



## Note

# Necrotizing pneumonia due to *Aspergillus* and *Salmonella* after immune checkpoint inhibitor treatment: An unusual case and review of the literature



Muhammed Cihan Işık<sup>a</sup>, Oğuz Karcıoğlu<sup>b</sup>, Gülşen Hazırolan<sup>c</sup>, Dolunay Gülmez<sup>c</sup>, Mehmet Ruhi Onur<sup>d</sup>, Mehmet Mahir Kunt<sup>e</sup>, Sevtap Arikani-Akdagli<sup>c</sup>, Gökhan Metan<sup>a,\*</sup>

<sup>a</sup> Department of Clinical Microbiology and Infectious Diseases, Hacettepe University Faculty of Medicine, Ankara, Turkey

<sup>b</sup> Department of Chest Diseases, Hacettepe University Faculty of Medicine, Ankara, Turkey

<sup>c</sup> Department of Medical Microbiology, Hacettepe University Faculty of Medicine, Ankara, Turkey

<sup>d</sup> Department of Radiology, Hacettepe University Faculty of Medicine, Ankara, Turkey

<sup>e</sup> Department of Emergency, Hacettepe University Faculty of Medicine, Ankara, Turkey

## ARTICLE INFO

### Article history:

Received 4 July 2022

Accepted 24 May 2023

Available online 14 September 2023

### Keywords:

Invasive pulmonary aspergillosis

*Salmonella* infection

Necrotizing pneumonia

Immune checkpoint inhibitors

Pembrolizumab

## ABSTRACT

**Background:** Immune checkpoint inhibitors (ICIs) are a promising new treatment for different types of cancer. The infectious complications in patients taking ICIs are rare.

**Case report:** A 58-year-old male who received chemotherapy consisting of pembrolizumab (PD-1 inhibitor) for esophagus squamous cell carcinoma one month before was admitted to the emergency room with shortness of breath soon after fiberoptic bronchoscopy, which was done for the inspection of the lower airway. A computed tomography of the chest revealed a progressive consolidation on the right upper lobe. *Salmonella* group D was isolated from the bronchoalveolar lavage (BAL) fluid culture. The fungal culture of the same clinical sample yielded *Aspergillus niger*; furthermore, a high titer (above the cut-off values) of *Aspergillus* antigen was found both in the BAL fluid and serum of the patient. Despite the effective spectrum and appropriate dose of antimicrobial treatment, the patient died due to disseminated intravascular coagulopathy.

**Conclusions:** Awareness of unusual pathogens in the etiology of pneumonia after ICI treatment may help to avoid underdiagnosis.

© 2023 Asociación Española de Micología. Published by Elsevier España, S.L.U. All rights reserved.

## Neumonía necrotizante por *Aspergillus* y *Salmonella* tras tratamiento con un inhibidor de puntos de control inmunitario: un caso inusual y revisión de la literatura

## RESUMEN

**Antecedentes:** Los fármacos inhibidores de puntos de control inmunitario (ICI) son una nueva y prometedora opción de tratamiento para diferentes tipos de cáncer. Las complicaciones infecciosas en pacientes que toman ICI son poco frecuentes.

**Caso clínico:** Un varón de 58 años que recibió quimioterapia con pembrolizumab (inhibidor de PD-1) para un carcinoma de células escamosas de esófago hacía un año, ingresó en Urgencias por dificultad respiratoria poco después de realizarse una broncoscopia de fibra óptica para una inspección de las vías aéreas inferiores. La tomografía computarizada de tórax reveló una consolidación progresiva en el lóbulo superior derecho. Se aisló *Salmonella* grupo D en el cultivo del líquido de lavado broncoalveolar (LBA). En el cultivo de hongos de la misma muestra creció *Aspergillus niger*; además, se detectó antígeno (por encima de los valores de corte) de *Aspergillus* tanto en la muestra del LBA como en el suero del paciente. A pesar del espectro eficaz y la dosis adecuada del antifúngico utilizado, el paciente falleció debido a una coagulopatía intravascular diseminada.

### Palabras clave:

Aspergilosis pulmonar invasiva

Infección por *Salmonella*

Neumonía necrotizante

Inhibidores de punto de control inmunitario

Pembrolizumab

\* Corresponding author.

E-mail address: [gokhanmetan@gmail.com](mailto:gokhanmetan@gmail.com) (G. Metan).

**Conclusiones:** El conocimiento de patógenos inusuales en la etiología de la neumonía tras el tratamiento con ICI puede ayudar a evitar el infradiagnóstico.

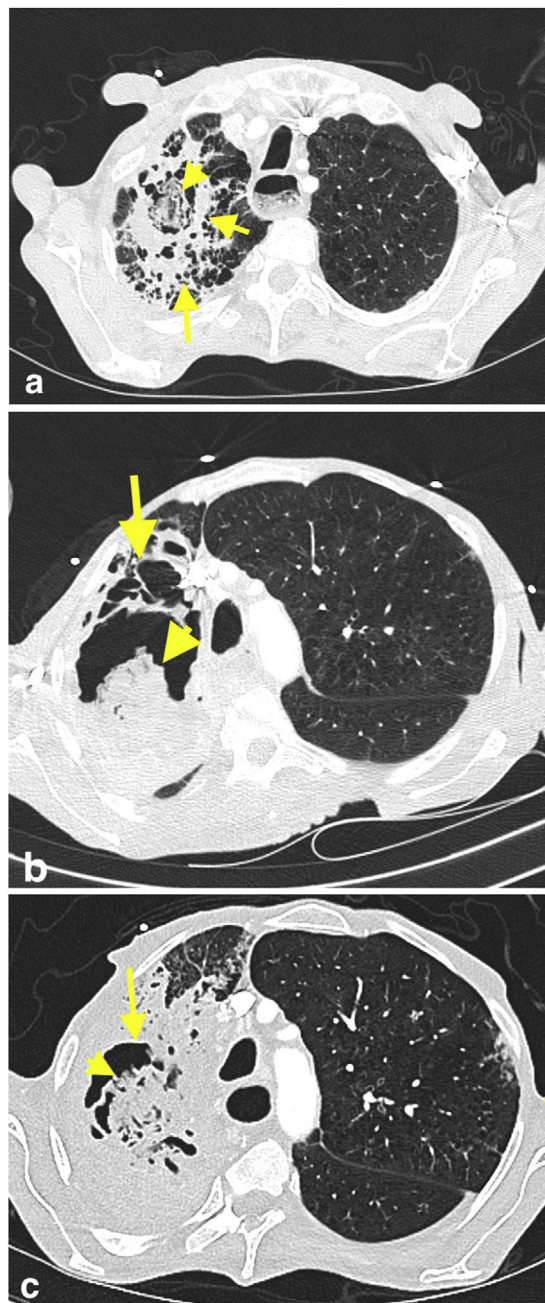
© 2023 Asociación Española de Micología. Publicado por Elsevier España, S.L.U. Todos los derechos reservados.

Cytotoxic T lymphocyte antigen 4 (CTLA-4) inhibitors, the programmed death-1 (PD-1), and programmed death-ligand (PD-L1) inhibitors are immune checkpoint inhibitors (ICIs), a promising option for treating different types of cancer.<sup>16</sup> ICIs block the apoptosis signal of cytotoxic T cells and lead to the destruction of malignant cells. Normal host tissues are exposed to autoimmune processes because of non-specific cytotoxic effects during ICIs treatment. This process is clinically named immune-related adverse events.<sup>4</sup> While ICI therapy is not considered a risk factor for opportunistic infections, the risk of uncommon infections increases when receiving immunosuppressant therapy for immune-related adverse events.<sup>1,14</sup> Herein, we report a fatal and unusual case of necrotizing pneumonia secondary to *Salmonella* and *Aspergillus* infection, after ICI therapy without a history of immunosuppressant therapy.

### Case presentation

A 58-year-old male was admitted to the emergency room (ER) with increasing shortness of breath, cough, and whitish sputum. His past medical history included esophagus squamous cell carcinoma with liver metastasis for which he started a treatment one year before with neoadjuvant chemotherapy and radiotherapy, right after an esophagectomy. The patient had received the fourth cycle of chemotherapy, consisting of m-FOLFOX (modified folinic acid, fluorouracil, and oxaliplatin), and pembrolizumab (PD-1 inhibitor) the month before, and had another ER visit due to shortness of breath and fever three weeks before the one in this case. Thoracic computed tomography (CT) revealed broad ground-glass opacities, nodular opacities, and fibrotic changes with a cavitory lesion that were not observed on thoracic CT the month before (Fig. 1a). With the diagnosis of community-acquired pneumonia, a treatment consisting of ampicillin-sulbactam (intravenous, 2 g ampicillin + 1 g sulbactam, q.i.d) and clarithromycin (oral 500 mg, b.i.d) was started; the patient had no antibiotic history in the previous three months. On the third day of the treatment, *Salmonella* group D (susceptible to ampicillin, and ciprofloxacin with a minimum inhibitory concentration (MIC) of 0.047 µg/mL) was isolated from the blood culture; antimicrobials were then changed to ciprofloxacin (intravenous, 400 mg b.i.d) and clindamycin (intravenous, 600 mg t.i.d). The culture from a blood sample obtained on the second day during the first treatment was negative. The treatment of the patient was completed in 10 days due to the clinical response and because procalcitonin concentration had decreased from 0.7 to 0.2 ng/mL. The patient was discharged, but an appointment for a fiberoptic bronchoscopy (FOB) was scheduled, since the patient could not provide a sputum sample for further examination.

When the FOB was done one week after the discharge, the patient was referred to the ER due to mild respiratory distress. Increased respiratory rate (30 breaths/min), bilateral coarse crackles, abdominal distention, and bilateral grade 3 pretibial edemas were detected at physical examination. Complete blood count showed a hemoglobin concentration of 8.7 g/dL (normal range 13.5–16.9 g/dL), a leukocyte count of 15 K/µL (normal range 4.3–10.3 K/µL) with neutrophils 85.5%, and a platelet



**Fig. 1.** Axial chest CT image obtained in patient's first visit to ER (a) shows a cavitory lesion (short arrow) that contains an oval-shaped opacity (arrowhead) in the upper lobe of the right lung. The cavitory lesion is surrounded by a thick wall and parenchymal consolidation (long arrow). Axial CT images of the patient obtained three weeks (b) and five weeks (c) after the first chest CT reveal progressive increase in size of the cavitory lesion (arrows), as well as its content with solid-like appearance (arrowheads). The consolidation around the cavitory lesion also progressed in the right upper lobe. CT findings were suggestive of fungus-ball in a cavity with surrounding consolidation.

**Table 1**

Clinical and demographic characteristics of the presented case and previously published cases of invasive aspergillosis after having immune-check point inhibitor treatment.

Author	Age and sex	Comorbidities	Immune status before the diagnosis of aspergillosis	Immune checkpoint inhibitor treatment	Diagnostic test	Microorganism	Diagnosis and treatment	Outcome
Malek et al. 2020 <sup>15</sup>	62-year-old male	Renal cell carcinoma (metastatic) Diabetes mellitus	<ul style="list-style-type: none"> <li>• Non-neutropenic</li> <li>• High dose of corticosteroids (2 mg/kg methyl-prednisolone then 60 mg prednisolone)</li> <li>• Mycophenolate mofetil</li> <li>• Rituximab for the treatment of autoimmune hepatitis associated with Nivolumab and/or Ipilimumab therapy</li> </ul>	Nivolumab plus Ipilimumab (3 cycles)	• Skin biopsy and culture	<i>Aspergillus fumigatus</i>	Fungal necrotizing skin infection, voriconazole	Improved
Gupta et al. 2019 <sup>11</sup>	63-year-old male	Lung squamous cell carcinoma COPD, hypertension, diabetes mellitus, chronic hepatitis C	<ul style="list-style-type: none"> <li>• Non-neutropenic</li> <li>• Prednisone 50 mg for 5 days for a COPD exacerbation (one month before admission)</li> </ul>	Durvalumab (4 cycles)	• Fungal cultures from the pleural fluid and decorticated lung tissue	<i>Aspergillus fumigatus</i>	Invasive aspergillosis, voriconazole	Improved
Oltolini et al. 2019 <sup>17</sup>	62-year-old female	Lung adenocarcinoma	<ul style="list-style-type: none"> <li>• Non-neutropenic</li> <li>• Dexamethasone 24 mg (&gt;2 weeks) for anti-edema therapy</li> </ul>	Pembrolizumab (2 cycles)	<ul style="list-style-type: none"> <li>• Thorax CT (macro-nodules with cavitations)</li> <li>• Galactomannan antigen from BAL fluid</li> <li>• <i>Aspergillus</i> PCR from BAL fluid</li> </ul>	Not recovered	Invasive pulmonary aspergillosis, voriconazole	Died Septicemia caused by Gram-negative bacteria
Taima et al. 2020 <sup>19</sup>	68-year-old male	Lung squamous cell carcinoma	<ul style="list-style-type: none"> <li>• Non-neutropenic</li> <li>• Prednisolone (0.5 mg/kg) for 2 weeks with suspicion of durvalumab related pneumonitis</li> </ul>	Durvalumab (2 cycles)	<ul style="list-style-type: none"> <li>• Thorax CT (cavitary lesion)</li> <li>• Culture from BAL fluid</li> </ul>	<i>Aspergillus fumigatus</i>	Invasive pulmonary aspergillosis, voriconazole followed by liposomal amphotericin B	Improved
Uchida et al. 2018 <sup>20</sup>	65-year-old male	Lung adenocarcinoma CPPA	<ul style="list-style-type: none"> <li>• Non-neutropenic</li> <li>• No immunosuppressants</li> </ul>	Nivolumab (20 cycles)	• Thorax CT (progression of a fungus ball)	Not recovered	Exacerbation of CPPA, voriconazole	Asymptomatic, stable size of the fungus ball
Inthasot et al. 2020 <sup>12</sup>	57-year-old female	Squamous cell lung cancer (locally advanced)	<ul style="list-style-type: none"> <li>• Non-neutropenic</li> <li>• No immunosuppressants</li> </ul>	Nivolumab (2 cycles)	<ul style="list-style-type: none"> <li>• Galactomannan antigen from BAL fluid</li> <li>• Thorax CT (excavated lesion)</li> </ul>	Not recovered	Invasive pulmonary aspergillosis, voriconazole	Good clinical response
Liu et al. 2020 <sup>13</sup>	55-year-old male	Lung squamous cell carcinoma Diabetes mellitus	• Not mentioned	Nivolumab (3 cycles)	<ul style="list-style-type: none"> <li>• Thorax CT (diffuse ground-glass opacities)</li> <li>• Next-generation sequencing assay of BAL fluid</li> </ul>	<i>Aspergillus fumigatus</i>	Invasive pulmonary aspergillosis, caspofungin	Improved
Uchida et al. 2022 <sup>21</sup>	70-year-old male	Malignant melanoma, diabetes mellitus, alcoholic liver disease, bronchial asthma	<ul style="list-style-type: none"> <li>• Non-neutropenic</li> <li>• Inhaled corticosteroids for chronic obstructive pulmonary disease</li> </ul>	Nivolumab (for two years)	<ul style="list-style-type: none"> <li>• Thorax CT (air-crescent sign)</li> <li>• <i>Aspergillus</i> antigen from serum <math>\geq 5.0</math></li> <li>• Diffuse pseudomembranous lesion on bronchoscopic view</li> </ul>	Not recovered	Invasive pulmonary aspergillosis, liposomal amphotericin B	Died Massive hemoptysis
Present case	58-year-old male	Esophagus squamous cell carcinoma	<ul style="list-style-type: none"> <li>• Non-neutropenic</li> <li>• No immunosuppressants</li> </ul>	Pembrolizumab (4 cycles)	<ul style="list-style-type: none"> <li>• Thorax CT (necrotizing pneumonia)</li> <li>• Galactomannan antigen from BAL fluid and serum</li> <li>• Culture from BAL fluid</li> </ul>	<i>Aspergillus niger</i>	Invasive pulmonary aspergillosis, voriconazole	Died Disseminated intravascular coagulopathy

COPD: chronic obstructive pulmonary disease; CPPA: chronic progressive pulmonary aspergillosis; CT: computed tomography; BAL: bronchoalveolar lavage; PCR: polymerase chain reaction; Nivolumab and Pembrolizumab: PD-1 inhibitors; Durvalumab: PD-L1 inhibitors; Ipilimumab: CTLA-4 inhibitors.

count of 179 K/ $\mu$ L (normal range 166–308 K/ $\mu$ L). Renal and liver function tests, lactate concentration, and arterial pH were reported in the normal range. The procalcitonin concentration was 0.852 ng/mL (normal range 0–0.1 ng/mL), and the C-reactive protein concentration was 19.23 mg/dL (normal range 0–0.5 mg/dL). Polymorphonuclear leukocytes and Gram-negative bacillus were reported on the Gram stain of bronchoalveolar lavage (BAL) fluid. The acid-fast bacterium stain was negative. Multiplex polymerase chain reaction (PCR) for respiratory infection (Bioeksen R&D Technologies Ltd, Turkey) and SARS-CoV-2 PCR were negative. Follow-up thorax CT revealed a progressed consolidation on the right upper lobe, and increased size of the cavitary lesion with solid-appearing content (Fig. 1b). Having the suspicion of necrotizing pneumonia, meropenem (intravenous, 1 g t.i.d) was started.

On day three, the growth of *Salmonella*, identified by matrix-assisted laser desorption/ionization time-of-flight mass spectrometry (MALDI-TOF MS) systems (Bruker, Germany), was reported in the BAL fluid culture; the isolate was serotyped in group D by using commercial antisera (Becton Dickinson, USA) and the Kauffmann–White scheme. The *Salmonella* isolate was subjected to an antimicrobial testing; it was resistant to ciprofloxacin, susceptible to ampicillin, ceftriaxone, and trimethoprim/sulfamethoxazole, according to European Committee on Antimicrobial Susceptibility Testing (EUCAST) guidelines.<sup>9</sup> The fungal culture of BAL fluid, which was requested as a part of an opportunistic pathogen investigation, yielded *Aspergillus niger* (MALDI-TOF MS Score 2.07). *Aspergillus* galactomannan antigen (Euroimmun Medizinische Labordiagnostika AG, Germany) optical density indices were 10.5 and 0.99 in both BAL fluid and serum.<sup>22</sup> Antifungal susceptibility testing was performed and interpreted according to EUCAST guidelines.<sup>2,6–8,10</sup> The MIC values were as follows: amphotericin B 0.5  $\mu$ g/mL (susceptible), voriconazole 0.5  $\mu$ g/mL (wild-type), posaconazole 0.25  $\mu$ g/mL (wild-type), and itraconazole 0.25  $\mu$ g/mL (wild-type).<sup>2,6–8,10</sup> With a diagnosis of invasive aspergillosis, intravenous voriconazole (6 mg/kg twice on day 1, followed by 4 mg/kg b.i.d) was added. Blood and urine cultures were negative. Results of transthoracic echocardiogram and renal ultrasonography were reported as non-pathological. The patient was intubated due to respiratory insufficiency. The antimicrobial regimen was de-escalated to ceftriaxone and clindamycin based on the results of susceptibility testing for *Salmonella*. Voriconazole concentration in serum was 3.7 mg/L (normal range 2–6 mg/L) on the third day of treatment. All the new cultures from blood, deep tracheal aspirate, urine, as well as *Mycobacterium tuberculosis* culture and PCR from BAL fluid, were negative. The patient passed away on the 20th day of admission due to disseminated intravascular coagulopathy.

## Discussion

The immune-related adverse events can affect multiple organs, including the lungs, in which potentially life-threatening pneumonitis may require rapid immunosuppressive treatment.<sup>12</sup> Opportunistic infections usually occur following immunosuppressant therapy for immune-related adverse events.<sup>1,14</sup> We found in the literature eight patients with invasive aspergillosis who were treated with ICIs<sup>11–13,15,17,19–21</sup>; four of them were receiving systemic corticosteroids, and one inhaled corticosteroids<sup>11,15,17,19,21</sup> (Table 1). Our patient did not have a history of prolonged neutropenia or any immunosuppressive therapy before the diagnosis of invasive aspergillosis. Since some cases of this mycosis induced by immunotherapy in the absence of immunosuppression have been reported,<sup>12,20,21</sup> hyperinflammatory dysregulated immune response after ICI therapy can be hypothesized as a cause of the pathogenesis in invasive aspergillosis.<sup>4,14</sup>

For the differential diagnosis, it is important to consider an extensive list of several opportunistic pathogens. Carrying out both bacterial and fungal cultures, as well as *Aspergillus* galactomannan antigen in BAL fluid, enabled, in our case, the diagnosis of an unusual coinfection. Consecutive *Aspergillus* and *Salmonella* infections were reported in two patients, one with sickle cell anemia and the other with chronic granulomatous disease.<sup>17,18</sup> In our case, there was neither history of hemoglobinopathy, nor recurrent or necrotizing infections. Pulmonary infections related to nontyphoidal *Salmonella* may present with lobar pneumonia and be complicated with lung abscess, empyema, and bronchopleural fistula formation.<sup>18</sup> A previous study showed that *Salmonella enterica* ser. *Typhimurium* can make biofilms on the hyphae of *A. niger*, while other bacteria cannot.<sup>5</sup> Moreover, a mutualistic interaction observed between *S. enterica* and *A. niger* might have favored the co-infection.<sup>3</sup> Concerning our case, we hypothesize that *Salmonella* spread to the lung by hematogenous route after the first admission of the patient, as 10 days of ciprofloxacin did not result in a microbiological cure. Then, the biofilm made by *Salmonella* might have protected *A. niger* conidia from the immune system, already impaired by ICI therapy with high-dose cytotoxic agents. This interaction might have caused an invasive co-infection. However, this hypothesis is only a speculation which requires to be proven in an animal model.

To the best of our knowledge, we report the first case of invasive *Salmonella* infection complicated by an invasive aspergillosis in a patient who received ICI treatment. Awareness of unusual pathogens in the etiology of pneumonia after an ICI treatment may help to avoid underdiagnosis.

## References

- Abers MS, Lionakis MS. Infectious complications of immune checkpoint inhibitors. *Infect Dis Clin North Am.* 2020;34:235–43.
- Arendrup MJ, Mouton JW, Lagrou K, Hamal P, Guinea J. Method for the determination of broth dilution minimum inhibitory concentrations of antifungal agents for conidia forming moulds. EUCAST antifungal MIC method for moulds [Internet]; 2020. Available from: [https://www.eucast.org/fileadmin/src/media/PDFs/EUCAST\\_files/AFST/Files/EUCAST\\_E\\_Def.9.3.2\\_Mould\\_testing\\_definitive\\_revised\\_2020.pdf](https://www.eucast.org/fileadmin/src/media/PDFs/EUCAST_files/AFST/Files/EUCAST_E_Def.9.3.2_Mould_testing_definitive_revised_2020.pdf)
- Balbotin R, Vlamakis H, Kolter R. Mutualistic interaction between *Salmonella enterica* and *Aspergillus niger* and its effects on *Zea mays* colonization. *Microb Biotechnol.* 2014;7:589–600.
- Bernardes M, Hohl TM. Fungal infections associated with the use of novel immunotherapeutic agents. *Curr Clin Microbiol Rep.* 2020;7:142–9.
- Brandl MT, Carter MQ, Parker CT, Chapman MR, Huynh S, Zhou Y. *Salmonella* biofilm formation on *Aspergillus niger* involves cellulose–chitin interactions. *PLoS One.* 2011;6:e25553.
- EUCAST. Posaconazole: rationale for the EUCAST clinical breakpoints, version 3.0. European Committee on Antimicrobial Susceptibility Testing [Internet]; 2020. Available from: [https://www.eucast.org/fileadmin/src/media/PDFs/EUCAST\\_files/Rationale\\_documents/Posaconazole\\_RD.v3.0.final.final\\_18\\_02.pdf](https://www.eucast.org/fileadmin/src/media/PDFs/EUCAST_files/Rationale_documents/Posaconazole_RD.v3.0.final.final_18_02.pdf)
- EUCAST. Breakpoint tables for interpretation of MICs for antifungal agents. European Committee on Antimicrobial Susceptibility Testing [Internet]; 2022. Available from: [https://www.eucast.org/fileadmin/src/media/PDFs/EUCAST\\_files/AFST/Clinical\\_breakpoints/AFST\\_BP.v10.0.200204.updatd.links.200924.pdf](https://www.eucast.org/fileadmin/src/media/PDFs/EUCAST_files/AFST/Clinical_breakpoints/AFST_BP.v10.0.200204.updatd.links.200924.pdf) [21.05.20].
- EUCAST. Voriconazole: rationale for the EUCAST clinical breakpoints, version 4.0. European Committee on Antimicrobial Susceptibility Testing [Internet]; 2020. Available from: [https://www.eucast.org/fileadmin/src/media/PDFs/EUCAST\\_files/Rationale\\_documents/Voriconazole\\_RD.v4.0.final\\_13\\_02.pdf](https://www.eucast.org/fileadmin/src/media/PDFs/EUCAST_files/Rationale_documents/Voriconazole_RD.v4.0.final_13_02.pdf)
- EUCAST. Breakpoint tables for interpretation of MICs and zone diameters. European Committee on Antimicrobial Susceptibility Testing [Internet]; 2021. Available from: [https://www.eucast.org/fileadmin/src/media/PDFs/EUCAST\\_files/Breakpoint\\_tables/v\\_11.0.Breakpoint.Tables.pdf](https://www.eucast.org/fileadmin/src/media/PDFs/EUCAST_files/Breakpoint_tables/v_11.0.Breakpoint.Tables.pdf)
- EUCAST. Itraconazole: rationale for the EUCAST clinical breakpoints, version 3.0. European Committee on Antimicrobial Susceptibility Testing [Internet]; 2021. Available from: [https://www.eucast.org/fileadmin/src/media/PDFs/EUCAST\\_files/Rationale\\_documents/Itraconazole\\_RD.v3.0.final.pdf](https://www.eucast.org/fileadmin/src/media/PDFs/EUCAST_files/Rationale_documents/Itraconazole_RD.v3.0.final.pdf)
- Gupta A, Tun A, Ticona K, Baqui A, Guevara E. Invasive aspergillosis in a patient with stage III (or 3a or 3b) non-small-cell lung cancer treated with durvalumab. *Case Rep Oncol Med.* 2019;2019:2178925.

12. Inthasot V, Bruyneel M, Muylle I, Ninane V. Severe pulmonary infections complicating nivolumab treatment for lung cancer: a report of two cases. *Acta Clin Belg.* 2020;75:308–10.
13. Liu Z, Liu T, Zhang X, Si X, Wang H, Zhang J, et al. Opportunistic infections complicating immunotherapy for non-small cell lung cancer. *Thorac Cancer.* 2020;11:1689–94.
14. Lu M, Zhang L, Li Y, Wang H, Guo X, Zhou J, et al. Recommendation for the diagnosis and management of immune checkpoint inhibitor related infections. *Thorac Cancer.* 2020;11:805–9.
15. Malek AE, Taremi M, Spallone A, Alvarez-Cardona JJ, Kontoyiannis DP. Necrotizing soft tissue invasive aspergillosis in a cancer patient treated with immunosuppressants due to checkpoint inhibitor-induced hepatitis. *J Infect.* 2020;80:232–54.
16. Morelli T, Fujita K, Redelman-Sidi G, Elkington PT. Infections due to dysregulated immunity: an emerging complication of cancer immunotherapy. *Thorax.* 2022;77:304–11.
17. Oltolini C, Ripa M, Andolina A, Brioschi E, Cilla M, Petrella G, et al. Invasive pulmonary aspergillosis complicated by carbapenem-resistant *Pseudomonas aeruginosa* infection during pembrolizumab immunotherapy for metastatic lung adenocarcinoma: case report and review of the literature. *Mycopathologia.* 2019;184:181–5.
18. Saeed NK. *Salmonella* pneumonia complicated with encysted empyema in an immunocompromised youth: case report and literature review. *J Infect Dev Ctries.* 2016;10:437–44.
19. Taima K, Tanaka H, Itoga M, Ishioka Y, Kurose A, Tasaka S. Destroyed lung due to sustained inflammation after chemoradiotherapy followed by durvalumab. *Respirol Case Rep.* 2020;8:e00580.
20. Uchida N, Fujita K, Nakatani K, Mio T. Acute progression of aspergillosis in a patient with lung cancer receiving nivolumab. *Respirol Case Rep.* 2018;6:e00289.
21. Uchida Y, Shimamura S, Ide S, Masuda K, Saiki M, Sogami Y, et al. A case of non-neutropenic invasive pulmonary aspergillosis under immune checkpoint inhibitor therapy for malignant melanoma. *Respir Med Case Rep.* 2022;37:101627.
22. Ullmann AJ, Aguado JM, Arikan-Akdagli S, Denning DW, Groll AH, Lagrou K, et al. Diagnosis and management of *Aspergillus* diseases: executive summary of the 2017 ESCMID-ECMM-ERS guideline. *Clin Microbiol Infect.* 2018;24:e1–38.