



## Concise reviews

# Immunotherapy in hepatocellular carcinoma

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## ABSTRACT

Hepatocellular carcinoma (HCC) is considered an immunogenic tumor that arises in chronically inflamed livers due to underlying chronic liver disease caused by viral and non-viral pathogenesis. This inflammation leads to tumor development and is associated to higher tumor immunogenicity.

For this reason immunotherapeutic approaches may be suitable therapeutic strategies for HCC. Indeed, several preclinical and clinical data support this hypothesis showing that immunotherapy and even more their combination may be a good alternative candidate for the treatment of HCC patients.

However, considering that the liver plays a central role in host defense as well as in the maintenance of self-tolerance, it is characterized by a strong intrinsic immune suppressive microenvironment as well as by a high immune evasion, which may represent a major impediment for an effective immune response against tumor. Furthermore, the low expression of tumor antigens on liver cancer cells leads to a lower T-cell activation and tumor infiltration, resulting in a less efficient control of the tumor growth and, consequently, in a worse clinical outcome.

For this reason, strategies should be developed to counteract the different factors in the HCC tumor microenvironment playing a major role in reducing the effects of immunotherapy.

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## 1. Introduction

Chronic inflammation and the intrinsic intra-hepatic immuno-suppressive microenvironment occurring during liver fibrosis contribute to Hepatocellular carcinoma (HCC) development. Every year, more than 800 Mio people die from HCC worldwide making HCC the sixth most common neoplasm and the third leading cause of cancer death [1].

Therapies for HCC are dependent on the stage of disease. In the early stages, surgery represents the standard treatment with a 5-year survival rate in 70% of treated patients. When surgery or liver transplantation are not applicable, loco-regional therapies (i.e., radiofrequency, thermal and non-thermal ablation, and transarterial chemoembolization) represent a second line of therapy, with highly variable 3–5-year survival rates [2,3]. In advanced unresectable HCC, the only approved systemic therapies are represented by the tyrosine-kinase inhibitors sorafenib and regorafenib (first and second line treatment, respectively) as well as by the inhibitor of vascular endothelial growth factor receptors 1–3 lenava-

tinib (as second line treatment). However, such systemic therapies provide only a very limited survival benefit [4–6]. In addition, the systemic chemotherapy has been reported to be unsuccessful in HCC patients because of the intrinsic chemoresistance of hepatocytes as well as the related severe toxicities [7].

Furthermore, very few clinical trials reported encouraging results of low dose metronomic chemotherapy in HCC patients, which has been shown to have anti-immunosuppressive as well as immune-stimulating effects without relevant toxicity [8–10].

In such an adverse scenario, immunotherapeutic approaches may be suitable strategies for HCC, which is considered an immunogenic cancer because it arises in chronically inflamed liver [11].

In the last years several preliminary preclinical and clinical data have supported such an intrinsic immunologic characteristic of HCC. In particular, these studies have shown in the tumor microenvironment (HCC-TME) the dual suppressive and activating role of immune cells and the chemokine axis as well as the proinflammatory cytokines release, which contribute either to tumor eradication or to tumor progression [12]. And it is exactly the balancing of these two sides of the same coin that may determine the immunotherapy efficacy and therefore the tumor progression. In this framework, immunotherapeutic interventions, including cancer vaccines, may represent a novel and effective therapeutic tool for HCC in order to counterbalance the HCC-TME

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immunosuppression as well as to improve the activation of intra-tumor effector cells.

In this review, different immunotherapy approaches and their combinations for HCC are summarized.

## 2. Tumor microenvironment in hepatocellular carcinoma

The liver is characterized by a strong intrinsic immune suppressive microenvironment which may represent a major barrier to an effective anti-tumor activity elicited by immunotherapeutic interventions.

The intrinsic immunologic composition of the liver plays a central role in host defense and even more in the maintenance of self-tolerance [13]. Several cells are involved in inducing such intra-hepatic tolerogenicity.

Liver sinusoidal endothelial cells (LSECs) represent the most potent scavenger system in the body, but have also an antigen-presenting cell (APC) function regulating the effector immune response in the liver [14]. In the physiological state, they prevent immune responses against bacterial antigens coming from the gut by inhibition of CD4<sup>+</sup> and CD8<sup>+</sup> T lymphocytes. Indeed, LSECs express high levels of the inhibitory molecule program death receptor ligand 1 (PD-L1) and low levels of the costimulatory molecules CD80 and CD86 on their surface [15,16]. Moreover, they also reduce the ability of dendritic cells (DCs) to activate T cells [17].

Similar to LSECs, Kupffer cells (KCs) promote immunological tolerance in the liver by removing from the circulation gut-derived materials as well as by producing inhibitory cytokines such as IL-10 and prostaglandins [18,19]. These cells have also a direct activation of inhibitory forkhead box P3 (FoxP3) in CD4<sup>+</sup> T cells leading to proliferation of inhibitory CD4<sup>+</sup> regulatory T cells (Tregs) [20,21].

Also the hepatic dendritic cells (HDCs) contribute to the tolerogenic microenvironment of the liver, being poor stimulators of effector CD4<sup>+</sup> T cells. Indeed, they express low levels of MHC II and co-stimulatory molecules, exhibit low endocytotic ability, produce the anti-inflammatory prostaglandin (PG) E2 which, in turn, increases IL-10 secretion and induces Tregs cells [22].

Such a physiological immune suppressive microenvironment is even more marked during the formation and progression of HCC. A progressive and persistent downregulated immune gene profile has been identified during HCC progression, which leads to a lower tumor immunity in advanced stages of disease [23]. In the same study has been shown that the liver fibrotic state favors tumor progression, representing a physical barrier made by collagens and ECM proteins to prevent CD8<sup>+</sup> CTL infiltration from peri-tumoral to intra-tumoral area. These observations suggest the possibility to use molecules able to disrupt the collagen/ECM structure in order to promote the intra-tumor infiltration by the CD8<sup>+</sup> CTL trapped in the peritumoral zone [24].

Overall, the immune suppressive microenvironment is important to induce self-tolerance in the normal liver. However, this represents a strong obstacle for the development of an anti-tumor immunity as well as for the efficacy of immune-based therapeutic strategies. Therefore, approaches to modulate such unfavorable tumor microenvironment are needed in combination with immunotherapies for HCC.

## 3. Immune evasion in hepatocellular carcinoma

In addition to the presence of cells with immune suppressive functions, the HCC TME is characterized by high expression of immune checkpoint molecules. The combination of the two leads to a markedly reduced activity of effector anti-tumor immune response and, consequently, to tumor immune evasion. Indeed, interaction between PD-1 and PD-L1 on tumor infiltrating

lymphocytes and tumor cells, respectively, contributes to T cell exhaustion, tumor-specific T-cell dysfunction and immune evasion by cancer cells. Exhausted T cells express additional inhibitory molecules directly correlating with severity of exhaustion, which may be reversed by combined PD-1/CTLA-4 blockade [25].

High expression of PD-L1 in HCC has been mainly found on Kupffer cells but also on tumor cells as well as on tumor infiltrating lymphocytes. Very often this is correlated with high PD-1 expression on CD8<sup>+</sup> T cells and is associated with higher risk of cancer recurrence or metastasis and cancer-related death [26,27]. Additional inhibitory immune checkpoint molecules have been identified in HCC and correlated with poor prognosis. High expression of T-cell immunoglobulin-and mucin-domain-containing molecule-3 (Tim-3) and lymphocyte-activation gene 3 (LAG-3) have been identified on tumor infiltrating CD8<sup>+</sup> T lymphocytes. In particular, Tim-3 is expressed only on T cells from tumor and not from the surrounding liver tissue. Moreover, it is expressed also on tumor-associated macrophages (TAM) [28].

The TIM-3 ligand Galectin-9 has been identified highly expressed on antigen presenting cells in HCC inducing T cell senescence and worse prognosis [29,30].

Similar to TIM-3, LAG-3 is expressed only on T cells from tumor and not from the surrounding liver tissue and treatment with specific blocking Abs has been shown to improve anti-tumor efficacy of anti-PD-1 blocking Abs [31].

In addition to the immune checkpoint pathway, also the chemokine axis plays a fundamental role in the tumor immune evasion, modulating the immune response in the TME and directly affecting HCC cell growth, invasion and migration properties. In particular, it has been demonstrated that in the HCC-TME the pro-inflammatory cytokines such as TNF, IFNG, and IL1 are significantly downregulated and are associated with increased levels of immunosuppressive cytokines (IL-4, IL-5, IL-8, and IL-10), thereby contributing to a higher aggressive tumor phenotype and poor prognosis [32,33]. Indeed, TNF and IFNG are involved in the activation of cytotoxic T lymphocytes to induce tumor killing, whereas elevated levels of IL4 and IL10 are associated with immune dysfunction and worse clinical outcome in cancer patients, including HCC [34–37].

## 4. Mutational landscape and neoantigen in HCC

The efficacy of specific antitumor immune response is based not only on the right balance of effectors and immunosuppressive cells in the TME, but also on the ability of malignant cells to present tumor antigens to Antigen-presenting cells (APC), which will promote the infiltration of cytotoxic T cells in tumor site and activate their antitumor response [38,39]. The latter event is strongly associated with the intra-tumoral expression levels of tumor associated antigens (TAAs) as well as mutated antigens (neoantigens) [33,40–44]. However, considering that TAAs are not tumor specific and could be a sub-optimal target for cytotoxic T cells, the best non-self immunological target is represented by real tumor-specific antigens deriving from public or personal mutations in cancer cells [45]. Indeed, infiltration of cytotoxic T cells into the TME has been reported to be directly correlated to tumor mutational burden (TMB) resulting in a better clinical response [44,46–48].

Given that the TMB is strongly related to the intra-tumor load of neoantigens, the latter represent a predictive biomarker for clinical response to immunotherapy treatment with immune checkpoint inhibitors (ICIs). Indeed, recent studies have shown that efficacy of ICIs correlates not only with the level of immune infiltration ("hot" or "cold" tumors) but also with the number of predicted neoantigens target of the infiltrating T cells [49–52].

Among others, HCC ranks as a medium variable tumor, with an average mutational burden of 5 somatic mutations per Mb, corresponding to approximately 60 non-synonymous substitutions within expressed genes, leading to generation of neoantigens targeted by tumor-infiltrating T cells [53].

Identification of naturally presented neoantigens on the surface of tumor cells by high sensitivity mass spectrometry, has proven to be much more cumbersome than the one of TAAs and needs to be improved for a broader application [54–56].

Currently, tumor neoantigens are predicted and validated through bioinformatics and experimental pipelines for which a general consensus has not been achieved yet. Indeed, a simple prediction step is not sufficient for identifying meaningful neoantigens which need to meet several parameters. The predicted affinity value to HLA molecules of neoantigen should be lower than 50 nM and greater than 10 times compared to the affinity of the corresponding wild type epitope (differential antigenicity index, DAI > 10). In addition, neoantigens should not share sequence homology with any wild type cellular self antigens, implying that a very limited fraction of cancer mutations give rise to mutated antigens which are immunologically relevant [48,57–60]. Neoantigens with such characteristics have been classified as “alternatively defined neopeptides” (ADNs) by Rech et al. [60]. Furthermore, if such neoantigens show homology with pathogen-derived epitopes, the pre-existing pathogen-specific immunity will respond faster and stronger to such neoantigens, resulting in a more efficient control of the tumor evolution and, consequently, in a better clinical outcome [58,60].

The only neoantigen discovery in HCC has been recently reported in a study by our group [48]. Neoantigens were classified as “true predicted neo-antigens” (TPNAs) only when their corresponding wt peptides showed a low prediction ranking in NetMHCStabPan. Moreover, TPNAs showed no or low sequence homology with corresponding wt peptides in the 4 aa residues of the epitope facing the TCR (p1, p4, p5, p8). Finally, when TPNAs showed homology with pathogen-derived antigens, HCC patients showed a significantly improved survival [48].

The overall results show that the quality more than the quantity of neoantigens expressed by cancer cells may predict clinical outcome as well as guide selection of the most appropriate target antigens for cancer immunotherapy.

## 5. Immunotherapy strategies for HCC

In view of the above, the immunotherapy strategies in HCC should be able to counteract the different mechanisms that underlie the HCC-TME, such as the immunosuppressive and immune evasion in TME, the effector T cell dysfunctions, the alterations in immune checkpoint molecules expression and deregulation of cytokine profiles. In this regards, therapeutic strategies based on immune modulatory strategies (e.g. chemotherapy and anti-checkpoint molecules), or active immunization with cancer vaccines, or their combination could be highly effective.

To date a limited number of immunotherapy trials for HCC have been conducted with yet modest results.

### 5.1. Immunotherapy trials based on Immune checkpoint inhibitors

To date only CTLA-4 and PD-1/PD-L1 inhibitors have been evaluated in HCC in four clinical trials. In particular, the anti-CTLA-4 Tremelimumab, was the first molecule to be clinically evaluated for safety and tumor response in HCC [61,62]. The phase II clinical trial involved 20 HCC patients infected with hepatitis C virus and not eligible for surgery or locoregional therapies, who were treated with

a suboptimal dose of the MAb. Three of the 17 evaluable patients showed partial response (PR) (17.6%) and an additional 10 patients (58.8%) were found to have stable disease (SD). Time to progression was 6.48 months and the overall survival reached 8.2 months. Decreased viral load was shown, suggesting an antiviral effect of immune checkpoint blockade and possible usefulness in patient with HCC related to viral etiology.

Subsequently, a phase I/II study by Duffy et al. described the potential of standard treatments (i.e. tumor ablation utilizing RFA and TACE) to enhance the efficacy of Tremelimumab, improving the infiltration of intratumoral effector CD8<sup>+</sup> T cells [63].

The last two clinical trials, based on anti-PD-1/PD-L1 blockade, have achieved more encouraging results.

An open-label phase I/II clinical trial has been conducted to evaluate the anti-PD-1 nivolumab in HCC patients with various etiologies irrespective of any previous treatment with sorafenib. Primary endpoints were safety, immunogenicity and antitumor activity (CheckMate 040). Complete response (CR) was reported in 3/214 (1.4%) patients and partial response (PR) in 39/214 (18.2%) patients, with a 83% overall survival (OS) at 6 months. Adverse events of grade 3–4 were observed in 25% of the study population [62]. More recently, very comparable results were observed in the KEYNOTE-224 trial which evaluated the anti-PD-1 Pembrolizumab in HCC patients who had progressed on sorafenib [64]. Complete response (CR) was reported in 1/104 (1%) patients and partial response (PR) in 17/104 (16%) patients, with a 54% overall survival (OS) at 12 months. Currently ongoing is a global phase III randomized control trial which is comparing nivolumab with sorafenib as first-line treatment in patients with advanced HCC (CheckMate 459) [ClinicalTrials.gov identifier: NCT02576509]. The overall safety profile of checkpoint inhibitors in HCC patients is manageable. Indeed, although HCC patients are very often characterized by liver dysfunction due to the underlying liver chronic disease, the substantial increase of AST and ALT induced by treatment with checkpoint inhibitors was not so relevant to cause discontinuation [65].

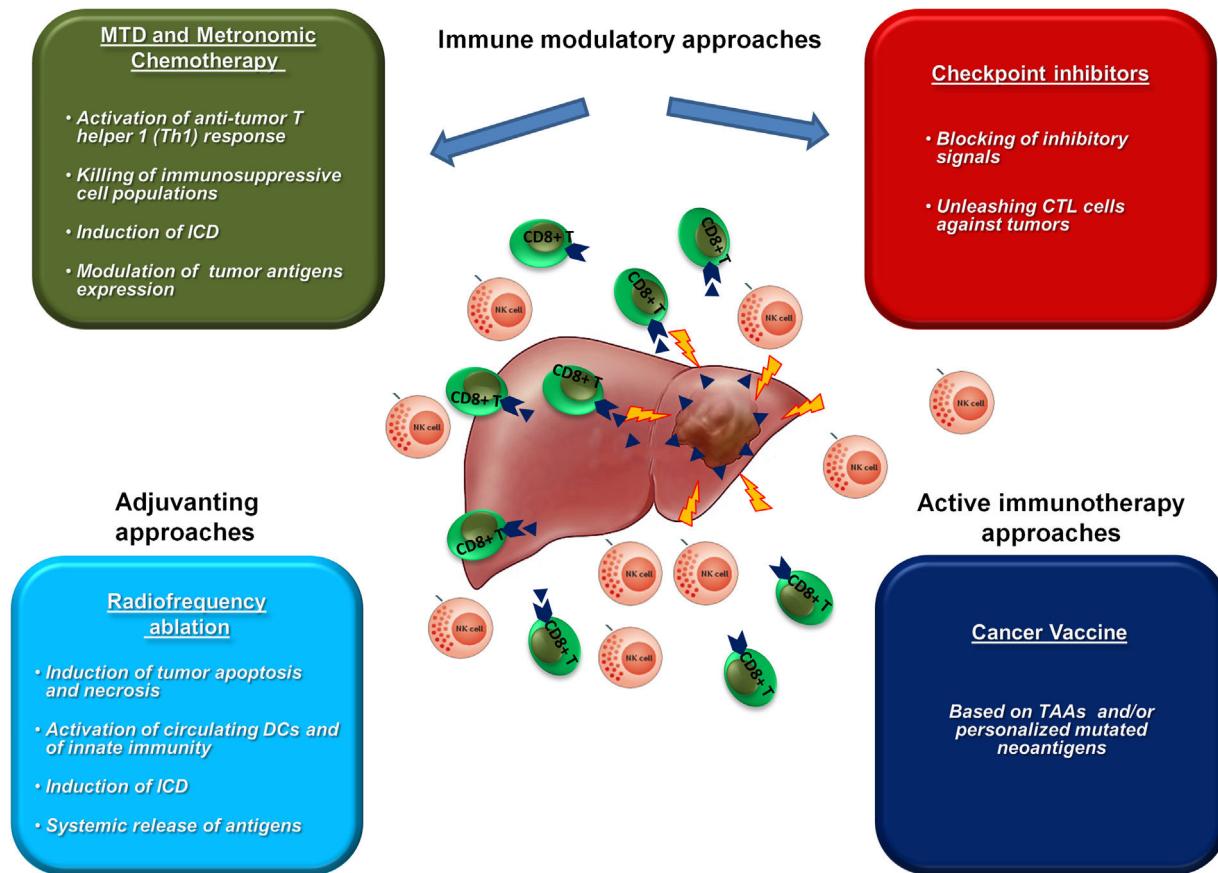
### 5.2. Active immunotherapy approaches: cancer vaccine strategies

Similar to ICIs clinical trials, also the number of cancer vaccine clinical trials in HCC are very limited showing only some degree of efficacy. A restricted number of TAAs has been identified in HCC and variable T cell response to them has been previously assessed [66]. Moreover, most of them are not specific to HCC and only few have been targeted in cancer vaccine clinical trials [67].

The first clinical trial was conducted in the early 2000', based on an alpha fetoprotein (AFP) derived peptide which induced a specific peptide T-cell response [68]. Vaccination with the same peptides presented by autologous DCs loaded ex vivo did not improve the results [69]. Subsequently, two vaccine approaches based on autologous DCs pulsed ex vivo either with a lysate of the autologous tumor or of the HepG2 cell line, have shown limited improvements in clinical outcomes [70–72].

An open label phase II clinical trial based on telomerase peptide did not lead to any complete or partial responses in advanced HCC patients [73], and similar results were reported in a phase I trial evaluating a GPC3 peptide [74].

A very innovative strategy is currently pursued for identification of shared “off-the-shelf” HCC-specific antigens within the HEPAVAC project ([www.hepavac.eu](http://www.hepavac.eu)) [75]. Novel HCC-associated antigens have been identified and a multi-epitope, multi-HLA peptide vaccine has been produced which is currently under evaluation for safety and immunogenicity in early-intermediate stage HCC patients undergoing surgical and/or loco-regional treatments (NCT03203005). The vaccination protocol will include also an



**Fig. 1.** Combinatorial approach for HCC treatment. The available immunological approaches in HCC therapy are shown. A combination of them is foreseen to produce a dramatic increase in clinical outcome in HCC patients.

actively personalized vaccine (APVAC) in a subset of vaccinees, based on patient-specific HCC neoantigens.

## 6. Immune modulatory approaches in HCC

The efficacy of immunotherapies in HCC can be significantly improved by combination with immunomodulatory approaches able to positively modulate the TME aiming at counterbalancing the strong immune suppressive setting (Fig. 1).

Chemotherapy is considered immune suppressive, however maximum tolerate dose (MTD) as well as low-dose metronomic chemotherapy may enhance antitumor immunity by several mechanisms. In particular, several studies have demonstrated not only the selective killing of immunosuppressive cell populations (e.g. MDSCs and Tregs), but also the induction of immunogenic cell death (ICD) in cancer cells with release of danger signals able to polarize DCs and activate an anti-tumor T helper 1 (Th1) response. Moreover, they can modulate the expression of tumor antigens and immune checkpoint molecules, modifying the TME and improving the efficacy of immunotherapy treatments [76–81].

Radiofrequency ablation (RFA) is considered the first line treatment option in early stage HCC patients not suitable for surgical therapies, resulting in tumor destruction by induction of tumor apoptosis and necrosis [82,83]. The release of TAAs as well as neoantigens induces significant intratumoral immune infiltrates and activation of immune response [84–87].

For the above reasons, such immunomodulatory treatments may significantly improve the efficacy of anticancer immune responses induced by cancer vaccines.

Several clinical trials have shown a better clinical outcome compared to individual treatments [88,89]. In particular, cytotoxic drugs can improve anti-tumor effects of cancer vaccines counteracting the immune-suppression, enhancing cross-presentation of tumor antigens and increasing the number of effector cells in the tumor microenvironment [90–94].

Systemic treatment with low-dose cyclophosphamide in patients with advanced HCC has been shown to be safe and to decrease the frequency and suppressor function of circulating regulatory T cells in peripheral blood, unmasking  $\alpha$ -fetoprotein-specific CD4 $^{+}$  T-cell responses [95]. However, a combination of such a treatment with a cancer vaccine based on hTERT peptide (GV1001) did not show antitumor efficacy in respect to tumor response and time-to-progression [73].

Combination of RFA and cancer vaccine has been evaluated in pre-clinical experimental settings, showing a significant enhancement of antitumor immunity with local and distal tumor regression [96,97]. A single clinical trial has shown that combination of RFA and a GPC3 peptide vaccine improved the 1-y recurrence rate in HCC patients with GPC3-positive tumors, compared to RFA alone [98].

Along the immunomodulatory effect of treatments, ICIs release the brake on the immune response, unleashing cytotoxic T cells against tumors, even those elicited by a cancer vaccine. Several pre-clinical studies have investigated combination strategies including cancer vaccines and checkpoint inhibitors, all of them showing significant enhancement of anti-tumor response associated with increased infiltration of effector CD8 $^{+}$  T cell [94,99,100].

Combination of vaccines and immune checkpoint inhibitors has been evaluated in clinical trials in different cancer settings

showing priming, expansion and boosting of an effective tumor immunotherapy [101–103]. None of such combinatorial strategies have been evaluated in HCC yet.

## 7. Conclusions

Alternative treatments, such as immunotherapies, are needed for HCC. Preliminary clinical trials with ICIs show a great potential in HCC as first and second line treatment. In addition, novel active immunotherapies (e.g. cancer vaccines) are currently developed and evaluated in clinical trials based on new TAAs as well as personalized mutated neoantigens.

Combination strategies including chemotherapy, RFA or checkpoint inhibitors together vaccines have been evaluated in several pre-clinical settings and in handful number of clinical trials. The latter strategy is predicted to be expanded in HCC in the coming years.

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## Competing interests

The authors declare no potential conflicts of interest.

## References

- [1] Forner A, Reig M, Bruix J. Hepatocellular carcinoma. Lancet 2018;391:1301–14.
- [2] Llovet JM, Real MI, Montana X, Planas R, Coll S, Aponte J, et al. Arterial embolisation or chemoembolisation versus symptomatic treatment in patients with unresectable hepatocellular carcinoma: a randomised controlled trial. Lancet 2002;359:1734–9.
- [3] Llovet JM, Bruix J. Systematic review of randomized trials for unresectable hepatocellular carcinoma: chemoembolization improves survival. Hepatology 2003;37:429–42.
- [4] Llovet JM, Ricci S, Mazzaferro V, Hilgard P, Gane E, Blanc JF, et al. Sorafenib in advanced hepatocellular carcinoma. N Engl J Med 2008;359:378–90.
- [5] Bruix J, Qin S, Merle P, Granito A, Huang YH, Bodoky G, et al. Regorafenib for patients with hepatocellular carcinoma who progressed on sorafenib treatment (RESORCE): a randomised, double-blind, placebo-controlled, phase 3 trial. Lancet 2017;389:56–66.
- [6] Kudo M, Finn RS, Qin S, Han KH, Ikeda K, Piscaglia F, et al. Lenvatinib versus sorafenib in first-line treatment of patients with unresectable hepatocellular carcinoma: a randomised phase 3 non-inferiority trial. Lancet 2018;391:1163–73.
- [7] Eatrides J, Wang E, Kothari N, Kim R. Role of systemic therapy and future directions for hepatocellular carcinoma. Cancer Control 2017;24, 1073274817729243.
- [8] Brandi G, de RF, Agostini V, di GS, Andreone P, Bolondi L, et al. Metronomic capecitabine in advanced hepatocellular carcinoma patients: a phase II study. Oncologist 2013;18:1256–7.
- [9] Ballardini P, Marri I, Margutti G, Aliberti C, Benea G, Manfredini R. Long-lasting response with metronomic capecitabine in advanced hepatocellular carcinoma. Tumori 2010;96:768–70.
- [10] Marinelli S, Granito A, Piscaglia F, Renzulli M, Stagni A, Bolondi L. Metronomic capecitabine in patients with hepatocellular carcinoma unresponsive to or ineligible for sorafenib treatment: report of two cases. Hepat Mon 2013;13, e11721.
- [11] Pardee AD, Butterfield LH. Immunotherapy of hepatocellular carcinoma: unique challenges and clinical opportunities. Oncoimmunology 2012;1:48–55.
- [12] Kapanadze T, Gamrelashvili J, Ma C, Chan C, Zhao F, Hewitt S, et al. Regulation of accumulation and function of myeloid derived suppressor cells in different murine models of hepatocellular carcinoma. J Hepatol 2013;59:1007–13.
- [13] Jenne CN, Kubes P. Immune surveillance by the liver. Nat Immunol 2013;14:996–1006.
- [14] Shetty S, Lalor PF, Adams DH. Liver sinusoidal endothelial cells – gatekeepers of hepatic immunity. Nat Rev Gastroenterol Hepatol 2018;15:555–67.
- [15] Carambia A, Frenzel C, Bruns OT, Schwinge D, Reimer R, Hohenberg H, et al. Inhibition of inflammatory CD4 T cell activity by murine liver sinusoidal endothelial cells. J Hepatol 2013;58:112–8.
- [16] Diehl L, Schurich A, Grochtmann R, Hegenbarth S, Chen L, Knolle PA. Tolerogenic maturation of liver sinusoidal endothelial cells promotes B7-homolog 1-dependent CD8+ T cell tolerance. Hepatology 2008;47:296–305.
- [17] Schildberg FA, Hegenbarth SI, Schumak B, Scholz K, Limmer A, Knolle PA. Liver sinusoidal endothelial cells veto CD8 T cell activation by antigen-presenting dendritic cells. Eur J Immunol 2008;38:957–67.
- [18] Dixon LJ, Barnes M, Tang H, Pritchard MT, Nagy LE. Kupffer cells in the liver. Compr Physiol 2013;3:785–97.
- [19] Thomson AW, Knolle PA. Antigen-presenting cell function in the tolerogenic liver environment. Nat Rev Immunol 2010;10:753–66.
- [20] You Q, Cheng L, Kedi RM, Ju C. Mechanism of T cell tolerance induction by murine hepatic Kupffer cells. Hepatology 2008;48:978–90.
- [21] Ormandy LA, Hillemann T, Wedemeyer H, Manni MP, Gretz TF, Korangy F. Increased populations of regulatory T cells in peripheral blood of patients with hepatocellular carcinoma. Cancer Res 2005;65:2457–64.
- [22] Dou L, Ono Y, Chen YF, Thomson AW, Chen XP. Hepatic dendritic cells, the tolerogenic liver environment, and liver disease. Semin Liver Dis 2018;38:170–80.
- [23] Okrah K, Tarighat S, Liu B, Koeppen H, Wagle MC, Cheng G, et al. Transcriptomic analysis of hepatocellular carcinoma reveals molecular features of disease progression and tumor immune biology. NPJ Precis Oncol 2018;2:25.
- [24] Neuzillet C, Tijeras-Raballand A, Cohen R, Cros J, Faivre S, Raymond E, et al. Targeting the TGFbeta pathway for cancer therapy. Pharmacol Ther 2015;147:22–31.
- [25] Wherry EJ, Kurachi M. Molecular and cellular insights into T cell exhaustion. Nat Rev Immunol 2015;15:486–99.
- [26] Calderaro J, Rousseau B, Amaddeo G, Mercey M, Charpy C, Costentin C, et al. Programmed death ligand 1 expression in hepatocellular carcinoma: relationship with clinical and pathological features. Hepatology 2016;64:2038–46.
- [27] Dai X, Xue J, Hu J, Yang SL, Chen GG, Lai PBS, et al. Positive expression of programmed death ligand 1 in peritumoral liver tissue is associated with poor survival after curative resection of hepatocellular carcinoma. Transl Oncol 2017;10:511–7.
- [28] Yan W, Liu X, Ma H, Zhang H, Song X, Gao L, et al. Tim-3 fosters HCC development by enhancing TGF-beta-mediated alternative activation of macrophages. Gut 2015;64:1593–604.
- [29] Li H, Wu K, Tao K, Chen L, Zheng Q, Lu X, et al. Tim-3/galectin-9 signaling pathway mediates T-cell dysfunction and predicts poor prognosis in patients with hepatitis B virus-associated hepatocellular carcinoma. Hepatology 2012;56:1342–51.
- [30] Sideras K, Biermann K, Verheij J, Takkenberg BR, Mancham S, Hansen BE, et al. PD-L1 galectin-9 and CD8(+) tumor-infiltrating lymphocytes are associated with survival in hepatocellular carcinoma. Oncoimmunology 2017;6:e1273309.
- [31] Zhou G, Sprengers D, Boor PPC, Doukas M, Schutz H, Mancham S, et al. Antibodies against immune checkpoint molecules restore functions of tumor-infiltrating T cells in hepatocellular carcinomas. Gastroenterology 2017;153:1107–19.
- [32] Budhu A, Forgues M, Ye QH, Jia HL, He P, Zanetti KA, et al. Prediction of venous metastases, recurrence, and prognosis in hepatocellular carcinoma based on a unique immune response signature of the liver microenvironment. Cancer Cell 2006;10:99–111.
- [33] Nishida N, Kudo M. Immunological microenvironment of hepatocellular carcinoma and its clinical implication. Oncology 2017;1(92 Suppl.):40–9.
- [34] Ganapathi SK, Beggs AD, Hodgson SV, Kumar D. Expression and DNA methylation of TNF, IFNG and FOXP3 in colorectal cancer and their prognostic significance. Br J Cancer 2014;111:1581–9.
- [35] Kim HD, Song GW, Park S, Jung MK, Kim MH, Kang HJ, et al. Association between expression level of PD1 by tumor-infiltrating CD8(+) T cells and features of hepatocellular carcinoma. Gastroenterology 2018;155:1936–50.
- [36] Zhao S, Wu D, Wu P, Wang Z, Huang J. Serum IL-10 predicts worse outcome in cancer patients: a meta-analysis. PLOS ONE 2015;10:e0139598.
- [37] Hattori E, Okumoto K, Adachi T, Takeda T, Ito J, Sugahara K, et al. Possible contribution of circulating interleukin-10 (IL-10) to anti-tumor immunity and prognosis in patients with unresectable hepatocellular carcinoma. Hepatol Res 2003;27:309–14.
- [38] Liu C, Lou Y, Lizée G, Qin H, Liu S, Rabinovich B, et al. Plasmacytoid dendritic cells induce NK cell-dependent, tumor antigen-specific T cell cross-priming and tumor regression in mice. J Clin Invest 2008;118:1165–75.
- [39] Chen DS, Mellman I. Oncology meets immunology: the cancer-immunity cycle. Immunity 2013;39:1–10.
- [40] Liang J, Ding T, Guo ZW, Yu XJ, Hu YZ, Zheng L, et al. Expression pattern of tumour-associated antigens in hepatocellular carcinoma: association with immune infiltration and disease progression. Br J Cancer 2013;109:1031–9.
- [41] Sideras K, Bots SJ, Biermann K, Sprengers D, Polak WG, Ijzermans JN, et al. Tumour antigen expression in hepatocellular carcinoma in a low-endemic western area. Br J Cancer 2015;112:1911–20.

- [42] Wang M, Li J, Wang L, Chen X, Zhang Z, Yue D, et al. Combined cancer testis antigens enhanced prediction accuracy for prognosis of patients with hepatocellular carcinoma. *Int J Clin Exp Pathol* 2015;8:3513–28.
- [43] Wang Z, Liu W, Chen C, Yang X, Luo Y, Zhang B. Low mutation and neoantigen burden and fewer effector tumor infiltrating lymphocytes correlate with breast cancer metastasization to lymph nodes. *Sci Rep* 2019;9:253.
- [44] Giannakis M, Mu XJ, Shukla SA, Qian ZR, Cohen O, Nishihara R, et al. Genomic correlates of immune-cell infiltrates in colorectal carcinoma. *Cell Rep* 2016;15:857–65.
- [45] Schumacher TN, Schreiber RD. Neoantigens in cancer immunotherapy. *Science* 2015;348:69–74.
- [46] Brown SD, Warren RL, Gibb EA, Martin SD, Spinelli JJ, Nelson BH, et al. Neoantigens predicted by tumor genome meta-analysis correlate with increased patient survival. *Genome Res* 2014;24:743–50.
- [47] Rooney MS, Shukla SA, Wu CJ, Getz G, Hacohen N. Molecular and genetic properties of tumors associated with local immune cytolytic activity. *Cell* 2015;160:48–61.
- [48] Petrizzo A, Tagliamonte M, Mauriello A, Costa V, Aprile M, Esposito R, et al. Unique true predicted neoantigens (TPNAs) correlates with anti-tumor immune control in HCC patients. *J Transl Med* 2018;16:286.
- [49] Kim JM, Chen DS. Immune escape to PD-L1/PD-1 blockade: seven steps to success (or failure). *Ann Oncol* 2016;27:1492–504.
- [50] Maleki VS. High and low mutational burden tumors versus immunologically hot and cold tumors and response to immune checkpoint inhibitors. *J Immunother Cancer* 2018;6:157.
- [51] Greillier L, Tomasini P, Barlesi F. The clinical utility of tumor mutational burden in non-small cell lung cancer. *Transl Lung Cancer Res* 2018;7:639–46.
- [52] Hendriks LE, Rouleau E, Besse B. Clinical utility of tumor mutational burden in patients with non-small cell lung cancer treated with immunotherapy. *Transl Lung Cancer Res* 2018;7:647–60.
- [53] Fujimoto A, Furuta M, Totoki Y, Tsunoda T, Kato M, Shiraishi Y, et al. Whole-genome mutational landscape and characterization of noncoding and structural mutations in liver cancer. *Nat Genet* 2016;48:500–9.
- [54] Bassani-Sternberg M, Braunlein E, Klar R, Engleitner T, Sinitcyn P, Audehm S, et al. Direct identification of clinically relevant neopeptides presented on native human melanoma tissue by mass spectrometry. *Nat Commun* 2016;7:13404.
- [55] Bassani-Sternberg M. Mass spectrometry based immunopeptidomics for the discovery of cancer neoantigens. *Methods Mol Biol* 2018;1719:209–21.
- [56] Bulik-Sullivan B, Busby J, Palmer CD, Davis MJ, Murphy T, Clark A, et al. Deep learning using tumor HLA peptide mass spectrometry datasets improves neoantigen identification. *Nat Biotechnol* 2018.
- [57] Duan F, Fujitama J, Al SS, Ayres CM, Corcelli SA, Pawashe AP, et al. Genomic and bioinformatic profiling of mutational neopeptides reveals new rules to predict anticancer immunogenicity. *J Exp Med* 2014;211:2231–48.
- [58] Balachandran VP, Luksz M, Zhao JN, Makarov V, Moral JA, Remark R, et al. Identification of unique neoantigen qualities in long-term survivors of pancreatic cancer. *Nature* 2017;551:512–6.
- [59] Luksz M, Riaz N, Makarov V, Balachandran VP, Hellmann MD, Solovyov A, et al. A neoantigen fitness model predicts tumour response to checkpoint blockade immunotherapy. *Nature* 2017;551:517–20.
- [60] Rech AJ, Balli D, Mantero A, Ishwaran H, Nathanson KL, Stanger BZ, et al. Tumor immunity and survival as a function of alternative neopeptides in human cancer. *Cancer Immunol Res* 2018.
- [61] Sangro B, Gomez-Martin C, de la MM, Inarraegui M, Garralda E, Barrera P, et al. A clinical trial of CTLA-4 blockade with tremelimumab in patients with hepatocellular carcinoma and chronic hepatitis C. *J Hepatol* 2013;59:81–8.
- [62] El-Khoueiry AB, Sangro B, Yau T, Crocenzi TS, Kudo M, Hsu C, et al. Nivolumab in patients with advanced hepatocellular carcinoma (CheckMate 040): an open-label, non-comparative, phase 1/2 dose escalation and expansion trial. *Lancet* 2017;389:2492–502.
- [63] Duffy AG, Ma C, Ulahannan SV, Rahma OE, Makarova-Rusher O, Cao L, et al. Phase I and preliminary phase II study of TRC105 in combination with sorafenib in hepatocellular carcinoma. *Clin Cancer Res* 2017;23:4633–41.
- [64] Zhu AX, Finn RS, Edeline J, Cattan S, Ogasawara S, Palmer D, et al. Pembrolizumab in patients with advanced hepatocellular carcinoma previously treated with sorafenib (KEYNOTE-224): a non-randomised, open-label phase 2 trial. *Lancet Oncol* 2018;19:940–52.
- [65] Brown ZJ, Heinrich B, Steinberg SM, Yu SJ, Greten TF. Safety in treatment of hepatocellular carcinoma with immune checkpoint inhibitors as compared to melanoma and non-small cell lung cancer. *J Immunother Cancer* 2017;5:93.
- [66] Mizukoshi E, Nakamoto Y, Arai K, Yamashita T, Sakai A, Sakai Y, et al. Comparative analysis of various tumor-associated antigen-specific t-cell responses in patients with hepatocellular carcinoma. *Hepatology* 2011;53:1206–16.
- [67] Buonaguro L. Developments in cancer vaccines for hepatocellular carcinoma. *Cancer Immunol Immunother* 2016;65:93–9.
- [68] Butterfield LH, Ribas A, Meng WS, Dissette VB, Amarnani S, Vu HT, et al. T-cell responses to HLA-A\*0201 immunodominant peptides derived from alpha-fetoprotein in patients with hepatocellular cancer. *Clin Cancer Res* 2003;9:5902–8.
- [69] Butterfield LH, Ribas A, Dissette VB, Lee Y, Yang JQ, De la RP, et al. A phase I/II trial testing immunization of hepatocellular carcinoma patients with dendritic cells pulsed with four alpha-fetoprotein peptides. *Clin Cancer Res* 2006;12:2817–25.
- [70] Lee WC, Wang HC, Hung CF, Huang PF, Lia CR, Chen MF. Vaccination of advanced hepatocellular carcinoma patients with tumor lysate-pulsed dendritic cells: a clinical trial. *J Immunother* 2005;28:496–504.
- [71] Palmer DH, Midgley RS, Mirza N, Torr EE, Ahmed F, Steele JC, et al. A phase II study of adoptive immunotherapy using dendritic cells pulsed with tumor lysate in patients with hepatocellular carcinoma. *Hepatology* 2009;49:124–32.
- [72] El AM, Mogawer S, Elhamid SA, Alwakil S, Aboelkasem F, Sabaawy HE, et al. Immunotherapy by autologous dendritic cell vaccine in patients with advanced HCC. *J Cancer Res Clin Oncol* 2013;139:39–48.
- [73] Greten TF, Forner A, Korangy F, N'Kontchou G, Barget N, Ayuso C, et al. A phase II open label trial evaluating safety and efficacy of a telomerase peptide vaccination in patients with advanced hepatocellular carcinoma. *BMC Cancer* 2010;10:209.
- [74] Sawada Y, Yoshikawa T, Nobuoka D, Shirakawa H, Kuronuma T, Motomura Y, et al. Phase I trial of a glycan-3-derived peptide vaccine for advanced hepatocellular carcinoma: immunologic evidence and potential for improving overall survival. *Clin Cancer Res* 2012;18:3686–96.
- [75] Buonaguro L. New vaccination strategies in liver cancer. *Cytokine Growth Factor Rev* 2017;36:125–9.
- [76] Zitvogel L, Apetoh L, Ghiringhelli F, Andre F, Tesniere A, Kroemer G. The anti-cancer immune response: indispensable for therapeutic success? *J Clin Invest* 2008;118:1991–2001.
- [77] Ghiringhelli F, Menard C, Puig PE, Ladoire S, Roux S, Martin F, et al. Metronomic cyclophosphamide regimen selectively depletes CD4+CD25+ regulatory T cells and restores T and NK effector functions in end stage cancer patients. *Cancer Immunol Immunother* 2007;56:641–8.
- [78] Ghiringhelli F, Apetoh L, Tesniere A, Aymeric L, Ma Y, Ortiz C, et al. Activation of the NLRP3 inflammasome in dendritic cells induces IL-1 $\beta$ -dependent adaptive immunity against tumors. *Nat Med* 2009;15:1170–8.
- [79] Tesniere A, Schlemmer F, Boige V, Kepp O, Martins I, Ghiringhelli F, et al. Immunogenic death of colon cancer cells treated with oxaliplatin. *Oncogene* 2010;29:482–91.
- [80] Apetoh L, Ghiringhelli F, Tesniere A, Obeid M, Ortiz C, Criollo A, et al. Toll-like receptor 4-dependent contribution of the immune system to anticancer chemotherapy and radiotherapy. *Nat Med* 2007;13:1050–9.
- [81] Lake RA, Robinson BW. Immunotherapy and chemotherapy – a practical partnership. *Nat Rev Cancer* 2005;5:397–405.
- [82] Lencioni R, Crocetti L. Local-regional treatment of hepatocellular carcinoma. *Radiology* 2012;262:43–58.
- [83] Chu KF, Dupuy DE. Thermal ablation of tumours: biological mechanisms and advances in therapy. *Nat Rev Cancer* 2014;14:199–208.
- [84] Nikfarjam M, Muralidharan V, Christophi C. Mechanisms of focal heat destruction of liver tumors. *J Surg Res* 2005;127:208–23.
- [85] Haen SP, Pereira PL, Salih HR, Rammensee HG, Gouttefangeas C. More than just tumor destruction: immunomodulation by thermal ablation of cancer. *Clin Dev Immunol* 2011;2011:160250.
- [86] Dromi SA, Walsh MP, Herby S, Traugher B, Xie J, Sharma KV, et al. Radiofrequency ablation induces antigen-presenting cell infiltration and amplification of weak tumor-induced immunity. *Radiology* 2009;251:58–66.
- [87] Zerbini A, Pilli M, Laccabue D, Pelosi G, Molinari A, Negri E, et al. Radiofrequency thermal ablation for hepatocellular carcinoma stimulates autologous NK-cell response. *Gastroenterology* 2010;138:1931–42.
- [88] Schlom J, Arlen PM, Gulley JL. Cancer vaccines: moving beyond current paradigms. *Clin Cancer Res* 2007;13:3776–82.
- [89] Wheeler CJ, Das A, Liu G, Yu JS, Black KL. Clinical responsiveness of glioblastoma multiforme to chemotherapy after vaccination. *Clin Cancer Res* 2004;10:5316–26.
- [90] Tagliamonte M, Petrizzo A, Napolitano M, Luciano A, Rea D, Barbieri A, et al. A novel multi-drug metronomic chemotherapy significantly delays tumor growth in mice. *J Transl Med* 2016;14:58.
- [91] Tagliamonte M, Petrizzo A, Napolitano M, Luciano A, Arra C, Maiolino P, et al. Novel metronomic chemotherapy and cancer vaccine combinatorial strategy for hepatocellular carcinoma in a mouse model. *Cancer Immunol Immunother* 2015;64:1305–14.
- [92] Audia S, Nicolas A, Cathelin D, Larmonier N, Ferrand C, Foucher P, et al. Increase of CD4+CD25+ regulatory T cells in the peripheral blood of patients with metastatic carcinoma: a phase I clinical trial using cyclophosphamide and immunotherapy to eliminate CD4+CD25+ T lymphocytes. *Clin Exp Immunol* 2007;150:523–30.
- [93] Laheru D, Lutz E, Burke J, Biedrzycki B, Solt S, Onners B, et al. Allogeneic granulocyte macrophage colony-stimulating factor-secreting tumor immunotherapy alone or in sequence with cyclophosphamide for metastatic pancreatic cancer: a pilot study of safety, feasibility, and immune activation. *Clin Cancer Res* 2008;14:1455–63.
- [94] Petrizzo A, Mauriello A, Luciano A, Rea D, Barbieri A, Arra C, et al. Inhibition of tumor growth by cancer vaccine combined with metronomic chemotherapy and anti-PD-1 in a pre-clinical setting. *Oncotarget* 2018;9:3576–89.
- [95] Greten TF, Ormandy LA, Fikuart A, Hochst B, Henschen S, Horning M, et al. Low-dose cyclophosphamide treatment impairs regulatory T cells and unmasks AFP-specific CD4+ T-cell responses in patients with advanced HCC. *J Immunother* 2010;33:211–8.
- [96] Gameiro SR, Higgins JP, Dreher MR, Woods DL, Reddy G, Wood BJ, et al. Combination therapy with local radiofrequency ablation and systemic vaccine enhances antitumor immunity and mediates local and distal tumor regression. *PLOS ONE* 2013;8:e70417.

- [97] Liu Q, Zhai B, Yang W, Yu LX, Dong W, He YQ, et al. Abrogation of local cancer recurrence after radiofrequency ablation by dendritic cell-based hyperthermic tumor vaccine. *Mol Ther* 2009;17:2049–57.
- [98] Sawada Y, Yoshikawa T, Ofuji K, Yoshimura M, Tsuchiya N, Takahashi M, et al. Phase II study of the GPC3-derived peptide vaccine as an adjuvant therapy for hepatocellular carcinoma patients. *Oncoimmunology* 2016;5:e1129483.
- [99] Karyampudi L, Lamichhane P, Scheid AD, Kalli KR, Shreeder B, Krempski JW, et al. Accumulation of memory precursor CD8 T cells in regressing tumors following combination therapy with vaccine and anti-PD-1 antibody. *Cancer Res* 2014;74:2974–85.
- [100] Ali OA, Lewin SA, Dranoff G, Mooney DJ. Vaccines combined with immune checkpoint antibodies promote cytotoxic T-cell activity and tumor eradication. *Cancer Immunol Res* 2016;4:95–100.
- [101] Collins JM, Redman JM, Gulley JL. Combining vaccines and immune checkpoint inhibitors to prime, expand, and facilitate effective tumor immunotherapy. *Expert Rev Vaccines* 2018;17:697–705.
- [102] Tran T, Blanc C, Granier C, Saldmann A, Tanckot C, Tartour E. Therapeutic cancer vaccine: building the future from lessons of the past. *Semin Immunopathol* 2019;41:69–85.
- [103] McArthur HL, Page F D.B. Immunotherapy for the treatment of breast cancer: checkpoint blockade, cancer vaccines, and future directions in combination immunotherapy. *Clin Adv Hematol Oncol* 2016;14:922–33.