

ORIGINAL ARTICLE

Pituitary tumor transforming gene and insulin-like growth factor 1 receptor expression and immunohistochemical measurement of Ki-67 as potential prognostic markers of pituitary tumors aggressiveness

Laura Sánchez-Tejada^{a,e}, Ruth Sánchez-Ortiga^{a,e}, Óscar Moreno-Pérez^a, Carmen Fajardo Montañana^b, Maria Niveiro^c, Nicholas A. Tritos^d, Antonio M. Picó Alfonso^{a,*}

^a Endocrinology Department, Research Unit, Hospital General Universitario Alicante, Alicante, Spain

^b Endocrinology Department, Hospital Universitario La Ribera, Spain

^c Department of Pathology, Hospital General Universitario Alicante, Alicante, Spain

^d Neuroendocrin Unit, Massachusetts General Hospital and Harvard Medical School, United States

Received 24 June 2012; accepted 28 September 2012

KEYWORDS

Pituitary adenoma; Pituitary tumor transforming gene protein; Humans; Ki-67 antigen; Insulin-like growth factor 1 receptor

Abstract

Introduction and objective: The ability to predict recurrence of pituitary adenoma (PA) after surgery may be helpful to determine follow-up frequency and the need for adjuvant treatment. The purpose of this study was to assess the prognostic capacity of pituitary tumor transforming gene (PTTG), insulin-like growth factor 1 receptor (IGFIR), and Ki-67. *Materials and methods*: In this retrospective study, the normalized copy number (NCN) of PTIG and IGFIR mRNA was measured using RT-PCR, and the Ki-67 index was measured by immunohistochemistry in 46 PA samples. Clinical data, histological subtype, and radiographic characteristics were collected to assess associations between variables and tumor behavior. Progression of tumor remnants and its association to markers was also studied in 14 patients with no adjuvant treatment after surgery followed up for 46 ± 36 months.

* Corresponding author.

E-mail address: antonio.pico@umh.es (A.M.P. Alfonso).

^e These authors contributed equally to this work.

1575-0922/\$ - see front matter © 2012 SEEN. Published by Elsevier España, S.L. All rights reserved. http://dx.doi.org/10.1016/j.endoen.2013.10.001

Abbreviations: PA, pituitary adenomas; PTTG, pituitary tumor transforming gene; IGFIR, insulin-like growth factor 1 receptor; RT-PCR, real time polymerase chain reaction; PI3K/Akt, phosphoinositide 3-kinase/Akt pathway; MAPK, mitogen-activated protein kinases; MRI, magnetic resonance imaging; FFPE, formalin-fixes and paraffin-embedded; FPA, functioning pituitary adenoma; NFPA, non-functioning pituitary adenoma; PRL, prolactin; ACTH, adrenocorticotropic hormone; GH, growth hormone; TSH, Thyroid stimulating hormone; DA, dopamine agonist; SSa, somatostatin analogs; ROC, receiver operating characteristics; LT, lactotrophic adenomas; CT, corticotrophic adenomas; TT, Thyrotrophic adenomas; ST, somatotrophic adenomas; GT, gonadotrophic adenomas; NC, null-cell adenomas.

Results: Extrasellar tumors had a lower PTTG expression as compared to sellar tumors (0.065 [1st-3rd quartile: 0.000-0.089] NCN vs. 0.135 [0.105-0.159] NCN, p = 0.04). IGFIR expression changed depending on histological subtype (p = 0.014), and was greater in tumor with remnant growth greater than 20% during follow-up (10.69 ± 3.84 NCN vs. 5.44 ± 3.55 NCN, p = 0.014). *Conclusions*: Our results suggest that the IGFIR is a more helpful molecular marker than PTTG in PA management. Ki-67 showed no association to tumor behavior. However, the potential of these markers should be established in future studies with standardized methods and on larger samples.

© 2012 SEEN. Published by Elsevier España, S.L. All rights reserved.

Expresión del gen transformador de tumores hipofisarios y del receptor del factor de crecimiento insulinoide 1 y determinación inmunohistoquímica de Ki-67 como posibles marcadores pronósticos de la agresividad de los tumores hipofisarios

Resumen

Introducción y objetivo: La capacidad de predecir recurrencia en los adenomas hipofisarios (AH) tras la cirugía puede ser útil para determinar la frecuencia de seguimiento y la necesidad de tratamientos adyuvantes. El objetivo del presente estudio fue valorar la capacidad pronóstica de gen transformador de tumores hipofisarios (*pituitary tumor transforming gene* [PTTG]), del receptor del factor de crecimiento insulinoide 1 (*insulin-like growth factor 1 receptor* [IGFIR]) y de Ki-67.

Material y métodos: En este estudio retrospectivo determinamos el número de copias normalizadas de ARNm (Cnn) de PTTG e IGFIR mediante RT-PCR y el índice Ki-67 mediante inmunohistoquímica en 46 muestras de AH. Los datos clínicos, el subtipo histológico y las características radiológicas se recogieron para determinar asociaciones entre las variables y el comportamiento tumoral. Además, estudiamos la progresión de los restos tumorales y su asociación con los marcadores en 14 pacientes sin tratamiento adyuvante posquirúrgico seguidos durante 46 \pm 36 meses.

Resultados: Los tumores extraselares mostraron una expresión de PTTG menor que los intraselares (0,065 [1.^{er}-3.^{er} cuartil: 0,000-0,089] Cnn frente a 0,135 [0,105-0,159] Cnn, p=0,04). La expresión de IGFIR varió en función del subtipo histológico (p=0,014), siendo mayor en los tumores que presentaron crecimiento de los restos mayor del 20% durante el seguimiento (10,69 \pm 3,84 Cnn frente a 5,44 \pm 3,55 Cnn, p=0,014).

Conclusiones: Nuestros resultados indican que IGFIR, en mayor medida que PTTG, es un marcador molecular útil en el manejo de los AH. Ki-67 no mostró asociación con el comportamiento tumoral. Sin embargo, el potencial de estos marcadores debe ser establecido en futuros estudios con una metodología estandarizada y una muestra mayor.

© 2012 SEEN. Publicado por Elsevier España, S.L. Todos los derechos reservados.

Introduction

Pituitary adenomas (PA) constitute 10–25% of intracranial neoplasms. They are almost always benign, but some show aggressive behavior with local invasion and recurrences. Several sporadic mutations of oncogenes and tumor-suppressor genes have been found in PA, but none has been found to be a reliable marker of poor outcome.^{1,2} In clinical practice, Ki-67 antigen expression is frequently used as a prognostic indicator because it has been associated with proliferative potential and invasiveness of several human malignancies, but it has shown discordant results in pituitary adenomas.^{3,4}

Thus, there is an increasing interest in finding specific prognostic markers. Pituitary tumor transforming gene (PTTG) encodes a protein that functions like securin and transcription factor.^{5,6} It is involved in the cell cycle regulation and may induce cellular proliferation and aneuploidy.^{6,7} Overexpression of PTTG has been described in several neoplasms including PA.^{8–10}

IGFIR is a tyrosine kinase receptor responsible for mediating IGF-I signaling, which plays a critical role in normal growth and has been associated with the early stages of tumor establishment. IGFIR stimulates the PI3K/Akt and MAPK pathways,¹¹ resulting in cell proliferation and apoptosis alteration. Its overexpression has been documented in many human malignancies¹² but it is not yet well studied in PA.

The aim of this study is to examine whether PTTG and IGFIR expression or Ki-67 index may have prognostic implications on pituitary adenomas.

Materials and methods

Subjects

This is a retrospective study performed at Hospital General Universitario de Alicante including patients who underwent

PALABRAS CLAVE Adenoma hipofisario; Proteína del gen transformador de tumores hipofisarios; Humanos; Antígeno Ki-67; Receptor del factor de crecimiento insulinoide 1

Table 1Patient characteristics.					
Age (mean ± SD) Men (n (%)) Largest diameter (mean ± SD)	53 ± 15 years 25 (54%) 30 ± 16 mm				
Histological classification (n (%)) Gonadotrophic adenomas Somatotrophic Corticotrophic Lactotrophic Thyrotrophic Null-cell adenoma	27 (58.7%) 10 (21.7%) 4 (8.7%) 3 (6.5%) 1 (2.2%) 1 (2.2%)				
Functioning status (n (%)) Functioning pituitary tumor Non-functioning pituitary tumor	13 (28.3%) 33 (71.7%)				
Extension according to Hardy's classij Grades I–II Grades III–IV	fication (n (%)) 4 (8.9%) 41 (91.9%)				
Pre-operative therapies (n) Dopamine agonist Somatostatin analogs	3 5				

surgery for PA between 1995 and 2008. We included cases that had complete hormonal tests and magnetic resonance imaging (MRI) upon diagnosis. Minimum follow-up needed was 3 months after surgery, with the exceptions of patients who died prior to this (2 patients died of postoperative complications and one due to cardiac arrest). Clinical data were retrieved from medical files. Tumor tissue samples were obtained from the formalin-fixed and paraffinembedded remaining fragments of pathological diagnosis. Overall, 46 patients were included in the study. This study was approved by the Ethics Committee of the Hospital General Universitario de Alicante.

Histological classification was performed according to WHO 2004 classification criteria (Table 1). We also assessed functional status, considering functioning pituitary tumors (FPA), those associated with hypersecretion of prolactin, ACTH, GH, TSH and specific hormone-related syndrome. Lack of a hormone related syndrome, whether associated with biologically inactive hypersecretion or not, was defined as a non-functioning pituitary adenoma (NFPA). Invasiveness was defined as optic chiasm compression or invasion of surrounding structures like cavernous or sphenoidal sinus. Other baseline characteristics are summarized in Table 1.

As this is a retrospective study, we considered patient follow-up to be the time between surgery and the last visit to the Endocrinology Unit. Following surgery, fourteen patients received adjuvant therapies: DA (1), SSa (9) or ketoconazole (2). Thirteen patients received radiotherapy and 12 underwent a second surgery during the follow-up period due to incomplete resection after first surgery (only one of the tumor remnants experienced significant growth before radiotherapy). Then, we studied the progression of PA after surgery in patients who did not receive early postoperative treatment. Fourteen patients, with a 46 ± 36 (mean \pm SD) months of follow-up, were included in this analysis. Aggressiveness was defined as the growth of tumor remnants greater than 20% in any of the tumor diameters measured.

We reviewed all the MRI scans performed during the followup period in order to establish the time of progression.

Immunohistochemical study: Ki-67 index

Immunohistochemical study was performed as previously described.¹³ The number of positive cells defines the Ki-67 index. We applied a Ki-67 index cut-off values of 3% associated with high proliferative activity.

RNA source, total RNA extraction and reverse transcription

We used 2 cylindrical cores (0.6 mm-thick) from pre-selected tumor areas of FFPE tissues, to extract and isolate total RNA by using RNAeasy FFPE Kit (QIAgen) according to the protocol and working under manufacturer-recommended conditions. We assessed the quality and quantity of RNA extracted from this using the NanoDrop. Reverse transcription of 2 μ g of RNA was performed in 20 μ L reaction volumes with random hexamer primers, with High Capacity cDNA Reverse Transcription Kit (Applied Biosystems[®]) and stored at -20 °C until used.

Real time Q-PCR

To select the control genes, we analyzed the expression of 32 housekeeping genes in three pools consisting of 10 sample mixes each, on a commercial endogenous plate based on TaqMan assay technologies (Applied Biosystems). The results, when analyzed with GeNorm software, showed that the most stable genes were GAPDH and YWHAZ.

To analyze the gene expression of PTTG and IGFIR genes, we chose the TaqMan Assays for each gene under manufacturer-recommended conditions. We constructed standard curves that were used to check that the amplification efficiency of the housekeeping genes selected and the target genes were similar (with a difference less than 10%) and estimated the number of copies of mRNA. Specific primers were designed to amplify the cDNA fragments detected by each TaqMan assay with Primer-BLAST. The primers chosen were subject to the Basic Local Alignment Search Tool (BLAST) database, which searches to find any similarities in sequence. We used commercial brain total RNA (Ambion[®]) as a template, which was transcribed together with adenoma samples. PCR amplification was performed in a 25 μ L reaction volume containing 2.5 mM MgCl₂, 0.5 µM of each primer, 1 unit of KAPA2G Robust HotStar Taq polymerase (KAPABiosystems[®]) and 25 ng of cDNA. PCR reactions were amplified for 35 cycles with an annealing temperature of 55 °C. The amplified fragments were purified by Qiaquick Purification Kit (Qiagen®). We confirmed the specificity of the primers and obtained the concentration of the amplified fragments using the 2100 Bioanalyzer (Agilent[®]). Dilutions were made for standards such as 10⁹, 10^{10} , 10^{11} , 10^{12} , 10^{13} and 10^{14} copies of mRNA, calculated from the total ng of the PCR product (according to the formula MW = [number of nucleotides \times 607.4] + 157.9). Real-time PCRs of serial dilutions were performed in triplicate in $12.5 \,\mu\text{L}$ reaction volumes containing $6.25 \,\mu\text{L}$ of TaqMan Gene Expression Master Mix (Applied Biosystems[®]), 0.625 μ L of TaqMan Gene Expression Assay, and 2.5 μ L of template in 7500 Real Time PCR System (Applied Biosystems) and were analyzed using SDS software (Applied Biosystems[®]). The R2 values for all standard curves generated were higher than 0.99 and all the efficiencies of the TaqMan assays were higher than 90%. We made an RT-minus control using Total Human DNA (Roche) to ensure that the TaqMan assays did not detect amplification of genomic DNA.

Analysis of the samples was performed in accordance with the same protocol. We included two dilutions of standard curves and no template control (NTC) on all QPCR plates to ensure that efficiencies were maintained and that they were free from contamination. All analyses were performed in duplicate. The results from each plate were only validated when the NTC was undetectable and the dCt maintained a standard deviation below 0.33 between duplicates and between both dilutions of standard curves and their initial results.

Since both housekeeping genes were highly correlated (Spearman rank correlation coefficient: r = 0.82; p < 0.000), all the results were normalized to YWHAZ. We used standard curves to estimate the copy number normalized (Cnn) of PTTG and IGFIR.

Statistical analyses

Data were tested for statistical significance using SPSS 11.0 software (SAS Institute®). Associations between the molecular assays and the clinical-pathological features were calculated. IGFIR showed a normal distribution, whereas the rest of molecular and IHC variables showed a non-normal distribution (Kolmogorov-Smirnov test). The Student's t-test and the Mann Whitney U-test were used for comparison between two groups of parametric and nonparametric data, respectively. The Chi-square test followed by Fisher's exact test, where appropriate, was used to identify correlations between categorical parameters. The Kruskal-Wallis test was used for comparison of more than two categories of nonparametric data. Receiver operating characteristics (ROC) curves were used to establish the optimal PTTG and IGFIR cut-offs for distinguishing between invasive and/or aggressive pituitary adenomas. Kaplan-Meier survival analysis and the log-rank test were used to compare progression-free interval curves. The effect of a variable was estimated by the odds ratio (OR) of the model and a 95% confidence interval (95% CI) was calculated for each OR. Data are shown as mean \pm standard deviation (normal distribution) or median and quartiles (non-normal distribution). p values < 0.05 were considered statistically significant.

Results

We investigated a series of 46 patients with pituitary adenomas. Tumors were classified according to WHO 2004. Table 1 summarizes the clinical-pathological features.

Expression of IGFIR mRNA, PTTG mRNA and Ki-67 index on histological subtypes of PA

IGFIR expression differed between histological subtypes (p=0.014) and had its greatest expression in NC, followed by GT. There were no statistically significant differences in

PTTG mRNA expression or Ki-67 index between different histological subtypes, although the Ki-67 index was higher in LT. We did not find significant difference in the expression of molecular variables between FPA and NFPA, although there was a trend toward more IGFIR expression in the NFPA subgroup (5.82 ± 6.43 Cnn vs. 9.85 ± 6.91 Cnn, p = 0.077) (Fig. 1).

Radiologic findings

Patients with extrasellar extension showed lower PTTG mRNA expression (0.065 $[P_{25}-P_{75}: 0.000-0.089]$ Cnn vs. 0.135 [0.105-0.159] Cnn, p=0.04) (Fig. 2). ROC analysis identified a significant cut-off point for PTTG predicting extrasellar extension (Fig. 3a). A cut-off of 0.129 PTTG mRNA Cnn or less predicted extrasellar extension with a sensitivity of 75% and specificity of 85%. Cases with less than 0.129 Cnn of PTTG mRNA had 17.5-fold risk of extrasellar extension (95% CI, 1.6-197.4, p=0.02). There were no statistically significant differences in clinical variables (age, sex, tumor size, histological classification, functioning status, extension according to Hardy's classification or preoperative therapies), IGFIR mRNA expression or the Ki-67 index (Fig. 2).

Association with growth

We studied the tumoral behavior of 14 patients who did not receive adjuvant postoperative therapies (Table 2). During follow-up period, 7 tumors grew more than 20% on at least one of the diameters measured. The average time free of progression was 26 ± 15 months. All tumors had extrasellar extension at diagnosis.

Only IGFIR mRNA Cnn showed an association with progression (10.69 \pm 3.84 Cnn vs. 5.44 \pm 3.55 Cnn, p = 0.014) (Fig. 4). Patients with more than 4.61 Cnn of IGFIR (25th percentile) had 2.3 (95% Cl 0.99–5.49, p = 0.07) times more risk of regrowth during the follow-up period. Through ROC curve analysis, this cut-off had a sensitivity of 100% and specificity of 75% (p = 0.010) (Fig. 3b). However, PTTG and Ki-67 index were not associated with tumor progression. Similarly, neither age nor sex showed association with progression.

On survival analysis, although without statistical significance, tumors without PTTG expression or with IGFIR expression less than 9 Cnn (50th percentile) had more time free of progression (32.3 ± 10.3 vs. 24.3 ± 7.36 months, p > 0.05; and 37.0 ± 0 vs. 24.0 ± 7.6 months, p > 0.05, respectively).

Discussion

Some PAs demonstrate aggressive behavior, which lends to invasiveness of adjacent local structures, or have early regrowth (recurrence) following surgery. Thus, it is very important to identify the tumors that will become aggressive early on, in order to intensify the follow-up or to consider adjuvant therapies.

Nowadays, the pathological index most used to predict the aggressive behavior of PAs has been the Ki-67 index. However, it is a semi-quantitative measurement that is

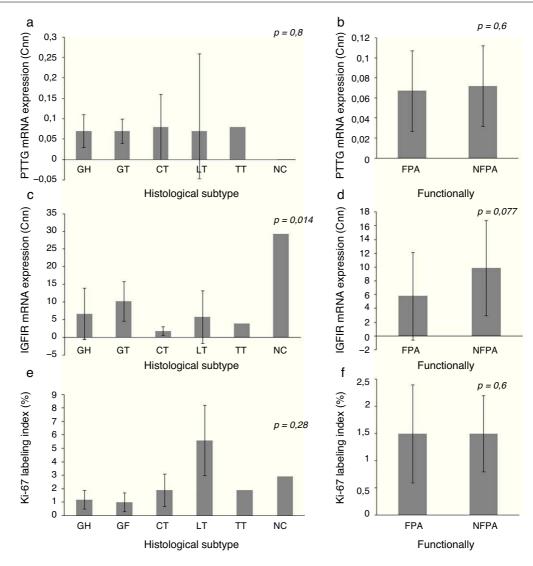


Figure 1 Pituitary tumor transforming gene (PTTG) and insulin-like growth factor I receptor (IGFIR) Cnn expression and Ki-67 index (%) according to histological subtypes (a, c, e) and functional status (b, d, f). Data reported as mean \pm S.E.M. Non-parametric Kuskal–Wallis and *U*-Mann Whitney tests were used with *p* values of 0.05 or less being considered significant. *Abbreviations*: GH, somatotrophic adenomas; GT, gonadotrophic adenomas; CT, corticotrophic adenomas; LT, lactotrophic adenomas; TT, Thyrotrophic adenomas; NC, null-cell adenomas; FPA, functioning pituitary adenoma; NFPA, non-functioning pituitary adenoma; Cnn, copy number normalized.

highly dependent on the observer and the prognostic capability of Ki-67 index in PA has been less predictive than in others tumors.⁴

In our series, we did not find any difference in Ki-67 index between invasive and non-invasive tumors. Nevertheless, higher Ki-67 index was found in LT than in other histological subtypes (5.6% [3.6–7.8]) and the three LT were classified as invasive. Several authors have demonstrated that Ki-67 index is significantly higher in FPA than in NFPA.⁴ When we analyzed LT, CT, TT and ST together vs. GT and the one NC in our series, we did not find any significant differences. This discrepancy could be attributed to the fact that 5/10 ST in our series had been treated previously with SSa and one of the three LT had received dopamine agonist. It has been suggested that preoperative treatment of ST and LT adenomas may decrease Ki-67 levels, resulting in values similar to those of NFPA.¹⁴ In any case, several studies have failed to show a significant difference among adenomas of varying hormonal subtypes or functional status.⁴

Different criteria used to define extension or invasiveness could explain the controversy on literature regarding the relationship between the invasiveness and Ki-67 index levels. Some authors consider invasion to be when there is histological evidence of dural invasion; but others, as us, base it on the radiologic findings on preoperative MRI. The discrepancy was greater when the invasion assessment was based on MRI imaging criteria. As an example, Wolfsberger et al.¹⁵ found a correlation between Ki-67 index and radiographic extension (1.9% in invasive adenomas vs. 1.7% in non-invasive adenomas) in adenomas 3 cm or less in diameter only. We did not find any significant difference in Ki-67 levels between invasive vs. non-invasive tumors (1.2

Please cite this article in press as: Sánchez-Tejada L, et al. Pituitary tumor transforming gene and insulin-like growth factor 1 receptor expression and immunohistochemical measurement of Ki-67 as potential prognostic markers of pituitary tumors aggressiveness. Endocrinol Nutr. 2013. http://dx.doi.org/10.1016/j.endonu.2012.09.005

Table 2 Clinical characteristics of patients without adjuvant therapies

No.	Age/sex	Histological subtype	Tumor size (mm)	Tumoral rest	Follow up (months)	Progression free time post surgery (months)	Ki-67 index (%)	PTTG (Cnn)	IGFIR (Cnn)
1	34/f	GT	15	Yes	60	11	1.0	0.042	13.071
2	70/f	GT	22	Yes	30	12	0.5	0.086	10.948
3	71/m	GT	20	Yes	36	21	2.2	0	17.257
4	67/f	GT	20	Yes	108	23	1.2	0	9.247
5	64/m	GT	32	Yes	48	37	1.0	0.101	5.049
6	67/m	GT	30	Yes	52	37	2.3	0.176	8.302
7	70/m	GT	50	Yes	88	53	2.3	0	10.979
8	43/m	GT	43	Yes	114	-	0.8	0.038	7.857
9	72/m	GT	28	Yes	3	-	2.0	0	3.087
10	50/f	LT	45	Yes	67	-	1.5	0	1.362
11	72/f	GT	15	Yes	20	-	4.5	0.152	10.266
12	67/f	GT	50	Yes	39	-	1.2	0.097	9.019
13	36/f	ST	42	No	24	-	1.2	0.064	2.298
14	63/f	GT	46	Yes	6	-	1.5	0.089	4.164

vs. 1.8), and we were unable to find a cut-off for Ki-67 index to predict the aggressive behavior of PAs that showed recurrence. Thus, we agree with Wolfsberger et al.¹⁵ having emphasized the limitation of relying solely on Ki-67 to predict tumor behavior. Moreover, only some studies demonstrated a correlation between Ki-67 and recurrence. These discrepancies may be attributable in part to variations in the definition of recurrence and the duration of follow-up. In our case, we considered aggressiveness as growth of primary tumor remnants and this situation has been found to be correlated to recurrence.¹⁶ With respect to follow up, we did not find a significant correlation between Ki-67 index and recurrence after 46 ± 36 months, similar to the findings of Scheithauer and Dubois that had follow-up over 9 years.^{1,17} Thus, it appears that Ki-67 alone is a poor predictor of tumor recurrence, which demonstrates the need to search for new markers of tumor behavior.

Many investigators have searched for other biological markers that could help with decision making. In recent years, IGFIR overexpression has been described in many types of cancer.^{11,12} However, the data on IGFIR in pituitary tumor pathology almost all come exclusively from animal pituitaries or established cell line derived from the pituitary and only a few authors have investigated this gene in human PA.

Otsuka et al.¹⁸ analyzed the mRNA expression of IGFIR, IGF-I, IGF-II and other growth factors in six LT, six ST and ten GT adenomas. They found lower IGFIR and IGF-I expression in ST than in LT and GT. In 2003, Kola et al.¹⁹ studied a series of 18 ST and other PAs and found that IGFIR mRNA levels were significantly lower in ST and tended to be lower in CT than in normal pituitary tissue. In our series, we also found different gene expression among histological subtypes, with the greatest expression in CT (Fig. 1), similar to the Otsuka et al. report.¹⁸ To date, no author has studied the relationship between IGFIR expression and the clinical behavior of PA, although it has become

apparent that it is associated with adverse prognosis in some type of tumors.²⁰ IGFIR overexpression has been associated with protecting cancer cells and giving the tumor capabilities such as anchorage-independent growth and surviving the process of detachment required for metastasis.^{21,22}

In our series, we did not find a relationship between IGFIR overexpression and the invasiveness of PA, possibly due to our arbitrary definition of invasion. However, we found a positive association between expression of the IGFIR gene and aggressiveness of the PA. In fact, IGFIR expression greater than 4.61 Cnn may distinguish tumors with significant growth during follow up with a sensitivity of 100% and specificity of 75%.

There are no references in literature with which to compare our results; however, they are consistent with studies carried out on the molecular biology of PAs. Thus, loss of E-cadherin expression or function and its nuclear translocation are well recognized as causing cell detachment and have been linked to pituitary tumor invasion.²³ The IGFIR pathway activation can result in the disruption of β -catenin/E-cadherin complexes, favoring the relocation of IRS-1/B-catenin complexes to the nucleus to enhance β -catenin transcriptional activity²⁴. Similarly, tumor cell motility and invasive potential are influenced by IGFinduced secretion of metalloproteinase, enzymes that can degrade and reorganize the extracellular matrix (ECM) and whose overexpression is usually associated with tumor invasion and metastasis.²⁵ Therefore, our results are consistent with the hypothesis that IGFIR expression can be related to the outcome of PA.

Another protein widely studied in regard to its relationship with tumorigenesis is PTTG. All studies that analyze normal pituitary tissue find higher PTTG expression in pituitary adenomas than in normal tissue. However, data from literature show a high discrepancy between the results of different studies of PTTG expression as marker of invasive and aggressive behavior of PA.¹⁰ Table 3 shows the conflicting views existing among different authors regarding the

Please cite this article in press as: Sánchez-Tejada L, et al. Pituitary tumor transforming gene and insulin-like growth factor 1 receptor expression and immunohistochemical measurement of Ki-67 as potential prognostic markers of pituitary tumors aggressiveness. Endocrinol Nutr. 2013. http://dx.doi.org/10.1016/j.endonu.2012.09.005

Author	Sample	Method	Results
Zhang et al. (1999) ²⁶	54 PA and normal pituitary tissue	RT-PCR	Higher levels of PTTG than normal pituitary tissues: 23/30 NPA, 9/10 LT, 13/13 ST, 1/1 CT. Higher expression in functioning pituitary adenomas with invasion of sinus sphenoid.
McCabe et al. (2003) ²⁷	121 PA: 92 NFPA, 16 ST, 5 LT, 5 CT, 5 TT. 10 normal pituitary tissue	qPCR Western Blot	5-Fold increased PTTG expression in PA vs. normal pituitary tissue. No correlation with radiological parameters or recurrence.
Filippella et al. (2006) ²⁹	45 PA: 18 NFPA, 8 ST, 14 LT, 6 CT. Normal pituitary tissue.	qPCR	Higher expression in PA vs. normal tissue (+89% of PA samples). No correlation with radiological parameters. Higher expression in recurrent tumors.
Wierinckx et al. (2007) ²⁸	25 LT Samples of rat LT	Immunohistochemistry	No correlation with clinical course.
Noh et al. (2009) ³⁰	35 NFPA: 26 with recurrence and 9 without	Immunohistochemistry	Association with early but no late recurrence.
Salehi et al. (2010) ³	89 PA	Immunohistochemistry	ST more expression than other tumors.
Our results	46 PA	qPCR	Inverse correlation with radiological parameters or recurrence

Table 3 Description of studies on relationship between pituitary tumor transforming gene (PTTG) expression and pituitary adenoma behavior.

Abbreviations: PA, pituitary adenoma; FPA, functioning pituitary tumors; NFPA, non-functioning pituitary tumors; GT, gonadotrophic adenoma; ST, somatotrophic adenoma; CT, corticotrophic adenoma; LT, prolactinoma; TT, thyrotrophic adenoma; NC, null-cell adenoma.

influence of PTTG on the behavior of a PA. Only Zhang et al.²⁶ find a correlation between its levels and the pre-operative extension of the FPA but not in NFPA.

Our results show lower PTTG expression in invasive tumors than in non-invasive tumors. When we studied only clinically functional adenomas, we found significantly higher PTTG expression in intrasellar vs. extrasellar tumors; however, we considered all types of extrasellar extension of PA, whereas Zhang et al.²⁶ found higher PTTG expression only in sphenoid sinus invasion. In addition, Zhang et al.²⁶ measured PTTG using a densitometry method on electrophoresis results after Reverse Transcription PCR, a semi-quantitative technique, while we used the current gold standard, realtime PCR with TaqMan® technology, which is a more sensitive and specific quantitative technique. On the whole, given the overall methodological differences, we feel that the results are not comparable. Although our results may suggest the role of PTTG as protective in FPA, the low number of patients studied in our series influences the strength of our results. Nevertheless, PTTG has demonstrated in vitro and in vivo activity as protooncogen, inducing cell proliferation and tumor growth, as well as securin, inhibiting proliferation; so this double function could explain the lower PTTG expression, with low securin activity, found in extrasellar tumors compared with intrasellar ones.⁹ After all, we agree with other authors^{27,28} that PTTG cannot be considered a biological marker of invasiveness on PA.

With respect to aggressiveness, Filippella et al.²⁹ in their series found that a PTTG score of 3.3% clearly distinguished between recurrent and non-recurrent pituitary adenomas with a sensitivity of 60% and specificity of 76% (ROC curve method); and Noh et al.³⁰ found early, no later than one year

after surgery, recurrence in tumors with the highest levels of PTTG. Paradoxically, to Filippella et al.²⁹ the fact that 18/45 patients in their series had a follow-up less than 1 year could dilute the relationship between PTTG and recurrence; therefore, they analyzed the results for 27 patients with more than 1 year of follow up separately. Thus, once again, the results from literature are conflicting. Moreover, McCabe et al.²⁷ using similar technology as us, real-time PCR, in the largest published PA series, did not observe a significant association between mRNA expression and the presence of recurrent tumor growth, although the authors did not report length of the follow up.

We demonstrated progression in 7/14 patients who did not receive post-operative adjuvant treatments during the follow-up (46 ± 36 months). To minimize bias, we performed an analysis of recurrence-free survival (Kaplan Meier). On the whole, we did not find a correlation between PTTG expression and aggressiveness; however, when we separated our sample by PTTG expression, the patients without PTTG expression show a tendency to have more time free of recurrence, although not reaching statistical significance.

Our main limitation was that we studied retrospectively a small cohort which could affect the strength of our results and our sample was a very heterogeneous group of tumors (different endocrine and non-endocrine types, in some cases only one representative of an adenoma type). Moreover, you should note that a comparison of the various tumor types in FPA is difficult since some types (e.g. prolactinomas) are treated surgically only when medical treatment fails. This selects for the more aggressive tumors and could lead to a selection bias. Furthermore, it has not been possible to use the relative quantification by qPCR using ddCT method

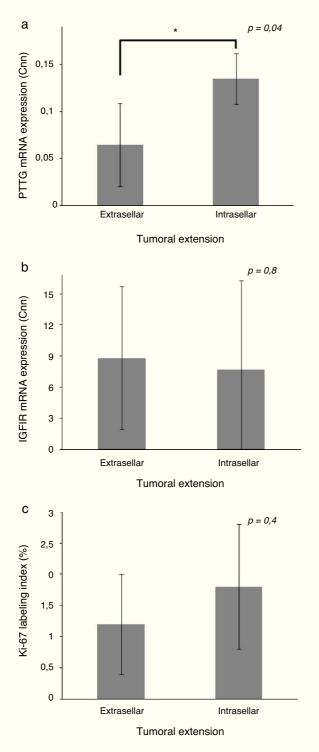


Figure 2 Comparison of pituitary tumor transforming gene (PTTG) (a) and insulin-like growth factor I receptor (IGFIR) (b) Cnn expression and Ki-67 index (%) (c) between intrasellar and extrasellar tumors. Data reported as mean \pm S.E.M. Non-parametric *U*-Mann Whitney test was used with *p* values of 0.05 or less being considered significant. *Abbreviations*: Cnn, copy number normalized.

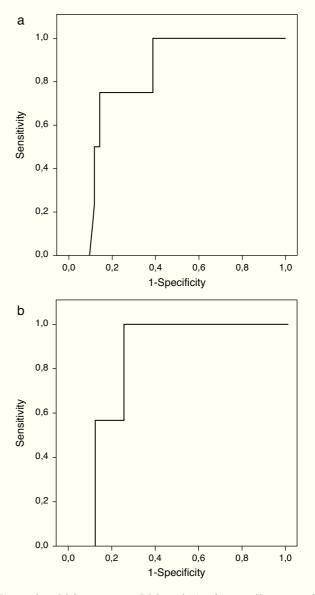


Figure 3 ROC curves: (a) ROC analysis of intrasellar tumoral extension and pituitary tumor transforming gene (PTTG) Cnn expression. (b) ROC analysis of tumoral progression on patients without adjuvant therapies and IGFIR Cnn expression. *Abbreviations*: Cnn, copy number normalized.

because it was not possible to obtain normal tissue. Nevertheless, we have adapted our protocol to estimate the number of copies of the different genes' transcripts and we have only compared the behavior of these genes among different subtypes of PA or among the different variables studied.

In conclusion, we have found that IGFIR is associated with progression of tumor rests and it can be new marker in PA outcome. In regard to PTTG, our data do not suggest clearly prognostic implications in PA according with some previous studies. Further work involving large series of patients and a prospective analysis needs to be performed in order to clarify the potential prognostic role of new biomarkers in clinical practice.

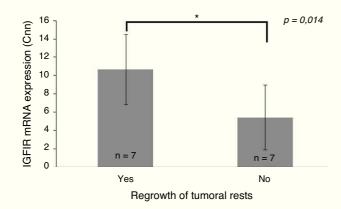


Figure 4 Differences between insulin-like growth factor I receptor (IGFIR) Cnn expression according to tumoral progression during the follow-up in tumors without post-surgical treatment. Data reported as mean \pm S.E.M. Non-parametric *U*-Mann Whitney test was used with *p* values of 0.05 or less being considered significant. *Abbreviations*: Cnn, copy number normalized.

Conflict of interest

This work was supported by Pfizer Grant.

References

- Scheithauer BW, Gaffey TA, Lloyd RV, Sebo TJ, Kovacs KT, Horvath E, et al. Pathobiology of pituitary adenomas and carcinomas. Neurosurgery. 2006;59:341–53, discussion 53.
- Vlotides G, Eigler T, Melmed S. Pituitary tumor-transforming gene: physiology and implications for tumorigenesis. Endocr Rev. 2007;28:165–86.
- Salehi F, Agur A, Scheithauer BW, Kovacs K, Lloyd RV, Cusimano M. Biomarkers of pituitary neoplasms: a review (part II). Neurosurgery. 2010;67:1790–8, discussion 8.
- Salehi F, Agur A, Scheithauer BW, Kovacs K, Lloyd RV, Cusimano M. Ki-67 in pituitary neoplasms: a review – part I. Neurosurgery. 2009;65:429–37, discussion 37.
- Zou H, McGarry TJ, Bernal T, Kirschner MW. Identification of a vertebrate sister-chromatid separation inhibitor involved in transformation and tumorigenesis. Science. 1999;285:418-22.
- 6. Tong Y, Eigler T. Transcriptional targets for pituitary tumortransforming gene-1. J Mol Endocrinol. 2009;43:179-85.
- Pei L. Identification of c-myc as a down-stream target for pituitary tumor-transforming gene. J Biol Chem. 2001;276: 8484-91.
- Pei L, Melmed S. Isolation and characterization of a pituitary tumor-transforming gene (PTTG). Mol Endocrinol. 1997;11:433-41.
- Salehi F, Kovacs K, Scheithauer BW, Lloyd RV, Cusimano M. Pituitary tumor-transforming gene in endocrine and other neoplasms: a review and update. Endocr Relat Cancer. 2008;15:721-43.
- Sanchez-Ortiga R, Sanchez Tejada L, Peiro Cabrera G, Moreno-Pérez O, Arias Mendoza N, Aranda López FI, et al. Rol of pituitary tumour-transforming gene (PTTG) in the pituitary adenomas. Endocrinol Nutr. 2010;57:28–34.
- 11. Hartog H, Wesseling J, Boezen HM, van der Graaf WT. The insulin-like growth factor 1 receptor in cancer: old focus, new future. Eur J Cancer. 2007;43:1895–904.

- Ouban A, Muraca P, Yeatman T, Coppola D. Expression and distribution of insulin-like growth factor-1 receptor in human carcinomas. Hum Pathol. 2003;34:803–8.
- Niveiro M, Aranda FI, Peiro G, Alenda C, Pico A. Immunohistochemical analysis of tumor angiogenic factors in human pituitary adenomas. Hum Pathol. 2005;36:1090–5.
- Jaffrain-Rea ML, Di Stefano D, Minniti G, Esposito V, Bultrini A, Ferretti E, et al. A critical reappraisal of MIB-1 labelling index significance in a large series of pituitary tumours: secreting versus non-secreting adenomas. Endocr Relat Cancer. 2002;9:103–13.
- Wolfsberger S, Wunderer J, Zachenhofer I, Czech T, Böcher-Schwarz HG, Hainfellner J, et al. Expression of cell proliferation markers in pituitary adenomas – correlation and clinical relevance of MIB-1 and anti-topoisomerase-IIalpha. Acta Neurochir (Wien). 2004;146:831–9.
- Nakabayashi H, Sunada I, Hara M. Immunohistochemical analyses of cell cycle-related proteins, apoptosis, and proliferation in pituitary adenomas. J Histochem Cytochem. 2001;49: 1193-4.
- 17. Dubois S, Guyetant S, Menei P, Rodien P, Illouz F, Vielle B, et al. Relevance of Ki-67 and prognostic factors for recurrence/progression of gonadotropic adenomas after first surgery. Eur J Endocrinol. 2007;157:141–7.
- Otsuka F, Tamiya T, Yamauchi T, Ogura T, Ohmoto T, Makino H. Quantitative analysis of growth-related factors in human pituitary adenomas. Lowered insulin-like growth factor-I and its receptor mRNA in growth hormone-producing adenomas. Regul Pept. 1999;83:31–8.
- Kola B, Korbonits M, Diaz-Cano S, Kaltsas G, Morris DG, Jordan S, et al. Reduced expression of the growth hormone and type 1 insulin-like growth factor receptors in human somatotroph tumours and an analysis of possible mutations of the growth hormone receptor. Clin Endocrinol (Oxf). 2003;59: 328–38.
- Chitnis MM, Yuen JS, Protheroe AS, Pollak M, Macaulay VM. The type 1 insulin-like growth factor receptor pathway. Clin Cancer Res. 2008;14:6364–70.
- Coppola D, Ferber A, Miura M, Sell C, D'Ambrosio C, Rubin R, et al. A functional insulin-like growth factor I receptor is required for the mitogenic and transforming activities of the epidermal growth factor receptor. Mol Cell Biol. 1994;14:4588–95.
- Samani AA, Brodt P. The receptor for the type I insulinlike growth factor and its ligands regulate multiple cellular functions that impact on metastasis. Surg Oncol Clin N Am. 2001;10:289–312, viii.
- Elston MS, Gill AJ, Conaglen JV, Clarkson A, Cook RJ, Little NS, et al. Nuclear accumulation of e-cadherin correlates with loss of cytoplasmic membrane staining and invasion in pituitary adenomas. J Clin Endocrinol Metab. 2009;94: 1436–42.
- Playford MP, Bicknell D, Bodmer WF, Macaulay VM. Insulin-like growth factor 1 regulates the location, stability, and transcriptional activity of beta-catenin. Proc Natl Acad Sci USA. 2000;97:12103-8.
- 25. Zhang D, Samani AA, Brodt P. The role of the IGF-I receptor in the regulation of matrix metalloproteinases, tumor invasion and metastasis. Horm Metab Res. 2003;35:802-8.
- Zhang X, Horwitz GA, Heaney AP, Nakashima M, Prezant TR, Bronstein MD, et al. Pituitary tumor transforming gene (PTTG) expression in pituitary adenomas. J Clin Endocrinol Metab. 1999;84:761–7.
- McCabe CJ, Khaira JS, Boelaert K, Heaney AP, Tannahill LA, Hussain S, et al. Expression of pituitary tumour transforming gene (PTTG) and fibroblast growth factor-2 (FGF-2) in human pituitary adenomas: relationships to clinical tumour behaviour. Clin Endocrinol (Oxf). 2003;58:141–50.

- 28. Wierinckx A, Auger C, Devauchelle P, Reynaud A, Chevallier P, Jan M, et al. A diagnostic marker set for invasion, proliferation, and aggressiveness of prolactin pituitary tumors. Endocr Relat Cancer. 2007;14:887–900.
- 29. Filippella M, Galland F, Kujas M, Young J, Faggiano A, Lombardi G, et al. Pituitary tumour transforming gene (PTTG) expression

correlates with the proliferative activity and recurrence status of pituitary adenomas: a clinical and immunohistochemical study. Clin Endocrinol (Oxf). 2006;65:536-43.

 Noh TW, Jeong HJ, Lee MK, Kim TS, Kim SH, Lee EJ. Predicting recurrence of nonfunctioning pituitary adenomas. J Clin Endocrinol Metab. 2009;94:4406–13.