



# Performance of Serum Glypican 3 in Diagnosis of Hepatocellular Carcinoma: A meta-analysis

Dahai Xu,<sup>\*,\*\*</sup> Chang Su,<sup>\*\*\*</sup> Liang Sun,<sup>\*</sup> Yuanyuan Gao,<sup>\*</sup> Youjun Li<sup>\*</sup>

<sup>\*</sup> Department of Human Anatomy, College of Basic Medical Sciences, Jilin University, Changchun, Jilin, China.

<sup>\*\*</sup> Department of Emergency, The First Hospital of Jilin University, Changchun, Jilin, China.

<sup>\*\*\*</sup> Department of Thyroid Surgery, The First Hospital of Jilin University, Changchun, Jilin, China.

## ABSTRACT

**Introduction and aim.** Serum glypican-3 (GPC3) has been explored as a non-invasive biomarker of hepatocellular carcinoma (HCC). However, controversy remains on its diagnostic accuracy. Therefore, we aimed to conduct a systematic review and meta-analysis to evaluate the differential diagnostic accuracy of serum GPC3 between HCC and liver cirrhosis (LC) cases. **Material and methods.** After the strict filtering and screening of studies from NCBI, PUBMED, Clinical Trials, Cochrane library, Embase, Prospero and Web of Science databases, 11 studies were selected. All studies provided the sensitivity and specificity of GPC3 and the alpha-fetoprotein (AFP) in the HCC and LC diagnosis. The sensitivity and specificity, and the area under the receiver operating characteristic curve (AUC) were determined and compared between GPC3 and AFP, which was set as a positive control. **Results.** Pooled sensitivity (95% CI) and specificity (95% CI) were 0.55 (0.52-0.58) and 0.58 (0.54-0.61) for GPC3, 0.54 (0.51-0.57) and 0.83 (0.80-0.85) for AFP, and 0.85 (0.81-0.89) and 0.79 (0.73-0.84) for GPC3 + AFP, respectively. The AUCs of GPC3, AFP and GPC3 + AFP were 0.7793, 0.7867 and 0.9366, respectively. GPC3 had a nearly similar sensitivity as AFP, while the specificity and AUC of GPC3 was lower than that of AFP. The combination of GPC3 and AFP yielded a better sensitivity and AUC than GPC3 or AFP. **Conclusion.** Serum GPC3 is inferior to AFP in the differential diagnosis between HCC and LC. However, the combination of GPC3 and AFP exhibited a much better performance.

**Key words.** Alpha fetoprotein. Biomarker. Liver cirrhosis. Sensitivity. Specificity.

## INTRODUCTION

Hepatocellular carcinoma (HCC) is the fifth most common cancer worldwide, and the second leading cause of site-specific cancer-related death. Usually, HCC is asymptomatic at the early stage. However, it is always detected at the advanced stage when diagnosed, which limits treatment options. Thus, the early diagnosis of HCC is of great significance to enable early therapeutic intervention and prolong the survival period.<sup>1</sup> In this context, the serological level of alpha-fetoprotein (AFP) has been used as a classical marker of HCC. However, AFP has modest sensitivity and specificity for HCC diagnosis,<sup>2,3</sup> which makes its application questionable. Indeed, AFP has been excluded from the present guidelines of the American Association for the Study of Liver Disease and the European Association for the Study of the Liver due to its modest

accuracy.<sup>4</sup> Therefore, there is an urgent need to explore a surrogate serological marker with higher sensitivity and specificity for HCC diagnosis.

Glypican-3 (GPC3) is a transmembrane proteoglycan anchored to the cell membrane by glycosylated phosphatidylinositol (GPI). GPC3 has been verified to be closely correlated to the growth, proliferation, invasion and metastasis of cancer cells.<sup>5,6</sup> Several studies have addressed the involvement of GPC3 in various types of tumors, including HCC.<sup>7-10</sup> Interestingly, increasing evidence indicates that approximately 40% of HCC patients are positive for GPC3 and negative for AFP.<sup>3</sup> This might indicate the promising role of GPC3 in HCC diagnosis and its superiority over AFP. Notably, there is no correlation between the level of GPC3 and AFP,<sup>11,12</sup> which indicates that both parameters are functionally independent. Given that increasing evidence has indicated the application of serum

GPC3 as a noninvasive marker for HCC diagnosis, many investigators have explored this issue in comprehensive settings with respect to the diagnostic efficiency of GPC3, compared to AFP.

Several liver diseases, such as viral hepatitis, autoimmune hepatitis, alcoholic hepatitis and primary biliary cirrhosis, have progressed to liver cirrhosis (LC), which is a major predisposing factor of HCC risk. It is noteworthy that although the vast majority of HCC cases developed from cirrhotic livers, not all cirrhotic livers always end with HCC. In previous meta-analysis studies,<sup>13-18</sup> no meta-analysis study employed cirrhotic patients as controls. Therefore, the novelty of the present systemic review and meta-analysis over all previous studies originates from the use of LC cases as the control for the study. Through this approach, we were able to precisely and comprehensively assess the accuracy of GPC3 in the differential diagnosis between HCC and LC, compared to AFP.

## MATERIAL AND METHODS

### Literature search strategy

Two independent investigators (Chang Su and Dahai Xu) conducted an electronic literature search on seven databases, which include the NCBI, PUBMED, Clinical Trials, Cochrane library, Embase, Prospero and Web of Science databases. The search was updated as of November 10, 2017. The entry terms used for the literature search were as follows:

- HCC. Liver neoplasm, hepatic neoplasm, hepatocellular cancer, hepatic cancer, and liver cancer.
- GPC3. Glypican, glypican 3, glypican3, and glypican-3.
- AFP. Alpha-Fetoprotein and alpha Fetoprotein. No limit was set on publication time, study design and publishing format, and only publications in the English language were searched.

### Inclusion and exclusion criteria

Inclusion criteria:

- Studies that accurately diagnosed the experimental group with HCC
- Studies that measured serum GPC3 and AFP protein.
- Studies that determined the HCC diagnostic sensitivity and specificity of GPC3 and AFP.

Exclusion criteria:

- Letters, reviews, case reports, abstracts, editorials, and expert opinions.

- Studies that lack sufficient data to obtain the sensitivity and specificity of GPC3 and AFP in HCC and LC.
- Studies on experimental models, such as laboratory animals and cultured cells.
- Studies that considered specimens other than blood.
- Studies that evaluated serum marker levels by messenger RNA, DNA, or DNA polymorphisms.
- Studies that focused on diseases other than primary hepatocellular carcinoma.

Different articles with the same authors and data were checked to avoid duplicates, and the most recent or most complete study was selected.

### Data extraction

After selecting all the eligible studies, two investigators (Liang Sun and Yuanyuan Gao) independently extracted the data. The collected data included the first author's name, the country of origin of the patients, publication year, name of the journal, study design, number of patients, age and gender, assay type, cut-off value, and raw data (True Positive, TP; False Positive, FP; False Negative, FN; True Negative, TN). For disagreements, a third investigator (Youjun Li) was consulted to make the judgment.

### Assessment of quality of the selected articles

Two independent investigators (Chang Su and Dahai Xu) assessed the quality of the included studies according to the Quality Assessment of Studies of Diagnostic Accuracy II (QUADAS-2).<sup>14</sup> Signaling questions on the risk of bias in the assessment check list were labeled as "yes", "no", or "unclear". Items that assessed applicability risk were labeled as "high", "low", or "unclear". If the study design was cross-sectional, the risk of bias of the patient selection domain was labeled as "high risk". A third investigator (Youjun Li) was consulted for disagreements.

### Statistical analysis

Statistical analyses were performed using Review Manager 5.3, Stata 12 and Meta-Disc 1.4 software. Sensitivity, specificity, positive likelihood ratio (LR+), negative likelihood ratio (LR-), diagnostic odds ratio (DOR), 95% confidence intervals (95% CIs) and the area under the receiver operating characteristic curve (AUC) were determined. Forest plots and receiver operating characteristic curves (ROCs) were used to determine the diagnostic performance. The heterogeneity of the retrieved data was evaluated using the I<sup>2</sup>-value. An I<sup>2</sup>-value < 50% and a

P-value > 0.1 was considered with insignificant heterogeneity. If the heterogeneity was not identified, the fixed-effects model was used for the meta-analysis, and if the heterogeneity was identified, the random-effects model (DerSimonian-Laird) was used. Publication bias was measured by Egger's test using Stata 12 (StataCorp LP, College Station, TX, USA). P < 0.05 was considered statistically significant. In order to analyze the source of heterogeneity, threshold analysis was conducted using Meta-Disc 1.4 and meta-regression was conducted using Stata 12.

## RESULTS

### Study selection

Initially, 938 potentially relevant articles were retrieved from the databases mentioned above. Then, due to duplication, 301 articles were excluded. Next, after reviewing the titles and abstracts, 10 abstracts, 21 case reports, 11 meta-analyses, 21 reviews and 442 irrelevant articles (subjects were not human and the specimens were not serum, or GPC3 was determined based of the expression level of its mRNA or DNA) were excluded. Subsequently, by reading the full text, 121 studies were excluded due irrelevant design or insufficient data to calculate the sensitivity and specificity of GPC3. Finally, a total of 11 studies were eligible<sup>8-11,19-25</sup> (Figure 1). All included studies were approved by the Ethics Committee, and an informed consent was obtained from all subjects.

### Features and methodology

The clinical features and methodology of these eligible studies are summarized in table 1.

### Quality assessment of the studies

QUADAS-2 quality assessment was conducted to evaluate the quality of the included studies (Figure 2). All studies were retrospectively designed, and none of these studies were randomized controlled trials (RCTs). Merely two studies were consecutive investigations.<sup>8,10</sup> Thus, the patient selection domain of other studies was labeled as 'high risk'. Since there was no fixed diagnostic standard in each study, the flow and timing domain was labeled as 'unclear', while most of the studies included the cut-off levels of either serum GPC3 or AFP.<sup>8-11,19,20,22,23,25</sup> Two studies<sup>21,24</sup> did not include the cutoff value for GPC3, while two studies<sup>21,24</sup> did not include the cutoff value for AFP. Notably, the baseline-pretreatment level of serum GPC3 was determined in four studies,<sup>8,9,19,24</sup> and the remaining seven studies did not mention whether the sam-

ple was collected before therapy.<sup>10,11,20-23,25</sup> On the other hand, some studies indicated that their thresholds were not pre-specific (the cut-off point was fixed before the test), because their cut-off values were determined based on the ROC analysis.<sup>8-10,19</sup> Hence, the index test was set as 'high risk'. Since the temperature of the serum sample storage is critical for assay accuracy, three different temperatures (-20°C, -70°C and -80°C) were considered.<sup>8,9,11,19,21,24</sup> Studies with high risk of bias were not excluded to investigate the heterogeneity in the following step.<sup>8-10,19,21-23,25,26</sup>

### Diagnostic accuracy of serum GPC3 and AFP for HCC diagnosis

Sensitivity, specificity, LR+, LR-, DOR and  $I^2$  were calculated for all the included studies (Table 2 and Figure 3). The sensitivity of GPC3 was almost the same as that of AFP, but the specificity of GPC3 was lower than that of AFP. Moreover, the AUC of GPC3 (0.7793) was lower than that of AFP (0.7867) and GPC3 + AFP (0.9366) (Figure 4).

### Investigation for heterogeneity

Based on Egger's test, the included studies did not have publication bias (Coefficient = 3.00, P = 0.052 for GPC3; Coefficient = 2.56, P = 0.065 for AFP) (Figure 5). Furthermore, the diagnostic odds ratio revealed significant

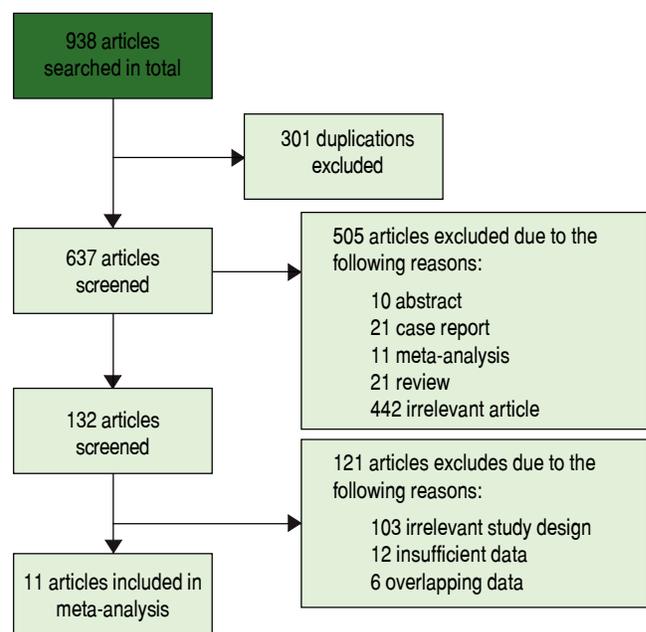


Figure 1. Study selection process.

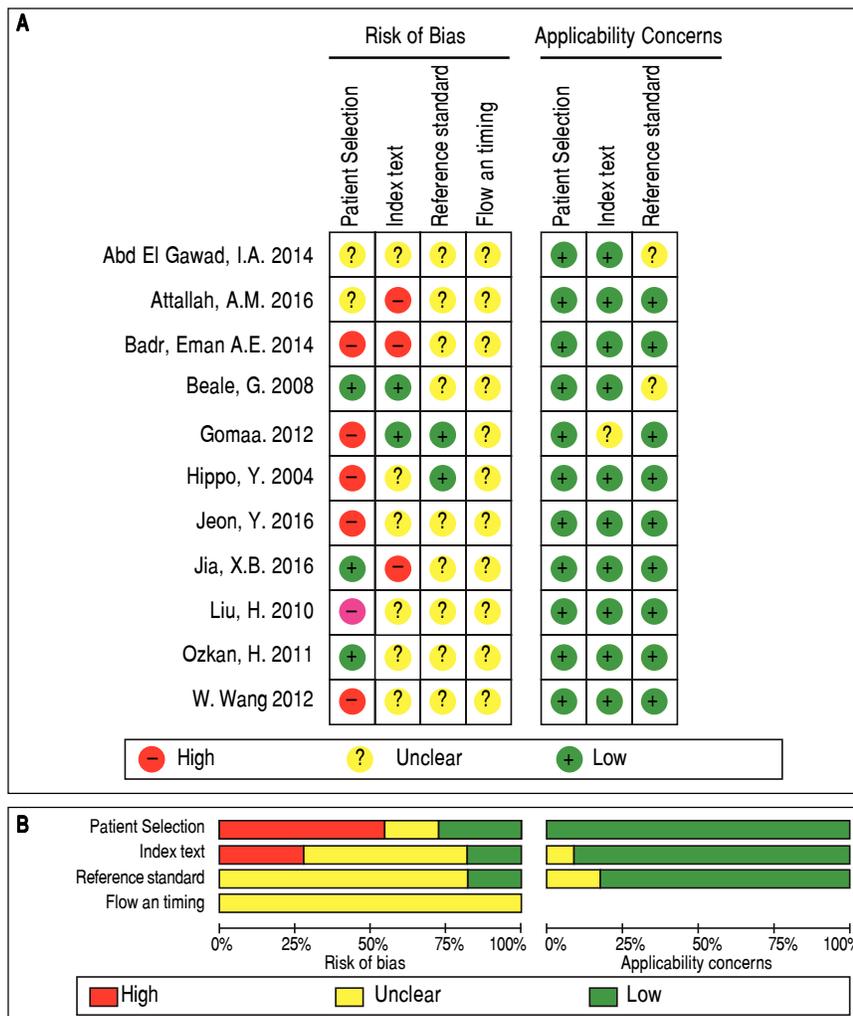
**Table 1.** Clinical features of the included studies

Author	Year	Country	HCC/LC	Gender (M/F)		Age (year)		Etiology		GPC3		AFP	
				HCC	LC	HCC	LC	HCC	LC Control	Cut-off value	Assay type	Cut-off value	Assay type
Jia, X. B.	2016	China	283/267	2.41	5.07	59 ± 9.6	52.9 ± 11.1	HBV 80.6%	HBV 80.9%	0.002 ng/mL	ELISA	20 ng/mL	ELISA
Jeon, Y.	2016	South Korea	157/156	4.23	1.36	60.8 ± 11.8	56.7 ± 10.8	HBV 67.5%, HCV 10.8%, ALD 10.8%	HBV 74.4%, HCV 23.7%, ALD 1.9%	0.61 ng/mL	ELISA	20 ng/mL	ELICA
Attallah, A. M.	2016	Egypt	138/56	NA	NA	54.9 ± 12.1	51.89 ± 8.9	HCV 100%	HCV 100%	6 ng/mL	ELISA	400 U/L	Chemiluminescence
Badr, Eman A.E.	2014	Egypt	30/30	5.00	NA	NA	NA	NA	HCV 100%	240 µg/mL	ELISA	200 ng/mL	ELISA
Abd, El Gawad, I.A.	2014	Egypt	40/10	4.00	4	59 (44-77)	57 (44-72)	HBV 17.5%, HCV 65%	NA	4.9 ng/mL	ELISA	20 ng/mL	MEIA
Gomaa, A H.O.	2012	Egypt	31/30	4.17	3.29	43-65	42-65	HBV 12.9%, HCV 87.1%	HBV 16.7%, HCV 83.3%	5.41 ng/mL	ELISA	42.32 ng/mL	NA
Wang, W.	2012	China	78/97	1.90	1.6	53.6 (46-66)	50.2 (38-68)	HBV 100% HBV 52%	HBV 100% HBV 29.1%	NA	ELISA	NA	ELISA
Ozkan, H.	2011	Turkey	75/55	2.57	1.5	63 ± 9.9	58 ± 12.7	HCV 22.7%, ALD 2.7%	HCV 16.4%, ALD 10.9%	3.9 pg/mL	NA	13 ng/mL	Chemiluminescence
Liu, H.	2010	China	75/32	2.75	3	55.4 ± 9.91	53.3 ± 5.81	HBV 84%, HCV 16%	HBV 78.1%, HCV 21.9%	300 ng/mL	ELISA	400 ng/mL	Chemiluminescence
Beale, G.	2008	England	50/41	4.00	2.15	67.5 ± 12.02	54.3 ± 9.62	ALD 60%, NAFLD 40%	ALD 80.5%, NAFLD 19.5%	NA	ELISA	NA	NA
Hippo, Y.	2004	Japan	69/38	NA	NA	NA	NA	NA	NA	NA	ELISA	NA	ELISA

NA: data are not available. HCC: hepatocellular carcinoma. LC: liver cirrhosis; NC: normal control. AH: atypical hyperplasia. HBV: hepatitis B virus. HCV: hepatitis C virus. ALD: alcoholic liver disease. NAFLD: non-alcoholic fatty liver disease. ELISA: enzyme-linked immunosorbant assay. ELICA: enzyme-linked chemiluminescent assay. MEIA: microparticle enzyme immunoassay.

heterogeneity ( $I^2 = 85.3\%$ ,  $P < 0.001$  for GPC3;  $I^2 = 86.8\%$ ,  $P < 0.001$  for AFP). The threshold effect test (Spearman's correlation coefficient was  $-0.209$ ,  $P = 0.537$ , for GPC3; Spearman's correlation coefficient was  $0.392$ ,  $P = 0.233$ , for AFP) indicated that the cut-off point was not the source of heterogeneity. In addition, the meta-regression analysis indicated that publication year, country, sample size, clinical characteristics methodology and the quality of articles were not correlated with heterogeneity. The sensitivity analysis indicated that the pooled estimates were stable and not influenced by a single study (Figure 6).

A subgroup analysis was conducted by publication year, country, methodology, clinical features, and study quality. However, subject size was the only possible source of heterogeneity found (whether the number of patients in both HCC and LC groups were  $\geq 40$  or not). For GPC3, in the subgroup that had a larger sample size was  $66.5\%$ , while the heterogeneity in another subgroup was  $77.2\%$ . The present meta-analysis did not include any RCTs. Other covariates were also potential sources of heterogeneity, such as the differences in operating protocol, enrollment period, pathological grades and tumor burden.



**Figure 2.** Quality evaluation of the included studies by QUADAS-2. **A.** Risk of bias and concerns graph by QUADAS-2. **B.** Risk of bias and concerns summary by QUADAS-2.

**Table 2.** Diagnostic values of GPC3 and/or AFP for HCC vs. LC

Serum biomarker	Sen. (95%CI)	Spe. (95%CI)	LR+ (95%CI)	LR- (95%CI)	DOR (95%CI)
GPC3	0.55 (0.52-0.58)	0.58 (0.54-0.61)	1.69 (1.20-2.39)	0.67 (0.50-0.90)	3.64 (1.74-7.60)
AFP	0.54 (0.51-0.57)	0.83 (0.80-0.85)	4.40 (2.39-8.08)	0.51 (0.42-0.63)	11.83 (5.02-27.88)
GPC3 + AFP	0.85 (0.81-0.89)	0.79 (0.73-0.84)	5.23 (1.75-15.65)	0.18 (0.07-0.47)	31.95 (5.75-177.65)

Sen.: Sensitivity. Spe.: Specificity. LR+: Positive likelihood Ratio. LR-: Negative likelihood Ratio. DOR: Diagnostic odds ratio.

## DISCUSSION

In the present study, we found that the sensitivity of GPC3 was nearly the same as that of AFP, while the specificity of GPC3 was lower than that of AFP. However, the combination of GPC3 and AFP can significantly in-

crease the sensitivity and AUC in the differential diagnosis between HCC and LC. However, the specificity of GPC3+AFP was lower than that of AFP.

Recently, GPC3 has become a focus in HCC studies. Many studies have explored the performance of GPC3 in liver tissues for HCC diagnosis,<sup>27-29</sup> and some clinical tri-

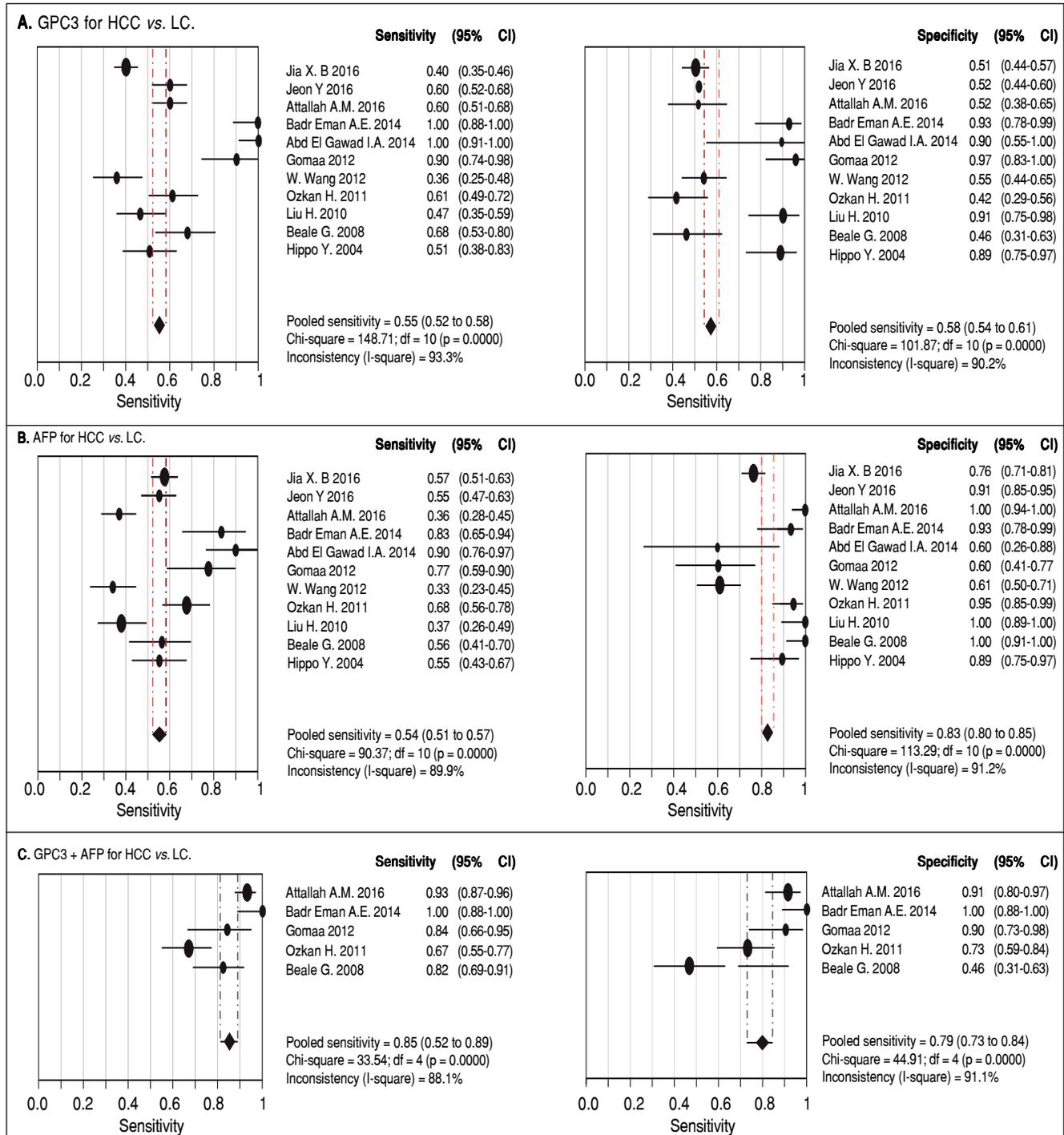
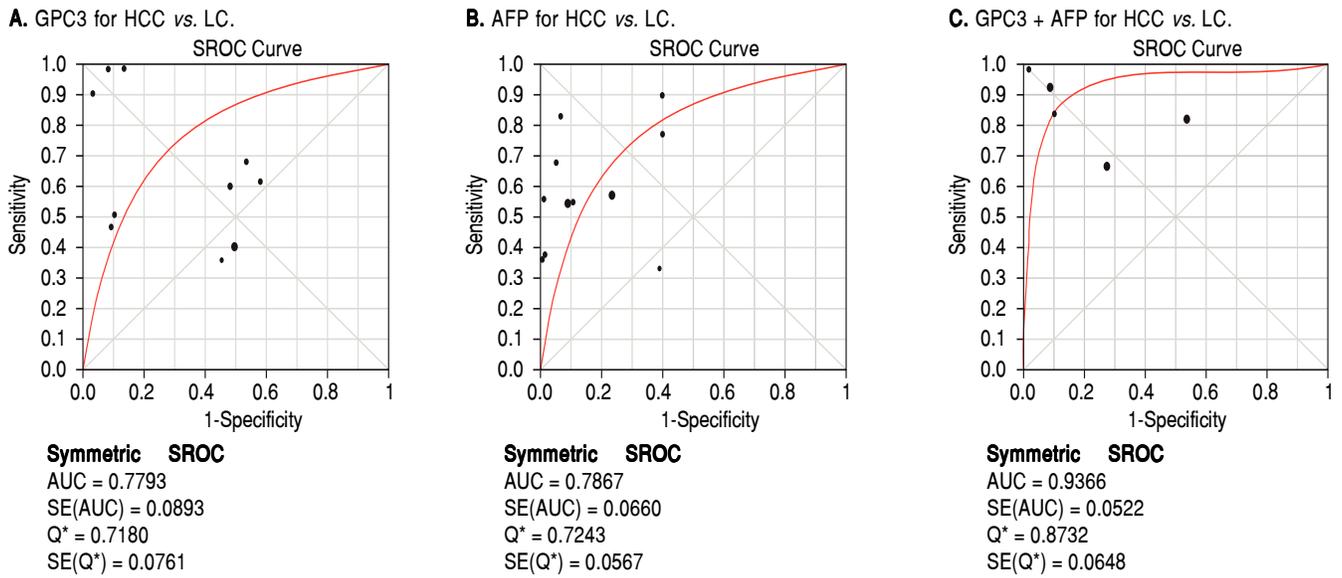
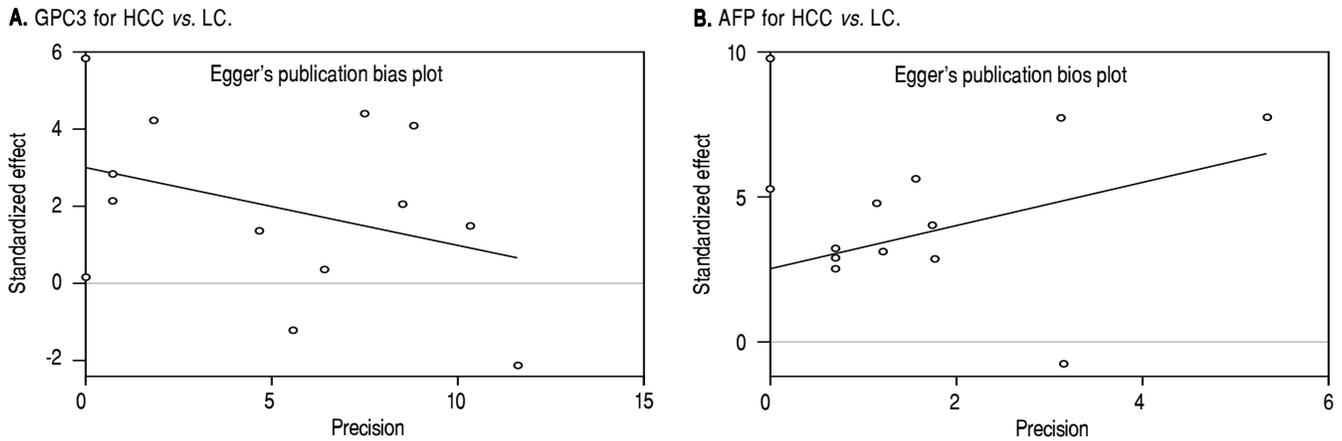


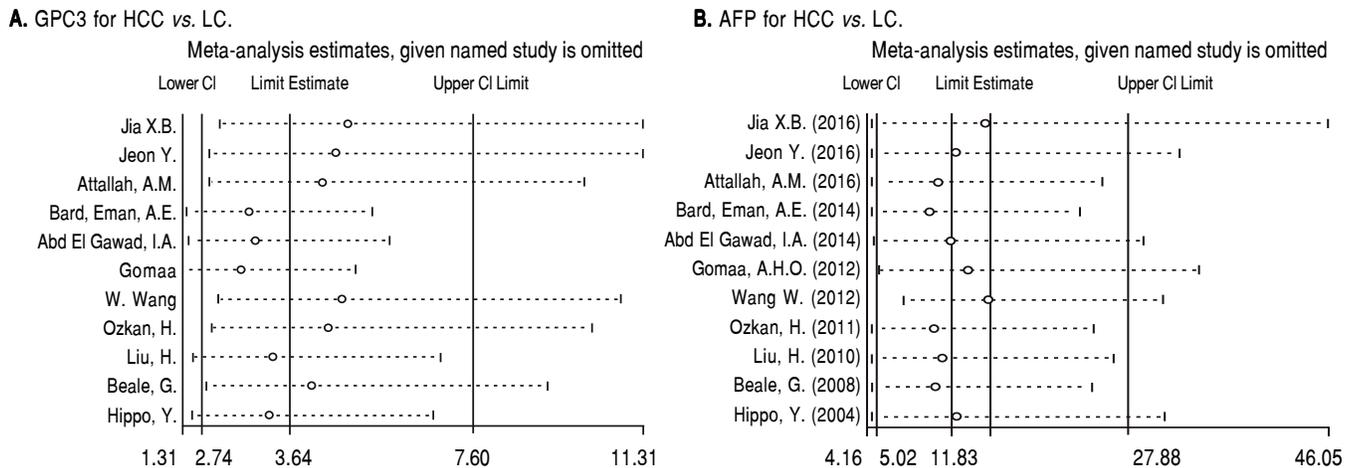
Figure 3. Sensitivity and specificity of GPC3 and/or AFP for HCC vs. LC.



**Figure 4.** ROC curves of GPC3 and/or AFP for the diagnosis between HCC and LC.



**Figure 5.** Egger's publication bias plot.



**Figure 6.** Sensitivity analysis plots.

als on therapeutic agents against GPC3 have also been conducted.<sup>30-33</sup> In the present investigation, we conducted a meta-analysis on studies that investigated the application of GPC3 as a diagnostic marker for HCC, and compared this with AFP. The novelty of this investigation originates from the use of LC patients as control subjects, which were compared to HCC. For the 11 eligible studies, the results infer that the performance of GPC3 to discriminate HCC from LC was unsatisfactory and lower than that of AFP. However, the combination of GPC3 and AFP produced a higher diagnostic accuracy than either of the makers when used separately.

GPC3 is implicated in cell growth, differentiation and migration.<sup>34</sup> GPC3 is greatly expressed in HCC tissues, fetal livers and most HCC cell lines, compared to other normal human tissues.<sup>8,20,35</sup> GPC3 has also been reported in other tumors, such as lung cancer, thyroid cancer and melanoma.<sup>36,37</sup> Notably, serum GPC3 levels were significantly higher in HCC than in normal controls, as well as in liver cirrhosis, colorectal cancer, esophageal cancer, gastric cancer and hepatitis cases.<sup>2,19,20</sup> Furthermore, GPC3 expression in HCC was positively associated with tumor size and pathological grade.<sup>3</sup> In addition, Ohno, *et al.* reported that HCC patients with high GPC3 level had poor prognosis.<sup>38-40</sup> Badr, *et al.* and Yeon, *et al.* found that the sensitivity of serum GPC3 in small size HCC was higher than that of AFP.<sup>19,41</sup> Moreover, Zhang QY, *et al.* reported that GPC3 is superior to AFP for HCC diagnosis, with higher sensitivity and specificity.<sup>42</sup> Generally, controversy remains on the application of GPC3 in HCC diagnosis, since several studies have either supported<sup>2,3,19,42,43</sup> or limited<sup>9,11,44,45</sup> the role of GPC3 in HCC diagnosis.

The results of the present meta-analysis support the observation of Jia, *et al.*,<sup>8</sup> since there was no significant difference in the serum level of GPC3 between HCC and LC patients, indicating that GPC3 is not efficiently valid as a HCC serum biomarker.

It is important to note that the sensitivity, specificity and AUC of GPC3 determined in the present meta-analysis differed from those reported in other studies.<sup>13-18</sup> This difference might be attributed to either the difference in the nature of the control cohort or the number of included studies.

To the best of our knowledge, the present meta-analysis is the first to set patients with LC as a control cohort. However, there were some limitations in the present study:

- A small number of included studies was included, which was attributed to the strict inclusion and exclusion criteria.
- There was an obvious heterogeneity, which might be contributed to the limited number of subjects.

- The quality of the included studies was unsatisfactory. Due to the absence of RCTs, the treatment effects may be overestimated.

Therefore, more high quality researches with different subgroups according to tumor etiology, tumor size and tumor stage are needed in the future to further verify the role of GPC3 in HCC diagnosis.

In conclusion, Serum GPC3 cannot be used as a substitute for AFP to differentially diagnose HCC and LC. However, the combination of both markers would be a better choice.

## ACKNOWLEDGMENTS

This work was supported by the National Natural Science Foundation of China (#81372456) YJL.

## CONFLICT OF INTEREST

The authors declares that there is no conflict of interest regarding the publication of this article.

## REFERENCES

1. Maluccio M, Covey A. Recent progress in understanding, diagnosing, and treating hepatocellular carcinoma. *CA: a Cancer Journal for Clinicians* 2012; 62: 394-9.
2. Zhao YS, Wang M, Cui CY, Zhang L, Liao F, Li HC, Wu X. Significance of combined tests of serum golgi glycoprotein 73 and other biomarkers in diagnosis of small primary hepatocellular carcinoma. *Cancer Biomark* 2015; 15: 677-83.
3. Yu JP, Xu XG, Ma RJ, Qin SN, Wang CR, Wang XB, Li M, et al. Development of a Clinical Chemiluminescent Immunoassay for Serum GPC3 and Simultaneous Measurements Alone With AFP and CK19 in Diagnosis of Hepatocellular Carcinoma. *J Clin Lab Anal* 2015; 29: 85-93.
4. Song DS, Bae SH. Changes of guidelines diagnosing hepatocellular carcinoma during the last ten-year period. *Clin Molec Hepatol* 2012; 18: 258-67.
5. Jiang ZW, Jiang XF, Chen SM, Lai YX, Wei XR, Li BH, Lin S, et al. Anti-GPEC3-CAR T Cells Suppress the Growth of Tumor Cells in Patient-Derived Xenografts of Hepatocellular Carcinoma. *Front Immunol* 2017; 7: 690
6. Zhu XT, Yuan JH, Zhu TT, Li YY, Cheng XY. Long noncoding RNA glypican 3 (GPC3) antisense transcript 1 promotes hepatocellular carcinoma progression via epigenetically activating GPC3. *Febs Journal* 2016; 283: 3739-54.
7. Sun B, Huang Z, Wang B, Yu YL, Lin SH, Luo L, Wang Y, et al. Significance of Glypican-3 (GPC3) Expression in Hepatocellular Cancer Diagnosis. *Med Sci Monit* 2017; 23: 850-5.
8. Jia XB, Gao YT, Zhai DK, Liu J, Cai JJ, Wang YJ, Jing L, et al. Assessment of the Clinical Utility of Glypican 3 as a Serum Marker for the Diagnosis of Hepatocellular Carcinoma. *Technol Cancer Res Treat* 2016; 15: 780-6.
9. Jeon Y, Jang ES, Choi YS, Kim JW, Jeong SH. Glypican-3 level assessed by the enzyme-linked immunosorbent assay is inferior to alpha-fetoprotein level for hepatocellular carcinoma diagnosis. *Clin Molec Hepatol* 2016; 22: 359-65.

10. Attallah AM, El-Far M, Omran MM, Abdelrazek MA, Attallah AA, Saeed AM, Farid K, et al. GPC-HCC model: a combination of glypican-3 with other routine parameters improves the diagnostic efficacy in hepatocellular carcinoma. *Tumour biology: the Journal of the International Society for Oncodevelopmental Biology and Medicine* 2016; 37: 12571-7.
11. Ozkan H, Erdal H, Kocak E, Tutkak H, Karaeren Z, Yakut M, Köklü S. Diagnostic and Prognostic Role of Serum Glypican 3 in Patients With Hepatocellular Carcinoma. *J Clin Lab Anal* 2011; 25: 350-3.
12. Tangkijvanich P, Chanmee T, Komtong S, Mahachai V, Wisedopas N, Pothacharoen P, Kongtawelert P. Diagnostic role of serum glypican-3 in differentiating hepatocellular carcinoma from non-malignant chronic liver disease and other liver cancers. *J Gastroenterol Hepatol* 2010; 25: 129-37.
13. Yang SL, Fang X, Huang ZZ, Liu XJ, Xiong ZF, Liu P, Yao HY, et al. Can serum glypican-3 be a biomarker for effective diagnosis of hepatocellular carcinoma? A meta-analysis of the literature. *Dis Markers* 2014; 2014: 127831.
14. Jia X, Liu J, Gao Y, Huang Y, Du Z. Diagnosis accuracy of serum glypican-3 in patients with hepatocellular carcinoma: a systematic review with meta-analysis. *Arch Med Res* 2014; 45: 580-8.
15. Xu C, Yan Z, Zhou L, Wang Y. A comparison of glypican-3 with alpha-fetoprotein as a serum marker for hepatocellular carcinoma: a meta-analysis. *J Cancer Res Clin Oncol* 2013; 139: 1417-24.
16. Liu JW, Zuo XL, Wang S. Diagnosis accuracy of serum Glypican-3 level in patients with hepatocellular carcinoma and liver cirrhosis: a meta-analysis. *Eur Rev Med Pharmacol Sci* 2015; 19: 3655-73.
17. Liu XF, Hu ZD, Liu XC, Cao Y, Ding CM, Hu CJ. Diagnostic accuracy of serum glypican-3 for hepatocellular carcinoma: a systematic review and meta-analysis. *Clin Biochem* 2014; 47: 196-200.
18. Fu WY, Lu HY, Li L, Wu KF, Li YP, Liu Y, Hu J, et al. Glypican-3 versus alpha-fetoprotein as a biomarker for hepatocellular carcinoma: A diagnostic meta-analysis. *Biocell* 2015; 39: 25-31.
19. Badr EAE, Korah TE, Ghani AA, El-Sayed S, Badr S. Role of serum glypican-3 in the diagnosis and differentiation of small hepatocellular carcinoma from hepatitis-C virus cirrhosis. *Alexandria J Med* 2014; 50: 221-6.
20. Abd El Gawad IA, Mossallam GI, Radwan NH, Elzawahry HM, Elhifnawy NM. Comparing prothrombin induced by vitamin K absence-II (PIVKA-II) with the oncofetal proteins glypican-3, Alpha feto protein and carcinoembryonic antigen in diagnosing hepatocellular carcinoma among Egyptian patients. *J Egypt Natl Canc Inst* 2014; 26: 79-85.
21. Wang W, Zhao LJ, Wang Y, Tao QY, Feitelson MA, Zhao P, Ren H, et al. Application of HBx-induced anti-URGs as early-warning biomarker of cirrhosis and HCC. *Cancer Biomark* 2012; 11: 29-39.
22. Gomaa AHO, Aboraia G, Attia H, Elezawy H, Nafie E, Elbaz S. The diagnostic value of peripheral blood glypican-3 in patients with hepatocellular carcinoma. *World J Med Sci* 2012; 7: 105-12.
23. Liu H, Li P, Zhai Y, Qu CF, Zhang LJ, Tan YF, Li N, et al. Diagnostic value of glypican-3 in serum and liver for primary hepatocellular carcinoma. *World J Gastroenterol* 2010; 16: 4410-5.
24. Beale G, Chattopadhyay D, Gray J, Stewart S, Hudson M, Day C, Trerotoli P, et al. AFP, PIVKAll, GP3, SCCA-1 and follistatin as surveillance biomarkers for hepatocellular cancer in non-alcoholic and alcoholic fatty liver disease. *Bmc Cancer* 2008; 8.
25. Hippo Y, Watanabe K, Watanabe A, Midorikawa Y, Yamamoto S, Ihara S, Tokita S, et al. Identification of soluble NH2-terminal fragment of glypican-3 as a serological marker for early-stage hepatocellular carcinoma. *Cancer Res* 2004; 64: 2418-23.
26. Yu J, Ma Q, Zhang B, Ma R, Xu X, Li M, Xu W, et al. Clinical application of specific antibody against glypican-3 for hepatocellular carcinoma diagnosis. *Science China Life Sciences* 2013; 56: 234-9.
27. Zhou R, Pan QJ, Wu K, Li R. Distribution and characterization of glypican-3 in hepatocellular carcinoma. *Int J Clin Exp Pathol* 2016; 9: 8631-3.
28. Sang W, Miao N, Cui WL, Li XX, Abulajiang G, Zhang W, Li Q, et al. Value of the combined detection arginase-1 and glypican-3 expression in distinguishing hepatocellular carcinoma from metastatic tumors. *Int J Clin Exp Pathol* 2016; 9: 7748-54.
29. Nguyen TB, Roncalli M, Di Tommaso L, Kakar S. Combined use of heat-shock protein 70 and glutamine synthetase is useful in the distinction of typical hepatocellular adenoma from atypical hepatocellular neoplasms and well-differentiated hepatocellular carcinoma. *Mod Pathol* 2016; 29: 283-92.
30. Abou-Alfa GK, Yen CJ, Hsu CH, O'Donoghue J, Beylertgil V, Ruan S, Pandit-Taskar N, et al. Phase Ib study of codrituzumab in combination with sorafenib in patients with non-curable advanced hepatocellular carcinoma (HCC). *Cancer Chemother Pharmacol* 2017; 79: 421-9.
31. Abou-Alfa GK, Puig O, Daniele B, Kudo M, Merle P, Park JW, Ross P, et al. Randomized phase II placebo controlled study of codrituzumab in previously treated patients with advanced hepatocellular carcinoma. *J Hepatol* 2016; 65: 289-95.
32. Ikeda M, Ohkawa S, Okusaka T, Mitsunaga S, Kobayashi S, Morizane C, Suzuki I, et al. Japanese phase I study of GC33, a humanized antibody against glypican-3 for advanced hepatocellular carcinoma. *Cancer science* 2014; 105: 455-62.
33. Sawada Y, Yoshikawa T, Nobuoka D, Shirakawa H, Kuronuma T, Motomura Y, Mizuno S, et al. Phase I trial of a glypican-3-derived peptide vaccine for advanced hepatocellular carcinoma: immunologic evidence and potential for improving overall survival. *Clinical Cancer Research: an Official Journal of the American Association for Cancer Research* 2012; 18: 3686-96.
34. Filmus J, Selleck SB. Glypicans: proteoglycans with a surprise. *J Clin Invest* 2001; 108: 497-501.
35. Chen IP, Ariizumi S, Nakano M, Yamamoto M. Positive glypican-3 expression in early hepatocellular carcinoma predicts recurrence after hepatectomy. *J Gastroenterol* 2014; 49: 117-25.
36. Chen M, Li GH, Yan J, Lu XZ, Cui JW, Ni ZX, Cheng W, et al. Reevaluation of glypican-3 as a serological marker for hepatocellular carcinoma. *Clin Chim Acta* 2013; 423: 105-11.
37. Ikuta Y, Nakatsura T, Kageshita T, Fukushima S, Ito S, Wakamatsu K, Baba H, et al. Highly sensitive detection of melanoma at an early stage based on the increased serum secreted protein acidic and rich in cysteine and glypican-3 levels. *Clin Cancer Res* 2005; 11: 8079-88.
38. Ohno A, Yorita K, Haruyama Y, Kondo K, Kato A, Ohtomo T, Kawaguchi M, et al. Aberrant expression of monocarboxylate transporter 4 in tumour cells predicts an unfavourable outcome in patients with hepatocellular carcinoma Akinobu Ohno. *Liver Int* 2014; 34: 942-52.
39. Xiao WK, Qi CY, Chen D, Li SQ, Fu SJ, Peng BG, Liang LJ. Prognostic significance of glypican-3 in hepatocellular carcinoma: a meta-analysis. *Bmc Cancer* 2014; 14: 104.

40. Li J, Gao JZ, Du JL, Wei LX. Prognostic and clinicopathological significance of glypican-3 overexpression in hepatocellular carcinoma: A meta-analysis. *World J Gastroenterol* 2014; 20: 6336-44.
41. Yeon JE, Suh SJ, Lee SJ, Yoon EL, Kang K, Yoo YJ, Lee HJ, et al. Clinical utility of plasma glypican-3 and osteopontin as biomarkers of hepatocellular carcinoma. *Gut and Liver* 2014; 8: 177-85.
42. Zhang QY, Xiao Q, Lin Z, Ying XT, Li ZJ, Lin JM. Development of a competitive radioimmunoassay for glypican-3 and the clinical application in diagnosis of hepatocellular carcinoma. *Clinical Biochemistry* 2010; 43: 1003-8.
43. Nassar A, Cohen C, Siddiqui MT. Utility of glypican-3 and survivin in differentiating hepatocellular carcinoma from benign and preneoplastic hepatic lesions and metastatic carcinomas in liver fine-needle aspiration biopsies. *Diagnostic Cytopathology* 2009; 37: 629-35.
44. Yasuda E, Kumada T, Toyoda H, Kaneoka Y, Maeda A, Okuda S, Yoshimi N, et al. Evaluation for clinical utility of GPC3, measured by a commercially available ELISA kit with Glypican-3 (GPC3) antibody, as a serological and histological marker for hepatocellular carcinoma. *Hepatology Research: the Official Journal of the Japan Society of Hepatology* 2010; 40: 477-85.
45. Wang Y, Yang H, Xu H, Lu X, Sang X, Zhong S, Huang J, et al. Golgi protein 73, not Glypican-3, may be a tumor marker complementary to alpha-Fetoprotein for hepatocellular carcinoma diagnosis. *J Gastroenterol Hepatol* 2014; 29: 597-602.

**Correspondence and reprint request:**

Youjun Li, M.D.

Department of Human Anatomy, College of Basic Medical Sciences, Jilin University, No. 126 Xinmin Street, Changchun, Jilin 130000, China. Tel.: +8614163577998; Fax: +86043185654528.

E-mail: yangzhouxux@sina.com