

The pathophysiology of DKA caused by SGLT2 inhibitors differs in several aspects from that of classical DKA^{2,3}:

- DKA occurs with lower glucose levels (blood glucose <250 mg/dL) due to the greater renal losses of glucose induced by SGLT2 inhibitors. The decrease in the amount of glucose available as an energy substrate is the main causal mechanism. The decreased renal elimination of ketone bodies due to the stimulation of their tubular reabsorption is also involved in the genesis of DKA.
- DKA develops with slightly decreased insulin levels (mild insulinopenia); these occur as compensation when blood glucose levels decrease (relative insulinopenia), and are not an essential pathophysiological element.
- Elevated glucagon levels appear upon the stimulation of glucagon secretion by pancreatic α -cells which express SGLT2 receptors. The participation of SGLT2 inhibitors in the regulation of glucagon gene expression and in gluconeogenesis has recently been shown.⁴ There is also an indirect mechanism, i.e. an increased insulin/glucagon ratio after the decrease in insulin levels.

To sum up, the reported patient with T1DM experienced moderate DKA associated with the use of canagliflozin after 13 days of treatment and triggered by alcohol consumption. Blood glucose levels were initially normal, but insulin discontinuation combined with decreased food intake promoted the subsequent development of DKA with overt hyperglycemia.

DKA is a serious complication. Few cases of DKA have been reported in patients treated with SGLT2 inhibitors, and the atypical presentation already discussed should be emphasized. Health care professionals should inform patients treated with SGLT2 inhibitors about the symptoms of DKA and the need for consulting their doctor if they occur. In these cases, the measurement of ketonemia should be performed irrespective of blood glucose levels.

False overt hyperthyroidism by interference in immunoassay[☆]



Falso hipertiroidismo franco por interferencia en inmunoanálisis

Despite the great advances in the sensitivity and specificity of laboratory methods, cases with significant clinical-biochemical discrepancies still occur in some patients. A case of technical interference in the laboratory is reported below.

This was a 38-year-old male patient with a history of X-linked adrenoleukodystrophy (XLAD), confirmed by genetic study and fatty acid tests. Laboratory tests and clinical

[☆] Please cite this article as: Simó-Guerrero O, Giménez-Pérez G, Recasens-Gracia A, Villà-Blasco C, Castells-Fusté I. Falso hipertiroidismo franco por interferencia en inmunoanálisis. Endocrinol Nutr. 2016;63:431–432.

The most recent statement issued by the EMA⁵ recommends for patients being treated with SGLT2 inhibitors the temporary interruption of treatment in the event of surgery or severe intercurrent disease. It also advises that patients be informed regarding the risk situations that may predispose them to ketoacidosis, such as intake restriction, severe dehydration, alcohol abuse, or increased insulin requirements due to infection.

References

1. Henry RR, Rosenstock J, Edelman S, Mudaliar S, Chalamdaris AG, Kasichayanula S, et al. Exploring the potential of the SGLT2 inhibitor dapagliflozin in type 1 diabetes: a randomized, double-blind, placebo-controlled pilot study. *Diabetes Care*. 2015;38:412–9.
2. Taylor SI, Blau JE, Rother KI. SGLT2 inhibitors may predispose to ketoacidosis. *J Clin Endocrinol Metab*. 2015;100: 2849–52.
3. Rosenstock J, Ferrannini E. Euglycemic diabetic ketoacidosis: a predictable, detectable, and preventable safety concern with SGLT2 inhibitors. *Diabetes Care*. 2015;38:1638–42.
4. Bonner C, Kerr-Conte C, Gmyr V, Queniat G, Moerman E, Thévenet J, et al. Inhibition of the glucose transporter SGLT2 with dapagliflozin in pancreatic alpha cells triggers glucagon secretion. *Nat Med*. 2015;21:512–7.
5. European Medicines Agency. Human medicines – SGLT2 inhibitors; 2016. Available from: http://www.ema.europa.eu/ema/index.jsp?curl=pages/medicines/human/referrals/SGLT2_inhibitors/human_referral_prac_000052.jsp&mid=WC0b01ac05805c516f

Marta Diéguez-Felechosa*, Lorena Suárez-Gutiérrez

Sección de Endocrinología y Nutrición, Hospital de Cabueñas, Gijón, Asturias, Spain

* Corresponding author.

E-mail address: marta.dieguez@sespa.prinast.es (M. Diéguez-Felechosa).

signs were also consistent with adrenal insufficiency. Since diagnosis the patient had received treatment with low-dose hydrocortisone, 15 mg daily in two divided doses, and was asymptomatic. He had been clinically monitored by the endocrinology department of our hospital since 2010. The patient had participated in different clinical trials for patients with XLAD, in which he received different antioxidant drugs. In January 2016, thyroid function tests showed a thyroid-stimulating hormone level of 0.07 mcU/mL (reference range: 0.30–5 mcU/mL) and a free thyroxine level of 3.89 ng/dL (reference range: 0.93–1.7 ng/dL). The patient denied any clinical symptom of hyperthyroidism, and the physical examination was normal.

Because of the clinical-biochemical discordance, the patient was asked about the treatment he was receiving, and reported that he was participating in a clinical trial with biotin (300 mg/day). The references consulted confirmed that biotin may interfere with laboratory tests using antibodies (Ab) or antigens (Ag) conjugated with biotin in their reactions. This was the case in the initial results of

TSH and FT4 obtained with TSH and FT4 II REAGENTS (Roche diagnostics®, Modular E170). It was therefore decided to repeat the thyroid function tests using an alternative chemiluminescence immunoassay for TSH and FT4 (Siemens, ADVIA Centaur®) that uses acridinium ester to label the reaction. The results obtained were: FT4 1.03 ng/dL (reference range: 0.89–1.76 ng/dL) and TSH 2.340 mcU/mL (reference range: 0.550–4.780 mcU/mL). Hyperthyroidism was ruled out, and technical laboratory interference was confirmed.

Vitamin B⁸, vitamin H, or biotin is a water soluble B-group vitamin occurring in small amounts in many foods. Biotin acts as a cofactor for carboxylase enzymes, including acetyl-CoA carboxylase, which is involved in fatty acid biosynthesis and elongation; pyruvate carboxylase, involved in gluconeogenesis; methylcrotonyl-CoA carboxylase, essential for leucine degradation; and propionyl-CoA carboxylase. Although mammals cannot synthesize biotin, biotin deficiency is very rare, because it is present in a wide variety of vegetable and animal foods.¹ The recommended daily doses in adults are 30 µg/day of biotin.² Biotin supplements are indicated in deficiency states and pregnancy. Some studies have related biotin supplementation to improved metabolic control in diabetic rats.^{1,3} Biotin is also used to treat alopecia.⁴ It is also used, as in the case reported, for the treatment of progressive multiple sclerosis and adrenomyeloneuropathy⁵ at high doses (300 mg/day), up to 10,000 times higher than the recommended daily dose for healthy adults.

Despite technological advances and the availability of increasingly sensitive laboratory tests, discrepancies between clinical and biochemical findings sometimes occur. Abs causing interference in immunoassays (heterophile Abs, rheumatoid factor, etc.) may cause discrepancies in the measurement of thyroid hormones^{6,7} or even of multiple hormones, at the same time.⁸

The investigation of interferences should usually include repeat measurements, the performance of tests with dilutions, the addition of immunoglobulins to block interfering antibodies, or the use of an alternative immunoassay; the latter is the easiest and fastest means of evaluating any potential interference.^{6,7}

In the case reported, the patient had values consistent with hyperthyroidism but no clinical signs of this condition and was taking high doses of biotin, which led us to suspect laboratory interference.

Biotin is used in some commercial assay kits because of its easy binding to Abs and, to a lesser extent, antigen. Biotin interference may occur in immunoassays using the vitamin.

TSH is measured using non-competitive or immunometric (sandwich) immunoassays. The latter use two different monoclonal Abs which bind to different parts of the Ag. One Ab is usually on a solid support, and the other Ab is labeled. The TSH and FT4 II immunoassay from Roche diagnostics for measuring TSH, uses biotinylated monoclonal TSH-specific Abs and monoclonal TSH-specific Abs labeled with ruthenium chelate, which react to form the complex (sandwich). After the addition of streptavidin, the complex binds to the solid part through the interaction between biotin and streptavidin. Microparticles are captured magnetically

by an electrode, and the application of a voltage induces chemiluminescence, which is the parameter measured. Biotin concentrations greater than 25 µg/L distort the result by competing with biotinylated Abs at their binding site with streptavidin, causing a false low signal (falsely low TSH levels). To measure FT4 levels, a competitive immunoassay with anti-thyroxine monoclonal Abs is used, in which T4 is biotinylated. High biotin concentrations cause a reduction in the light generated, which is inversely proportional to the concentration of the free part of the analyte. Thus, high biotin concentrations cause positive interference with FT4.⁹

When an immunoassay using no biotin (such as the Advia Centaur®, Siemens) is used, the interference disappears, showing the actual thyroid function of the patient.

Interference with immunoassays requires constant vigilance and good communication between laboratory professionals and clinicians. Although such situations are exceptional, they should be aware of any potential interference and of the need to verify the consistency of the results with the clinical signs before starting unnecessary interventions or treatments.

References

1. Valdés-Ramos R, Guadarrama-López A, Martínez-Carrillo E, Benítez-Arciniega A. Vitamins and type 2 diabetes mellitus. *Endocr Metab Immune Disord Drug Targets*. 2015;1:54–63.
2. Zempleni J, Kuroishi T. Biotin. *Adv Nutr*. 2012;3:213–4.
3. Xiang X, Liu Y, Zhang X, Wang Z. Effects of biotin on blood glucose regulation in type 2 diabetes rat model. *Wei Sheng Yan Jui*. 2015;44:185–9.
4. Famenini S, Goh C. Evidence for supplemental treatments in androgenetic alopecia. *J Drugs Dermatol*. 2014;13:809–12.
5. Sedel F, Bernard D, Mock DM, Tourbah A. Targeting demyelination and virtual hypoxia with high-dose biotin as treatment for progressive multiple sclerosis. *Neuropharmacology*. 2015, <http://dx.doi.org/10.1016/j.neuropharm.2015.08.028>. S0028-3908(15)30073-3.
6. Kellogg MD, Law T, Huang S, Rifai N. A Girl with goiter and inappropriate thyroid-stimulating hormone secretion. *Clin Chem*. 2008;54:1242–7.
7. Van der Watt G, Haarburger D, Berman P. Euthyroid patient with elevated serum free thyroxine. *Clin Chem*. 2008;54: 1239–47.
8. Gulbahar O, Konca Degertekin C, Akturk M, Yalcin MM, Kalan I, Atikeler GF, et al. A case with immunoassay interferences in the measurement of multiple hormones. *J Clin Endocrinol Metab*. 2015;100:2147–53.
9. Kwok JS, Chan IH, Chan MH. Biotin interference on TSH and free thyroid hormone measurement. *Pathology*. 2012;44:278–80.

Olga Simó-Guerrero^{a,*}, Gabriel Giménez-Pérez^b, Assumpta Recasens-Gracia^b, Carme Villà-Blasco^a, Ignasi Castells-Fusté^b

^a Servicio de Laboratorio Clínico, Hospital General de Granollers, Granollers, Barcelona, Spain

^b Unidad de Diabetes, Endocrinología y Nutrición, Hospital General de Granollers, Granollers, Barcelona, Spain

* Corresponding author.

E-mail address: osimo@fhag.es (O. Simó-Guerrero).