



REVIEW ARTICLE

## Current state and perspectives of stem cell therapy for stroke



D.M. Martínez-Garza<sup>a</sup>, O.G. Cantú-Rodríguez<sup>a,\*</sup>, J.C. Jaime-Pérez<sup>a</sup>,  
C.H. Gutiérrez-Aguirre<sup>a</sup>, J.F. Góngora-Rivera<sup>b</sup>, D. Gómez-Almaguer<sup>a</sup>

<sup>a</sup> Hematology Service, Hospital Universitario "Dr. José Eleuterio González," Universidad Autónoma de Nuevo León, Monterrey, Mexico

<sup>b</sup> Neurology Service, Hospital Universitario "Dr. José Eleuterio González," Universidad Autónoma de Nuevo León, Monterrey, Mexico

Received 21 July 2016; accepted 21 July 2016

Available online 3 September 2016

### KEYWORDS

Stroke;  
Stem cell therapy;  
Mesenchymal stem cells;  
Neural stem cells;  
Hematopoietic stem cells;  
CD34

**Abstract** Stroke represents a public health enemy. Currently, and in spite of multiple clinical trials, thrombolysis remains as the only approved therapy. Most preclinical trials and animal trials employing stem cell-based therapies have shown very promising evidence of benefits. The aim of this review is to provide a landscape of what has been done in human clinical trials, and what are the possible ways that stem cell therapy may enhance functional recovery in stroke patients.

© 2016 Universidad Autónoma de Nuevo León. Published by Masson Doyma México S.A. This is an open access article under the CC BY-NC-ND license (<http://creativecommons.org/licenses/by-nc-nd/4.0/>).

### Introduction

Acute stroke is the sixth most common cause of death among Mexicans and the third cause in elderly people; just in 2014

it was responsible for 30,000 deaths.<sup>1</sup> Though these numbers may even be sub-registered.<sup>2</sup> More startling is the fact that stroke is the leading cause of disability in this country and worldwide.<sup>3</sup> This is because in spite of the high mortality, 75% of patients survive a stroke with some kind of sequel; this is 258 patients daily.<sup>4</sup>

The main mid-term and long-term sequels of stroke are dysphagia, fatigue, muscle weakness, paralysis, visual problems, incontinence, chronic pain, seizures, insomnia, depression, dementia, aphasia, amnesia among many others.<sup>5</sup>

The annual incidence of stroke is 232/100,000 people, with a prevalence of 8/1000.<sup>6</sup> With a population of 112

\* Corresponding author at: Hematology Service, Hospital Universitario "Dr. José Eleuterio González", Universidad Autónoma de Nuevo León (UANL), Ave. Madero y Gonzalitos s/n, Colonia Mitras Centro, Monterrey, Nuevo León C.P. 64460, Mexico. Tel.: +52 81 83487871; fax: +52 81 86756717.

E-mail address: [ogcantur@yahoo.com.mx](mailto:ogcantur@yahoo.com.mx) (O.G. Cantú-Rodríguez).

million Mexicans, this means almost 900,000 people survive with some type of disability or sequel. Worse, after a year almost half of them survive with significant sequels that affect their quality of life and their capacity to perform daily life activities (Modified Rankin Scale [mRS] between 2 and 5).<sup>7</sup>

As if that were not enough, the economic impact is notorious as well. Among hospital expenses, studies, treatments, doctors' fees, work incapacities, early retirement, rehabilitation programs and more, each patient surviving stroke yearly spends around \$25,000 dollars.<sup>4</sup> This is of course without taking into consideration the social and emotional expenses.

Stroke prevalence is on the rise worldwide,<sup>8</sup> and our country is not the exception.<sup>9</sup> In Mexico the median age of a first stroke event is of 68 years (ranges 52–84).<sup>7</sup> Epidemiology studies project that by 2050, the group conformed by people aging between 55 and 80 will almost triple, to 28%.<sup>10</sup> Therefore, by that year there will be more than 2 million stroke survivors; 1 in every 60 Mexicans.

Even though there have been many attempts to develop new stroke cures, a thrombolytic approach with tissue plasminogen activator for an acute ischemic stroke remains as the only approved therapy.<sup>11</sup> It is imperative to find or develop a new treatment to enhance recovery and restore, at least partially, the lost neurological functions.

## Physiopathology of stroke

### Etiology

Stroke can be divided into ischemic stroke, constituting 85% of cases (atherothrombotic, cardioembolic, small vessel disease), and hemorrhagic stroke (subarachnoid hemorrhage, intraparenchymal hemorrhage) which accounts for the remainder 15%.<sup>12</sup> The later one has an overall worse prognosis.<sup>13</sup> The primary risk factors for stroke include, but are not limited to, hypertension, diabetes mellitus, smoking, dyslipidemia and atrial fibrillation.<sup>14</sup>

Cerebral ischemia is graded depending on the cerebral blood flow (CBF; which would normally be of 50–55 ml/100 g/min). Anatomically, stroke lesions can be divided into an ischemic penumbra (CBF of 15 ml/100 g/ml), with functionally impaired neurons, though still reversible with acute stroke therapy, and an ischemic central core (CBF of 6 ml/100 g/ml) with irreversible neuronal death.<sup>15</sup>

### Natural history

Lack of energy substrates quickly leads to dysfunction of energy-dependant ion transport pumps and depolarization of neurons and glia.<sup>16</sup> This depolarization releases excitatory neurotransmitters, primary glutamate, which amplify the damage by the release of free radicals and interruption of the electron chain transport.<sup>17</sup> The following oxidative stress contributes to neuronal death by disruption of the cell membrane.<sup>18</sup> Apoptosis also mediates many of the lost neurons, predominantly in the penumbra region if no acute treatment is installed.<sup>19</sup>

Afterwards, astrocytes concentrate along ischemic lesions, and produce proteoglycans to form a glial scar,

which act as both a physical and biochemical barrier to axonal regeneration and sprouting, limiting the reconnection of neural circuits and contributing to many of the long-term sequels of stroke.<sup>20,21</sup>

Most important for the matter of this review it is the robust inflammatory reaction following cerebral ischemia. Inflammatory molecules (e.g. interleukin-1 [IL-1], interleukin-6 [IL-6] and tumoral necrotic factor- $\alpha$  [TNF- $\alpha$ ]) are predominantly deleterious in the early phase after an ischemic stroke<sup>22</sup> and paradoxically promote brain regeneration and neurovascular remodeling in the later or chronic phases.<sup>23</sup>

Finally, stroke releases many chemotactic molecules (e.g. interleukin 8 [IL-8], monocyte chemoattractant protein-1 [MCP-1]) both for leucocytes and stem cells.<sup>24</sup> Particularly stromal-derived factor 1a (SDF1a) is released by activated endothelial cells after hypoxic injuries, and its receptor (CXC chemokine receptor-4 [CXCR4]) is up-regulated as well.<sup>25</sup> They both acts as chemoattractants that mediate neural<sup>26</sup> and bone marrow<sup>27</sup> stem cell migration to injured areas, which is critical for stem cell-based therapies.

## Neurorestorative therapies

After neuroprotection (acute) therapies have failed, and scarring, inflammation and edema have installed, the approach must be shifted then to a neurorestorative therapy rather than on preventing the extension of a damage that has already been well established. This type of therapy focuses on orchestrating through all type of parenchymal cells (i.e. neuroblasts, immune cells, astrocytes, oligodendrocytes and neurons), the enhancement of endogenous neurogenesis, angiogenesis, axonal sprouting and synaptogenesis in affected brain tissue.<sup>28</sup>

Neurorestorative therapies include, but are not limited to, stem cells. There are also ongoing pharmacological investigations<sup>29</sup> and other type of treatments such as electromagnetic stimulation, device-based strategies, repetitive training and task-oriented strategies.<sup>30</sup> Rehabilitation could exploit the combination of functional reorganization and adaptation after stroke.<sup>31</sup> Of all, currently only constraint-induced therapy has evidenced some type of efficacy.<sup>32</sup>

## Stem cells

### Types of stem cells

Since their first descriptions in 1963 by Till and McCulloch,<sup>33</sup> stem cells have been used as an alternative to many diseases, especially those without any other treatment options. Neurologic conditions have not been the exception. In fact they represent the fifth most common cause for ongoing clinical trials based just on mesenchymal stem cell therapy (after bone, heart, gastrointestinal and autoimmune disorders).<sup>34</sup>

A stem cell is considered as such when two properties are met: capacity for long-term self-renewal without senescence and the ability to differentiate into one or more specialized cell types. Based on their transdifferentiation

spectrum stem cells are classified as *totipotent* if they can form any cell in the body (including placental cells), as *pluripotent* if they are able to divide into any cell of the three germ-lines of early embryogenesis, and as *multipotent* if they are already committed to only one germ-line (e.g. most bone marrow stem cells can only differentiate into mesoderm derived cells).<sup>35</sup>

Stem cells are also classified based on their origin as embryonic or adult type. Embryonic stem cells (ESC) are pluripotent cells that are obtained by manipulating embryos before implantation.<sup>36</sup> Adult (somatic) stem cells are multipotent (or pluripotent) cells obtained from mature differentiated tissues such as bone marrow, umbilical cord, human olfactory mucosa, fat tissue and brain.<sup>37</sup>

Finally, another type of stem cell is the induced pluripotent stem cells (iPSC). In this technique, differentiated mouse fibroblasts are reprogrammed to an embryonic-like state (pluripotency) by transfer of nuclear contents into oocytes or by fusion with embryonic stem cells.<sup>38</sup>

### Neural stem cell

Contrary to old assumptions, evidence of neurogenesis in the adult human brain has been demonstrated.<sup>39</sup> Neural stem cells (NSC) are a multipotent variant of stem cells present in the brain.<sup>40</sup> These cells are located in the sub-ventricular zone (SVZ) of the third ventricle<sup>41</sup> and in the subgranular zone (SGZ) of dentate gyrus,<sup>42</sup> and respond to brain insults that cause neuronal death such as stroke,<sup>43</sup> Huntington's disease,<sup>44</sup> and Alzheimer's disease.<sup>45</sup> NSC not only proliferate but also migrate to areas of lesion even in elderly patients.<sup>46</sup> NSC can be cultured *in vitro* for stem cell therapies, and even if administered intravenously, have the capacity to migrate into ischemic areas.<sup>47</sup>

It is been documented that after a stroke NSC expand and mature into well differentiated neurons and integrate functionally into neuronal circuits.<sup>48</sup> After a stroke, the brain's environment holds a rise in many growth factors that induce changes in NSC's mitotic cell cycle such as reduction of G1 phase<sup>49</sup> which boosts mitotic rate up to a 12-fold increase in number<sup>42</sup> as well as activation of phosphatidylinositol 3-kinases-Akt signaling pathway which enhances cell survival, proliferation, differentiation and migration.<sup>50,51</sup> Stroke also activates many genes involved in neurogenesis during embryonic development, especially those of transforming growth factor-beta [TGF- $\beta$ ] superfamily (bone morphogenic protein 8 [BMP2], bone morphogenetic protein type 1 receptors [BMPR1] and growth differentiation factor 2 [GDF2]).<sup>52</sup> These newly formed neurons differentiate into the phenotype of most of the neurons that were lost during ischemia, in an attempt to regenerate lost circuits and recover lost functions.<sup>46</sup>

Discouragingly, one of the setbacks is their slim capacity to migrate into areas of the cortex where higher mental functions lie.<sup>46</sup> What is more, after a couple of weeks 80% of these newly formed neurons die and actually just 0.2% of dead tissue is replaced.<sup>46</sup> We hypothesize that if the percentage of incorporated renewed cells could be increased somehow, (e.g. neurotrophic, or angiogenic factors) restoration of neurological functions would be much greater as well.

### Bone marrow stem cells

Bone marrow stem cells (BMSC) are an array of different type of multipotent and pluripotent cells homed in the spongy tissue of almost all bones. Two basic lineages prevail: hematopoietic stem cells (HSC, PBSC if obtained peripherally) and mesenchymal stem cells (MSC). HCS give rise to all the type of blood cells and are typically CD34+, CD133+ and negative for all markers of differentiation or further lineage commitment (CD13-, CD71-, CD19-, CD61-).<sup>53</sup> MSC lie on the stroma of the bone marrow, and contrary to HSC, they can differentiate into a broader variety of cell types, such as osteoblasts, chondrocytes, myocytes and adipocytes and even neurons.<sup>54</sup> MSC are usually CD34-.<sup>55</sup>

BMSC actually have limited cellular differentiation ability in comparison to other type of stem cells; evidence suggests instead that the beneficial properties are due to immunomodulatory mechanisms, as they migrate to sites of inflammation (by the mechanisms explained before)<sup>56</sup> and secrete many bioactive molecules.<sup>57</sup> This is supported by the fact that PBSC are also used with efficacy in the autologous therapy of non-hematopoietic tissues like neurons,<sup>58</sup> skeletal muscle<sup>59</sup> and heart.<sup>60</sup> In multiple sclerosis and amniotrophic lateral sclerosis for instance, immunomodulatory effects and improvements were observed just 24h after intrathecal delivery of MSC, which would be an irrational time frame for differentiation and rather backs up the hypothesis of a bystander effect instead.<sup>61</sup> Furthermore, six months later, evidence of integration or even survival of these cells was very poor.<sup>62</sup> In an animal model, CD34+ cells (HSC) were tracked by magnetic resonance, where they prove they migrate to lesion sites but just persisted for about 3 to 4 weeks.<sup>63</sup>

Even though there is a very low rate of transdifferentiation into neurons, there is still clinical recovery, motor evoked potential improvements, as well as reconstruction of the ischemic tissue.<sup>64</sup> As stated before, the benefits of BMSC would be by enhancing endogenous neurogenesis rather than cellular lineage reprogramming. The mechanisms involved appear to be paracrine secretion of bioactive molecules and upgrade regulation of receptors that reinforce and augment the natural recovery processes implemented by the brain; subsequently increasing the number of new functional neurons derived from endogenous neuroblasts.

It has been proved that exogenous administration of brain-derived neurotrophic factor (BDNF) stimulates neurogenesis,<sup>65</sup> therefore, endogenous secretion of BDNF and similar trophic factors by stem cells would aid in such purposes. BMSC increases concentration of SDF1a as well as expression of the SDF-1 receptor, CXCR4 in the perischemic area.<sup>66</sup> There is also promotion of basic fibroblast growth factor (bFGF) and other trophic factor like  $\beta$ -nerve growth factor ( $\beta$ -NGF)<sup>67</sup> which would not only promote proliferation, but will reduce apoptosis as well.<sup>68</sup>

BMSC increase the number of oligodendrocyte progenitors and increase axonal density around the ischemic lesion, extending and orienting axons parallel to the boundary of the penumbra.<sup>69</sup> They do this by reducing expression of axonal growth inhibitory proteins, such as reticulon and neurocan, enabling axonal and neurite outgrowth.<sup>70</sup>

MSC also share the properties of secreting many trophic factors (BDNF, SDF-1, NGF, bFGF, and VEGF) and

promoting neurogenesis<sup>71</sup> with the added benefit of a greater potential than regular HSC to transdifferentiate into neurons themselves.<sup>72,73</sup> MSCs carry the benefit of being readily obtained from bone marrow and easily expanded by culture in vitro, though this involves a time frame of 4 to 5 weeks before being delivered back to patients.<sup>74</sup>

MSC are pretty safe. Because of their low major histocompatibility complex proteins they are considered immune privileged and cause no immunogenicity, neither acute or chronic.<sup>75</sup> In a recent meta-analysis, there was no association between MSC and neoplastic potential, infection, embolism or zoonosis; in fact the only side effect was transient low-grade fever (OR: 16.82).<sup>76</sup>

Angiogenesis also plays a critical role in functional recovery. As in neurogenesis, angiogenesis is induced by several growth factors present in the penumbra 3 to 4 days after a stroke.<sup>77</sup> It is so relevant, that patients who have a high density of blood vessels after stroke survive longer than those who do not.<sup>78</sup> Animal models with denser vascularization have a better functional outcome as well.<sup>79</sup> This density is determined by the presence of vascular growth factors, for there is a correlation between greater concentration gradients of them and increased blood vessel neoformation.<sup>80</sup> Interestingly, neurogenesis actually enhances symbiotically angiogenesis by secreting the same factors.<sup>81,82</sup> Given that BMSC up-regulate expression and paracrine secretion of angiogenic growth factors such as vascular endothelial growth factor (VEGF) and its receptor (VEGFR2) as well as angiopoietins 1 and 2 and their receptor (TIE-1 and TIE-2),<sup>83</sup> it is hypothesized that they would magnify the beneficial properties of neurogenesis and angiogenesis along with improving clinical outcomes and survival rate.<sup>84</sup>

Deciding which is better among HSC, PBSC or MSC is still unachieved. The only human clinical trial comparing HSC and expanded MSC found out that patients had better clinical outcomes (Barthel Index [BI]) with HSC.<sup>85</sup>

## Other type of stem cells

Besides BMSC and NSC, other type of stem cells may be used for stroke and other diseases. Pluripotent stem cells (e.g. ESC), with a wider transdifferentiation spectrum, have the theoretical advantage over multipotent cells in its use for regenerative medicine; however, the downside is the accompanying increased risk of developing malignancies as well.<sup>86</sup> In addition to this ESC bear ethical, technical and legal issues regarding the use of human embryos.<sup>87</sup>

As promising as they might be, iPSC have not been approved yet to be used in any clinical trial involving humans as concerns involving tumorigenicity abound.<sup>88</sup> iPSC display more genetic and epigenetic abnormalities than other type of stem cells.<sup>89</sup> Actually, their capability of developing pluripotent malignancies, such as teratoma surpasses that of ESC.<sup>90</sup>

## Stem cell therapy for stroke

### Design of clinical trial

The first attempt to treat stroke using stem cells was more than 15 years ago.<sup>91</sup> Hitherto there is still no optimum model

for a clinical trial. With stroke being so diverse and many aspects of stem cell therapy still unexplored, many variables have to be thrown into the equation. We will discuss each of these variables separately.

### Selection of patient

Stroke lesion sizes and locations are broadly heterogeneous in addition to normal neuroanatomical variations among individuals. Besides, depending on the etiology, stroke outcomes and prognosis vary hugely as well.<sup>9</sup> Therefore, stroke outcomes are extraordinarily diverse among patients; even without intervention, most patients exhibit limited spontaneous recovery, though a subgroup will remain severely impaired.<sup>92</sup> On the other hand, even with effective thrombolysis most patients will still have neurological deficits.<sup>93</sup>

So without a control or a placebo group, it is difficult to distinguish whether improvements are stem cell therapy-based or just the natural history of the disease. Ideally, and specially to address efficacy, inclusion criteria should be the most homogeneously possible (in age, etiology and comorbidities) to avoid confounding biases, even if this comes at the cost of shortening the number of patients.<sup>94</sup>

Selecting patients with little to no predicted natural recovery may highlight the benefits of cell therapy, though this represents an obstacle given that most patients do not exhibit explicit recovery until 3–6 months after stroke, a time frame which limit most of the clinical trials that advocate for administration of stem cells much earlier.<sup>95</sup> The expected recovery can be anticipated early (within days after stroke) by the use of specialized techniques of neuroimaging (e.g. fiber numbers asymmetry)<sup>96</sup> and neurophysiological assessments (e.g. motor-evoked potentials),<sup>97</sup> which would help us select patients with the worst prognoses to treat them in acute phases; though these are not yet used routinely.<sup>28</sup>

Double blinding enhances statistical power to the clinical trial, but this may not be fitting for the more invasive interventions, such as intrathecal or intracerebral approaches.

### Dosage

A consensus regarding dosage has not been met. Nonetheless, there is clear relation between more cells administered and better outcomes.<sup>67,98</sup> Therefore, given the safety profile of autologous stem cells, efforts to recollect the highest number cells possible must be done. This of course would not apply for allogenic stem cells, where the risk of graft versus host disease and rejection are much greater with higher doses.<sup>99</sup>

Another matter regarding dosage concerns the use of granulocyte colony stimulating factor (GCS-F) either as an attempt to increase number of available stem cells for collection, or even as a mean of treatment itself. GCS-F, as an up-regulator of hematopoiesis has demonstrated to increase exponentially the number of PBSC and could theoretically work as if these have been exogenously administered (i.e. migrate to penumbra and enhance recovery).<sup>100</sup> Safety of GCS-F has been established in hyperacute stages of stroke (24–48 h after onset),<sup>101</sup> which would carry an enormous advantage over stem cells, given that these would be

difficult to have at hand that much early, especially in unstable patients. Although the trend is toward better outcomes,<sup>102</sup> efficacy of GCS-F has not been thoroughly proven and is yet to be determined if they could be used as an alternative therapy alone or even as a coadjuvant of stem cell therapy.

### Time of intervention

If any natural recovery is going to happen is not seen until around 3–6 months after the onset of stroke,<sup>103</sup> and waiting that much could limit many of the immunomodulatory effects of stem cells. Besides most data points toward better functional outcomes if stem cells are administered much earlier.<sup>104</sup> In one animal model, only the rats receiving BMSC intravenously 7 days after artery occlusion exhibited decreased ischemic lesion volume in contrast to those who received them at days 14 and 28. Interestingly though, all three groups displayed clinically significant better functional outcomes compared to the placebo group, suggesting that 1 month after stroke might be a suitable time-frame.<sup>66,105</sup>

Nevertheless, patients with much more chronic stroke still exhibit improvements in neurological functions.<sup>106</sup> Considering the importance of inflammatory chemotaxis for stem cell therapy, this hints to the hypothesis that stroke constitutes a state of very chronic state of inflammation. Therefore, independent of the time of administration, stem cells will always migrate to some degree to the areas of lesion or even to the scarring tissue. Evidence suggests that stem cells in early stages work as anti-inflammatory molecules and in chronic stages aid in endogenous recovery and neurorestoration.<sup>23</sup>

Administration time should also be decided depending on the feasibility of the route of administration as patients with hyperacute stroke (<72 h) are usually neurologically unstable and cannot tolerate invasive procedures.<sup>107</sup>

### Route of administration

The American Stroke Association in its recommendation for future stem cell research states that the safest and most effective route of cell delivery should be defined using preclinical trials.<sup>95</sup> There are four major possible routes: intravenous (IV), intra-arterial (IA), intrathecal (IT) and intracerebral (IC).<sup>108,109</sup> (See Fig. 1)

It is clear due to the many clinical trials, that the IV route is the safest and most feasible for administering stem cells.<sup>74,85,110,111</sup> Unfortunately, the most effective route is yet to be determined.

The basis for peripheral and non-invasive approaches (IV and IA) is the phenomenon of the selective permeability of the blood-brain barrier (BBB). After a brain insult, specially a hypoxic one, the tight junctions between the endothelial cells of the capillaries loosen up, therefore increasing its permeability and allowing income of many molecules including inflammatory cells.<sup>112</sup> This breakdown may even persist for weeks or even months after the original insult, justifying stem cell therapies in chronic stages.<sup>113</sup> Despite this, it remains unclear whether systemically infused stem cells are able to cross in a significant extent the BBB under both normal and pathological conditions.<sup>114</sup>

IA administration of stem cells theoretically increments the number of cells delivered to the lesion area in comparison to IV; yet stem cells through these route, especially in high doses, may cause recurrent stroke.<sup>115</sup> On the other hand, another study that displayed safety of IA infusion in a window time of 3–7 days, though this was just made in 4 patients.<sup>116</sup> Even though considered not as an invasive approach as others, safety of IA route must be reevaluated in larger clinical trials.

Another major pitfall of peripheral routes is the substantial loss of cells in other parts of the body.<sup>117</sup> When injected IV, stem cells are distributed all around the body and are homed in other organs like the liver, kidney, lungs and spleen.<sup>118</sup> Nonetheless, perhaps it isn't strictly necessary for the cells to be homed in the penumbra area for them to perform their anti-inflammatory properties. Some types of stem cells may not even enter the central nervous system and may instead promote stroke recovery by acting on peripheral organs.<sup>28</sup> Whether this contributes significantly to clinical improvements remains an interrogatory.

In one animal model of human umbilical cord blood-derived MSC,<sup>119</sup> it was observed that there was a major migration and concentration of stem cells in the areas of hypoxia after IT administration compared to an IV approach, as well as a longer survival period of these giving a great advantage to this approach. Additionally, IT administration of stem cells for other neurological disorders has been found to be safe and well tolerated; with the main side effects being headache and transient low-grade fever.<sup>120–122</sup> Nevertheless, some serious adverse effects have been described as well. Case reports of inflammatory hypertrophic cauda equine,<sup>123</sup> demyelinating encephalomyelitis,<sup>124</sup> and spinal myoclonus<sup>125</sup> following intrathecal injection of stem cells (combination of ESC, MSC and HSC) for different diseases have been published. However, these interventions were performed in stem cell therapy clinics, with no more information given.

When implanted IC via stereotaxis, grafted MSC cells are visualized prominently just 24 h after implantation and are homed almost exclusively in the affected site, positioning this route as the most effective in terms of cell concentration. Though as with animal models, evidence of these cells by neuroimaging (hypointensity on T2) became smaller gradually in the following 4 weeks until finally disappearing.<sup>64</sup> One must weight the risk and benefits when considering this invasive approach, particularly when using BMSC where benefits may just be temporal. Stereotaxis may be more justifiable if using exogenous NSC where there is a reasonable expectation of functional engraftment and a permanent incorporation to neural circuits.<sup>126</sup> One must take into account that IC administration is unfeasible in acute and in unstable patients.

### Randomized clinical trials

To our knowledge, there are only 7 randomized clinical trials published that have used stem cell therapy for the treatment of acute stroke (Table 1).

The first ever randomized clinical trial<sup>74</sup> used ex vivo cultured autologous MSC and then infused them IV twice (weeks 4 and 8). All patients from the MSC group ( $n = 5$ ) had

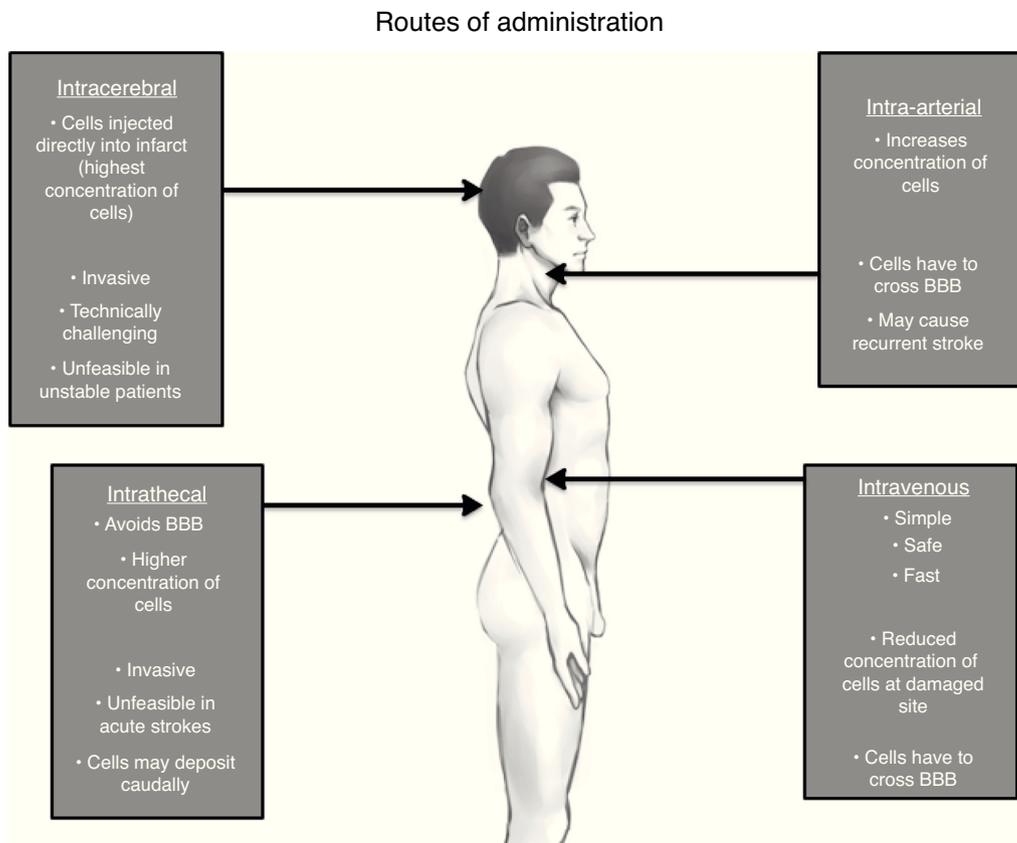
**Table 1** Randomized clinical trials.

	Title	Country	Year	Type of stroke	GCS-F	Type of Cell	Route of administration	<i>N (cases)</i>	Results
1	Intravenous autologous bone marrow mononuclear stem cell therapy for ischemic stroke	India	2014	Acute (10 d to 28 d)	No	HSC ( $3 \times 10^6$ CD34+)	Intravenous	120 (60)	No changes
2	Intracerebral implantation of autologous peripheral blood stem cells in stroke patients: a randomized phase II study	Taiwan	2014	Chronic (6 m to 5 y)	Yes	PBSC ( $3-8 \times 10^6$ CD34+)	Intracerebral	30 (15)	Significant improvements
3	Intra-arterial bone marrow mononuclear cells in ischemic stroke: a pilot clinical trial	Spain	2012	Acute (5 d to 9 d)	No	HSC ( $1.5 \times 10^6$ CD34+)	Intra-arterial	20 (10)	Non-significant improvements
4	Stem cell therapy: a clinical trial of stroke	India	2012	Chronic (3 m to 24 m)	No <sup>a</sup>	HSC & MSC ( $5 \times 10^6$ ) <sup>b</sup>	Intravenous	40 (20)	Significant improvements
5	A long-term follow up study of intravenous autologous mesenchymal stem cell transplantation in patients with ischemic stroke	South Korea	2010	Acute (4 wk)	No <sup>a</sup>	MSC ( $1 \times 10^8$ )	Intravenous	52 (16)	Significant improvements
6	Autologous mesenchymal stem cell transplantation in stroke patients	South Korea	2005	Acute (4 wk to 5 wk)	No <sup>a</sup>	MSC ( $1 \times 10^8$ )	Intravenous	30 (5)	Significant improvements
7	Neurotransplantation for patients with subcortical motor stroke: a phase 2 randomized trial	USA	2005	Chronic (1 yr to 6 yr)	No	Human neuronal cells (LBS®) $5-10 \times 10^6$	Intracerebral	18 (4)	Non-significant improvements

**Abbreviations:** GCS-F, Granulocyte colony stimulating factor; d, days; wk, weeks; m, months; yr, years; HSC, Hematopoietic stem cells; PBSC, Peripheral blood stem cells, MSC, Mesenchymal stem cells, LBS, Layton BioScience.

<sup>a</sup> MSC were expanded in vitro.

<sup>b</sup> For both CD34+ & MSC.



**Figure 1** Possible routes of administration of stem cells for stroke.

cerebral infarcts that involved the middle cerebral artery territory. They reported significant improvements in BI (3 and 6 months) and mRS (3 months). One year outcomes were not significant. The main limitations of this study are the small treatment group and a short follow-up period. A similar approach was used by the same authors but with a larger population (MSC group  $n = 16$  and control group  $n = 36$ ) and a longer follow-up (5 years).<sup>110</sup> Significant improvement of the mRS were reported in the MSC group ( $p = 0.046$ ), in contrast with the control group ( $p = 0.257$ ). The mortality rate in the MSC group was lower than in the control group (Log rank:  $p = 0.059$ ), and there was no difference in comorbidities during the follow-up period. Notably, there was also a correlation between higher levels of the SDF-1 $\alpha$  and clinical improvements, emphasizing the crucial role played by chemoattractants in this type of therapies.

An IC approach has been used twice. Kondziolka et al. implanted stereotactically 5 or 10 million allogenic neuronal cells cultivated from human embryonic carcinoma-derived cells (LBS<sup>®</sup>) to 14 patients with chronic stroke.<sup>104</sup> They demonstrated safety of the procedure, as no serious adverse effects occurred after a 5 year follow-up. Non-significant improvements were found, especially in those having an ischemic stroke. Regarding neuropsychological testing, marked improvement was seen,<sup>127</sup> as well as improved F-fluorodeoxyglucose (FDG) uptake in hypoxic areas. The other clinical trial also used stereotaxis in 15 chronic stroke patients, implanting 3–8 million CD34+ cells after stimulated (with G-CSF) PBSC where recollected by apheresis.<sup>64</sup>

The treated group showed a significant improvement in the National Institute of Health Stroke Scale (NIHSS), European stroke scale (ESS) and ESS motor subscale (EMS). Further, there were reductions in fiber number asymmetry of the damaged corticospinal tracts as well as restoration of motor evoked potentials response, both correlating with better functional outcomes. These changes were not observed in the control group. Safety end-points were acknowledged.

The only IA clinical trial conducted showed functional improvements, though these were not significant.<sup>67</sup> Ten patients were injected with  $1.5 \times 10^8$  autologous HSC between 5 and 9 days after ischemic stroke. No serious adverse events, stroke recurrence (clinical or by image), nor tumor formation were observed during the follow-up period (6 months). Interestingly, there was a trend toward better clinical outcome when higher numbers of CD34+ cells were injected.

The most recent, and by far, the larger clinical trial performed (60 cases and 60 controls) infused intravenously  $2.9 \times 10^6$  CD34+ obtained by HSC in subacute stroke patients (median of 18.5 days after stroke).<sup>111</sup> Even though safety was met, no changes were observed as to functional improvements. This contrasts with results obtained by the same research group a few years back, where they used either HSC or MSC in chronic patients ( $n$  treated = 20), and found statically significant improvements in BI, as well as increased number of cluster activation in motor cortex area, suggesting neuroplasticity.<sup>85</sup>

## Follow-up

A biological marker that can assess selectively improvements of neurological functions after an ischemic or hemorrhagic event is in great need,<sup>94</sup> but that as it may, there is currently no validated marker for such purposes.<sup>128</sup> Therefore, we must rely on other tools such as clinical scales and neuroimaging.

On this matter, more extensive, objective and specific neurological outcomes that measure beyond the classical NIHSS, BI, mRS, ESS, or EMS need to be developed and implemented for restorative treatments.<sup>129</sup>

Magnetic resonance imaging (MRI) is an invaluable resource to gauge more objectively improvements after stem cell therapy. The focus should not be on the reduction of stroke volume size or edema, as these do not translate directly into better functional outcomes, and should remain then as secondary endpoints.<sup>28</sup> Rather, restructuring of white matter tracts, neurogenesis and angiogenesis can be better used as to monitor recovery, which can be evaluated through more sensitive MRI techniques such as anisotropy<sup>130,131</sup> and magnetically labeled cells.<sup>132</sup>

## Future & perspective

The feasibility and safety of stem cells in stroke patients have both been roundly confirmed. But in spite of the remarkable improvements observed in animal models, translation to clinical scenarios has not been achieved so far. The overall results of stem cell therapy for stroke have been inconclusive, at best. Yet, the tendency seems to lean toward better functional outcomes. Many unsolved issues remain regarding timing, dosage, type of cell, and route of administration. And until these are not addressed, conclusions concerning efficacy should not be given at all. Therefore, larger double-blind randomized clinical trials with homogenous selection criteria and domain-specific end points are strongly encouraged to clarify this matter. Certainly, a predictive marker of which patients would benefit the most from cell therapy would be of immense aid.

Given the magnitude of the physical, emotional and economic burden that stroke survivors have to endure, and its colossal impact on society as a whole, efforts to find the appropriate stem cell therapy for neurorestoration should not surcease but be encouraged.

## Funding

No financial support was provided.

## Conflict of interest

The authors have no conflict of interest to declare.

## Disclosure statement

The authors have nothing to disclose.

## References

1. Principales causas de mortalidad por residencia habitual, grupos de edad y sexo del fallecido. Instituto Nacional de Estadística y Geografía; 2014. <http://www.inegi.org.mx/est/contenidos/proyectos/registros/vitales/mortalidad/tabulados/ConsultaMortalidad.asp>
2. Góngora-Rivera F. Perspective on stroke in Mexico. *Med Univ.* 2016;17:184–7, <http://dx.doi.org/10.1016/j.rmu.2015.04.001>.
3. Norrving B. The global burden of stroke and need for a continuum of care. *Neurology.* 2013;80 Suppl 2:5–12, <http://dx.doi.org/10.1212/WNL.0b013e3182762397>.
4. Ruiz JL. Costos de la enfermedad vascular cerebral en México. *AMEVASC;* 2014. <http://amevasc.mx/costo-de-la-enfermedad-vascular-cerebral-en-mexico/>
5. Teasell R. Long-term sequelae of stroke. *Can Fam Phys.* 1992;38:381–8.
6. Cantú-Brito C, Majersik JJ, Sánchez BN, et al. Door-to-door capture of incident and prevalent stroke cases in Durango, Mexico: the Brain Attack Surveillance in Durango Study. *Stroke.* 2011;42:601–6, <http://dx.doi.org/10.1161/STROKEAHA.110.592592>.
7. Cantú-Brito C, Ruiz-Sandoval JL, Murillo-Bonilla LM, et al. Acute care and one-year outcome of Mexican patients with first-ever acute ischemic stroke: the PREMIER study. *Rev Neurol.* 2010;51:641–9.
8. Donnan GA, Fisher M, Macleod M, et al. *Stroke Lancet.* 2008;371:1612–23, [http://dx.doi.org/10.1016/S0140-6736\(08\)60694-7](http://dx.doi.org/10.1016/S0140-6736(08)60694-7).
9. Marquez-Romero JM, Arauz A, Góngora-Rivera F, et al. The burden of stroke in México. *Int J Stroke.* 2015;10:251–2, <http://dx.doi.org/10.1111/ijs.12189>.
10. Proyecciones de la población 2010–2050. Consejo Nacional de la Población; 2010. <http://www.conapo.gob.mx/es/CONAPO/Proyecciones>
11. Cheng YD, Al-Khoury L, Zivin JA. Neuroprotection for ischemic stroke: two decades of success and failure. *NeuroRx.* 2004;1:36–45, <http://dx.doi.org/10.1602/neurorx.1.1.36>.
12. Morales-Vidal S. Stroke pathophysiology. *Futur Med.* 2001:6–20, <http://dx.doi.org/10.2217/EBO.12.443>.
13. Ruiz-Sandoval JL, Cantú C, Chiquete E, et al. Aneurysmal subarachnoid hemorrhage in a Mexican multicenter registry of cerebrovascular disease: the RENAMEVASC Study. *J Stroke Cerebrovasc Dis.* 2009;18:48–55, <http://dx.doi.org/10.1016/j.jstrokecerebrovasdis.2008.09.019>.
14. Nayak SD, Sarma PS, Radhakrishnan K. Incidence, types, risk factors, and outcome of stroke in a developing country. *Stroke.* 2009;40:1212–9, <http://dx.doi.org/10.1161/STROKEAHA.108.531293>.
15. Yoo AJ, Copen WA, Gonza RG. Cerebral blood flow thresholds for tissue infarction in patients with acute ischemic stroke treated with intra-arterial revascularization therapy depend on timing of reperfusion. *Am J Neuroradiol.* 2011;32:846–51, <http://dx.doi.org/10.3174/ajnr.A2415>.
16. Lloyd HGE, National A. The early events of oxygen and glucose deprivation: setting the scene for neuronal death? *Trends Neurosci.* 1994;17:251–7.
17. Mehta SL, Manhas N, Raghubir R. Molecular targets in cerebral ischemia for developing novel therapeutics. *Brain Res.* 2007;54:34–66, <http://dx.doi.org/10.1016/j.brainresrev.2006.11.003>.
18. Koehler C. Oxygen radical mechanisms of brain injury following ischemia and reperfusion. *J Appl Physiol.* 1991:1185–95.
19. Broughton BRS, Reutens DC, Sobey CG. Apoptotic mechanisms after cerebral ischemia. *Stroke.* 2009;40:331–9, <http://dx.doi.org/10.1161/STROKEAHA.108.531632>.

20. Carmichael ST. Cellular and molecular mechanisms of neural repair after stroke: making waves. *Ann Neurol*. 2006;59:735–42, <http://dx.doi.org/10.1002/ana.20845>.
21. Stoll G, Jander S, Schroeter M. Inflammation and glial responses in ischemic brain lesions. *Prog Neurobiol*. 1998;56:149–71, [http://dx.doi.org/10.1016/S0301-0082\(98\)00034-3](http://dx.doi.org/10.1016/S0301-0082(98)00034-3).
22. Jin R, Yang G, Li G. Inflammatory mechanisms in ischemic stroke: role of inflammatory cells. *J Leukoc Biol*. 2010;87(May):779–89, <http://dx.doi.org/10.1189/jlb.1109766>.
23. Amantea D, Nappi G, Bernardi G, et al. Post-ischemic brain damage: pathophysiology and role of inflammatory mediators. *FEBS J*. 2009;276:13–26, <http://dx.doi.org/10.1111/j.1742-4658.2008.06766.x>.
24. Minami M, Katayama T, Satoh M. Brain cytokines and chemokines: roles in ischemic injury and pain. *J Pharmacol Sci*. 2006;100:461–70, <http://dx.doi.org/10.1254/jphs.CRJ06005X>.
25. Ohab JJ, Fleming S, Blesch A, et al. A neurovascular niche for neurogenesis after stroke. *J Neurosci*. 2006;26:13007–16, <http://dx.doi.org/10.1523/JNEUROSCI.4323-06.2006>.
26. Bajetto A, Bonavia R, Barbero S, et al. Chemokines and their receptors in the central nervous system. *J Neurochem*. 2001;22:1311–29, <http://dx.doi.org/10.1006/frne.2001.0214>.
27. Wang Y, Deng Y, Zhou GQ. SDF-1a/CXCR4-mediated migration of systemically transplanted bone marrow stromal cells towards ischemic brain lesion in a rat model. *Brain Res*. 2008;1195:104–12, <http://dx.doi.org/10.1016/j.brainres.2007.11.068>.
28. Zhang ZG, Chopp M. Neurorestorative therapies for stroke: underlying mechanisms and translation to the clinic. *Lancet Neurol*. 2009;8:491–500, [http://dx.doi.org/10.1016/S1474-4422\(09\)70061-4](http://dx.doi.org/10.1016/S1474-4422(09)70061-4).
29. Zhang R, Wang Y, Zhang L, et al. Sildenafil (Viagra) induces neurogenesis and promotes functional recovery after stroke in rats. *Stroke*. 2002;33:2675–80, <http://dx.doi.org/10.1161/01.STR.0000034399.95249.59>.
30. Cramer SC. Repairing the human brain after stroke. II. Restorative therapies. *Ann Neurol*. 2008;63:549–60, <http://dx.doi.org/10.1002/ana.21412>.
31. Langhorne P, Bernhardt J, Kwakkel G, et al. Stroke rehabilitation. *Lancet*. 2011;377:1693–702, [http://dx.doi.org/10.1016/S0140-6736\(11\)60325-5](http://dx.doi.org/10.1016/S0140-6736(11)60325-5).
32. Wolf SL, Winstein CJ, Miller JP, et al. Effect of constraint-induced movement therapy on upper extremity function 3 to 9 months after stroke: the EXCITE randomized clinical trial. *JAMA*. 2015;296:2095–104, <http://dx.doi.org/10.1001/jama.296.17.2095>.
33. Becker AJ, McCulloch EA, Till JE. Cytological demonstration of the clonal nature of spleen colonies derived from transplanted mouse marrow cells. *Nature*. 1963;197:452–4, <http://dx.doi.org/10.1038/197452a0>.
34. Trounson A, Thakur RG, Lomax G, et al. Clinical trials for stem cell therapies. *BMC Med*. 2011;9:52, <http://dx.doi.org/10.1186/1741-7015-9-52>.
35. Rippon H, Bishop A. Embryonic stem cells. *Cell Prolif*. 2004;37:23–34.
36. Thomson JA, Itskovitz-eldor J, Shapiro SS, et al. Embryonic stem cell lines derived from human blastocysts. *Science*. 1998;282:1145–8.
37. Young HE, Black ASAC. Adult stem cells. *Anat Rec Part A*. 2004;276A:75–102, <http://dx.doi.org/10.1002/ar.a.10134>.
38. Takahashi K, Yamanaka S. Induction of pluripotent stem cells from mouse embryonic and adult fibroblast cultures by defined factors. *Cell*. 2006;126:663–76.
39. Eriksson PS, Perfilieva E, Björk-Eriksson T, et al. Neurogenesis in the adult human hippocampus. *Nat Med*. 1998;4:1313–7, <http://dx.doi.org/10.1038/3305>.
40. Gage FH. Mammalian neural stem cells. *Science*. 2000;287:1433–8, <http://dx.doi.org/10.1126/science.287.5457.1433>.
41. Zhang RL, Zhang ZG, Zhang L, et al. Proliferation and differentiation of progenitor cells in the cortex and the subventricular zone in the adult rat after focal cerebral ischemia. *Neuroscience*. 2001;105:33–41, [http://dx.doi.org/10.1016/S0306-4522\(01\)00117-8](http://dx.doi.org/10.1016/S0306-4522(01)00117-8).
42. Liu J, Solway K, Messing RO, et al. Increased neurogenesis in the dentate gyrus after transient global ischemia in gerbils. *J Neurosci*. 1998;18:7768–78.
43. Parent JM, Vexler ZS, Gong C, et al. Rat forebrain neurogenesis and striatal neuron replacement after focal stroke. *Ann Neurol*. 2002;52:802–13, <http://dx.doi.org/10.1002/ana.10393>.
44. Curtis MA, Penney EB, Pearson AG, et al. Increased cell proliferation and neurogenesis in the adult human Huntington's disease brain. *Proc Natl Acad Sci U S A*. 2003;100:9023–7, <http://dx.doi.org/10.1073/pnas.1532244100>.
45. Jin K, Peel AL, Mao XO, et al. Increased hippocampal neurogenesis in Alzheimer's disease. *Proc Natl Acad Sci U S A*. 2004;101:343–7, <http://dx.doi.org/10.1073/pnas.2634794100>.
46. Arvidsson A, Collin T, Kirik D, et al. Neuronal replacement from endogenous precursors in the adult brain after stroke. *Nat Med*. 2002;8:963–70, <http://dx.doi.org/10.1038/nm747>.
47. Iskander A, Knight RA, Zhang ZG, et al. Intravenous administration of human umbilical cord blood-derived AC133+ endothelial progenitor cells in rat stroke model reduces infarct volume: magnetic resonance imaging and histological findings. *Stem Cells Transl Med*. 2013;70:3–714, <http://dx.doi.org/10.5966/sctm.2013-0066>.
48. Hou SW, Wang YQ, Xu M, et al. Functional integration of newly generated neurons into striatum after cerebral ischemia in the adult rat brain. *Stroke*. 2008;39:2837–44, <http://dx.doi.org/10.1161/STROKEAHA.107.510982>.
49. Zhang RL, Zhang ZG, Roberts C, et al. Lengthening the G(1) phase of neural progenitor cells is concurrent with an increase of symmetric neuron generating division after stroke. *J Cereb Blood Flow Metab*. 2008;28:602–11, <http://dx.doi.org/10.1038/sj.jcbfm.9600556>.
50. Vojtek AB, Taylor J, DeRuiter SL, et al. Akt regulates basic helix-loop-helix transcription factor-coactivator complex formation and activity during neuronal differentiation. *Mol Cell Biol*. 2003;23:4417–27, <http://dx.doi.org/10.1128/MCB.23.13.4417>.
51. Katakowski M, Zhang ZG, Chen J, et al. Phosphoinositide 3-kinase promotes adult subventricular neuroblast migration after stroke. *J Neurosci Res*. 2003;74:494–501, <http://dx.doi.org/10.1002/jnr.10775>.
52. Liu XS, Zhang ZG, Zhang RL, et al. Stroke induces gene profile changes associated with neurogenesis and angiogenesis in adult subventricular zone progenitor cells. *J Cereb Blood Flow Metab*. 2007;27:564–74, <http://dx.doi.org/10.1038/sj.jcbfm.9600371>.
53. Birbrair A, Frenette PS. Niche heterogeneity in the bone marrow. *Ann NY Acad Sci*. 2016;1370:82–96, <http://dx.doi.org/10.1111/nyas.13016>.
54. Cogle CR, Yachnis AT, Laywell ED, et al. Bone marrow transdifferentiation in brain after transplantation: a retrospective study. *Lancet*. 2004;363:1432–7, [http://dx.doi.org/10.1016/S0140-6736\(04\)16102-3](http://dx.doi.org/10.1016/S0140-6736(04)16102-3).
55. Dominici M, Blanc K, Le Mueller I, et al. Minimal criteria for defining multipotent mesenchymal stromal cells. *Int Soc Cell Therapy Position Statement*. 2006;8:315–7, <http://dx.doi.org/10.1080/14653240600855905>.

56. Li Y, Chen J, Chen XG, et al. Human marrow stromal cell therapy for stroke in rat: neurotrophins and functional recovery. *Neurology*. 2002;59:514–23, <http://dx.doi.org/10.1212/WNL.59.4.514>.
57. Caplan A. Why are MSCs therapeutic? New data: new insight. *J Pathol*. 2009;217:318–24, <http://dx.doi.org/10.1002/path>.
58. Sigurjonsson OE, Perreault M-C, Egeland T, et al. Adult human hematopoietic stem cells produce neurons efficiently in the regenerating chicken embryo spinal cord. *Proc Natl Acad Sci U S A*. 2005;102:5227–32, <http://dx.doi.org/10.1073/pnas.0501029102>.
59. Doyonnas R, LaBarge MA, Sacco A, et al. Hematopoietic contribution to skeletal muscle regeneration by myelomonocytic precursors. *Proc Natl Acad Sci U S A*. 2004;101:13507–12, <http://dx.doi.org/10.1073/pnas.0405361101>.
60. Orlic D, Kajstura J, Chimenti S, et al. Mobilized bone marrow cells repair the infarcted heart, improving function and survival. *Proc Natl Acad Sci U S A*. 2001;98:10344–9, <http://dx.doi.org/10.1073/pnas.181177898>.
61. Martino G, Marco B, Peruzzotti-Jametti L. Therapeutic stem cell plasticity orchestrates tissue plasticity. *Brain*. 2011;134:1585–7, <http://dx.doi.org/10.1093/brain/awr115>.
62. Karussis D, Karageorgiou C, Vaknin-Dembinsky A, et al. Safety and immunological effects of mesenchymal stem cell transplantation in patients with multiple sclerosis and amyotrophic lateral sclerosis. *Arch Neurol*. 2010;67:1187–94, <http://dx.doi.org/10.1001/archneurol.2010.248>.
63. Jendelova P, Fales I, Andersson B, et al. Magnetic resonance tracking of human CD34+ progenitor cells separated by means of immunomagnetic selection and transplanted into injured rat brain. *Cell Transplant*. 2005;14:173–82.
64. Lin S-Z, Shyu W-C, Liu S-P, et al. Intracerebral implantation of autologous peripheral blood stem cells (CD34) in old ischemic stroke patients. *Cell Transplant*. 2014;23:1599–612, <http://dx.doi.org/10.3727/096368914X678562>.
65. Pencea V, Bingaman KD, Wiegand SJ, et al. Infusion of brain-derived neurotrophic factor into the lateral ventricle of the adult rat leads to new neurons in the parenchyma of the striatum, septum, thalamus, and hypothalamus. *J Neurosci*. 2001;21:6706–17.
66. Shen LH, Li Y, Chen J, et al. Therapeutic benefit of bone marrow stromal cells administered 1 month after stroke. *J Cereb Blood Flow Metab*. 2007;27:6–13.
67. Moniche F, Gonzalez A, Gonzalez-Marcos JR, et al. Intra-arterial bone marrow mononuclear cells in ischemic stroke: a pilot clinical trial. *Stroke*. 2012;43:2242–4, <http://dx.doi.org/10.1161/strokeaha.112.659409>.
68. Chen J, Li Y, Katakowski M, et al. Intravenous bone marrow stromal cell therapy reduces apoptosis and promotes endogenous cell proliferation after stroke in female rat. *J Neurosci Res*. 2003;73:778–86, <http://dx.doi.org/10.1002/jnr.10691>.
69. Li Y, Chen J, Zhang CL, et al. Gliosis and brain remodeling after treatment of stroke in rats with marrow stromal cells. *Glia*. 2005;49:407–17, <http://dx.doi.org/10.1002/glia.20126>.
70. Shen LH, Li Y, Gao Q, et al. Down-regulation of neurocan expression in reactive astrocytes promotes axonal regeneration and facilitates the neurorestorative effects of bone marrow stromal cells in the ischemic rat brain. *Glia*. 2008;56:1747–54, <http://dx.doi.org/10.1002/glia.20722>.
71. Pavlichenko N, Sokolova I, Vijde S, et al. Mesenchymal stem cells transplantation could be beneficial for treatment of experimental ischemic stroke in rats. *Brain Res*. 2008;1233:203–13, <http://dx.doi.org/10.1016/j.brainres.2008.06.123>.
72. Carter BD, Dobrowsky RT, Chao MV, et al. Marrow stromal cells as stem cells for nonhematopoietic tissues. *Science*. 1997;276(April):71–4.
73. Terada N, Hamazaki T, Oka M, et al. Bone marrow cells adopt the phenotype of other cells by spontaneous cell fusion. *Nature*. 2002;416:542–5, <http://dx.doi.org/10.1038/nature730>.
74. Bang OY, Lee JS, Lee PH, et al. Autologous mesenchymal stem cell transplantation in stroke patients. *Ann Neurol*. 2005;57:874–82, <http://dx.doi.org/10.1002/ana.20501>.
75. Le Blanc K, Tammik C, Rosendahl K, et al. HLA expression and immunologic properties of differentiated and undifferentiated mesenchymal stem cells. *Exp Hematol*. 2003;31:890–6.
76. Lalu MM, McIntyre L, Pugliese C, et al. Safety of cell therapy with mesenchymal stromal cells (SafeCell): a systematic review and meta-analysis of clinical trials. *PLoS ONE*. 2012;7, <http://dx.doi.org/10.1371/journal.pone.0047559>.
77. Krupinski J, Kaluza J, Kumar P, et al. Role of angiogenesis in patients with cerebral ischemic stroke. *Stroke*. 1994;25:1794–8, <http://dx.doi.org/10.1161/01.STR.25.9.1794>.
78. Krupinski J, Kaluza JPK, Wan M, et al. Prognostic value of blood vessel density in ischaemic stroke. *Lancet*. 1993;342:742.
79. Wang L, Zhang Z, Wang Y, et al. Treatment of stroke with erythropoietin enhances neurogenesis and angiogenesis and improves neurological function in rats. *Stroke*. 2004;35:1732–7, <http://dx.doi.org/10.1161/01.STR.0000132196.49028.a4>.
80. Zhang ZG, Zhang L, Tsang W, et al. Correlation of VEGF and angiopoietin expression with disruption of blood-brain barrier and angiogenesis after focal cerebral ischemia. *J Cereb Blood Flow Metab*. 2002;22:379–92, <http://dx.doi.org/10.1097/00004647-200204000-00002>.
81. Teng H, Zhang ZG, Wang L, et al. Coupling of angiogenesis and neurogenesis in cultured endothelial cells and neural progenitor cells after stroke. *J Cereb Blood Flow Metab*. 2008;28:764–71, <http://dx.doi.org/10.1038/sj.jcbfm.9600573>.
82. Wang L, Chopp M, Gregg SR, et al. Neural progenitor cells treated with EPO induce angiogenesis through the production of VEGF. *J Cereb Blood Flow Metab*. 2008;28:1361–8, <http://dx.doi.org/10.1038/jcbfm.2008.32>.
83. Chen J, Zhang ZG, Li Y, et al. Intravenous administration of human bone marrow boundary zone after stroke in rats. *Circ Res*. 2003;92:692–9, <http://dx.doi.org/10.1161/01.RES.0000063425.51108.8D>.
84. Chen J, Li Y, Zhang R, et al. Combination therapy of stroke in rats with a nitric oxide donor and human bone marrow stromal cells enhances angiogenesis and neurogenesis. *Brain Res*. 2004;1005:21–8, <http://dx.doi.org/10.1016/j.brainres.2003.11.080>.
85. Bhasin A, Srivastava MVP, Mohanty S, et al. Stem cell therapy: a clinical trial of stroke. *Clin Neurol Neurosurg*. 2013;115:1003–8, <http://dx.doi.org/10.1016/j.clineuro.2012.10.015>.
86. Knoepfler PS. Deconstructing stem cell tumorigenicity: a roadmap to safe regenerative medicine. *Stem Cells*. 2009;27:1050–6, <http://dx.doi.org/10.1002/stem.37>.
87. Baldwin T. Mortality and human embryo research. *EMBO Rep*. 2009;10:299–300, <http://dx.doi.org/10.1038/embor.2009.37>.
88. Okano H, Nakamura M, Yoshida K, et al. Steps toward safe cell therapy using induced pluripotent stem cells. *Circ Res*. 2013;112:523–33, <http://dx.doi.org/10.1161/CIRCRESAHA.111.256149>.
89. Fujita K. The dark side of induced pluripotency. *Nature*. 2011;471:46–7, <http://dx.doi.org/10.1029/1999GL006075>.
90. Gutierrez-Aranda I, Ramos-Mejía V, Bueno C, et al. Human induced pluripotent stem cells develop teratoma more efficiently and faster than human embryonic stem cells regard-

- less the site of injection. *Stem Cells*. 2010;28:1568–70, <http://dx.doi.org/10.1002/stem.471>.
91. Kondziolka D, Wechsler L, Goldstein S, et al. Transplantation of cultured human neuronal cells for patients with stroke. *Neurology*. 2000;55:565–9, <http://dx.doi.org/10.1212/WNL.55.4.565>.
  92. Prabhakaran S, Zarahn E, Riley C, et al. Inter-individual variability in the capacity for motor recovery after ischemic stroke. *Neurorehabil Neural Repair*. 2007;22(1):64–71, <http://dx.doi.org/10.1177/1545968307305302>.
  93. Cramer SC, Chopp M. Recovery recapitulates ontogeny. *Trends Neurosci*. 2000;23:265–71, [http://dx.doi.org/10.1016/S0166-2236\(00\)01562-9](http://dx.doi.org/10.1016/S0166-2236(00)01562-9).
  94. Savitz SI, Chopp M, Deans R, et al. Stem cell therapy as an emerging paradigm for stroke (STEPS) II. *Stroke*. 2011;42:825–9, <http://dx.doi.org/10.1161/STROKEAHA.110.601914>.
  95. Savitz SI, Cramer SC, Wechsler L, et al. Stem cells as an emerging paradigm in stroke 3: enhancing the development of clinical trials. *Stroke*. 2014;45:634–9, <http://dx.doi.org/10.1161/STROKEAHA.113.003379>.
  96. Lindenberg R, Renga V, Zhu LL, et al. Structural integrity of corticospinal motor fibers predicts motor impairment in chronic stroke. *Neurology*. 2010;74:280–7, <http://dx.doi.org/10.1212/WNL.0b013e3181ccc6d9>.
  97. Lee SY, Lim JY, Kang EK, et al. Prediction of good functional recovery after stroke based on combined motor and somatosensory evoked potential findings. *J Rehabil Med*. 2010;42:16–20, <http://dx.doi.org/10.2340/16501977-0475>.
  98. Chen J, Li Y, Wang L, et al. Therapeutic benefit of intracerebral transplantation of bone marrow stromal cells after cerebral ischemia in rats. *J Neurol Sci*. 2001;189:49–57, [http://dx.doi.org/10.1016/S0022-510X\(01\)00557-3](http://dx.doi.org/10.1016/S0022-510X(01)00557-3).
  99. Atkinson K, Horowitz MM, Gale RP, et al. Risk factors for chronic graft-versus-host disease after HLA-identical sibling bone marrow transplantation. *Blood*. 1990;75:2459–64.
  100. Sprigg N, Bath PM, Zhao L, et al. Granulocyte-colony-stimulating factor mobilizes bone marrow stem cells in patients with subacute ischemic stroke: the stem cell trial of recovery enhancement after stroke (STEMS) pilot randomized, controlled trial (ISRCTN 16784092). *Stroke*. 2006;37:2979–83, <http://dx.doi.org/10.1161/01.STR.0000248763.49831.c3>.
  101. Alasheev AM, Belkin AA, Leiderman IN, et al. Granulocyte-colony-stimulating Factor for Acute Ischemic Stroke: A Randomized Controlled Trial (STEMTHER). *Transl Stroke Res*. 2011;2:358–65, <http://dx.doi.org/10.1007/s12975-011-0091-3>.
  102. England TJ, Abaei M, Auer DP, et al. Granulocyte-colony stimulating factor for mobilizing bone marrow stem cells in subacute stroke: the stem cell trial of recovery enhancement after stroke 2 randomized controlled trial. *Stroke*. 2012;43:405–11, <http://dx.doi.org/10.1161/STROKEAHA.111.636449>.
  103. Jørgensen HS, Nakayama H, Raaschou HO, et al. Outcome and time course of recovery in stroke. Part II: time course of recovery. *The Copenhagen Stroke Study*. *Arch Phys Med Rehabil*. 1995;76:406–12.
  104. Kondziolka D, Steinberg GK, Wechsler L, et al. Neurotransplantation for patients with subcortical motor stroke: a phase 2 randomized trial. *J Neurosurg*. 2005;103:38–45, <http://dx.doi.org/10.3171/jns.2005.103.1.0038>.
  105. Komatsu K, Honmou O, Suzuki J, et al. Therapeutic time window of mesenchymal stem cells derived from bone marrow after cerebral ischemia. *Brain Res*. 2010;1334:84–92, <http://dx.doi.org/10.1016/j.brainres.2010.04.006>.
  106. Sharma A, Sane H, Gokulchandran N, et al. Autologous bone marrow mononuclear cells intrathecal transplantation in chronic stroke. *Stroke Res Treat*. 2014;2014:1–9, <http://dx.doi.org/10.1155/2014/234095>.
  107. Savitz SI, Misra V, Kasam M, et al. Intravenous autologous bone marrow mononuclear cells for ischemic stroke. *Ann Neurol*. 2011;70:59–69, <http://dx.doi.org/10.1002/ana.22458>.
  108. Smith HK, Gavins FN. The potential of stem cell therapy for stroke: is PISCES the sign? *FASEB J*. 2012;26:2239–52, <http://dx.doi.org/10.1096/fj.11-195719>.
  109. Rodríguez-Frutos B, Otero-Ortega L, Gutiérrez-Fernández M, et al. Stem cell therapy and administration routes after stroke. *Transl Stroke Res*. 2016;1–10, <http://dx.doi.org/10.1007/s12975-016-0482-6>.
  110. Lee JS, Hong JM, Moon GJ, et al. A long-term follow-up study of intravenous autologous mesenchymal stem cell transplantation in patients with ischemic stroke. *Stem Cells*. 2010;28:1099–106, <http://dx.doi.org/10.1002/stem.430>.
  111. Prasad K, Sharma A, Garg A, et al. Intravenous autologous bone marrow mononuclear stem cell therapy for ischemic stroke: a multicentric, randomized trial. *Stroke*. 2014;45:3618–24, <http://dx.doi.org/10.1161/STROKEAHA.114.007028>.
  112. Kaur C, Ling EA. Blood brain barrier in hypoxic-ischemic conditions. *Curr Neurovasc Res*. 2008;5:71–81, <http://dx.doi.org/10.2174/156720208783565645>.
  113. Greenwood J. Mechanisms of blood-brain barrier breakdown. *Neuroradiology*. 1991;33:95–100.
  114. Liu L, Eckert MA, Riazifar H, et al. From blood to the brain: can systemically transplanted mesenchymal stem cells cross the blood-brain barrier? *Stem Cells Int*. 2013;2013:1–7, <http://dx.doi.org/10.1155/2013/435093>.
  115. Vulliet PR, Greeley M, Halloran SM, et al. Intracoronary arterial injection of mesenchymal stromal cells and microinfarction in dogs. *Lancet*. 2004;363:783–4, [http://dx.doi.org/10.1016/S0140-6736\(04\)15695-X](http://dx.doi.org/10.1016/S0140-6736(04)15695-X).
  116. Friedrich MAG, Martins MP, Araújo MD, et al. Intra-arterial infusion of autologous bone marrow mononuclear cells in patients with moderate to severe middle cerebral artery acute ischemic stroke. *Cell Transplant*. 2012;21 Suppl 1:13–22, <http://dx.doi.org/10.3727/096368912X612512>.
  117. Battistella V, Freitas GR De, Dias V, et al. Safety of autologous bone marrow mononuclear cell transplantation in patients with nonacute ischemic stroke. *Futur Med*. 2011;6:45–52.
  118. Lu D, Mahmood A, Wang L, et al. Adult bone marrow stromal cells administered intravenously to rats after traumatic brain injury migrate into brain and improve neurological outcome. *Neuroreport*. 2001;12:559–63.
  119. Lim JY, Jeong CH, Jun JA, et al. Therapeutic effects of human umbilical cord blood-derived mesenchymal stem cells after intrathecal administration by lumbar puncture in a rat model of cerebral ischemia. *Stem Cell Res Ther*. 2011;2:38, <http://dx.doi.org/10.1186/scrt79>.
  120. Mancias-Guerra C, Marroquín-Escamilla AR, González-Llano O, et al. Safety and tolerability of intrathecal delivery of autologous bone marrow nucleated cells in children with cerebral palsy: an open-label phase I trial. *Cytherapy*. 2014;16:810–20, <http://dx.doi.org/10.1016/j.jcyt.2014.01.008>.
  121. Yang W-Z, Zhang Y, Wu F, et al. Safety evaluation of allogeneic umbilical cord blood mononuclear cell therapy for degenerative conditions. *J Transl Med*. 2010;8:75, <http://dx.doi.org/10.1186/1479-5876-8-75>.
  122. Mehta T, Feroz A, Thakkar U, et al. Subarachnoid placement of stem cells in neurological disorders. *Transplant Proc*. 2008;40:1145–7, <http://dx.doi.org/10.1016/j.transproceed.2008.03.026>.
  123. Hurst RW, Peter Bosch E, Morris JM, et al. Inflammatory hypertrophic cauda equina following intrathecal

- neural stem cell injection. *Muscle Nerve*. 2013;48:831–5, <http://dx.doi.org/10.1002/mus.23920>.
124. Alderazi YJ, Coons SW, Chapman K. Catastrophic demyelinating encephalomyelitis after intrathecal and intravenous stem cell transplantation in a patient with multiple sclerosis. *J Child Neurol*. 2012;27:632–5, <http://dx.doi.org/10.1177/0883073811422831>.
  125. Kawarai T. Spinal myoclonus resulting from intrathecal administration of human neural stem cells. *Mov Disord*. 2011;26:1353–4, <http://dx.doi.org/10.1002/mds.22959>.
  126. Qiao L-Y, Huang F-J, Zhao M, et al. A two-year follow-up study of cotransplantation with neural stem/progenitor cells and mesenchymal stromal cells in ischemic stroke patients. *Cell Transplant*. 2014;23 Suppl 1:65–72, <http://dx.doi.org/10.3727/096368914X684961>.
  127. Stilley CS, Ryan CM, Kondziolka D, et al. Changes in cognitive function after neuronal cell transplantation for basal ganglia stroke. *Neurology*. 2004;132:0–1322, <http://dx.doi.org/10.1212/01.WNL.0000140700.44904.53>.
  128. Saenger AK, Christenson RH. Stroke Biomarkers: progress and challenges for diagnosis, prognosis, differentiation, and treatment. *Clin Chem*. 2010;56:21–33, <http://dx.doi.org/10.1373/clinchem.2009.133801>.
  129. Cramer SC, Koroshetz WJ, Finklestein SP. The case for modality specific outcome measures in clinical trials of stroke recovery promoting agents. *Stroke*. 2007;38:1393–5, <http://dx.doi.org/10.1161/01.STR.0000260087.67462.80>.
  130. Ding G, Jiang Q, Li L, et al. Magnetic resonance imaging investigation of axonal remodeling and angiogenesis after embolic stroke in sildenafil treated rats. *J Cereb Blood Flow Metab*. 2008;28:1440–8, <http://dx.doi.org/10.1038/jcbfm.2008.33>.
  131. Jiang Q, Zheng GZ, Guang LD, et al. Investigation of neural progenitor cell induced angiogenesis after embolic stroke in rat using MRI. *Neuroimage*. 2005;28:698–707, <http://dx.doi.org/10.1016/j.neuroimage.2005.06.063>.
  132. Li L, Jiang Q, Zhang L, et al. Ischemic cerebral tissue response to subventricular zone cell transplantation measured by iterative self-organizing data analysis technique algorithm. *J Cereb Blood Flow Metab*. 2006;26:1366–77, <http://dx.doi.org/10.1038/sj.jcbfm.9600288>.