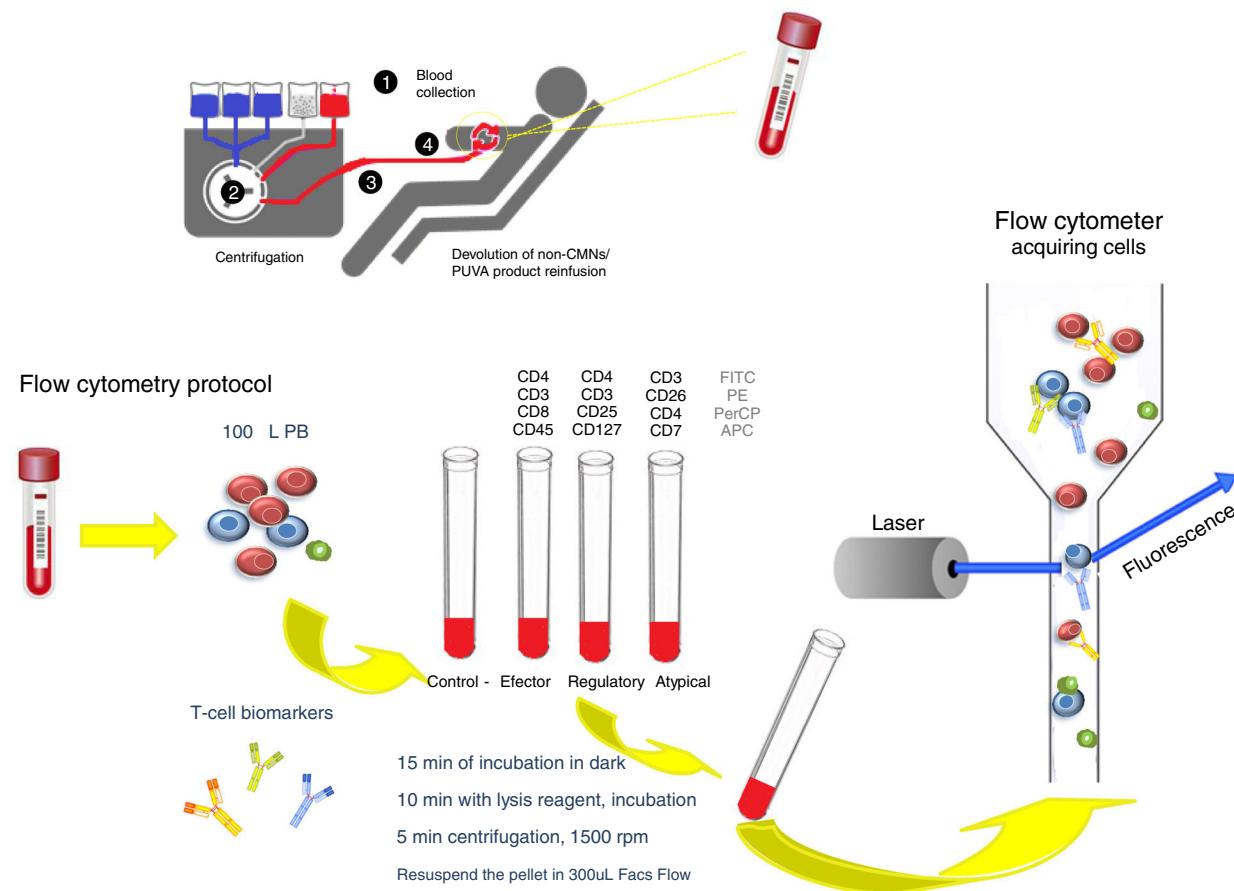
We describe a 56 years-old Caucasian male with SS stage III diagnosed in 2014 (according to TNMB classification of the ISCL/EORTC). The patient presented erythroderma, multiple palpable adenomegalies and lymphocytosis. He was initially treated with a combination of prednisolone, interferon- $\alpha$  (IFN- $\alpha$ ) and ECP (bi-weekly); after, it was added systemic bexarotene. The pruritus was controlled with hydroxyzine and loratadine and the cutaneous lesions with local measures; a complex of iron, folic acid, vitamins D and B12 was also prescribed. Two years later (Fig. 1), we observe a positive clinical response with an improvement of the cutaneous exfoliation affecting mainly the back and the thorax without cracks

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### Apheresis procedure



**Fig. 1.** Schematic representation of the flow cytometry protocol.

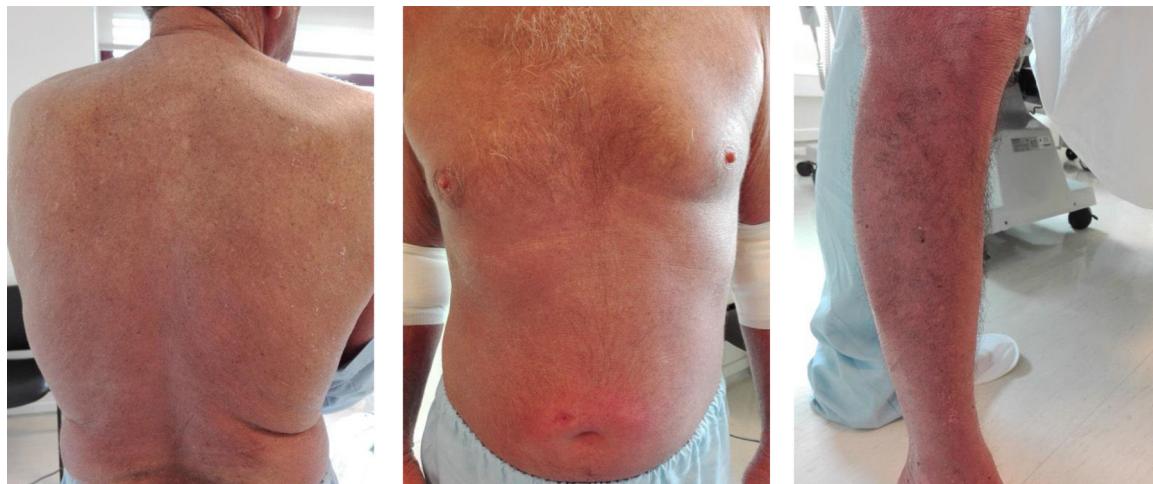
or maceration; the dark-red color of skin also shows some decrease. However, he maintains itching specially in the face and lower limbs.

The PB was analyzed before and after each ECP by an automated hematology analyzers and through flow cytometry (FACSCanto II) using a biomarkers panel to characterize effector, regulatory and atypical T-cell populations. The protocol is schematically represented in Fig. 2. We also evaluated results obtained from a healthy control.

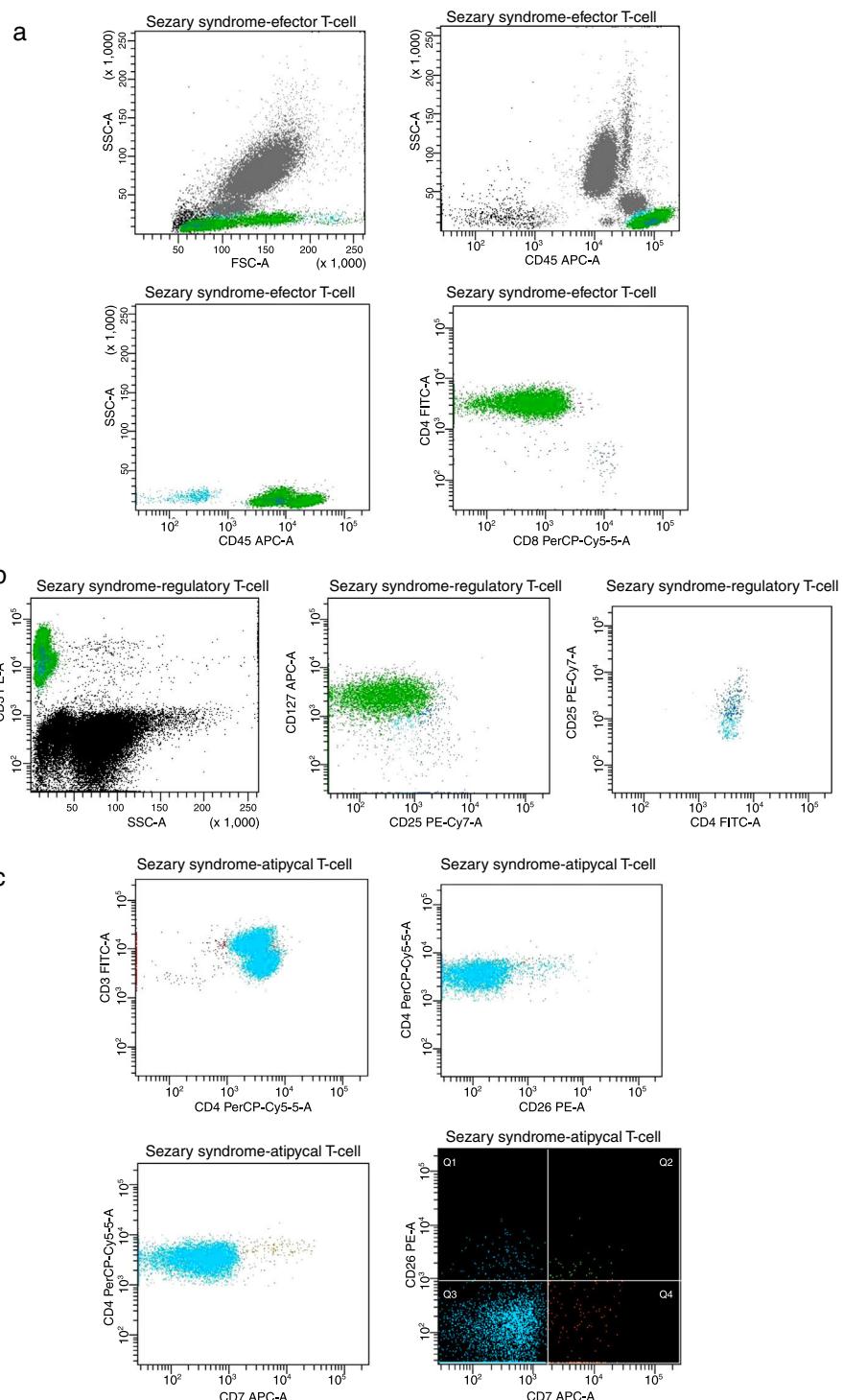
### Results and discussion

ECP has a recognized long-term immunomodulatory effect. In that way we observe, 2 years after, some improvement in patient's skin and a gradual decrease of lymphocytosis; however, the low value of hemoglobin has no change was well as the neutropenia (mostly post-session) (Table 1).

The circulating T-cells express an aberrant and stable immunophenotype over the ECP treatment, as represented in Fig. 3.



**Fig. 2.** The skin of a Sézary syndrome patient with exfoliative erythroderma in back, trunk and lower limbs.



**Fig. 3.** Plots of T-cell expression in peripheral blood from the studied patient after the last session. (A) Lymphocytes (cyan events), CD4+ effector T-cells showing CD4+ and CD8+ populations (green and navy, respectively). (B) Regulatory T-cells CD4+CD25<sup>high</sup> showing phenotype CD25+CD127<sup>-low</sup> (navy). (C) Atypical cell clusters majority CD3+CD4+ (green) displaying CD7<sup>-</sup> (cyan) and CD26<sup>-</sup> (orange).

**Table 1**  
The complete blood counts (average of values during the treatment).

Hematological parameters	Patient	Healthy control
Hemoglobin [g/dL]	8.3	15.4
Leukocytes [ $10^3/\mu\text{L}$ ]	4.5	7.7
Lymphocytes [ $10^3/\mu\text{L}$ ]	2.4	1.7
Platelets [ $10^3/\mu\text{L}$ ]	240.6	209.0
Neutrophils [ $10^3/\mu\text{L}$ ]	1.3	5.4
Monocytes [ $10^3/\mu\text{L}$ ]	0.6	0.5

The lymphocyte CD3+ population represents more than 60% of the total number of leucocytes (Fig. 3A). The effector T-cells display an abnormal proportion, around 99% are CD4+ and only 1% CD8+, resulting in a high CD4/CD8 ratio (>10, with 88% >100).<sup>8–10</sup> A high fraction (80%) of activated regulatory T-cells (Treg) CD4+CD25<sup>high</sup> are also detected (Fig. 3B), which is similar to the healthy control.<sup>3,11,12</sup> In the last twenty sessions, we have observed a slight increase of CD8+ associated with Treg cells reduction, so the CD4/CD8 ratio also decreases. We speculate

that this immunotolerance state is promoted by the downregulation of circulating cells, induced by ECP treatment and/or IFN- $\alpha$  action.<sup>6</sup>

We investigated the presence of Sézary cells using CD7 and CD26 biomarkers. Considering the thresholds in subsets recommended by the ISCL/EORTC (CD4+CD7+ >40% and CD4+CD26+ >30%), we detect >1000 neoplastic cells/ $\mu$ L.<sup>3,7</sup> The atypical T-cells exhibit an aberrant arrangement which is often expressed by malignant T-cell clones (Fig. 3C). An overexpression of CD3+CD4+CD7+ (98%) and CD3+CD4+CD26+ (95%) was demonstrated, without evidence of loss or alteration of these T-cell antigens.<sup>8,9</sup> However, the high expression of biomarkers typically related to the severity of Sézary's disease suggest that more studies should be performed to corroborate these insights, especially to better understand the Sézary typical cells relevance.

The multiparameter flow cytometry assay can be considered as an advanced tool during the follow-up of Sézary disease, including the aberrant phenotypes. According to the best of our knowledge, PB analysis should be performed after the second session, once there is no significant difference between pre- and post-assay. Based on the tumor-associated immunophenotype identified, we suggest the use of CD3, CD4, CD7, CD8, CD25, CD26, CD45, CD127 biomarkers to monitorize circulating neoplastic T-cells and assess the ECP response in these patients with LTCL.

### Conflicts of interest

The authors declare no conflicts of interest.

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### References

- Boon SE, et al. Evaluation of immunophenotypic and Sézary syndrome molecular biomarkers using standard operating procedures: a multicenter study of 59 patients. *J Investig Dermatol*. 2016;136:1364–72.
- Novelli M, et al. Blood flow cytometry in Sézary syndrome new insights on prognostic relevance and immunophenotypic changes during follow-up. *Am J Clin Pathol*. 2015;143:57–69.
- Olsen EA, et al. Clinical end points and response criteria in mycosis fungoides and Sézary syndrome: a consensus statement of the International Society for Cutaneous Lymphomas. *J Clin Oncol*. 2011;29:598–2607.
- Atta M, Papanicolaou N, Tsirigotis P. Transfusion and apheresis science the role of extracorporeal photopheresis in the treatment of cutaneous T-cell lymphomas. *Transfus Apher Sci*. 2012;46:195–202.
- Willems R, et al. Primary cutaneous lymphomas: ESMO Clinical Practice Guidelines for diagnosis, treatment and follow-up – clinical practice guidelines. *Ann Oncol*. 2013;24:1–6.
- Andy CH, et al. Expression of helper T cell master regulators in inflammatory dermatoses and primary cutaneous T-cell lymphomas: diagnostic implications. *Am Acad Dermatol*. 2014;18:159–67.
- Kelemen K, Guitart J, Kuzel TM, Goolsby CL, Peterson LC. The usefulness of CD26 in flow cytometric analysis of peripheral blood in Sézary syndrome. *Am Soc Clin Pathol*. 2016;129:146–56.
- Krejsgaard T, Odum N, Geisler C, Wasik MA, Woetmann A. Sézary syndrome regulatory T cells and immunodeficiency in mycosis fungoides and Sézary syndrome. *Leukemia*. 2011;26:424–32.
- Horna P, et al. Flow cytometric identification of immunophenotypically aberrant T-cell clusters on skin shave biopsy specimens from patients with mycosis fungoides. *Am J Clin Pathol*. 2015;143:785–96.