



ELSEVIER

Gastroenterología y Hepatología

www.elsevier.es/gastroenterologia



REVIEW IN GASTROENTEROLOGY

Chronic diarrhoea: Definition, classification and diagnosis[☆]

Fernando Fernández-Baños^{a,i,*}, Anna Accarino^{b,i}, Agustín Balboa^c,
Eugenio Domènech^{d,i}, María Esteve^{a,i}, Esther García-Planella^e, Jordi Guardiola^f,
Xavier Molero^{b,i}, Alba Rodríguez-Luna^g, Alexandra Ruiz-Cerulla^f, Javier Santos^{b,i},
Eva Vaquero^{h,i}

^a Servicios de Digestivo, Hospital Universitari Mútua de Terrassa, Terrassa, Barcelona, Spain

^b Servicios de Digestivo, Hospital Universitari Vall d'Hebron, Barcelona, Spain

^c Servicios de Digestivo, Hospital Teknon, Barcelona, Spain

^d Servicios de Digestivo, Hospital Germans Trias i Pujol, Badalona, Barcelona, Spain

^e Servicios de Digestivo, Hospital de Sant Pau, Barcelona, Spain

^f Servicios de Digestivo, Hospital Universitari de Bellvitge, Hospitalet de Llobregat, Barcelona, Spain

^g Medicina de Familia, CAP Sur, Terrassa, Barcelona, Spain

^h Servicios de Digestivo, Hospital Clínic, Barcelona, Spain

ⁱ Centro de Investigaciones Biomédicas en Red de Enfermedades Hepáticas y Digestivas (CIBEREHD), Spain

Received 19 June 2015; accepted 30 September 2015

Available online 28 July 2016

KEYWORDS

Chronic diarrhoea;
GRADE system;
Evidence-based
recommendations;
Consensus document

Abstract Chronic diarrhoea is a common presenting symptom in both primary care medicine and in specialised gastroenterology clinics. It is estimated that >5% of the population has chronic diarrhoea and nearly 40% of these patients are older than 60 years. Clinicians often need to select the best diagnostic approach to these patients and choose between the multiple diagnostic tests available. In 2014 the Catalan Society of Gastroenterology formed a working group with the main objective of creating diagnostic algorithms based on clinical practice and to evaluate diagnostic tests and the scientific evidence available for their use. The GRADE system was used to classify scientific evidence and strength of recommendations. The consensus document contains 28 recommendations and 6 diagnostic algorithms. The document also describes criteria for referral from primary to specialised care.

© 2015 Elsevier España, S.L.U., AEEH and AEG. All rights reserved.

[☆] Please cite this article as: Fernández-Baños F, Accarino A, Balboa A, Domènech E, Esteve M, García-Planella E, et al. Diarrea crónica: definición, clasificación y diagnóstico. Gastroenterol Hepatol. 2016;39:535–559.

* Corresponding author.

E-mail address: ffbanares@mutuaterrassa.es (F. Fernández-Baños).

PALABRAS CLAVE

Diarrea crónica;
Sistema GRADE;
Recomendaciones
basadas en la
evidencia;
Documento
de consenso

Diarrea crónica: definición, clasificación y diagnóstico

Resumen La diarrea crónica es un síntoma de presentación frecuente, tanto en las consultas de medicina de familia como en las de digestivo. Se estima que >5% de la población sufre diarrea crónica y que cerca del 40% de estos sujetos son mayores de 60 años. El clínico se enfrenta con frecuencia a la necesidad de decidir cuál es el mejor enfoque diagnóstico de estos pacientes y elegir entre las múltiples pruebas diagnósticas existentes. En 2014 la Societat Catalana de Digestología creó un grupo de trabajo con el objetivo principal de crear algoritmos diagnósticos en base a la práctica clínica y evaluar las pruebas diagnósticas disponibles y la evidencia científica para su utilización. Para clasificar la evidencia científica y la fuerza de las recomendaciones se utilizó el sistema GRADE. Se han establecido 28 recomendaciones y 6 algoritmos diagnósticos. Se describen los criterios de derivación desde medicina primaria a digestivo de un paciente con diarrea crónica.

© 2015 Elsevier España, S.L.U., AEEH y AEG. Todos los derechos reservados.

Introduction

Purpose of the consensus document

Chronic diarrhoea is a common complaint seen by both primary care doctors and gastroenterologists. According to estimates, chronic diarrhoea has a prevalence of over 5%, with over 40% of cases occurring in the over-60 age group.¹ The list of possible causes is long (Table 1), and various diagnostic tests are usually needed before reaching a definitive diagnosis.¹⁻⁵ Clinicians are often faced with the challenge of deciding the best diagnostic approach in these patients, and must choose between the broad array of diagnostic tests currently available. A definitive diagnosis all too often proves elusive, and many patients are diagnosed with functional or idiopathic diarrhoea.

Several clinical practice guidelines have been published for the purpose of establishing the best investigation protocol in patients with chronic diarrhoea.^{2,4,6} The aim of these recommendations is to maximise positive diagnoses while minimising examinations. These guidelines need to be updated and adapted to current clinical practice, as in recent years a series of diseases with an underlying organic cause presenting with the characteristics of a "functional" disease have been identified. In fact, some authors have challenged the very existence of functional diarrhoea *per se*.^{7,8}

In 2014, with the aim of helping clinicians diagnose chronic diarrhoea, the *Societat Catalana de Digestología* proposed creating a working group to draw up a consensus document on the issue. The document was presented at the association's annual conference in January 2015, and an abbreviated version is available online (http://www.scdigestologia.org/index.php?link=docs_posicio). The primary aim of the group was to create diagnostic algorithms based on clinical practice, and to evaluate existing diagnostic tests and the clinical evidence supporting their use.

Scientific evidence and strength of recommendation were classified according to the Grading of Recommendations Assessment, Development and Evaluation (GRADE) system (<http://www.gradeworkinggroup.org/>). Table 2

shows the different categories used to grade certainty (or quality) of the evidence (CE) and strength of recommendation (SR).⁹

Definition of chronic diarrhoea

Chronic diarrhoea is defined as the passage of loose or liquid stools, urgent need to evacuate or feelings of abdominal discomfort, or increased frequency of these, lasting more than 4 weeks.^{2,5} Stool consistency is determined by the relationship between faecal water and the water-holding capacity of insoluble faecal solids. As stools consist predominantly of water (60–85%), consistency is difficult to quantify, and for this reason stool weight is used as a reasonable indirect estimation of consistency. Diarrhoea, therefore, can be defined by the weight or volume of stools measured over 24–72 h (on average, 2–3 days). The normal weight of stool output over a 24-h period in children and adults is less than 200 g; thus, stool weight >200 g/24 h is an objective definition of diarrhoea. However, it is important to note that up to 20% of patients with liquid diarrhoea, and thus a lower stool weight, are excluded from this definition.

A pragmatic definition incorporates the following elements: passage of loose or liquid stools more than 3 times daily and/or an output of 200 g/day of loose or liquid stools.

Patient history and classification

A detailed medical history and physical examination are essential in the assessment of patients with chronic diarrhoea.³⁻⁵ When taking the medical history, clinicians should first evaluate the patient's family history of diseases such as coeliac disease or inflammatory bowel disease, both of which have a familial component, their history of travel to regions where diarrhoea is endemic, engagement in risky sexual practices, history of systemic diseases (for example, diabetes mellitus, systemic or neurological diseases, amyloidosis, etc.) and gastrointestinal surgery (for example, cholecystectomy, intestinal resection), use of medicinal

Table 1 Classification of chronic diarrhoea.**Chronic watery diarrhoea****1. Osmotic**

- Osmotic laxatives (Mg^{+2} , PO^- , SO_4^{-2})
- Carbohydrate malabsorption
- Excessive consumption of poorly absorbed carbohydrates
- Lactulose
- Sorbitol and mannitol ("sugar-free" chewing gum)
- Fructose (fruit, soft drinks)

2. Secretory

- Congenital chloride diarrhoea
- Bacterial enterotoxins
- Bile acid malabsorption
- Inflammatory bowel disease
- Ulcerative colitis
- Crohn disease
- Microscopic colitis
- Vasculitis
- Laxative abuse
- Drugs
- Food allergies
- Heavy metal poisoning
- Dysmotility
- Post-vagotomy diarrhoea
- Post-sympathectomy diarrhoea
- Diabetic autonomic neuropathy
- Irritable bowel syndrome
- Faecal impaction
- Faecal incontinence
- Endocrine disorders
- Addison disease
- Hyperthyroidism
- Gastrinoma
- VIPoma
- Somatostatinoma
- Carcinoid syndrome
- Mastocytosis
- Other neoplasms
- Colorectal cancer
- Small bowel lymphoma
- Secretory villous adenoma of the rectum
- Idiopathic secretory diarrhoea
- Others: amyloidosis

Chronic inflammatory diarrhoea**Inflammatory bowel disease**

- Ulcerative colitis
- Crohn disease
- Diverticulitis
- Ulcerative jeunoileitis

Infectious diseases

- Bacteria: *Shigella*, *Salmonella*, *Campylobacter*, *Yersinia*, *Clostridium difficile*
- Viruses: herpes simplex, CMV
- Parasites: amoebiasis, strongyloides

Ischaemic colitis**Radiation colitis****Neoplasms**

- Colorectal cancer
- Lymphoma

Table 1 (Continued)**Chronic diarrhoea with steatorrhoea****Enteric causes**

- Mucosal diseases: coeliac, Whipple, giardiasis, lymphoma, Crohn, radiation enteritis, gastrointestinal lymphangiectasia, amyloidosis, eosinophilic gastroenteritis, tropical sprue, sprue, collagenous colitis.

Short bowel syndrome**Bacterial overgrowth****Chronic mesenteric ischaemia****Maldigestion syndromes**

- Exocrine pancreatic insufficiency

- Low bile acid levels in the intestinal lumen

products that could cause diarrhoea, or use of chewing gum or sweets with a high sorbitol content.

The differential diagnosis of diarrhoea has traditionally been based on its causative mechanisms. Diarrhoea is associated with 4 pathophysiological mechanisms: osmotic, secretory, exudative and altered motility. This classification, however useful from an academic perspective, is impractical in routine practice, because, in addition to other considerations, more than 1 mechanism is often present. From a practical standpoint, it is more useful to classify patients presenting with symptoms of diarrhoea according to "functional" or "organic" characteristics (Algorithm 1, Fig. 1).

Accordingly, the first step in the diagnostic process involves evaluating signs, symptoms and analytical tests suggestive of organic disease (Table 3). Alarm symptoms or abnormal findings in blood tests point to an organic cause. In these cases, diarrhoea can be characterised as inflammatory, malabsorption (steatorrhoea), or watery diarrhoea. The medical history will often help locate the intestinal segment causing the diarrhoea. The presence of large quantities of liquid or pasty stools with a shiny appearance,

Table 2 GRADE system.**Quality of the evidence (CE)**

- | | |
|----------|---|
| High | Further research is very unlikely to change our confidence in the estimate of effect. |
| Moderate | Further research is likely to have an important impact on our confidence in the estimate of effect and may change the estimate. |
| Low | Further research is very likely to have an important impact on our confidence in the estimate of effect and is likely to change the estimate. |
| Very low | Any estimate of effect is very uncertain. |

Grade of recommendation (SR)

- | | |
|--------|---|
| Strong | Benefits clearly outweigh risk and burdens, or vice versa. |
| Weak | Benefits closely balanced with risks and burdens, or some uncertainty in the estimates of benefits and risks. |

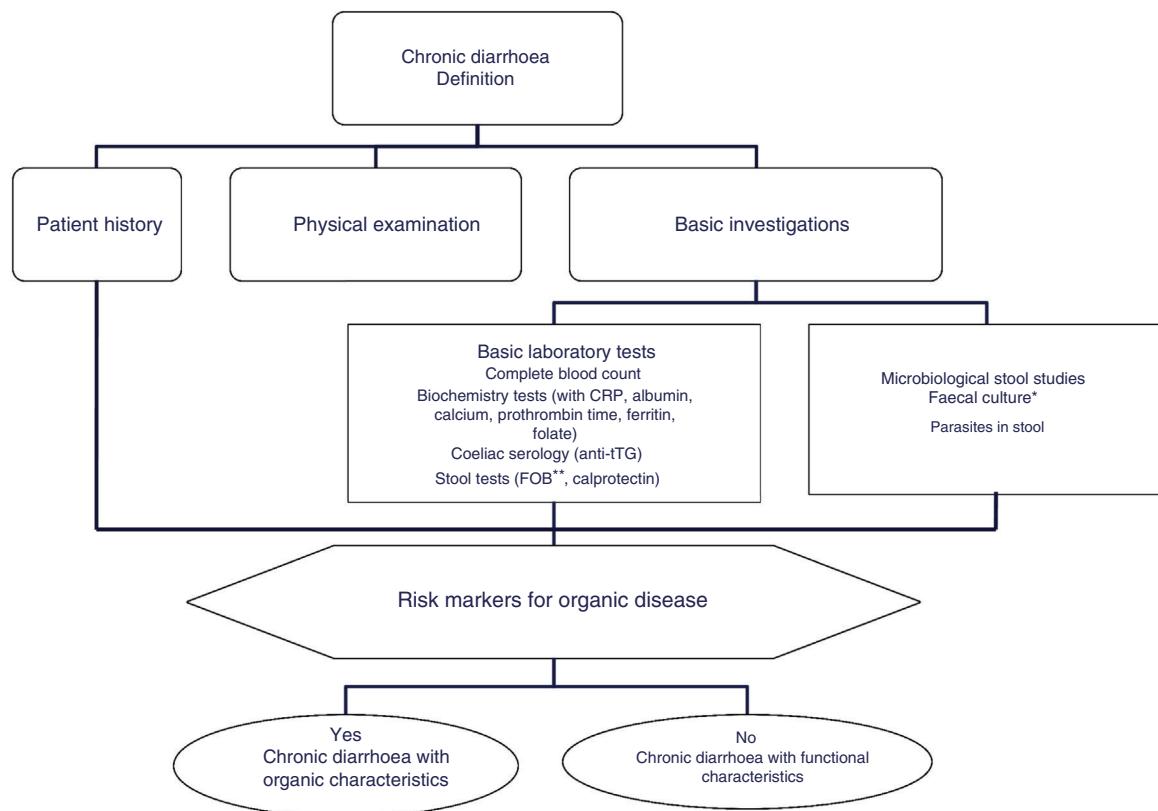


Figure 1 Algorithm 1. Initial approach in patients with chronic diarrhoea.

*Faecal culture is only indicated in immunocompromised patients or patients receiving immunosuppressants (in many cases these tests will have been performed during the acute phase of the disease, before the duration of symptoms suggests a chronic process).

**The faecal occult blood test has high sensitivity for intestinal inflammation.

CRP: C-reactive protein; FOB: faecal occult blood; tTG: tissue transglutaminase IgA test.

Table 3 Chronic diarrhoea: signs, symptoms and analytical findings suggestive of an organic cause.

Blood in stools
Fever
Recent weight loss (>5 kg) (in absence of concomitant depression)
Recent onset of symptoms, or change in the characteristics of previous symptoms
Onset at an advanced age (≥ 50 years)
Family history of colorectal cancer or polyps
Nocturnal diarrhoea
Diarrhoea persists after fasting
High-volume stool output or steatorrhoea
Weight of stool output over 24 h >400 g/day
Abnormalities on physical examination (pallor, hepatosplenomegaly, adenopathies, abdominal mass, etc.)
Anaemia, macrocytosis, hypoprothrombinaemia, hypoalbuminaemia or other laboratory findings (e.g. elevated ESR or C-reactive protein)
Positive faecal occult blood test, elevated faecal calprotectin

accompanied by cramp-like pain in the umbilical area (suggestive of malabsorption diarrhoea), would point to an origin in the proximal small intestine or pancreas. Small quantities of loose or liquid stools, however, mixed with blood, mucus or pus, associated with urgent evacuation or tenesmus and pain in the hypogastrium or sacrum, are more suggestive of an origin in the left colon and/or rectum (inflammatory diarrhoea).

The "functional" diarrhoea group, meanwhile, includes some entities with an organic origin and functional diarrhoea per se; these will be differentiated in the section on functional diarrhoea. These patients frequently present chronic non-bloody diarrhoea in either recurrent form, with frequent watery stools interspersed with episodes of normal bowel function, or in a persistent form, with passage of loose or liquid stools. The frequency of bowel movement will vary in these cases, but some patients may report up to 10–15 movements daily. Other common symptoms are urgency and faecal incontinence, which can prevent the patient from engaging in their normal activity and diminish their quality of life. Patients can also present moderate weight loss secondary to reduced food intake associated with an astringent or restrictive diet designed to prevent diarrhoea.

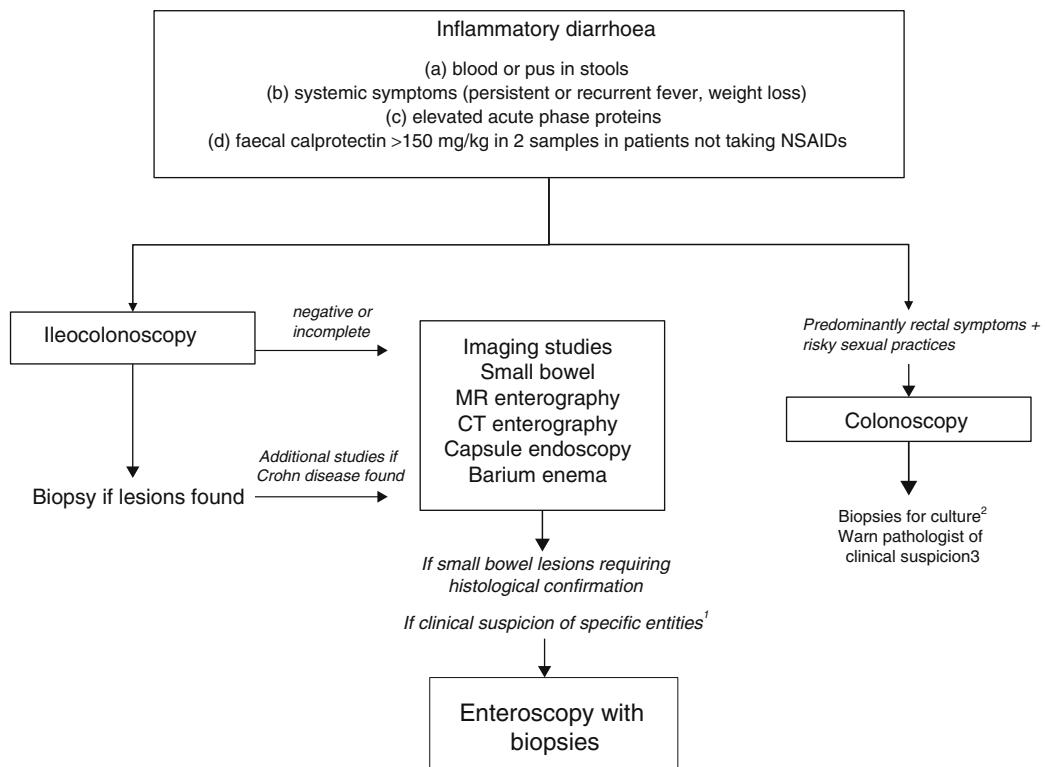


Figure 2 Algorithm 2. Diagnosis of chronic inflammatory diarrhoea

¹Certain entities may be hard to diagnose without initial clinical suspicion (for example, Whipple disease). In these cases, enteroscopy is indicated to obtain biopsy samples from the small bowel.

²Biopsy for gonorrhoea culture.

³Tissue PCR for lymphogranulomavenerum and herpes simplex.

Chronic diarrhoea with organic characteristics

Inflammatory diarrhoea

Chronic inflammatory diarrhoea is traditionally defined as the presence of white blood cells in stools. As these tests are not performed in most centres, a more practical definition is the following: (a) blood or pus in faeces; (b) accompanied by systemic symptoms (persistent or recurring fever, weight loss) or extra intestinal inflammatory manifestations (mainly affecting the joints, skin or eyes); and (c) elevated acute phase reactants (c-reactive protein, erythrocyte sedimentation rate, platelet count) or faecal calprotectin >150 mg/kg in 2 samples taken at different times in patients not taking NSAIDs.^{10,11}

As evaluation of the intestinal mucosa and histological confirmation are required for the diagnosis of most entities causing chronic inflammatory diarrhoea (Table 1), colonoscopy should initially be performed (with or without ileoscopy, depending on symptoms and colonoscopic findings as far as the caecum) (Algorithm 2, Fig. 2). If colonoscopy (with or without ileoscopy) is incomplete, or if findings are unremarkable, other small bowel imaging techniques should be used. Of these, magnetic resonance (MR) enterography, which is similar to CT enterography as regards diagnostic accuracy but with no harmful radiation, is the technique of choice. Alternatively, abdominal ultrasound with or without

IV contrast medium can be performed. This technique is similar in terms of diagnostic accuracy to both MR and CT, but is less costly, available in most centres, and does not irradiate the patient.¹² Because of this, abdominal ultrasound is an attractive choice for a preliminary study, particularly in paediatric patients. Ultrasound, however, can only accurately explore the terminal ileum, and is operator-dependent, two drawbacks that considerably limit its usefulness in our setting. If ultrasound, MR or CT are unavailable, a barium enema can be performed, although this technique has several drawbacks: lower diagnostic accuracy than MR, because it cannot detect extra intestinal complications; considerable radiation; and lack of training in this technique among the younger generation of radiologists. For all these reasons, barium enemas should be avoided. Abnormal findings from any of the foregoing will indicate the need for further studies, such as gastroduodenoscopy, enteroscopy, capsule endoscopy or specific imaging scans.

Chronic inflammatory diarrhoea secondary to infection is rarely found except in the immuno compromised patient population; the principle causes are summarised in Table 4. Faecal cultures, therefore, are only indicated in immuno compromised patients or those receiving immuno suppressants, and are typically tested for *Salmonella*, *Shigella*, *Yersinia* and *Campylobacter*. Serial sampling is not necessary, and samples can be refrigerated. Several methods can be used to test for *Clostridium difficile* (stool toxin assay,

Table 4 Causative agents of chronic infectious diarrhoea.^a

Bacteraemia (rare)	Protozoa	Viruses (Immunocompromised)
<i>Salmonella</i>	<i>Giardia</i>	<i>Norovirus</i>
<i>Shigella</i>	<i>Cryptosporidium</i>	<i>Citomegalovirus</i>
<i>Yersinia</i>	<i>Blastocystis hominis</i>	
<i>Campylobacter</i>	<i>Entamoebahistolytica</i>	
<i>Aeromonas</i>	<i>Dientamoeba</i>	
<i>Escherichia coli</i> (enteroinvasive)	<i>Helmintos</i> (<i>Strongyloides</i>)	
<i>Clostridium difficile</i>		

^a Faecal culture is only indicated in immunocompromised patients or patients receiving immunosuppressants.

faecal culture, cytotoxin assay, and glutamate dehydrogenase tests), although none has been shown to be clearly superior¹³; no single test will give maximum sensitivity and specificity while minimising costs and analysis time, and diagnostic algorithms recommend basing diagnosis on 2 or 3 consecutive tests.

Testing for parasites must be performed on 2 fresh samples taken on alternate days. If fresh samples cannot be obtained, patients can be given an appropriate container and medium for providing "fixed" refrigerated faeces samples. Parasites (trophozoite, cysts and eggs) are identified under direct microscopy, and the sensitivity of the test will depend on the intensity of colonisation, the freshness of the sample, and the expertise of laboratory personnel.

Special mention should be made of sexually transmitted gastrointestinal infections,¹⁴ which are usually characterised by rectal symptoms (rectal discharge, tenesmus, or faecal incontinence). These infections are associated with specific aetiological agents, they can mimic inflammatory lesions or tumours on endoscopy, and require specific diagnostic methods (Table 5).

Table 5 Diagnosis of sexually transmitted proctitis.

Aetiology	Diagnostic test of choice
Gonorrhoea	Biopsy culture
Lymphogranulomavenereum	Tissue PCR can identify serotypes, but is not widely available. Serology is useful for diagnosing patients with compatible symptoms and endoscopic findings, but given the high prevalence of <i>Chlamydia</i> infection in sexually active patients, the test is not specific for acute infection
Syphilis	Dark field microscopy findings of treponema in rectal exudate has low sensitivity but high specificity (valid in early stages of infection) Positive serology from 2 or 3 weeks after first infection
Herpes simplex	PCR ulcer biopsy

Recommendations

- 1 Determination of faecal calprotectin is recommended as a useful biomarker for chronic diarrhoea caused by inflammation (CE, high; SR, strong).
- 2 Initial investigation in a patient with suspected chronic inflammatory diarrhoea is colonoscopy (CE, high; SR, strong).
- 3 If colonoscopy is negative, incomplete, or ileoscopy could not be performed, other studies, preferably MR enterography, should be performed to evaluate the small intestine (CE, high; SR, strong).

Chronic diarrhoea due to malabsorption

Maldigestion is typically defined as decreased intraluminal hydrolysis of foodstuffs, and malabsorption as the reduced mucosal absorption of nutrients. Although this distinction is useful in terms of pathophysiology, maldigestion and malabsorption have similar clinical presentations and complications. For this reason, only the term malabsorption will be used in these guidelines.

In clinical practice, it is important to differentiate between diarrhoea due to enteropathy, diarrhoea due to bacterial overgrowth, and diarrhoea due to pancreatic disease. Diagnosis of each of these entities is described in a specific algorithm (Algorithms 3 and 4, Figs. 3 and 4).

Diarrhoea due to enteropathy

This is caused by malabsorption of nutrients secondary to enterocyte dysfunction. Coeliac disease is the paradigm of chronic diarrhoea due to enteropathy, and is by far the most frequent cause of villous atrophy, although it can also be caused by other entities (Table 6).¹⁵⁻²⁰ Duodenal biopsy (6 samples: 2 from the duodenal bulb and 4 from the distal duodenum) will usually yield a diagnosis, or at least confirm the existence of enteropathy, irrespective of its aetiology. In some cases, such as amyloidosis or Whipple disease, a duodenal biopsy is diagnostic due to the characteristic histopathological appearance of the tissue.²¹ Intestinal lesions caused by coeliac disease, however, are non-specific (for any grade of lesion, from lymphocytic enteritis to atrophy) (Tables 6 and 7).²²⁻²⁵ Diagnosis should be complemented or confirmed with other analytical tests (serology, genetic studies, lymphocyte subset tests) and clinical evaluations (good response to gluten-free diet).²⁶ As none of the foregoing criteria are in themselves sufficient for a diagnosis of coeliac disease, the use of score-based tests, such as the "4 out of 5" rule, has been suggested (Table 8).²⁷ Nonetheless, when diarrhoea due to enteropathy is suspected, an intestinal biopsy must be performed to either confirm suspicion or determine the extent of the lesion. The clinical context (epidemiological and/or family history, radiation enteritis caused by radiation therapy, bone marrow transplant in the case of graft-versus-host disease, pharmaceutical therapy, etc.) will guide diagnosis. Table 6 shows the main causes of enteropathy-related diarrhoea, together with diagnostic methods and the quality of the supporting evidence. It also describes the diagnostic

Table 6 Principle causes of enteropathy that can cause diarrhoea, and diagnosis.

Pathological entity	Histopathological findings	Special stains/molecular biology	Other diagnostic methods	CE
Coeliac disease	Lymphocytic enteritis to atrophy (Classification Table 7) <i>/Non-specific lesion</i>	CD3 immunostaining (CE, M)	Serology (Se, 85%-99%; Sp, 91%-100%) ^a Genetic testing (S, 93% Sp, 77%) ^a Flow cytometry study of lymphocyte subsets (Se, 85%; Sp, 100%) 4 out of 5 rule (Table 8)	H M M M GS
Giardiasis	Presence of <i>Giardia</i> / <i>Specific lesion</i> Normal mucosa (53%-96%) Atrophy (3%) Nodular lymphoid hyperplasia (35%) Eosinophilic infiltrate (35%)	No	Parasites detected in stool Formalin-ethyl acetate concentration (FEAC) Direct vision of eggs or cysts	GS
Tropical sprue	Lymphocytic enteritis to atrophy (more frequently subtotal atrophy) <i>/Non-specific lesion</i>	CD3 immunostaining (CE, M)	Therapeutic response to antibiotics, folic acid and vitamin B ₁₂ Negative coeliac serology Epidemiological exposure	L
Collagenous sprue	Lymphocytic enteritis to atrophy (more frequently atrophy) Collagenous band > 10 µm <i>/Specific lesion</i>	Trichrome Tenascin	No response to gluten-free diet Negative coeliac serology Response to steroids Possible association with refractory coeliac disease and lymphoma (see below)	L
Refractory coeliac disease (RCD):				
Type I	Atrophy <i>/Non-specific lesion</i>	CD3 immunostaining Normal phenotype	Ensure gluten-free diet Flow cytometry study of lymphocyte subsets: no aberrant phenotype	M
Type II	Atrophy <i>/Non-specific lesion</i>	Immunostaining: Aberrant phenotype (CE, M) Clonality testing (PCR) (CE, M)	Flow cytometry: aberrant immunophenotype CD3ε+CD8– and monoclonality Negative coeliac serology (previously positive)	M
Ulcerative jejunitis	Lymphocytic enteritis to atrophy <i>/Non-specific lesion</i>	CD3 immunostaining (CE, M)	Capsule endoscopy	L
Enteropathy-associated T-cell lymphoma	Atrophy Atypical lymphocytic infiltrate <i>/Specific lesion</i>	Immunostaining: aberrant phenotype (CE, M) Clonality testing (PCR) (CE, M)	Flow cytometry: aberrant immunophenotype CD3ε+CD8– and monoclonality Capsule endoscopy	M L
Other B and T lymphomas ^b (e.g. MALT)	Lymphoid hyperplasia or atypical lymphocytic infiltrate <i>/Lesion not always specific or not detectable with biopsy forceps</i>	Immunostaining ^b Subsets, proliferation and apoptosis	Imaging techniques Capsule endoscopy Double-balloon enteroscopy Cross-sectional radiography	L
Drug-induced enteropathy - olmesartan	Lymphocytic enteritis to atrophy <i>/Non-specific lesion</i>	CD3 immunostaining (CE, M)	Negative coeliac serology Positive genetic testing for coeliac disease 50%-60% Mucosa normalised after withdrawal of therapy	L L H

Table 6 (Continued)

Pathological entity	Histopathological findings	Special stains/molecular biology	Other diagnostic methods	CE
Whipple disease	Isolation of macrophages with PAS+ stain around <i>Tropherymawhipplei</i> /Specific lesion	Immunostaining <i>T. whipplei</i> -specific PCR (CE, M)	Isolation of <i>T. whipplei</i> in fluids or other involved tissues (synovial, CSF, stools, etc.)	M
Amyloidosis	Amyloide in submucosal vessels and possibly in mucosa /Specific lesion	Polarized light (Congo red) (CE, M)	Immunohistochemistry for: AL proteins (primary A) AA proteins (secondary A)	M

CE: quality of the evidence (H, high; M, moderate; L, low); GS: gold standard.

^a Coeliac serology: anti-endomysial, anti-tissue transglutaminase, anti-deamidated gliadin peptide antibodies. Sensitivity (Se) and specificity (Sp) levels generally refer to coeliac disease with atrophy. In lymphocytic enteritis due to coeliac disease, sensitivity can be as low as 15%.

^b B- and T-cell lymphoma classification: WHO 2008.³⁰

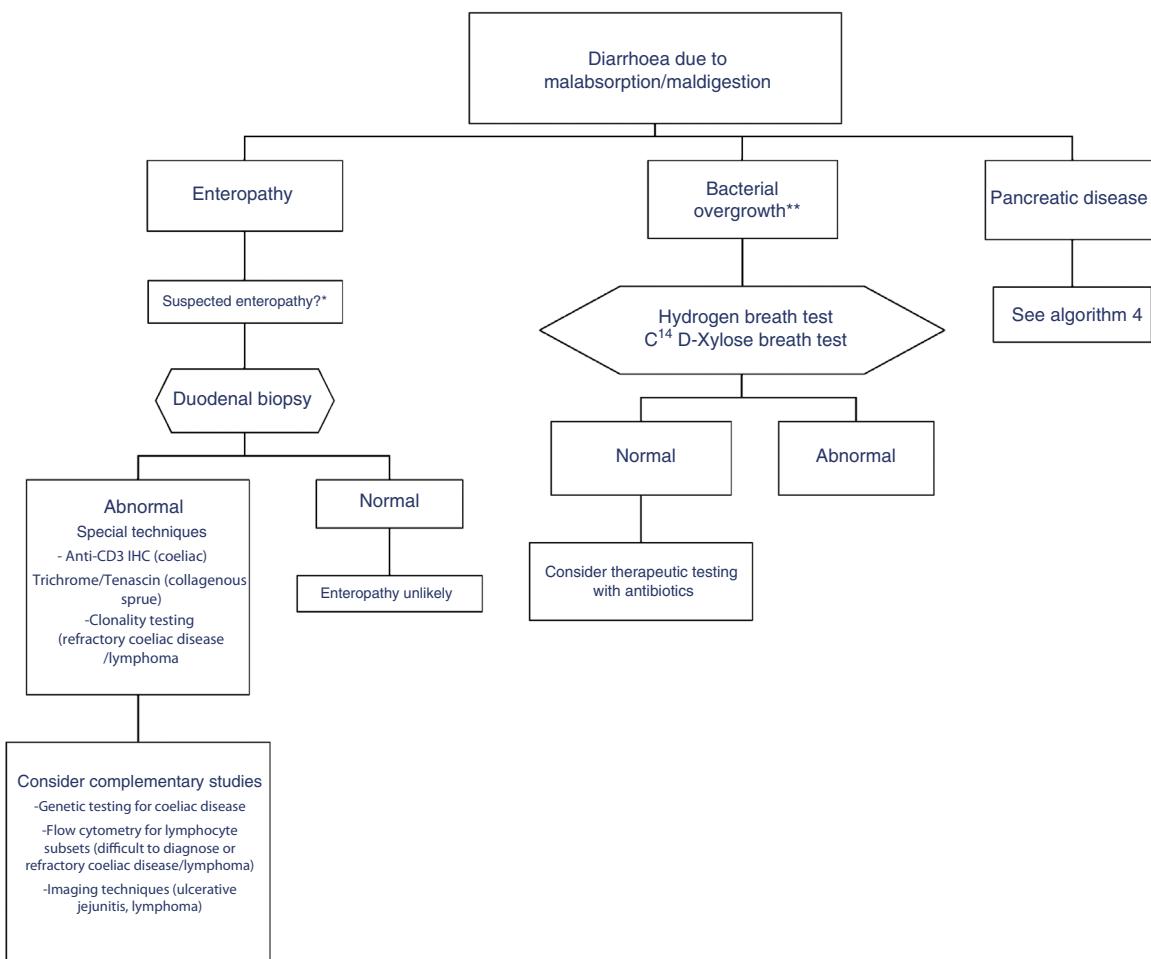


Figure 3 Algorithm 3. Diagnosis of chronic diarrhoea due to enteropathy and bacterial overgrowth

*Suspected enteropathy: risk group for coeliac disease (family members, Down syndrome, organ-specific systemic autoimmune diseases, compatible symptoms, etc.), visits to tropical countries, poorly controlled coeliac disease, olmesartan therapy, etc.

**Suspected bacterial overgrowth: structural changes in the small bowel (e.g. by-pass, stenosis) or motility disorder.

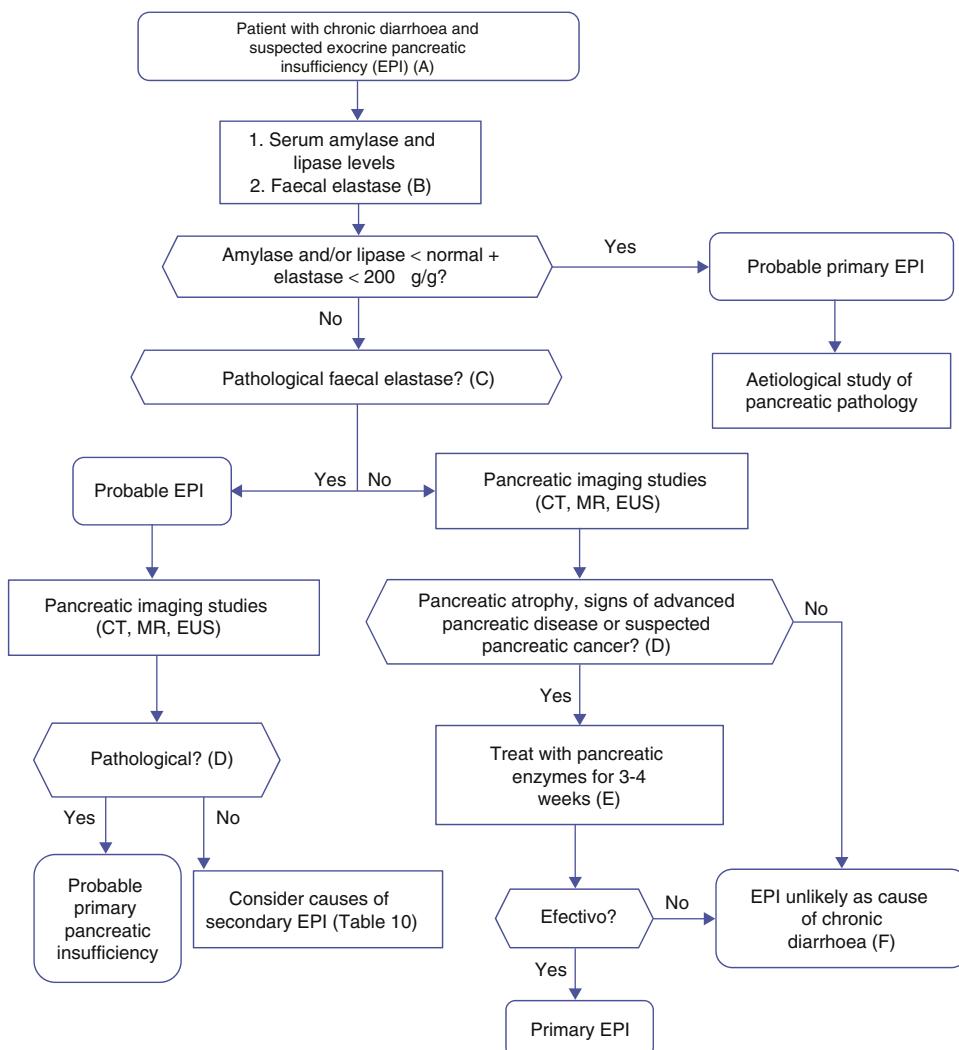


Figure 4 Algorithm 4. Diagnosis of chronic diarrhoea due to exocrine pancreatic insufficiency.

(A) **Strong suspicion.** Patients with heavy alcohol consumption and smoking habit, history of acute or recurrent pancreatitis, chronic pancreatic-type abdominal pain, suspicion of mutation in the cystic fibrosis gene (bronchiectasis, male sterility, family history of cystic fibrosis), suspicion or diagnosis of pancreatic cancer, previous pancreatectomy.

Suspicion. Occasional smoker, diabetes mellitus, gastrectomy, advanced age, diarrhoea improves with fasting, no anorexia.

(B) Faecal elastase is the most widely used pancreatic function test, and is available in most hospitals. It is performed on a solid stool sample. Liquid stool samples can give falsely low elastase levels.

(C) Concentrations below 100 µg/g are definitely pathological. Concentrations between 100 and 200 µg/g are also pathological, but with lower sensitivity and specificity; they should be viewed with caution. In patients with pancreatectomy, concentrations <200 µg/g can indicate exocrine insufficiency.

(D) Although it is a cause of primary pancreatic insufficiency, pancreatic atrophy can in itself be primary (due to pancreatic disease) or secondary to other factors, such as prolonged malnutrition or advanced age.

(E) Response to enzyme therapy should be evaluated following administration of an appropriate dose, and in the absence of bacterial overgrowth or other causes of enzyme inactivation or that contribute to the diarrhoea. Recommended initial lipase dose: 25000 IU with small meals, and 50000 IU with large meals (\pm proton pump inhibitor).

(F) Therapeutic testing can be used when no other cause for diarrhoea is found, and suspicion of exocrine insufficiency persists.

process for some specific entities, such as tropical sprue,²⁸ refractory coeliac disease and gastrointestinal lymphoproliferative disorders.^{20,29,30}

In addition to the foregoing, other less common diseases, such as abetalipoproteinemia, hypobetalipoproteinemia or hypogammaglobulinemia, can also cause diarrhoea.

Diagnosis is generally made in childhood, and suspicion is rarely guided by the enteropathy caused by these conditions, but rather by the clinical context and laboratory results (low levels of apoprotein B, triglycerides and cholesterol, in the former, and hypogammaglobulinemia with no B lymphocytes in the latter).

Table 7 Histopathological classification systems in coeliac disease.

Marsh 1992 ²²	Oberhuber 1999 ²³	Corazza 2005 ²⁴	Ensari 2010 ²⁵
Type 1 Infiltrative lesion	Type 1 Infiltrative lesion	Grade A	Type 1
Type 2 Crypt hyperplasia	Type 2 Crypt hyperplasia	Removed. Added to Grade A	Removed. Added to Type 1
Type 3 Atrophy	Type 3 Atrophy Type 3A. Partial Type 3B. Subtotal Type 3C. Total	Atrophy Grade B1 Grade B1 Grade B2	Atrophy Type 2 Type 2 Type 3
Type 4 Destructive lesion	Type 4 Destructive lesion	Obsolete	Obsolete

Table 8 The gold standard 4 out of 5 rule for the diagnosis of coeliac disease.

1. Typical symptoms (e.g., diarrhoea, growth delay, anaemia, etc.)
2. Positive for high titres of class A antibodies (IgG accepted in case of IgA deficiency, IgG anti-deamidated gliadin peptide antibodies strengthen diagnosis)
3. Positive for HLA-DQ2 and/or HLA-DQ8 genotypes
4. Enteropathy on intestinal biopsy (includes Marsh 1 –positive serology or associated with subepithelial deposits – to Marsh 3)
5. Clinical and serological improvement with gluten-free diet (histological improvement in sero-negative patients)

Source: adapted from Catassi and Fasano.²⁷

Recommendations

(See Table 6 for the quality of evidence of each diagnostic study.)

- 1 Determination of IgA anti-tissue transglutaminase antibody levels is the test of choice to screen for coeliac disease in patients with chronic diarrhoea (CE, high; SR, strong).
- 2 If blood tests are negative for coeliac disease, the presence of HLA-DQ2.5 should be determined. If positive, intestinal biopsies should be taken to rule out coeliac disease (CE, moderate; SR, strong).
- 3 In the case of suspected malabsorption syndrome, distal duodenal biopsy is indicated to diagnose coeliac disease or other types of enteropathy (CE, moderate; SR, strong).

Diarrhoea due to bacterial overgrowth

Diarrhoea due to bacterial overgrowth (BO) is caused by malabsorption of foodstuffs, including fats and carbohydrates.^{31,32} Certain situations involving high pH values, stasis or delayed gastric emptying can give rise to qualitative and quantitative changes in the bacteria colonising the proximal intestine, with a significant impact on the gut microbiota. The usual gut flora, consisting of lactobacilli, enterococci, and gram-positive facultative anaerobes, may be partially replaced with colic bacteria

(coliforms and anaerobes: *Bacteroides* and *Clostridium*), causing an increase in certain enzymes, such as colilamidase and protease. Colilamidases deconjugate bile salts, which are then more easily absorbed by passive transport in the proximal jejunum. This hampers micelle production, leading to malabsorption of fats. The increased proteases reduce villous surface enzymes, such as the disaccharidases, thus causing carbohydrate malabsorption. Small bowel bacterial overgrowth (SBBO) can be caused by a number of diseases, and should be suspected in any situation associated with impaired motility due to either structural (stenosis, by-pass, etc.) or functional (pseudo-obstruction, diabetic enteropathy, etc.) disorders. Increased pH in the duodenum or proximal jejunum (caused by proton pump inhibitor [PPI] therapy, gastrectomy or atrophic gastritis) increases the risk of SBBO due to absence of the bacteriostatic effect of the acid environment. The role of SBBO in the pathophysiology of irritable bowel syndrome (particularly associated with diarrhoea and/or distension) is still unclear, although it could be a causative factor in some patient subgroups. There is insufficient conclusive evidence to recommend the routine study of SBBO in these patients. Table 9 summarises the studies recommended for detecting bacterial overgrowth. Small-bowel aspiration and quantitative culture is considered the gold standard, but the complexity of the test rules out its use in routine clinical practice. As an alternative, the ¹⁴C/¹³C D-xylose breath test has been shown to be the most accurate, but has less supporting evidence than hydrogen breath tests.^{33,34}

Some authors have proposed therapeutic testing with antibiotics to detect SBBO when pre-test probability is high (clear predisposing cause and compatible clinical presentation).³³ Nevertheless, it is important to bear in mind that SBBO should usually be treated cyclically, and thus calls for an objective diagnostic test.

Recommendations

- 1 The hydrogen breath test has low sensitivity but adequate specificity for diagnosing SBBO, and is useful when results are positive. Hydrogen breath testing with glucose is recommended (CE, high; SR, strong).
- 2 A therapeutic test with antibiotics can be used to diagnose SBBO when pre-test probability is high (clear predisposing cause and compatible clinical presentation), and

Table 9 Recommended tests for bacterial overgrowth.

	Technique	Diagnostic accuracy	Level of evidence
Quantitative culture of jejunal fluid aspirate	Complex technique. Special sample collection techniques for anaerobic bacteria, avoiding contamination from pharyngeal bacteria, and rapid plating for anaerobic and aerobic bacteria. Bacterial overgrowth $>10^5$ CFU/ml in the proximal jejunum False positives: collection of samples from a diverticulum False negatives: collection of samples proximal to the structural abnormality	Gold standard	
Hydrogen breath test	50 g glucose challenge: bacterial overgrowth >20 ppm of H ₂ for 2 h after administration 10 g lactulose challenge: bacterial overgrowth >20 ppm of H ₂ in the first 90 min (early peak), or presence of a double peak, or sustained increase >10 ppm over baseline value False negatives: Non-H ₂ producing flora False positives: Very rapid oro-caecal transit (e.g., gastrectomy)	Se 62.5% Sp 81.8%	H
C ¹⁴ D-xylose breath test	Administration of 1 g of ¹⁴ C-labelled D-xylose Bacterial overgrowth: detection of ¹⁴ C in exhaled breath Fewer false positives for D-xylose. Absorbed in small bowel and does not reach colon	Se and Sp, $\sim 85\text{--}90\%$	H

CFU: colony forming units; Se: sensitivity; Sp: specificity.

the hydrogen breath test is negative or not available (CE, low; SR, weak).

Chronic diarrhoea caused by exocrine pancreatic insufficiency

Pancreatic enzyme deficiency results in malabsorption of foodstuffs (particularly fats), which results in loose or liquid stools and increased daily stool output. Steatorrhoea is defined as the elimination of >7 g/day fat with a dietary intake of 100 g fat/day (fat absorption coefficient $<93\%$). Pancreatic steatorrhoea only occurs when the functional reserve capacity of the pancreas is severely depleted.³⁵

Moderate steatorrhoea might not be associated with diarrhoea. Constipation is a common symptom in patients with chronic pancreatitis or cystic fibrosis and severe pancreatic insufficiency. Diarrhoea secondary to primary pancreatic disease is not usually particularly voluminous, hardly ever watery (can be oily), and improves with fasting. Patients may suffer weight loss without anorexia, unless insufficiency is due to pancreatic cancer.

Pancreatic insufficiency can be primary (due to pancreatic disease) or secondary. In the latter, suboptimal enzyme levels are only partially responsible for the diarrhoea. However, administration of enzymes can improve symptoms in some cases, such as gastrectomy, diabetes mellitus and pancreatic cancer. Table 10 summarises the causes of pancreatic insufficiency. The most common primary cause is chronic pancreatitis, followed by cystic fibrosis and pancreatic cancer.

Table 10 Causes of exocrine pancreatic insufficiency.

Primary causes

1. Chronic pancreatitis
2. Cystic fibrosis
3. Pancreatic cancer
4. Acute pancreatitis (generally transient)
5. Duct obstruction (including ampulloma, cysts and MPTs)
6. Pancreatectomy
7. Senile pancreatic atrophy, acquired (persistent severe malnutrition) or congenital (BDS, Johanson-Blizzard, pancreatic lipomatosis, dorsal pancreatic agenesis, MODY) atrophy

Secondary causes

1. Coeliac disease and other enteropathies (Crohn disease, common variable immune deficiency with intestinal involvement, eosinophilic enteritis, etc.)
2. Gastrectomy – pancreatic stent (postprandial asynchrony)
3. Zollinger-Ellison syndrome
4. Drugs (long-term octreotide therapy)

Of uncertain origin

1. Diabetes mellitus
2. Irritable bowel syndrome
3. Chronic renal failure

Tests for exocrine pancreatic insufficiency

We will address the diagnosis of pancreatic insufficiency as a cause of chronic diarrhoea, and not as a factor in the diagnosis of chronic pancreatitis. More than 20 tests for pancreatic insufficiency have been described, but none give optimal results in clinical practice. Some are non-diagnostic (cerulein-secretin stimulation test), or are no longer in use (faecal chymotrypsin). As far as common clinical studies are concerned, the sensitivity, specificity and predictive value of the tests will depend to a large extent on the chosen reference value (steatorrhoea vs chronic pancreatitis) and the underlying disease (cystic fibrosis, pancreatectomy, chronic pancreatitis, etc.).

Tests that yield relevant clinical information

- 1 *Serum amylase, lipase and trypsin levels.* Due to their low sensitivity, these tests are rarely used. However, low levels of these enzymes are indicative and specific (>90%) for primary exocrine insufficiency, particularly in patients with cystic fibrosis, advanced chronic pancreatitis, pancreatic cancer and Shwachman–Diamond syndrome. As diarrhoea only occurs with severely impaired pancreatic function, pancreatic enzymes could have adequate sensitivity in cases of chronic diarrhoea; however, this hypothesis has not been tested.
- 2 *Evidence.* In paediatric patients with cystic fibrosis, low lipase and trypsin levels correlate with steatorrhoea,³⁶ with a sensitivity of 95% and 93%, respectively, and a specificity of 86% and 92%.³⁷ In patients with chronic pancreatitis, low trypsin or amylase levels have a sensitivity of 70–85% for steatorrhoea³⁸, but less than 50% for mild exocrine insufficiency measured by the secretin stimulation test.³⁹ Specificity for pancreatic steatorrhoea is between 90% and 100%.⁴⁰ Low or undetectable serum lipase levels are typical of congenital lipase deficiency, a disease that presents with steatorrhoea and normal faecal elastase.
- 3 *Faecal pancreatic elastase determination.* This is the gold standard for detecting pancreatic insufficiency, and is recommended in British guidelines for chronic diarrhoea.⁵ The test determines levels of human pancreatic elastase (but not enzymes) in stool. Results are not affected by pancreatic enzyme therapy. It is a simple, inexpensive and reproducible test (15% individual variability),⁴¹ but can yield false positives when used with liquid stool samples. Concentrations of >200 µg/gelastase per g of stool are normal, while concentrations of <100 indicate severe pancreatic insufficiency. Concentrations of between 100 and 200 are suggestive of pancreatic disease, but should be considered in the general context of the case. Evidence for sensitivity and specificity levels is conflicting, but it is generally considered a useful screening test for severe pancreatic insufficiency, which is the clinical situation associated with chronic diarrhoea, in the absence of other factors (for example, bacterial overgrowth). In a preliminary analysis, a cut off value of <200 µg/g was found to have a sensitivity of 63% for mild and 100% for moderate to severe insufficiency (compared with the cerulein-secretin test), with a specificity of 93% and significant correlation with enzyme and bicarbonate secretion.⁴¹ Values under 100 µg/g have a sensitivity and specificity for steatorrhoea of 93% and 81%, respectively.⁴² In patients with pancreaticoduodenectomy, faecal elastase concentrations of <200 µg/g are accompanied by steatorrhoea.⁴² Sensitivity for detecting steatorrhoea in these cases is 91%, but specificity is only 35%.⁴³ The difference in these cases arises because exocrine insufficiency is accompanied by anatomical abnormalities that prevent optimal mixing of nutrients with bile salts and with the enzymes and bicarbonate secreted by the pancreatic remnant (postprandial asynchrony).
- 4 *Faecal chymotrypsin.* Enzyme levels <3 U/g have a specificity of between 49% and 100% and a sensitivity of between 50% and 90% for chronic pancreatitis. The specificity for pancreatic disease is extremely high (90–100%) for determining both the origin of steatorrhoea⁴⁴ and for detecting pancreatic insufficiency in patients with cystic fibrosis.⁴⁵ The test has a sensitivity of 69% and a specificity of 89% for detecting pancreatic insufficiency in patients with chronic pancreatitis.⁴¹ When used with watery stools, it can yield false positives. Test kits are not widely available.
- 5 *¹³Cmixedtriglyceride breath test.* In this test, concentrations of ¹³C in exhaled breath are measured following intake of triglycerides labelled with ¹³C. It has a sensitivity of 89% and a specificity of 81% for the diagnosis of pancreatic steatorrhoea.⁴⁶ Compared with direct secretion tests (secretin-cerulein) and with faecal elastase and chymotrypsin, it has a sensitivity of 85% and a specificity of 100% for diagnosing severe pancreatic insufficiency, but only 69% and 46%, respectively, for moderate insufficiency. The test is not superior to the sensitivity and specificity of the faecal elastase test.⁴⁷ Presence of bacterial overgrowth can give false negatives, particularly in patients with pancreatectomy.⁴⁸ Other factors that can distort results are a diet rich in foodstuffs containing ¹³C, abnormal gastric emptying, physical exercise, and basal CO₂ production. Drawbacks include the duration of the test (between 6 and 8 h), the requirement for patients to fast and abstain from exercise, and the need for specially trained personnel.
- 6 *Faecal fat and coefficient of fat absorption.* Quantification of faecal fat is a good marker of exocrine insufficiency in patients with a known pancreatic parenchymal lesion. The total fat content in stool samples taken over 72 h is measured. Patients are required to eat a diet containing 100 g fat/day for 2 days prior to the start of the test. Faecal fat >7 g/day indicates steatorrhoea (coefficient of fat absorption <93%). Although this is the gold standard for steatorrhoea screening,⁴³ it is not specific for pancreatic disease.
- 7 *Endoscopic pancreatic function test.* This test is only performed in some hospitals. Pancreatic secretions in response to secretin or cerulein stimulation are aspirated through the endoscope at certain time intervals. The test has a sensitivity of 83% and a specificity of 87% for diagnosis of chronic pancreatitis.⁴⁹ A reduced version of the test, in which secretions are aspirated over 15 min, also gives good results.⁵⁰

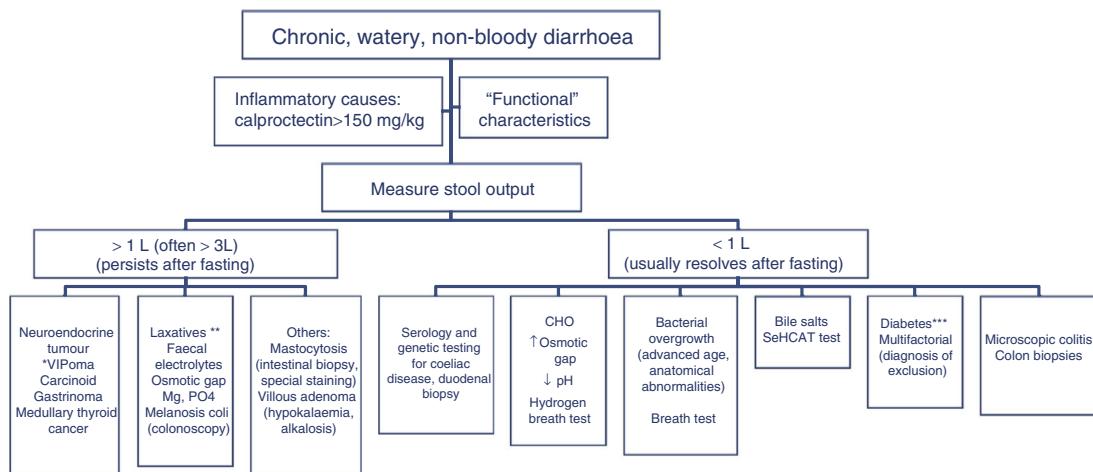


Figure 5 Algorithm 5. Diagnosis of chronic watery non-bloody diarrhoea.

Other causes of <1 L/day: pharmacological therapy, hyperthyroidism, alcohol.

*Serum peptide panels have a false positive rate of 45% (positive predictive value <1%); only request these when there are signs and symptoms of neoplasm (Table 11) or when CT/MR or octreoscan findings indicate a tumour.

**Can be high- or low-volume, depending on the dose administered; can respond to fasting. A finding of melanosis coli on colonoscopy suggests regular use of anthraquinone laxatives, such as senna and cascara sagrada.

***In diabetic patients, diarrhoea can have multifactorial causes: excess fructose consumption, bacterial overgrowth, increased risk of coeliac disease, and microscopic colitis in type 1 DM, some pharmacological therapies (metformin, acarbose), autonomic neuropathy.

CHO: carbohydrates (lactose, fructose and/or sorbitol).

- 8 *Secretin-enhanced magnetic resonance cholangiopancreatography*. The test quantifies duodenal secretions in response to secretin infusion. Imaging findings can discriminate patients (chronic pancreatitis) from healthy volunteers,⁵¹ and correlate with faecal elastase test findings.⁵² The test only measures volume of secretions, and is not standardised.
- 9 *Morphological investigations*. The greater the morphological abnormalities in imaging studies, the greater the pancreatic dysfunction. Pancreatic insufficiency rarely occurs in patients with normal pancreatic morphology. Correlation between imaging and direct function test results is good, but not perfect.^{53,54} In patients with partial pancreatectomy (in whom pancreatic function testing is difficult or inaccurate), the main duct diameter/parenchymal thickness ratio correlates well with exocrine insufficiency.⁵⁵
- 10 *Therapeutic testing with pancreatic enzymes*. Therapeutic testing is used when there is a strong suspicion of pancreatic insufficiency and the results of other tests have proved inconclusive, and is a useful confirmatory test. For example, in a group of patients with diarrhoea-predominant irritable bowel syndrome, 6% had faecal elastase levels <100 mg/g, and symptoms improved after enzyme therapy.⁵⁶

Recommendations

- 1 In chronic diarrhoea investigations, determination of faecal elastase in non-watery stool samples is the gold standard for evaluating exocrine pancreatic insufficiency (CE, high; SR, strong). The labelled triglyceride test can

be a good alternative in hospitals where this test has been validated (CE, moderate; SR, weak).

- 2 If the findings of the former test are inconclusive, morphological abnormalities on imaging studies can indicate exocrine pancreatic insufficiency (CE, low; SR, strong).
- 3 If diagnostic tests are inconclusive, or when pancreatic insufficiency is merely a contributing factor to chronic diarrhoea, therapeutic testing with pancreatic enzymes may be indicated (CE, low; SR, weak).

Watery, non-bloody diarrhoea with organic characteristics

In patients with chronic watery, non-bloody diarrhoea with organic characteristics (Table 3), diagnosis is based on the strategy outlined in algorithm 5 (Fig. 5). These usually involve secretory diarrhoea characterised by large-volume, watery stools, often more than 1 L per day; persistence of diarrhoea after fasting; and stool-water tests that show measured osmolality to be identical to that calculated from its electrolyte concentration (see below).⁵⁷ Agents causing secretions that can lead to chronic diarrhoea include various hormones and substances produced by neuroendocrine tumours (Table 11).^{57–59} However, watery, secretory diarrhoea is only caused by such tumours in 1 in 5000 to 1 in 500,000 patients with chronic diarrhoea, depending on the type of tumour.

Some patients present chronic secretory diarrhoea due to abuse of laxatives. The presence or absence of large-volume watery stools will depend on the laxative dose taken, and symptoms will improve with fasting. This aetiology should be suspected in the following circumstances^{60,61}:

Table 11 Neuroendocrine tumours that cause diarrhoea, and their markers.

Tumour	Characteristic symptoms	Tumour marker
Gastrinoma	Zöllinger-Ellison syndrome: peptic ulcer, diarrhoea, steatorrhoea	Gastrin
VIPoma	Verner-Morrison syndrome: diarrhoea (>3 L/day), hypokalaemia, hypochlorhydria	VIP
Medullary thyroid cancer	Thyroid nodule	Calcitonin, prostaglandins
Carcinoid	Flushing, bronchospasm, right-sided valve disease	Serotonin, kinins
Somatostatinoma	Diabetes mellitus, gall stones, steatorrhoea	Somatostatin
Glucagonoma	Rash (necrolytic migratory erythema), diabetes	Glucagon
Mastocytosis	Flushing, urticariapigmentosa, abdominal pain, vomiting	Histamine

(a) bulimic patients (usually young or adolescent women concerned about their body weight, or with confirmed eating disorders); (b) patients with an ulterior motive (economic, manipulation of family members); (c) Munchhausen syndrome (patients with a psychological need to be a diagnostic challenge); and (d) Munchhausen's syndrome by proxy (children or dependent adults given laxatives by their guardian or carer in order to seek personal benefit).

In patients with chronic secretory diarrhoea associated with hypokalaemia and metabolic alkalosis, large villous adenomas should be suspected

Diagnosis

Routine testing for gastrointestinal peptides in blood or urine has a false positive rate of 45%, and is not recommended in the diagnosis of patients with chronic diarrhoea.⁶² Considering, moreover, that the pre-test probability of diagnosing a neuroendocrine tumour as the cause of chronic secretory diarrhoea is extremely low, the positive predictive value of these tests is less than 1%. Serum determination of these peptides or their urinary metabolites should therefore only be performed in patients with chronic diarrhoea and signs and symptoms consistent with neoplasia (Table 11) or with evidence of a neuroendocrine tumour on imaging studies (CT, MR, EUS). Most neuroendocrine tumours secrete chromogranin A; however, routine testing for this protein contributes little to diagnosis, as high levels have been described in other types of cancer (pancreatic, prostate, small cell lung cancer), renal failure, diarrhoea-predominant IBS, inflammatory bowel disease, and collagenous colitis, as well as in atrophic gastritis and PPI therapy, probably due to enterochromaffin cell hyperplasia.^{63,64} The specificity of this marker for diagnosis of neuroendocrine tumours is only 10–35%, with a sensitivity of around 60%.⁶³ An octreotide scan, or octreoscan, can be useful for identifying peptide-producing neuroendocrine tumours.⁶⁵

A finding of melanosis coli on colonoscopy suggests regular use of anthraquinone laxatives, such as senna and cascara sagrada. However, a histological finding of pseudomelanosis coli in biopsy samples from a macroscopically normal colon is non-specific, and has been associated with increased epithelial apoptosis secondary to use of these laxatives and other drugs.⁶⁶ Abuse of laxatives derived

from magnesium sulphate, phosphates and other sulphates can be detected with electrolyte and stool osmotic gap panels.⁶⁷ Normal stool osmotic gap values are between 50 and 125 (the osmotic gap is the results of subtracting stool osmolality [290mOsm] from $2^* \text{Na}^{+} + \text{K}^{+}$ in stool water). In secretory diarrhoea, the osmotic gap is less than 50, while in osmotic diarrhoea it increases to >125. Patients with diarrhoea secondary to Mg^{2+} present high osmotic gap values and stool $\text{Mg}^{2+} > 50 \text{ mmol/L}$. Diarrhoea caused by intake of sodium sulphate (Na_2SO_4) and disodium sulphate (Na_2PO_4) mimic secretory diarrhoea and can be diagnosed by findings of low Cl^{-} concentration in faecal water (usually <20 mmol/L).

In factitious diarrhoea, attempts to tamper with findings can involve the addition of water or urine to stool samples. This can be detected with stool osmolality testing, and is shown as reduced values (<25 mOsm) in the case of contamination with hypotonic urine or water, or increased values (>375 mOsm) in the case of contamination with concentrated urine. If suspicion is high, spectrofluorimetry or chromatography studies may be needed to determine the presence of laxatives in urine or stool samples.

Recommendations

- 1 Serum peptide panels to screen for a neuroendocrine tumour in patients with chronic diarrhoea have a positive predictive value of less than 1%, and routine use is discouraged (CE, moderate; SR, strong).
- 2 CT and MR scans are useful for diagnosing and staging neuroendocrine tumours. An octreotide scan, or octreoscan, can be useful for identifying functioning neuroendocrine tumours (CE, high; SR, strong).
- 3 Routine determination of chromogranin A in patients with chronic watery diarrhoea has low specificity and is of little diagnostic value (CE, high; SR strong).
- 4 A histological finding of pseudomelanosis coli in biopsy samples from a macroscopically normal colon is non-specific and should not be taken as a marker of laxative abuse, as it has also been associated with administration of other drugs (CE, moderate; SR, strong).
- 5 Stool osmotic gap measurements can be useful in the diagnosis of chronic large-volume watery diarrhoea (CE, moderate; SR, strong).

Table 12 Differential diagnosis of functional diarrhoea.*Drug-induced diarrhoea**Sugar malabsorption*

Lactose

Fructose-sorbitol

*Bile acid malabsorption**Microscopic colitis**Celiac disease**Giardiasis**Bacterial overgrowth**Exocrine pancreatic insufficiency**Inflammatory bowel disease***Chronic diarrhoea with functional characteristics****Functional diarrhoea**

Chronic functional diarrhoea is defined as the persistent or recurrent passage of loose or liquid stools for more than 4 weeks with no obvious organic cause. If symptoms have lasted for at least 6 months, and occur in more than 75% of bowel movements in the previous 3 months, a diagnosis of functional diarrhoea can be established according to Rome III diagnostic criteria for functional gastrointestinal disorders. According these criteria, diarrhoea accompanied by abdominal pain that improves or is associated with loose stools indicates a diagnosis of diarrhoea-predominant irritable bowel syndrome.⁶⁸

Functional diarrhoea may affect around 5% of the general population,⁶⁸ and differential diagnosis must consider a wide range of disorders that can cause chronic diarrhoea (Table 12) (Algorithm 6, Fig. 6). In young patients with no alarm signs or symptoms, an unremarkable physical examination, and mild diarrhoea with no night symptoms and little impact on the patient's daily activity, general laboratory tests including serology for coeliac disease and a stool parasite test should be performed. If symptoms persist, are incapacitating, or significantly impact the patient's quality of life, other investigations will be needed to rule out an organic aetiology (Table 13).^{7,69–72} Some organic diseases, such as microscopic colitis, choleretic diarrhoea or sugar malabsorption diarrhoea, can mimic functional diarrhoea, and are discussed below.

Microscopic colitis

Microscopic colitis (MC) is a generic term used mainly to describe 2 entities: collagenous colitis (CC) and lymphocytic colitis (LC). MC describes a form of chronic, recurrent inflammatory bowel disease characterised by (a) chronic or intermittent passage of non-bloody watery stools; (b) macroscopically normal or nearly normal colonic mucosa on colonoscopy; and (c) characteristic histopathological findings (Table 14).⁷³

In both these entities, symptoms are usually similar to functional diarrhoea or irritable bowel syndrome, and general laboratory tests are usually negative. Both CC and LC are rare entities, and many clinicians and pathologists are

Table 13 Recommended investigations in patients with suspected functional diarrhoea.

Clinical situation	Investigations
All patients with chronic diarrhoea	Basic blood tests with serology for coeliac disease Stool parasite test
Mainly postprandial diarrhoea	Therapeutic testing with cholestyramine SeHCAT test (if available)
Diarrhoea with abdominal distension	Hydrogen or methane breath test with lactose Hydrogen or methane breath test with fructose-sorbitol Hydrogen or methane breath test with glucose
Nocturnal diarrhoea	Faecal calprotectin test Total colonoscopy with multiple biopsies
Diarrhoea in patients over 50 years of age	Total colonoscopy with multiple biopsies
Diarrhoea in patients with family history of colorectal cancer	Apply colorectal cancer prevention protocol, taking multiple colon biopsies
Diarrhoea in patients with family history of inflammatory bowel disease	Faecal calprotectin test
Diarrhoea in patients with family history of coeliac disease	Genetic testing for coeliac disease: if positive - gastroscopy with duodenal biopsies
Refractory diarrhoea	Genetic testing for coeliac disease Gastroscopy with duodenal biopsies Total colonoscopy with multiple biopsies Faecal elastase

Table 14 Histopathological diagnostic criteria for microscopic colitis.*Collagenous colitis*

Subepithelial collagenous band >10 µm

Total intraepithelial lymphocytes >7 per 100 epithelial cells

Epithelial lesion (detachment, flattening)

Chronic inflammation of the lamina propria

Lymphocytic colitis

Total intraepithelial lymphocytes >20 per 100 epithelial cells

Epithelial lesion (detachment, flattening)

Chronic inflammation of the lamina propria

Subepithelial collagenous band <10 µm

unfamiliar with their diagnosis. A recent epidemiological study carried out in Spain reported a mean annual incidence of MC of 4.8/10⁵ inhabitants/year.⁷⁴ Although it can present in young patients, incidence peaks in women aged 60 years or more, and is higher than Crohn disease and ulcerative

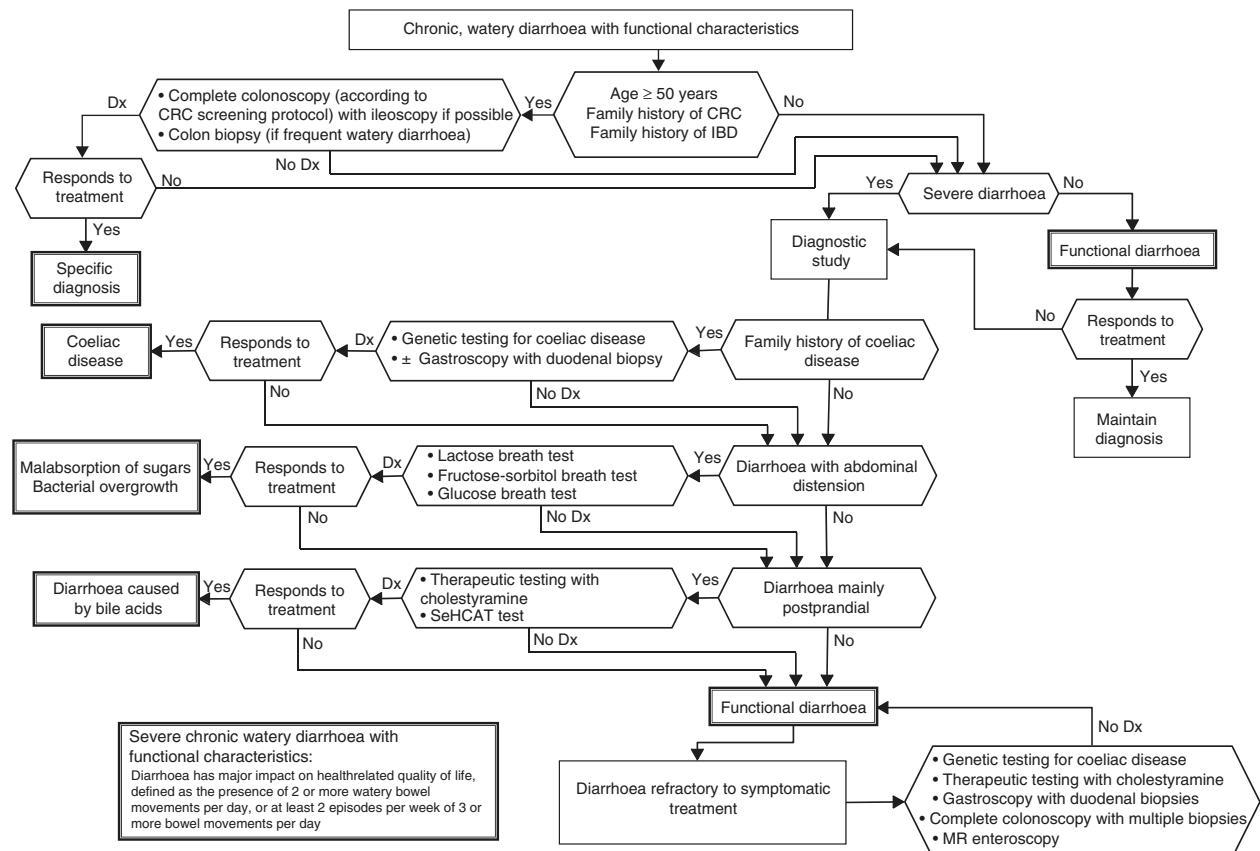


Figure 6 Algorithm 6. Diagnosis of watery diarrhoea with functional characteristics

All patients with chronic diarrhoea with no signs or symptoms of alarm, no abnormal findings on laboratory tests (including serology for coeliac disease) and no parasites in stool samples, are *a priori* diagnosed with functional diarrhoea. If such patients are aged 50 years or over, or have a family history of colorectal cancer (CRC), a complete colonoscopy should be performed as part of the screening protocol for individuals at moderate risk of CRC. During colonoscopy, biopsies should be taken from various sites along the colon to rule out microscopic colitis in patients with frequent passage of watery stools. A family history of inflammatory bowel disease is also an indication for total colonoscopy with ileoscopy. If no pathology is found, the protocol for patients aged under 50 years with no family history should be applied; in other words, if the diarrhoea is not serious, a diagnosis of functional diarrhoea should be established and the patient treated accordingly. In patients with severe diarrhoea, it is advisable to perform diagnostic tests. Therefore, in patients with a family history of coeliac disease, it is advisable to perform genetic testing for coeliac disease (HLA-DQ2/8). If positive, duodenal biopsy should be performed. In patients presenting diarrhoea with abdominal distension, it is important to bear in mind that, although distension can be caused by various functional gastrointestinal disorders, it is advisable to perform a hydrogen breath test to rule out intolerance to sugars (lactose or fructose and sorbitol) or bacterial overgrowth. In patients with mainly postprandial diarrhoea, it should be borne in mind that although postprandial onset is common in functional gastrointestinal disorders, it is essential to rule out bile acid malabsorption, ideally using the SeHCAT test. If this is not available, then therapeutic testing with cholestyramine should be performed. If these tests are not diagnostic, or if they are diagnostic but with no therapeutic response, the diagnosis of functional diarrhoea should be maintained. All patients with undiagnosed severe functional diarrhoea that does not respond to symptomatic treatment should undergo a comprehensive study to rule out bile acid malabsorption, coeliac disease, microscopic colitis or inflammatory bowel disease.

colitis in this population. MC is a common diagnosis in this age group, and should be one of the first options in the differential diagnosis of chronic non-bloody watery diarrhoea.

Diagnostic process

Biopsies from different sites along a macroscopically normal colon is the gold standard for diagnosis of CC and LC in patients with chronic non-bloody watery diarrhoea.⁷³ This will confirm a diagnosis of MC in between 8% and 16% of patients with chronic non-bloody watery diarrhoea referred for colonoscopy,^{74–78} in up to 17% of women aged 50 years or

over, and in 23% of men aged 70 years or over.⁷⁸ Histopathological diagnostic criteria are consistent and reproducible, with excellent inter-individual and intra-individual agreement to discriminate between MC, normal histology and others (inter-observer agreement >90%),^{79,80} with slightly less agreement to discriminate between CC, LC, and incomplete MC (70–80% agreement).⁸⁰

A recent study suggests that at least 2 diagnostic biopsies from 2 different segments of the colon are needed to diagnose CC and LC, with at least mild chronic inflammation of the lamina propria in the other biopsies and segments.⁸¹ The

study in question showed that a diagnostic collagenous band ($>10\text{ }\mu\text{m}$) was found in 5 segments in only 47% of patients. It also showed that the diagnostic yield of biopsies taken from the ascending, transverse and descending colon was 96.2% higher than other sampling strategies. Therefore, it is recommended to take 2 biopsies each from, at least, the right, transverse and left colon, and store them in separate containers.

Recommendations

- 1 Biopsies from different sites along a macroscopically normal colon is the gold standard for diagnosis of CC and LC in patients with chronic non-bloody watery diarrhoea (CE, moderate; SR, strong).
- 2 It is recommended to take 2 biopsies each from, at least, the right, transverse and left colon, and store them in separate containers (CE, moderate; SR, strong).

Diarrhoea caused by bile acids

Bile acids are synthesised in the liver, secreted by the gall bladder and released in the duodenum, where they facilitate absorption of liposoluble vitamins and aid in fat digestion. They are then reabsorbed in the terminal ileum, from where enterohepatic circulation returns them to the liver. Less than 5% reach the colon.⁸² Elevated bile acid levels in the colon increase colonic motility and the transport of water and electrolytes in the intestinal lumen, causing diarrhoea.⁸³ Symptoms are secretory, watery diarrhoea that can be associated with abdominal pain, distension, faecal urgency, incontinence, nocturnal defecation and, rarely, steatorrhoea.^{82,83}

Bile acid malabsorption (BAM) is a common though little-known cause of chronic diarrhoea.⁸² Between 20% and 50% of patients with chronic functional-type diarrhoea (functional diarrhoea, diarrhoea-predominant or alternating irritable bowel syndrome) present BAM. BAM associated with a structurally normal ileum is usually called type 2, primary, or idiopathic BAM. In contrast, BAM associated with disease or ileectomy is known as type 1 or secondary BAM. It is very common in patients with Crohn disease (30%) or radiation enteritis (>50%), and nearly always presents after ileectomy (90%).^{84,85} BAM is also often associated with microscopic colitis (10–60%),⁸⁶ and is the primary cause of postcholecystectomy chronic diarrhoea. Finally, BAM has also been associated with miscellaneous factors, such as peptic ulcer surgery, coeliac disease, chronic pancreatitis, diabetes mellitus, cystic fibrosis, and the use of some drugs (for example, NSAIDs, colchicine or olsalazine). When BAM presents with any of the foregoing, it is known as type 3 BAM or BAM secondary to gastrointestinal disorders. BAM has also been reported to be a common, and often under-diagnosed, cause of gastrointestinal symptoms associated with cancer therapies.⁸⁷

Diagnosis

The most common diagnostic technique used in Europe (but not available in the USA) is the ^{75}Se -homotaurocholic Acid (SeHCAT) test, which uses scintigraphy to evaluate abdominal retention of an orally-administered bile acid analogue

labelled with selenium (^{75}Se -homotaurocholic acid). The test is highly accurate, and abnormal retention values are predictive of good response to therapy with bile acid sequestrants. A retention value of less than 10% at 7 days is highly sensitive (80–90%) and specific (70–100%) for BAM.^{72,82,88} The drawbacks of the test are radiation exposure (equivalent to a chest X-ray) and the need for special equipment.

Although therapeutic testing with bile acid sequestrants (cholestyramine) is widely used, the diagnostic accuracy of the test has not been evaluated in the literature, and a definitive diagnosis must be established for a disorder such as BAM, which often requires long-term therapy.^{82,88,89} If therapeutic testing is performed, it is recommended to start treatment with low-dose bile acid sequestrants, and gradually uptitrate while monitoring patient response.

Recommendations

- 1 BAM should be evaluated in patients with functional-type chronic diarrhoea (functional diarrhoea, diarrhoea-predominant irritable bowel syndrome), in intestinal diseases associated with the ileum (Crohn disease and radiation enteritis), and in microscopic colitis (CE, moderate; SR, strong).
- 2 BAM is a ubiquitous feature of chronic diarrhoea following cholecystectomy or ileectomy, and in these cases empirical treatment is advised (CE, low; SR, strong).
- 3 The recommended test for BAM is SeHCAT (CE, moderate; SR, strong).
- 4 If SeHCAT is unavailable, the best option is therapeutic testing with cholestyramine (CE, very low; SR, weak).

Sugar malabsorption diarrhoea

Malabsorption of carbohydrates can cause digestive intolerance symptoms, such as diarrhoea, abdominal pain, nausea and meteorism. The most common culprits are lactose, fructose and sorbitol. In a small percentage of cases, malabsorption is caused by rare congenital disorders that cause deficiency of disaccharidases (lactase, sucrase-isomaltase, trehalase, aldolase B) or affect intestinal transport (for example, congenital glucose-galactosemal absorption caused by a mutation in the *SLC5A1* gene).

Any sugars that are not absorbed in the small intestine produce symptoms caused by osmotic imbalance and fermentation by the intestinal microbiota, producing short-chain fatty acids and gases (hydrogen, methane, carbon dioxide). Due to the rapid transit of carbohydrates through the gastrointestinal tract, symptoms will start shortly after sugar intake, and can persist for 6–9 h. Typically, symptoms only occur when the patient ingests the offending carbohydrate. Symptoms are highly nonspecific, and differential diagnosis with other digestive pathologies is essential.

Lactose intolerance

Lactose intolerance is the body's symptomatic response to lactose ingestion caused by hypolactasia or decreased β -galactosidase levels in intestinal villi⁹⁰; this enzyme hydrolyses lactose into monosaccharides (glucose and

galactose) that can be absorbed by the jejunum. Hypolactasia is classed as primary (acquired or congenital) or secondary. The most common form is acquired hypolactasia, which is also called primary lactase deficiency or lactase non-persistence. Persistence or non-persistence of lactase is determined by autosomal-recessive inheritance of polymorphisms in the regulatory region of the lactase gene.⁹¹ In lactase non-persistent individuals, the activity of the enzyme gradually declines after weaning. It is a highly prevalent disorder that affects 70% of the world's population, although it is more predominant in certain regions. Most adults in Africa, Asia, Latin America and the Mediterranean region are lactase non-persistent, while most individuals in north and central Europe are lactase-persistent.

Lactose malabsorption is not always associated with intolerance and symptoms. Only 30–50% of individuals with lactose malabsorption are lactose intolerant. However, subjective perception of lactose intolerance does not always indicate lactose malabsorption,^{92–94} and therefore lactose malabsorption and lactose intolerance are not synonymous. For this reason, the patient history contributes little to a diagnosis of lactose intolerance, and diagnostic confirmation is needed before imposing life-long lactose restriction. The gold standard test is the determination of lactase levels in samples from intestinal biopsies,⁹⁵ although it is rarely used because of its invasive nature, high cost, and limited results due to the patchy distribution of lactase in the gut.⁹⁶ The most widely used, simplest, non-invasive, inexpensive and highly accurate diagnostic techniques are the H₂ breath test with lactose challenge³⁴ (sensitivity, 73%; specificity, 86%) and the gaxilose test^{97,98} (sensitivity, 93%; specificity, 92%). The breath test measures increased levels of H₂ in exhaled breath (>20 ppm) following intake of 50 g of lactose. False positives can be caused by bacterial overgrowth and rapid intestinal transit. The test can trigger digestive symptoms but yield false negative results in individuals with non-H₂-producing bacterial flora (15–20% of the population) or due to a placebo effect (44% of individuals with a negative breath test).⁹⁹ The gaxilose test measures the total amount of xylose in urine collected over 5 h following oral administration of 4-galactosylxylose, with xylose levels of <37.87 mg being diagnostic for hypolactasia.

Genetic testing evaluates the existence of polymorphisms (C/T-13910, G/A-22018) associated with lactase non-persistence. CC and GG genotypes are associated with non-persistence, while CT, TT, GA and AA genotypes are associated with lactase persistence, although some polymorphic variants can affect the accuracy of the test.¹⁰⁰ The test has a high negative predictive value (98%), so the possibility of CT and TT-1390 genotypes presenting primary lactase deficiency is extremely remote.¹⁰¹ Agreement with a positive finding in a breath test is 100%.^{101–103} Genetic testing, however, is of no use in patients with suspected secondary hypolactasia, it is more costly than the gaxilose test or breath test, and is only available in a few hospitals.

Fructose and sorbitol intolerance

Fructose is a monosaccharide found in fruit and some vegetables, and is widely used as a sweetening agent in the

food industry. Fructose is absorbed by a number of pathways that facilitate transport across the intestinal epithelium. When fructose is present in excess of glucose (also called "free fructose"), the excess is absorbed by a low-capacity transporter (GLUT-5), so the greater the fructose overload, the greater the likelihood of malabsorption. Glucose facilitates fructose absorption along a more efficient pathway which uses the GLUT-2 transport system (facilitated glucose transporter).^{104,105}

Sorbitol is a sugar alcohol found in some fruit that is widely used as a sugar substitute, and is found in cakes, jams, chewing gum and low-calorie products. It is absorbed in the small intestine by passive transport, and acts as an inhibitor of the GLUT-5 fructose transporter. Consumption of fructose and sorbitol together, therefore, worsens symptoms caused by fructose malabsorption.

Fructose intolerance is defined as the onset of symptoms following intake of <25–30 g of fructose¹⁰⁴. The H₂ breath test with 25 g fructose challenge is used to diagnose fructose malabsorption, while sorbitol malabsorption is confirmed by the H₂ breath test 5 g sorbitol challenge. Some authors, however, prefer to evaluate malabsorption of a combination of fructose and sorbitol, instead of fructose alone (20–25 g fructose + 3.5–5 g sorbitol),¹⁰⁴ as these sugars are frequently ingested together and, as already described, sorbitol obstructs absorption of fructose. Nevertheless, the main problem encountered in the diagnosis of both fructose and sorbitol malabsorption is uncertainty surrounding the normal absorption capacity of these sugars in healthy subjects.^{94,106} The correct dosage and concentration of fructose and/or sorbitol needed to differentiate between normal or impaired absorption remains unclear.

Other carbohydrates

The term "fermentable oligosaccharides, disaccharides, monosaccharides and polyols" (FODMAP) has recently been introduced to define a group of poorly absorbed short-chain carbohydrates that were previously thought to be unrelated, but have since been found to exhibit similar behaviour in the small and large intestine.^{104,107} The term includes malabsorbed fructose and lactose, different polyalcohol sugars (sorbitol, maltitol, xylitol, etc.) that are usually poorly absorbed, and fructans and galactans (galacto-oligosaccharides, such as raffinose and stachyose), which are always malabsorbed. All FODMAPs exert a similar osmotic effect in the colon, and are rapidly fermented by the gut microflora. The combination of fructose and fructans can aggravate symptoms in a similar way as the aforementioned dual action of fructose and sorbitol. This factor should be borne in mind when drawing up diets designed to restrict intake of malabsorbed sugars.

Recommendations

- 1 Subjective manifestations of lactose intolerance do not always indicate lactose malabsorption. If diarrhoea secondary to primary lactase deficiency is suspected, diagnostic confirmation is needed before imposing life-long lactose restriction (CE, high; SR, strong).

- 2 The H₂ breath test with lactose challenge, the gaxilose test, and genetic testing are recommended to confirm diagnosis of lactose malabsorption.
- 3 When fructose and/or sorbitol intolerance are suspected, a breath test using these sugars is advised (CE, low; SR, weak).

Chronic diarrhoea secondary to intestinal motility disorders

Motility abnormalities can be an aetiopathogenic factor shared by several different digestive disorders in which the primary symptom is chronic diarrhoea. In these entities, changes in motility are more a contributing factor than the underlying mechanism causing diarrhoea. This section will focus on chronic diarrhoea caused by gastrointestinal neuromuscular diseases in which the condition arises as a result of changes in motor function *per se* (for example, diabetes mellitus), or secondary to bacterial overgrowth (for example, scleroderma). Intestinal neuromuscular diseases (chronic intestinal pseudo-obstruction and intestinal dysmotility) are uncommon disorders that are rarely the cause of chronic diarrhoea. As such, they should only be investigated when other, more common, causes of chronic diarrhoea have been ruled out.^{108,109}

Classification

Neuromuscular diseases of the gastrointestinal tract are divided into:

- 1 Primary neuromuscular disease. This is characterised by:
 - Chronic and recurrent subocclusive episodes or abdominal distension.
 - Persistent or intermittent daytime or nocturnal diarrhoea interspersed with constipation, associated with weight loss and laboratory test findings of malnutrition/malabsorption. Bacterial overgrowth is a common feature (see corresponding section above), and the diarrhoea may improve or resolve after normalisation of bacterial colonisation.
 - Endoscopic and radiological studies (CT, transit, MR enterography) used to rule out other structural diseases are usually unremarkable or show dilated bowel loops.
 - There may also be extra intestinal symptoms due to involvement of other organs or systems (most often the urological, cardiovascular and autonomic nervous systems).
- 2 Secondary to general disease with intestinal involvement:
 - Connective tissue diseases: scleroderma, dermatomyositis, systemic lupus erythematosus.
 - Endocrine and metabolic disorders: diabetes mellitus, hyperthyroidism.
 - Neurological disorders: dysautonomia, MNGIE disease, Parkinson disease.
 - Muscular diseases: muscular dystrophy, Steinert disease.
 - Infiltrative diseases: amyloidosis.
 - After gastric surgery with vagotomy.

Diagnosis

Diagnosis is based on: (a) compatible clinical history: suggestive symptoms, with other organic pathology ruled out by radiographic studies. Given the rarity of intestinal neuromuscular diseases, it is important to first rule out other, more common, causes of chronic diarrhoea. In milder cases, difficulties can be encountered in differentiating these diseases from other functional pathologies (IBS/functional diarrhoea); (b) test for bacterial overgrowth (see corresponding section above); and (c) test for motor dysfunction using small bowel manometry, which is the gold standard for studying small bowel motility.¹¹⁰⁻¹¹²

Small bowel manometry has high specificity but low sensitivity. Internationally accepted manometric diagnosis criteria are available.¹¹⁰ Manometry can be used to detect motor dysfunction and differentiate between myopathic and neuropathic disorders. It is a complex technique that requires skilled interpretation, and is only available in specialised centres.

Motor abnormalities correlate poorly with oro-caecal transit time, and the latter cannot be taken as an indication of motor dysfunction.

Recommendations

Intestinal manometry can be useful in the diagnosis of neuromuscular disorders in patients with recurrence of diarrhoea in an appropriate clinical context (CE, moderate; SR, weak).

Chronic diarrhoea secondary to food allergies

Adverse reactions to certain foods can be divided into 2 major subgroups: (a) food allergies, which are a subgroup of hypersensitivity reactions caused by immune mechanisms derived from IgE-dependent mast cell activation, or mediated by other immune mechanisms, often type 4 or delayed-type hypersensitivity reactions,¹¹³⁻¹¹⁵ and (b) food intolerance, which includes a number of reactions mediated by non-immune mechanisms and derived from enzyme or transport deficiencies, or the pharmacological properties of certain food components.¹¹⁶ Food allergies are more common in the paediatric population, and usually decrease with age; according to estimates based on diagnoses made using accepted techniques, prevalence ranges between 3.5% for cow's milk and 0.5–1.3% for other common food allergies (eggs, nuts, cereals, fruit, fish and seafood).^{114,117} The prevalence of food intolerance, in contrast, is estimated at between 15% and 20%, and is particularly associated with gastrointestinal functional diseases, such as irritable bowel syndrome and functional dyspepsia.¹¹⁸ The subgroup of food intolerance disorders includes both common and less frequent conditions that can cause diarrhoea and other gastrointestinal manifestations, such as flatulence, abdominal distension, nausea, vomiting and abdominal and/or epigastric pain. Sugar malabsorption, non-coeliac gluten sensitivity and FODMAP are among the most common disorders, and have been discussed elsewhere in this report. Less common disorders include intolerance to salicylates, biogenic amines, caffeine and glutamate, toxic reactions to

microbial or fungal products, such as aflatoxins, and psychological reactions.¹¹⁶ It is important to differentiate between allergies and intolerance, as the gastrointestinal manifestations of food allergies are often comparable to symptoms of intolerance. Allergies, however, are commonly associated with other specific extra intestinal manifestations, such as rash or angioedema, which usually appear immediately after exposure or intake of the offending allergen. In addition, unlike food intolerance, in which diarrhoea can persist while the individual continues to consume, albeit unwittingly, the offending foodstuff, food allergies can cause acute diarrhoea, but are rarely the direct cause of chronic diarrhoea. A particular case in point is diarrhoea-predominant irritable bowel syndrome, in which it can be difficult to establish the role of food allergies and intolerance in the pathogenesis of the diarrhoea, as both entities often combine in these patients, and can even be associated with psychological adverse food reactions. In recent years, some authors have implicated IgG- or IgG4-mediated delayed food hypersensitivity reactions¹¹⁹⁻¹²¹ in the pathogenesis and severity of clinical manifestation, including diarrhoea. The association is still under debate, and no consensus has yet been reached.

Diagnosis

The first and most important step in diagnosing food allergies and differentiating them from food intolerance is to compile a comprehensive clinical and dietary history, focussing in particular on the food consumed by the patient in the 2 h prior to the onset of symptoms. It is important to take into account levels of tolerance before and after ingestion of the offending foodstuff: good tolerance after ingestion will exclude the presence of an IgE-mediated mechanism, except when food allergies are facilitated or aggravated by co-factors (exercise, NSAIDs, stress or alcohol). Once the clinical suspicion has been confirmed, tests must be performed to establish whether the allergic symptoms are caused by an IgE-mediated hypersensitivity reaction. These include¹²²: (a) skin prick tests using commercial extracts, or prick-prick tests using fresh foodstuffs; (b) determination of food-specific IgE in serum (UniCAP100, ImmunoCAP250, Immulite System, HYTEC-288 system); and to a lesser extent (c) the use of purified or recombinant molecules (component separation) to accurately identify the allergens causing reaction in each patient (microarray chip, ISAC; UNICAP, ThermoFisher). These tests are mainly used to differentiate between true allergic reactions or cross reactivity; (d) screening tests for foods or pollens, particularly useful in primary care, that contain a mixture of the most common allergens (multiallergen screen IgE test, Phadiatop); and (e) other blood tests: tryptase, eosinophil cationic protein, carboxypeptidase, cathepsin G and mast cell chymase, IFN- γ , TNF- α , FGF, IL-4, IL-5, IL-6, IL-13, histamine, chondroitin sulphate, heparin, leukotriene C4, PGD2, total IgE and free IgE. Table 15 shows the specificity and sensitivity of the most widely used complementary tests for specific target foods.¹²³ The double-blind, placebo-controlled oral food challenge is considered the gold standard diagnostic test. However, it is impractical, because it is time consuming and resource-intensive, and can cause anaphylaxis.

Table 15 Approximate diagnostic accuracy of the main complementary food allergytests.

Food	Technique	Diagnostic accuracy
Egg	s-IgE	Se, 93% (82–98); Sp, 49% (40–58)
	SPT	Se, 92% (80–97); Sp, 58% (49–67)
Cow's milk	s-IgE	Se, 87% (79–94); Sp, 48% (36–59)
	SPT	Se, 88% (76–94); Sp, 68% (56–77)
Wheat	s-IgE	Se, 83% (69–92); Sp, 43% (20–69)
	SPT	Se, 73% (56–85); Sp, 73% (48–89)
Soy	s-IgE	Se, 83% (64–93); Sp, 38% (24–54)
	SPT	Se, 55% (33–75); Sp, 68% (52–80)
Nuts	s-IgE	Se, 96% (92–98); Sp, 59% (45–72)
	SPT	Se, 95% (88–98); Sp, 61% (47–74)
Fish	s-IgE	Se, -% (67–94); Sp, -% (65–88)
	SPT	Se, 91%–100% (-); Sp, 57% (-)
Seafood	s-IgE	Se, 100% (80–100); Sp, 45% (23–68)
	SPT	Se, 100% (-); Sp, -% (32–50)

Se: sensitivity; Sp: specificity; SPT: Skin prick test.

Source: adapted from Soares-Weiser et al.¹²³

Any methods that have not proved effective and safe in controlled trials should be classed as “unproven methods” or “non-validated methods”, and are therefore not recommended for the purpose of diagnosis.¹²⁴ Such tests include: (a) controversial in vivo tests: subcutaneous, sublingual and intradermal provocation tests; the *ricercaintolleranza alimentaria* (DRIA) test, based on the sublingual administration of an allergenic extract and on measurement of muscle strength with an ergonometer; electroacupuncture and electrodermal testing (Voll method); applied kinesiology (manual measurement of muscle strength), and bioresonance; and (b) controversial in vivo tests: determination of IgG in blood samples using ELISA (A200 test, IADM test, and food intolerance screening [FIS]), and testing for IgG4 against foods (IgG4 Screen Nutritional); DNA analysis of hair samples to determine food intolerance, and measurement of white cells using the Coulter counter, either with or without incubation with food extracts (ALCAT test). There is currently insufficient evidence to recommend the use of detection of IgG or IgG4 against food components to guide the dietary management of chronic diarrhoea.

A recent recommendation indicates ruling out wheat allergy before diagnosing a patient with non-coeliac gluten sensitivity.¹²⁵ Nevertheless, wheat allergy is rare and usually causes respiratory symptoms, above all rhinitis, particularly in association with co-factors such as exercise and stress. Diagnosis is based on the skin prick test (whole wheat, gluten and gliadin) and also by determination of IgE against ω_5 -recombinant gliadin.

Recommendations

- 1 Taking a comprehensive medical history will help differentiate between diarrhoea associated with an immune reaction or with other mechanisms more typical of food intolerance.
- 2 If food allergy is suspected, the patient should be referred to an allergist for specific IgE determination and a skin

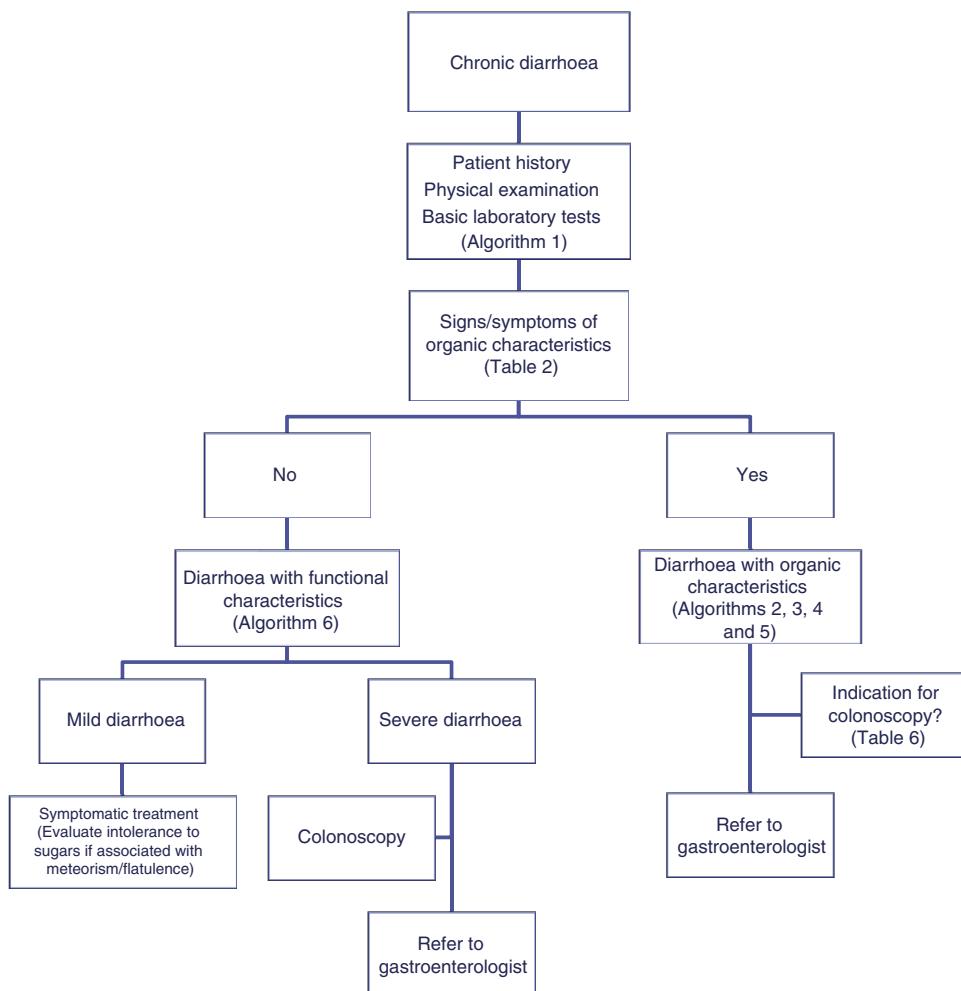


Figure 7 Algorithm 7. Patients with chronic diarrhoea referred from primary to specialist care.

prick test, both of which have good sensitivity but low specificity (CE, high; SR, strong).

3 The double-blind, placebo-controlled oral food challenge is considered the gold standard diagnostic test. However, it is impractical, because it is time consuming and resource-intensive, and can cause anaphylaxis (CE, high, SR, strong).

Diagnosis and care of chronic diarrhoea patients in primary care

Primary care doctors make the initial evaluation of patients presenting with chronic diarrhoea (Algorithm 7, Fig. 7). It is important to determine whether the patient meets diagnostic criteria for chronic diarrhoea, to take a comprehensive medical history, and order basic blood and stool tests. It is essential to evaluate the presence of organic factors, to decide whether to refer the patient to a gastroenterologist, and whether to order a colonoscopy (Table 16).

In many patients presenting with recurrent but mild (less than 2 episodes per week) watery diarrhoea, empirical therapy to treat symptoms (with loperamide or racecadotril) is often appropriate. In these cases, it is important to evaluate whether the complaint is associated mainly with symptoms

Table 16 Indications for colonoscopy in patients with chronic diarrhoea.

Recent onset chronic diarrhoea in patients aged > 50 years
Chronic diarrhoea and history of colorectal cancer in first degree relatives
Chronic inflammatory diarrhoea (see definition in text)
Chronic non-bloody diarrhoea with positive faecal occult blood
Chronic non-bloody diarrhoea with calprotectin > 150 mg/kg
Chronic non-bloody diarrhoea with elevated C-reactive protein
Chronic watery diarrhoea with severe functional diarrhoea criteria or signs and symptoms and test findings suggestive of organic origin (see text)

of meteorism and abdominal distension. If so, the presence of sugar malabsorption (lactose and/or fructose or sorbitol) must be ruled out.

Conflict of interests

The authors declare they have no conflicts of interest.

References

1. Headstrom PD, Surawicz CM. Chronic diarrhea. *Clin Gastroenterol Hepatol.* 2005;3:734–7.
2. Schiller LR, Pardi DS, Spiller R, Semrad CE, Surawicz CM, Giannella RA, et al. *Gastro* 2013 APDW/WCOG Shanghai working party report: chronic diarrhea: definition, classification, diagnosis. *J Gastroenterol Hepatol.* 2014;29:6–25.
3. Sandhu DK, Surawicz C. Update on chronic diarrhea: a run-through for the clinician. *Curr Gastroenterol Rep.* 2012;14:421–7.
4. Fine KD, Schiller LR. AGA technical review on the evaluation and management of chronic diarrhea. *Gastroenterology.* 1999;116:1464–86.
5. Thomas PD, Forbes A, Green J, Howdle P, Long R, Playford R, et al. Guidelines for the investigation of chronic diarrhoea, 2nd edition. *Gut.* 2003;52, v1–15.
6. Gentile NM, Khanna S, Loftus EV Jr, Smyrk TC, Tremaine WJ, Harmsen WS, et al. The epidemiology of microscopic colitis in Olmsted County from 2002 to 2010: a population-based study. *Clin Gastroenterol Hepatol.* 2014;12:838–42.
7. Fernández-Bañares F, Esteve M, Salas A, Alsina M, Farré C, González C, et al. Systematic evaluation of the causes of chronic watery diarrhea with functional characteristics. *Am J Gastroenterol.* 2007;102:2520–8.
8. Habba SF. Diarrhea predominant Irritable Bowel Syndrome (IBS-D): fact or fiction. *Med Hypotheses.* 2011;76:97–9.
9. Marzo-Castillejo M, Alonso-Coello P, Rotaeche del Campo R. How are the quality of evidence and the strength of recommendations to be classified? *Aten Prim.* 2006;37:5–8.
10. Waugh N, Cummins E, Royle P, Kandala NB, Shyangdan D, Arasaradnam R, et al. Faecal calprotectin testing for differentiating amongst inflammatory and non-inflammatory bowel diseases: systematic review and economic evaluation. *Health Technol Assess.* 2013;17, xv–xix, 1–211.
11. Van Rheenen PF, van de Vijver E, Fidler V. Faecal calprotectin for screening of patients with suspected inflammatory bowel disease: diagnostic meta-analysis. *BMJ.* 2010;341, 3369.
12. Panes J, Bouhnik Y, Reinisch W, Stoker J, Taylor SA, Baumgart DC, et al. Imaging techniques for assessment of inflammatory bowel disease: joint ECCO and ESGAR evidence-based consensus guidelines. *J Crohns Colitis.* 2013;7:556–85.
13. Rodríguez-Pardo D, Mirelis B, Navarro F. Infections caused by *Clostridium difficile*. *Enferm Infect Microbiol Clin.* 2013;31:254–63.
14. Cone MM, Whitlow CB. Sexually transmitted and anorectal infectious diseases. *Gastroenterol Clin North Am.* 2013;42:877–92.
15. Koot BG, ten Kate FJ, Juffrie M, Rosalina I, Taminiau JJ, Benninga MA. Does *Giardia lamblia* cause villous atrophy in children? A retrospective cohort study of the histological abnormalities in giardiasis. *J Pediatr Gastroenterol Nutr.* 2009;49:304–8.
16. Ianiro G, Bibbò S, Montalto M, Ricci R, Gasbarrini A, Cammarota G. Systematic review: sprue-like enteropathy associated with olmesartan. *Aliment Pharmacol Ther.* 2014;40:16–23.
17. Brown IS, Bettington A, Bettington M, Rosty C. Tropical sprue: revisiting an underrecognized disease. *Am J Surg Pathol.* 2014;38:666–72.
18. Haghghi P, Wolf PL. Tropical sprue and subclinical enteropathy: a vision for the nineties. *Crit Rev Clin Lab Sci.* 1997;34:313–41.
19. Pallav K, Leffler DA, Tariq S, Kabbani T, Hansen J, Peer A, et al. Noncoeliac enteropathy: the differential diagnosis of villous atrophy in contemporary clinical practice. *Aliment Pharmacol Ther.* 2012;35:380–90.
20. Rubio-Tapia A, Murray JA. Classification and management of refractory coeliac disease. *Gut.* 2010;59:547–57.
21. Lagier JC, Lepidi H, Raoult D, Fenollar F. Systemic *Tropheryma whipplei*: clinical presentation of 142 patients with infections diagnosed or confirmed in a reference center. *Medicine (Baltimore).* 2010;89:337–45.
22. Marsh MN. Gluten, major histocompatibility complex, and the small intestine. A molecular and immunobiologic approach to the spectrum of gluten sensitivity ('celiac sprue'). *Gastroenterology.* 1992;102:330–54.
23. Oberhuber G, Granditsch G, Vogelsang H. The histopathology of coeliac disease: time for a standardized report scheme for pathologists. *Eur J Gastroenterol Hepatol.* 1999;11:1185–94.
24. Corazza GR, Villanacci V. Coeliac disease. *J Clin Pathol.* 2005;58:573–4.
25. Ensari A. Gluten-sensitive enteropathy (celiac disease): controversies in diagnosis and classification. *Arch Pathol Lab Med.* 2010;134:826–36.
26. Rubio-Tapia A, Hill ID, Kelly CP, Calderwood AH, Murray JA. Gastroenterology ACo. ACG clinical guidelines: diagnosis and management of celiac disease. *Am J Gastroenterol.* 2013;108:656–76, quiz 677.
27. Catassi C, Fasano A. Celiac disease diagnosis: simple rules are better than complicated algorithms. *Am J Med.* 2010;123:691–3.
28. Langenberg MC, Wismans PJ, van Genderen PJ. Distinguishing tropical sprue from celiac disease in returning travellers with chronic diarrhoea: a diagnostic challenge? *Travel Med Infect Dis.* 2014;12:401–5.
29. Liu H, Brais R, Lavergne-Slove A, Jeng Q, Payne K, Ye H, et al. Continual monitoring of intraepithelial lymphocyte immunophenotype and clonality is more important than snapshot analysis in the surveillance of refractory coeliac disease. *Gut.* 2010;59:452–60.
30. O’Malley DP, Goldstein NS, Banks PM. The recognition and classification of lymphoproliferative disorders of the gut. *Hum Pathol.* 2014;45:899–916.
31. Gabrielli M, d’Angelo G, di Renzo T, Scarpellini E, Ojetto V. Diagnosis of small intestinal bacterial overgrowth in the clinical practice. *Eur Rev Med Pharmacol Sci.* 2013;17:30–5.
32. Miazga A, Osiński M, Cichy W, Zaba R. Current views on the etiopathogenesis, clinical manifestation, diagnostics, treatment and correlation with other nosological entities of SIBO. *Adv Med Sci.* 2015;60:118–24.
33. Saad RJ, Chey WD. Breath testing for small intestinal bacterial overgrowth: maximizing test accuracy. *Clin Gastroenterol Hepatol.* 2014;12:1964–72, quiz e119–20.
34. Gasbarrini A, Corazza GR, Gasbarrini G, Montalto M, Di Stefano M, Basilico G, et al. Methodology and indications of H2-breath testing in gastrointestinal diseases: the Rome Consensus Conference. *Aliment Pharmacol Ther.* 2009;29:1–49.
35. DiMagno EP. Medical treatment of pancreatic insufficiency. *Mayo Clin Proc.* 1979;54:435–42.
36. Bollbach R, Becker M, Rothauwe HW. Serum immunoreactive trypsin and pancreatic lipase in cystic fibrosis. *Eur J Pediatr.* 1985;144:167–70.
37. Cleghorn G, Benjamin L, Corey M, Forstner G, Dati F, Durie P. Serum immunoreactive pancreatic lipase and cationic trypsinogen for the assessment of exocrine pancreatic function in older patients with cystic fibrosis. *Pediatrics.* 1986;77:301–6.
38. Ammann RW, Bühl H, Pei P. Comparative diagnostic accuracy of four tubeless pancreatic function tests in chronic pancreatitis. *Scand J Gastroenterol.* 1982;17:997–1002.
39. Lankisch PG, Koop H, Otto J. Estimation of serum pancreatic isoamylase: its role in the diagnosis of exocrine pancreatic insufficiency. *Am J Gastroenterol.* 1986;81:365–8.

40. Pezzilli R, Talamini G, Gullo L. Behaviour of serum pancreatic enzymes in chronic pancreatitis. *Dig Liver Dis.* 2000;32:233–7.
41. Löser C, Möllgaard A, Fölsch UR. Faecal elastase 1: a novel, highly sensitive, and specific tubeless pancreatic function test. *Gut.* 1996;39:580–6.
42. Benini L, Amodio A, Campagnola P, Agugiaro F, Cristofori C, Micciolo R, et al. Fecal elastase-1 is useful in the detection of steatorrhea in patients with pancreatic diseases but not after pancreatic resection. *Pancreatology.* 2013;13:38–42.
43. Halloran CM, Cox TF, Chauhan S, Raray MG, Sutton R, Neoptolemos JP, et al. Partial pancreatic resection for pancreatic malignancy is associated with sustained pancreatic exocrine failure and reduced quality of life: a prospective study. *Pancreatology.* 2011;11:535–45.
44. Goldberg DM. Proteases in the evaluation of pancreatic function and pancreatic disease. *Clin Chim Acta.* 2000;291:201–21.
45. De Pedro C, Codoceo R, Vazquez P, Hernanz A. Fecal chymotrypsin levels in children with pancreatic insufficiency. *Clin Biochem.* 1986;19:338–40.
46. Vantrappen GR, Rutgeerts PJ, Ghoos YF, Hiele MI. Mixed triglyceride breath test: a noninvasive test of pancreatic lipase activity in the duodenum. *Gastroenterology.* 1989;96: 1126–34.
47. Heikius B, Niemelä S, Lehtola J, Karttunen T, Lähde S. Pancreatic duct abnormalities and pancreatic function in patients with chronic inflammatory bowel disease. *Scand J Gastroenterol.* 1996;31:517–23.
48. Muniz CK, dos Santos JS, Pfrimer K, Ferrioli E, Kemp R, Marchini JS, et al. Nutritional status, fecal elastase-1, and 13C-labeled mixed triglyceride breath test in the long-term after pancreaticoduodenectomy. *Pancreas.* 2014;43:445–50.
49. Conwell DL, Zuccaro G, Morrow JB, van Lente F, Obuchowski N, Vargo JJ, et al. Cholecystokinin-stimulated peak lipase concentration in duodenal drainage fluid: a new pancreatic function test. *Am J Gastroenterol.* 2002;97:1392–7.
50. Erchinger F, Engjom T, Tjora E, Hoem D, Hausken T, Gilja OH, et al. Quantification of pancreatic function using a clinically feasible short endoscopic secretin test. *Pancreas.* 2013;42:1101–6.
51. Sanyal R, Stevens T, Novak E, Veniero JC. Secretin-enhanced MRCP: review of technique and application with proposal for quantification of exocrine function. *AJR Am J Roentgenol.* 2012;198:124–32.
52. Manfredi R, Perandini S, Mantovani W, Frulloni L, Faccioli N, Pozzi Mucelli R. Quantitative MRCP assessment of pancreatic exocrine reserve and its correlation with faecal elastase-1 in patients with chronic pancreatitis. *Radiol Med.* 2012;117:282–92.
53. Stevens T, Dumot JA, Parsi MA, Zuccaro G, Vargo JJ. Combined endoscopic ultrasound and secretin endoscopic pancreatic function test in patients evaluated for chronic pancreatitis. *Dig Dis Sci.* 2010;55:2681–7.
54. Albasir S, Bronner MP, Parsi MA, Walsh RM, Stevens T. Endoscopic ultrasound, secretin endoscopic pancreatic function test, and histology: correlation in chronic pancreatitis. *Am J Gastroenterol.* 2010;105:2498–503.
55. Nakamura H, Murakami Y, Uemura K, Hayashidani Y, Sudo T, Ohge H, et al. Reduced pancreatic parenchymal thickness indicates exocrine pancreatic insufficiency after pancreaticoduodenectomy. *J Surg Res.* 2011;171:473–8.
56. Leeds JS, Hopper AD, Sidhu R, Simonette A, Azadbakht N, Hoggard N, et al. Some patients with irritable bowel syndrome may have exocrine pancreatic insufficiency. *Clin Gastroenterol Hepatol.* 2010;8:433–8.
57. Fabian E, Kump P, Krejs GJ. Diarrhea caused by circulating agents. *Gastroenterol Clin North Am.* 2012;41:603–10.
58. Jensen RT. Overview of chronic diarrhea caused by functional neuroendocrine neoplasms. *Semin Gastrointest Dis.* 1999;10:156–72.
59. Brelian D, Tenner S. Diarrhoea due to pancreatic diseases. *Best Pract Res Clin Gastroenterol.* 2012;26:623–31.
60. Roerig JL, Steffen KJ, Mitchell JE, Zunker C. Laxative abuse: epidemiology, diagnosis and management. *Drugs.* 2010;70:1487–503.
61. Abraham BP, Sellin JH. Drug-induced, factitious, & idiopathic diarrhoea. *Best Pract Res Clin Gastroenterol.* 2012;26:633–48.
62. Schiller LR, Rivera LM, Santangelo WC, Little KH, Fordtran JS. Diagnostic value of fasting plasma peptide concentrations in patients with chronic diarrhea. *Dig Dis Sci.* 1994;39:2216–22.
63. Modlin IM, Kidd M, Bodei L, Drozdov I, Aslanian H. The clinical utility of a novel blood-based multi-transcriptome assay for the diagnosis of neuroendocrine tumors of the gastrointestinal tract. *Am J Gastroenterol.* 2015;110:1223–32.
64. Marotta V, Nuzzo V, Ferrara T, Zuccoli A, Masone M, Nocerino L, et al. Limitations of Chromogranin A in clinical practice. *Biomarkers.* 2012;17:186–91.
65. Van Essen M, Sundin A, Krenning EP, Kwekkeboom DJ. Neuroendocrine tumours: the role of imaging for diagnosis and therapy. *Nat Rev Endocrinol.* 2014;10:102–14.
66. Byers RJ, Marsh P, Parkinson D, Haboubi NY. Melanosis coli is associated with an increase in colonic epithelial apoptosis and not with laxative use. *Histopathology.* 1997;30:160–4.
67. Eherer AJ, Fordtran JS. Fecal osmotic gap and pH in experimental diarrhea of various causes. *Gastroenterology.* 1992;103:545–51.
68. Longstreth GF, Thompson WG, Chey WD, Houghton LA, Mearin F, Spiller RC. Functional bowel disorders. *Gastroenterology.* 2006;130:1480–91.
69. Money ME, Camilleri M. Review: management of postprandial diarrhea syndrome. *Am J Med.* 2012;125:538–44.
70. Macaigne G, Lahmek P, Locher C, Lesgourgues B, Costes L, Nicolas MP, et al. Microscopic colitis or functional bowel disease with diarrhea: a French prospective multicenter study. *Am J Gastroenterol.* 2014;109:1461–70.
71. Fan X, Sellin JH. Review article: small intestinal bacterial overgrowth, bile acid malabsorption and gluten intolerance as possible causes of chronic watery diarrhoea. *Aliment Pharmacol Ther.* 2009;29:1069–77.
72. Wedlake L, A'Hern R, Russell D, Thomas K, Walters JR, Andreiev HJ. Systematic review: the prevalence of idiopathic bile acid malabsorption as diagnosed by SeHCAT scanning in patients with diarrhoea-predominant irritable bowel syndrome. *Aliment Pharmacol Ther.* 2009;30:707–17.
73. Magro F, Langner C, Driessens A, Ensari A, Geboes K, Mantzaris GJ, et al. European consensus on the histopathology of inflammatory bowel disease. *J Crohns Colitis.* 2013;7:827–51.
74. Fernandez-Banares F, Salas A, Esteve M, Pardo L, Casalots J, Forne M, et al. Evolution of the incidence of collagenous colitis and lymphocytic colitis in Terrassa, Spain: a population-based study. *Inflamm Bowel Dis.* 2011;17:1015–20.
75. Fernandez-Banares F, Salas A, Forne M, Esteve M, Espinos J, Viver JM. Incidence of collagenous and lymphocytic colitis: a 5-year population-based study. *Am J Gastroenterol.* 1999;94:418–23.
76. Guagnazzi D, Lucendo AJ, Angueira-Lapena T, Gonzalez-Castillo S, Tenias Burillo JM. Prevalence and incidence of microscopic colitis in patients with diarrhoea of unknown aetiology in a region in central Spain. *Dig Liver Dis.* 2012;44:384–8.
77. Pardi DS, Loftus EV Jr, Smyrk TC, Kammer PP, Tremaine WJ, Schleck CD, et al. The epidemiology of microscopic colitis: a population based study in Olmsted County, Minnesota. *Gut.* 2007;56:504–8.

78. Olesen M, Eriksson S, Bohr J, Jarnerot G, Tysk C. Microscopic colitis: a common diarrhoeal disease. An epidemiological study in Örebro, Sweden, 1993–1998. *Gut*. 2004;53:346–50.
79. Limsui D, Pardi DS, Smyrk TC, Abraham SC, Lewis JT, Sanderson SO, et al. Observer variability in the histologic diagnosis of microscopic colitis. *Inflamm Bowel Dis*. 2009;15:35–8.
80. Fiehn AM, Bjornbak C, Warnecke M, Engel PJ, Munck LK. Observer variability in the histopathologic diagnosis of microscopic colitis and subgroups. *Hum Pathol*. 2013;44:2461–6.
81. Aust DE, Münch A, Olesen M, Vieth M, Bonderup OK, Bohr J, et al. Su1183 topographical distribution of collagenous colitis – a pooled histological analysis of 2 European prospective multicenter trials. *Gastroenterology*. 2013;144:S-421.
82. Wilcox C, Turner J, Green J. Systematic review: the management of chronic diarrhoea due to bile acid malabsorption. *Aliment Pharmacol Ther*. 2014;39:923–39.
83. Khalid U, Lalji A, Stafferton R, Andreyev J. Bile acid malabsorption: a forgotten diagnosis? *Clin Med*. 2010;10:124–6.
84. Nyhlin H, Merrick MV, Eastwood MA. Bile acid malabsorption in Crohn's disease and indications for its assessment using SeHCAT. *Gut*. 1994;35:90–3.
85. Lenicek M, Duricova D, Komarek V, Gabrysova B, Lukas M, Smerhovsky Z, et al. Bile acid malabsorption in inflammatory bowel disease: assessment by serum markers. *Inflamm Bowel Dis*. 2011;17:1322–7.
86. Ung KA, Gillberg R, Kilander A, Abrahamsson H. Role of bile acids and bile acid binding agents in patients with collagenous colitis. *Gut*. 2000;46:170–5.
87. Phillips F, Muls AC, Lalji A, Andreyev HJ. Are bile acid malabsorption and bile acid diarrhoea an important cause of diarrhoea complicating cancer therapy? *Colorectal Dis*. 2015;17:730–4.
88. Vijayvargiya P, Camilleri M, Shin A, Saenger A. Methods for diagnosis of bile acid malabsorption in clinical practice. *Clin Gastroenterol Hepatol*. 2013;11:1232–9.
89. Barkun AN, Love J, Gould M, Pluta H, Steinhart H. Bile acid malabsorption in chronic diarrhea: pathophysiology and treatment. *Can J Gastroenterol*. 2013;27:653–9.
90. Suchy FJ, Brannon PM, Carpenter TO, Fernandez JR, Gilsanz V, Gould JB, et al. NIH consensus development conference statement: lactose intolerance and health. *NIH Consens State Sci Statements*. 2010;27:1–27.
91. Enattah NS, Kuokkanen M, Forsblom C, Natah S, Oksanen A, Jarvela I, et al. Correlation of intestinal disaccharidase activities with the C/T-13910 variant and age. *World J Gastroenterol*. 2007;13:3508–12.
92. Casellas F, Aparici A, Casaus M, Rodríguez P, Malagelada JR. Subjective perception of lactose intolerance does not always indicate lactose malabsorption. *Clin Gastroenterol Hepatol*. 2010;8:581–6.
93. Jellema P, Schellevis FG, van der Windt DA, Kneepkens CM. Lactose malabsorption and intolerance: a systematic review on the diagnostic value of gastrointestinal symptoms and self-reported milk intolerance. *QJM*. 2010;103:555–72.
94. Fernández-Bañares F. Reliability of symptom analysis during carbohydrate hydrogen-breath tests. *Curr Opin Clin Nutr Metab Care*. 2012;15:494–8.
95. Newcomer AD, McGill DB, Thomas PJ, Hofmann AF. Prospective comparison of indirect methods for detecting lactase deficiency. *N Engl J Med*. 1975;293:1232–6.
96. Maiuri L, Rossi M, Raia V, Garipoli V, Hughes LA, Swallow D, et al. Mosaic regulation of lactase in human adult-type hypolactasia. *Gastroenterology*. 1994;107:54–60.
97. Aragón JJ, Hermida C, Martínez-Costa OH, Sánchez V, Martín I, Sánchez JJ, et al. Noninvasive diagnosis of hypolactasia with 4-Galactosylxylose (Gaxilose): a multicentre, open-label, phase IIB-III nonrandomized trial. *J Clin Gastroenterol*. 2014;48:29–36.
98. Hermida C, Martínez-Costa OH, Corrales G, Teruel C, Sánchez V, Sánchez JJ, et al. Improvement and validation of D-xylose determination in urine and serum as a new tool for the non-invasive evaluation of lactase activity in humans. *J Clin Lab Anal*. 2014;28:478–86.
99. Vernia P, di Camillo M, Foglietta T, Avallone VE, De Carolis A. Diagnosis of lactose intolerance and the 'nocebo' effect: the role of negative expectations. *Dig Liver Dis*. 2010;42:616–9.
100. Swallow DM. Genetics of lactase persistence and lactose intolerance. *Annu Rev Genet*. 2003;37:197–219.
101. Pohl D, Savarino E, Hersberger M, Behlis Z, Stutz B, Goetze O, et al. Excellent agreement between genetic and hydrogen breath tests for lactase deficiency and the role of extended symptom assessment. *Br J Nutr*. 2010;104:900–7.
102. Krawczyk M, Wolska M, Schwartz S, Gruenhage F, Terjung B, Portincasa P, et al. Concordance of genetic and breath tests for lactose intolerance in a tertiary referral centre. *J Gastrointest Liver Dis*. 2008;17:135–9.
103. Mattar R, Monteiro MS, Villares CA, dos Santos AF, Carriño FJ. Single nucleotide polymorphism C/T(-13910), located upstream of the lactase gene, associated with adult-type hypolactasia: Validation for clinical practice. *Clin Biochem*. 2008;41:628–30.
104. Fernández-Bañares F, Esteve M, Viver JM. Fructose-sorbitol malabsorption. *Curr Gastroenterol Rep*. 2009;11:368–74.
105. Frieling T, Kuhlbusch-Zicklam R, Kalde S, Heise J, Hülsdonk A, Kreysel C. Fructose malabsorption: how much fructose can a healthy subject tolerate? *Digestion*. 2011;84:269–72.
106. Kyaw MH, Mayberry JF. Fructose malabsorption: true condition or a variance from normality. *J Clin Gastroenterol*. 2011;45:16–21.
107. Gibson PR, Varney J, Malakar S, Muir JG. Food components and irritable bowel syndrome. *Gastroenterology*. 2015;148, 1158.e4–1174.e4.
108. Stanghellini V, Cogliandro RF, de Giorgio R, Barbara G, Morselli-Labate AM, Cogliandro L, et al. Natural history of chronic idiopathic intestinal pseudo-obstruction in adults: a single center study. *Clin Gastroenterol Hepatol*. 2005;3:449–58.
109. Knowles CH, Lindberg G, Panza E, de Giorgio R. New perspectives in the diagnosis and management of enteric neuropathies. *Nat Rev Gastroenterol Hepatol*. 2013;10:206–18.
110. Camilleri M, Bharucha AE, di Lorenzo C, Hasler WL, Prather CM, Rao SS, et al. American Neurogastroenterology and Motility Society consensus statement on intraluminal measurement of gastrointestinal and colonic motility in clinical practice. *Neurogastroenterol Motil*. 2008;20:1269–82.
111. Camilleri M, Hasler WL, Parkman HP, Quigley EM, Soffer E. Measurement of gastrointestinal motility in the GI laboratory. *Gastroenterology*. 1998;115:747–62.
112. Parkman HP, Jones MP. Tests of gastric neuromuscular function. *Gastroenterology*. 2009;136:1526–43.
113. Johansson SG, Hourihane JO, Bousquet J, Bruijnzeel-Koomen C, Dreborg S, Haahrtela T, et al. A revised nomenclature for allergy. An EAACI position statement from the EAACI nomenclature task force. *Allergy*. 2001;56:813–24.
114. Chafen JJ, Newberry SJ, Riedl MA, Bravata DM, Maglione M, Suttorp MJ, et al. Diagnosing and managing common food allergies: a systematic review. *JAMA*. 2010;303:1848–56.
115. Turnbull JL, Adams HN, Gorard DA. Review article: the diagnosis and management of food allergy and food intolerances. *Aliment Pharmacol Ther*. 2015;41:3–25.
116. Lomer MC. Review article: the aetiology, diagnosis, mechanisms and clinical evidence for food intolerance. *Aliment Pharmacol Ther*. 2015;41:262–75.

117. Nwari BI, Hickstein L, Panesar SS, Roberts G, Muraro A, Sheikh A, et al. Prevalence of common food allergies in Europe: a systematic review and meta-analysis. *Allergy*. 2014;69:992–1007.
118. Böhn L, Störsrud S, Törnblom H, Bengtsson U, Simrén M. Self-reported food-related gastrointestinal symptoms in IBS are common and associated with more severe symptoms and reduced quality of life. *Am J Gastroenterol*. 2013;108:634–41.
119. Aydinlar EI, Dikmen PY, Tiftikci A, Saruc M, Aksu M, Gunsoy HG, et al. IgG-based elimination diet in migraine plus irritable bowel syndrome. *Headache*. 2013;53:514–25.
120. Atkinson W, Sheldon TA, Shaath N, Whorwell PJ. Food elimination based on IgG antibodies in irritable bowel syndrome: a randomised controlled trial. *Gut*. 2004;53:1459–64.
121. Ligaarden SC, Lydersen S, Farup PG. IgG and IgG4 antibodies in subjects with irritable bowel syndrome: a case control study in the general population. *BMC Gastroenterol*. 2012;12:166.
122. Hamilton RG. Clinical laboratory assessment of immediate-type hypersensitivity. *J Allergy Clin Immunol*. 2010;125:S284–96.
123. Soares-Weiser K, Takwoingi Y, Panesar SS, Muraro A, Werfel T, Hoffmann-Sommergruber K, et al. The diagnosis of food allergy: a systematic review and meta-analysis. *Allergy*. 2014;69:76–86.
124. Ortolani C, Bruijnzeel-Koomen C, Bengtsson U, Bindslev-Jensen C, Björkstén B, Høst A, et al. Controversial aspects of adverse reactions to food. European Academy of Allergology and Clinical Immunology (EAACI) reactions to Food Subcommittee. *Allergy*. 1999;54:27–45.
125. Fasano A, Sapone A, Zevallos V, Schuppan D. Nonceliac gluten sensitivity. *Gastroenterology*. 2015;148:1195–204.