Original article

Cryptococcosis in an Infectious Diseases Hospital of Buenos Aires, Argentina. Revision of 2041 cases: Diagnosis, clinical features and therapeutics

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A R T I C L E I N F O

A B S T R A C T

Background: Cryptococcosis is still a life-threatening mycosis that continues to be of serious concern in Latin American countries, especially among HIV+positive population. However, there is not any reliable information about the prevalence of this disease in this region.
Aims: The aim of this study is to report data of 2041 patients with cryptococcosis that were attended at the Infectious Diseases Hospital F. J. Muñiz over a 30 year-period.
Methods: Information about demographic and clinical data, survival time and the applied treatment, was taken from the Mycology Unit database. Mycological exams from different clinical samples were performed. Cryptococcal capsular antigen in serum and cerebrospinal fluid was detected through the latex agglutination technique. *Cryptococcus* isolates were phenotypically identified and the genotype was determined in some of them. Susceptibility tests were carried out following M27-A3 document.
Results: Seventy five percent of HIV+positive patients and 50% of the HIV-negative population were males. Mean ages were 34.1 in HIV+positive patients and 44.8 in the HIV-negative. Cryptococcosis was associated with AIDS in 98% of the cases. Meningeal compromise was seen in 90% of the patients. Although cerebrospinal fluid rendered more positive results, blood culture was the first diagnostic finding in some cases. Cryptococcal antigen showed positive results in 96.2% of the sera samples and in the 93.1% of the cerebrospinal fluid samples. Most of the isolates were *Cryptococcus neoformans* and belonged to genotype VNI. Minimal inhibitory concentration values were mostly below the epidemiological cutoff values.
Conclusions: We observed that thanks to a high level of clinical suspicion, early diagnosis, combined therapy and intracranial pressure control by daily lumbar punctures, the global mortality rate has markedly decreased through the years in the analyzed period.

Criptococosis en un hospital de enfermedades infecciosas de Buenos Aires, Argentina. Revisión de 2.041 casos: aspectos diagnósticos, clínicos y terapéuticos

R E S U M E N

Antecedentes: La criptococosis es una micosis grave y un motivo de preocupación en América Latina, en especial en los pacientes positivos para el VIH. Sin embargo, no existen aún datos regionales fiables acerca de la prevalencia de la enfermedad.
Objetivos: Presentar los datos de 2.041 pacientes con criptococosis atendidos en la Unidad de Micología del Hospital de Infecciosas F. J. Muñiz de Buenos Aires, recogidos en un período de 30 años.

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Métodos: Se presentan datos demográficos, diagnósticos, clínicos y el tiempo de supervivencia de los pacientes, obtenidos de la base de datos de la Unidad de Micología. Se realizaron exámenes micológicos de diversas muestras clínicas, además de antigenemia y antigenorraquia por aglutinación de látex para *Cryptococcus* en el momento del diagnóstico y durante el seguimiento. Se llevó a cabo la identificación fenotípica de los aislamientos y en numerosos casos también se efectuó la genotipificación. La determinación de los valores de concentración mínima inhibitoria frente a diversos antifúngicos se realizó según el documento M27-A3 (CLSI).

Resultados: El 75% de los pacientes positivos para el VIH y el 50% de los no portadores eran varones; la media de edad fue 34,1 años para los positivos para el VIH y 44,8 para los no portadores. La criptococosis se asoció con el sida en el 98% de los casos y el 90% de ellos presentó compromiso meníngeo. Aunque la muestra clínica con mayor porcentaje de resultados positivos fue el LCR, en numerosas ocasiones el hemocultivo fue el primer elemento diagnóstico. La antigenemia fue positiva en el 96,2% de los casos y la antigenorraquia en el 93,1%. La mayor parte de las cepas era *Cryptococcus neoformans* y pertenecía al genotipo VNI, y la concentración mínima inhibitoria en las pruebas de sensibilidad a los antifúngicos de la mayoría de ellos mostró valores inferiores al punto de corte epidemiológico.

Conclusiones: Observamos que un alto nivel de sospecha clínica, el diagnóstico temprano, el tratamiento combinado y el control de la presión intracraneal mediante punciones lumbares diarias han permitido disminuir la mortalidad global a lo largo de los años en el período analizado.

From the onset of the AIDS' pandemic, cryptococcosis has been one of the opportunistic infections associated with this condition, being the third most frequent mycoses after oropharyngeal candidiasis and pulmonary pneumocystosis.[54] Cryptococcosis annual incidence in industrialized countries reaches 2–3% and goes higher in developing countries such as Argentina, where it is about 8–10% in HIV-positive patients requiring hospitalization. Approximately one million new cases of cryptococcal meningitis are registered annually and 650,000 of them die as a result of this mycosis.[60] Although these figures seem to have decreased slightly,[63] in Latin American countries the amount of cases continues to be of serious concern. There is not available information about the global prevalence of this disease in the region, and there is only isolated data from the national surveillance in some countries like Colombia.[30,31]

The first case of cryptococcosis associated with AIDS at the Infectious Diseases Hospital F. J. Muñiz was diagnosed in 1983. In the nineties of the last century approximately 3 new cases per week were attended, and in the last 5 years 60 new patients on average have been diagnosed yearly in this institution. At the present time, it is still the most frequent systemic mycosis in HIV-infected patients in the former hospital followed by pneumocystosis and histoplasmosis.

As cryptococcosis notification is not mandatory in Argentina there is not available information about its incidence and prevalence. According to unpublished statistical data of Buenos Aires Mycology Net, near 50% of AIDS related cryptococcosis diagnosed and treated in Buenos Aires City belong to F. J. Muñiz Hospital. The aim of this study is to show the Mycology Unit experience in the diagnosis of cryptococcosis, and clinical features, therapeutics and progress of patients attended at the F. J. Muñiz Hospital in the last 30 years.

Materials and methods

Patients

Mycology Unit database was used to obtain the information about cryptococcosis in patients diagnosed or attended at the F. J. Muñiz Hospital between January 1986 and December 2015. When available, a retrospective analysis of the demographic data, such as underlying conditions, lesion localization, clinical and images features, CD_{4+} counts, clinical samples used for diagnosis, cryptococcal polysaccharide capsular antigen titers (in serum and cerebrospinal fluid – CSF), molecular identification of the isolates and their antifungal susceptibility, treatment schemes and survival time, was carried out.

Diagnostic methods

Direct examination and cultures were made on the following clinical samples: CSF, blood, muco-cutaneous scrapings, urine, skin and other organ biopsies, lymph node and bone marrow aspirations, bronchoalveolar lavages, sputa or other bronchial secretions, peritoneal and pleural fluids, and other clinical samples (Table 3). All the samples were processed according to the standard methodology of the Mycology Unit.[5,8–10,37] Alcian blue or mucicarmin stains were used in histopathological preparations.[17,22]

Phenotypical differentiation between *Cryptococcus neoformans* and *Cryptococcus gattii* was carried out by seeding on glycine-canavanin-bromothimol blue agar (GCB) and glycine-cycloheximide-phenol red agar (Salkin medium).[43,65] Molecular identification through PCR-RFLP of URA5 gen was carried out in some of the isolates. PCR products were subjected to a double enzymatic digestion with *Sau*96I and *Hha*I, and restriction fragments were separated by electrophoresis in agarose gel and compared with the patterns obtained from the following reference strains: *C. neoformans* var. *grubii*: CBS 10085 VNI, CBS 10084 VNII; *C. neoformans* hybrid AD: CBS 10080 VNIII; *C. neoformans* var. *neoformans*: CBS 10079 VNIV; and *C. gattii*: CBS 10078 VGI, CBS 10082 VGII, CBS 10081 VGIII, and CBS 10101 VGIV.[16,51,52]

Presence and titer of capsular polysaccharide antigen (CrAg) in serum and CSF were determined by the latex agglutination technique (LA) (IMMY, Immunomycologics, Norman Kew Surrey, OK, USA) at diagnosis and during the follow up. Lateral flow chromatography (LFC) (IMMY, Immunomycologics) was just used in the last three years (209 samples) in patients with a clinical suspicion of the disease; whenever the test was positive, the titer was determined by LA, and CSF and other samples were taken to confirm the cryptococcosis. In order to determine the CrAg titer in HIV patients the following serum and CSF dilutions were used: 1:10, 1:100, 1:1000, 1:5000 and 1:10,000. In the case of HIV-negative patients the standard dilutions ($1:2^n$) were tested.[5]

Minimal inhibitory concentration (MIC) by means of the broth microdilution technique according to M27-A3 and M27S4 documents of the Clinical Laboratory Standard Institute – USA, were assessed to study the antifungal susceptibility of *Cryptococcus*

Table 1
Cases of cryptococcosis diagnosed from January 1986 to December 2015 divided into 6 quinquennia.

Quinquennium	Years included	Number of patients	% of total patients
1	1986–1990	27	1.3
2	1991–1995	357	17.5
3	1996–2000	527	25.8
4	2001–2005	374	18.3
5	2006–2010	393	19.3
6	2011–2015	363	17.8

Table 2
Mean age of HIV patients with cryptococcosis.

Quinquennium (years)	Mean age ± SD (years)
1 (1986–1990)	26.7 ± 6.0
2 (1991–1995)	30.1 ± 6.9
3 (1996–2000)	32.3 ± 7.6
4 (2001–2005)	35.4 ± 9.2
5 (2006–2010)	37.3 ± 8.9
6 (2011–2015)	38.3 ± 9.0

Table 3
Clinical samples in HIV+positive patients.

Sample	Number of samples	Positive results	
		Nr.	%
CSF			
Indian ink	1862	1411	75.8
Culture	1848	1661	89.9
Blood culture	1248	780	62.5
Urine	316	90	28.5
Broncho-alveolar lavage	66	55	83.3
Sputum	31	16	51.6
Skin scraping or biopsy	31	22	71.0
Bone marrow aspiration	14	11	78.6
Lymph node aspiration or biopsy	10	8	80.0

Other positive samples: pleural effusion (7), ascitic fluid (4), panniculitis (1), palate lesion (1), knee lesion (1), and biopsies from brain (1), lung (4), and duodenum (1).

isolates to amphotericin B (AMB) (Sigma–Aldrich, USA), fluconazole (FCZ) and voriconazole (VCZ) (Pfizer, UK), itraconazole (ITZ) (Panalab, Argentina), posaconazole (PCZ) (Schering-Plough, USA), and albaconazole (ABZ) (Uriach, Spain).[19,20] As clinical cutoff-values for *Cryptococcus* are not still determined, the epidemiological cutoffs (ECV) were used as reference.[32]

Statistical analysis

Data from continuous variables were expressed as average with its standard deviation or as median with its interquartile range as appropriate. MIC values were also presented as geometric means and ranges. The categorical variables were expressed as percentages. For continuous variables, analysis of variance or Student's *t* test were used to evaluate statistical differences; for the categorical data Z-test (for proportions) or χ^2 test were employed. The difference was considered significant when the *p*-value was less than 0.05. The statistical analysis was performed with the Statistix® 8.0 software.

Results

Patients

The data of 2041 patients were included. Two thousand individuals were HIV+positive and only 41 were HIV-negative; 1923 were attended at the F. J. Muñiz Hospital from the onset of the mycosis (1800 cases) or were received at the already mentioned institution after being treated in other hospitals (123 cases). Clinical samples from the remaining 118 cases were just referred to the Mycology Unit for diagnosis, identification of the isolate, capsular antigen detection and antifungal susceptibility test determination. The number and percentage of the patients included are presented in Table 1.

a. Characteristics of HIV-positive patients (2000 cases)

The mean age of this group of patients was 34.1 ± 8.1 years (median age: 33; range: 12–68 years), and there was not statistical difference between both sexes ($p = 0.4413$) with respect to the age. On the other hand a statistically significant increase in the patients' age along the six five-year periods considered was observed ($p = 0.0006$) (Table 2). One thousand fifty hundred and sixteen patients were males (75.8%) and 484 (24.2%) were females (rate 3:1 during the six periods). The median of the CD_{4+}

lymphocyte subset count at diagnosis of 625 patients was 35 cells/μl and the interquartiles range 15–70 cells/μl.

Diagnosis

Direct microscopy observation with Indian ink was performed on 1862 CSF sediments; mycological culture was performed in only 1848 CSF samples. Both tests were carried out in 1819 CSF samples with the following results: Indian ink and culture were positive in 1331 samples (73.2%), in 306 samples (16.8%) only the culture was positive, and in 47 samples (2.6%) Indian ink preparation was positive and the culture was negative; these samples were collected from patients previously treated in other institutions. In 135 (7.4%) both determinations were negative, especially in individuals without meningeal compromise. Considering the 6 quinquennia analyzed, the amount of CSF positive cultures only showed statistical differences between the second (94.3%) and the fifth period (84.9%) ($p = 0.0001$). A total of 1248 blood cultures were processed at diagnosis, and criptococcemia was detected in 780/1248 (62.5%) cases. Variations among the 6 quinquennia ranged from 52.8% (5th period) to 66.3%, (2nd period) ($p = 0.015$). *C. neoformans* was recovered from both CSF and blood cultures in 648/1166 (55.6%) cases in which the two samples were studied at diagnosis. Another 493 specimens from different lesions were also cultured. The performance in diagnosing cryptococcal infection of all the analyzed samples is shown in Table 3. *C. neoformans* was found in 4074/5451 (74.7%) clinical samples at the onset of the mycosis.

Isolates identification

Isolates from 1916 patients were phenotypically identified, and the causal species were *C. neoformans* in 1914 and *C. gattii* in two. One hundred and thirty two *C. neoformans* isolates were genotyped and 121 (91.7%) belonged to VNI genotype, 4 (3%) to VNII, 5 (3.8%) to VNIII and 2 (1.5%) to VNIV. Both *C. gattii* isolates were identified only phenotypically.

Capsular polysaccharide antigen (CrAg) detection in serum and CSF at diagnosis

A total of 1508 serum samples allowed us to detect CrAg by LA in 1450 (96.2%) of the patients. CrAg screening in CSF was carried out in 1756 samples and the test was positive in 1635 (93.1%) (Table 4). LFA was performed at diagnosis in 26 sera and 14 CSF samples of patients with cryptococcosis. The results were coincident with those of LA except for one CSF sample with a positive result by LFA and a negative one by LA; in another case only LFA could be done due to the high CSF protein concentration, which hindered the performance of LA test. LFA was also carried out in 112 sera and 57 CSF samples of patients without cryptococcosis, with a negative result in all the cases.

Table 4
Results of cryptococcal capsular antigen detection by latex agglutination test.

Sample	Nr.+/total (%)	CrAg[a] level	Nr. (% of total)		Median titer
		High	Medium	Low	
Serum	1450/1508 (96.2)	576 (38.2)	667 (44.2)	207 (13.7)	1:1000
CSF	1635/1756 (93.1)	321 (18.3)	927 (52.8)	387 (22.0)	1:100

Relation between mycological exams from 1688 CSF samples with their CrAg level and 1043 blood cultures with the corresponding CrAg serum level at diagnosis					
CSF		CrAg CSF level			
Culture	Indian ink	High	Medium	Low	Negative
Negative	Negative	2	10	40	66
	Positive	4	23	11	3
Positive	Negative	13	97	143	31
	Positive	296	775	169	5
Blood culture		CrAg serum level			
Negative	97	213	92	23	
Positive	354	218	39	7	

[a] CrAg: cryptococcal capsular antigen. Low titer ($\leq$1:10), medium titer (1:100 to 1:1000), high titer ($\geq$1:5000).

In order to compare the culture results and the data of CrAg titers by LA in CSF and serum, CrAg titers were divided in three categories: low ($\leq$1:10), medium (1:100 to 1:1000) and high ($\geq$1:5000). The relation between CrAg concentration in CSF and blood culture of 1688 CSF samples and 1043 blood samples are presented in Table 4. Samples with positive Indian ink and CSF culture assembled 94% (296/315) of high CrAg in CSF. On the other hand, 89.8% of negative CSF samples (Indian ink and culture) showed low level or negative CSF CrAg titers. Serum CrAg titers $\geq$1:5000 from 354/451 (78.5%) cases corresponded to patients with positive blood cultures, and 115/161 (71.4%) with low or negative CrAg serum level belonged to patients with negative blood cultures. CrAg test both in blood and CSF at diagnosis was determined in 1364 individuals. High CrAg levels were simultaneously found in serum and CSF in 166 (12.1%) patients, and 63.3% of them also had positive CSF and blood cultures ($p < 0.0001$), 23 had negative blood culture and in 32 patients a blood culture was not performed. Medium CrAg titer in CSF and serum was found in 384 patients (28.2%), and low level or negative CrAg in both fluids in 167 patients (12.2%). CrAg high titers were found in 64.7% of the sera and 39.3% of CSF in patients who died within the first week after cryptococcosis was diagnosed. Conversely, those patients who survived showed high CrAg levels only in 31.6% of the sera and 15.6% of CSF samples ($p < 0.0001$).

Clinical data

Meningeal cryptococcosis in AIDS patients was the most frequent clinical presentation (90%). Clinical information was recovered from the Mycology Unit clinical records in 1012 cases. Signs and symptoms of the patients are presented in Table 5. Fever (72.3%) and headaches (71.2%) were the most habitual symptoms. Patients suffering meningoencephalitis presented incomplete meningeal syndrome, vomiting, photophobia, visual alterations, blindness,[25] diarrhea, anorexia and asthenia. Seizures and focal signs were much less frequent. Four hundred an ten patients (40.5%) had respiratory involvement, and hepatosplenomegaly was observed in 249 (24.6%). Hepatosplenomegaly was recognized in all these patients by ultrasonography. The etiology of this clinical finding was not established, and may be due not only to the cryptococcosis but to several other diseases that these patients suffer simultaneously, including the advanced HIV infection.

The central nervous system images most frequently observed were cerebral atrophy, ventricular enlargement, space-occupying mass in brain, and cerebral edema. There were no evident lesions in the brain CT scan or even in magnetic resonance imaging in many

Table 5
Main signs and symptoms in HIV patients.

	Symptom	Number of cases (%)
General condition	Fever	732 (72.3)
	Weight loss	351 (34.7)
	Hepatosplenomegaly	249 (24.6)
	Lymphadenopathy	94 (9.3)
	Anemia	68 (6.7)
	Other manifestations	122 (11.1)
Neurological involvement	Headache	721 (71.2)
	Vomiting	331 (32.7)
	Meningeal signs	193 (19.1)
	Seizures	66 (6.5)
	Focal signs	14 (1.4)
	Other manifestations	310 (30.6)
Respiratory tract involvement	Cough	410 (40.5)
	Dyspnea	78 (7.7)
	Interstitial infiltrates	270 (26.7)
	Pleural effusion	17 (1.7)
	Other manifestations	63 (6.2)

cases. The opening pressure was informed in 185 cases and it was above the normal value in 78.9% of these patients.

Interstitial infiltrates were the most prevalent pulmonary images followed by micro nodular lesions and lung nodules; cavity images were rare. Associated diseases were registered in 490 cases, and several infections were concomitantly diagnosed. Among the mycoses, *Pneumocystis jirovecii* pneumonia was diagnosed in 69 patients and histoplasmosis in 31. Oral and esophageal candidiasis, and even dermatophytes infections, were also recorded. The high incidence of histoplasmosis can be explained by the fact that this mycosis is endemic in Buenos Aires city and its disseminated form is often observed in patients with advanced HIV infection.[23,56,60] Prior to or concurrently with cryptococcosis, tuberculosis and other mycobacterial infections were diagnosed in 51.7% (193/373). Mycobacterial infections are often detected in HIV-infected patients with low CD$_{4+}$ cell counts, especially tuberculosis which presents a high prevalence in Argentina. Other bacterial pathologies were observed in 95 cases. The most frequent viral infection was hepatitis (B and C in 116 cases). Cytomegalovirus, herpes virus, *Molluscum contagiosum* virus and JC virus infections were also diagnosed in 99 patients. Chagas disease and toxoplasmosis (47 cases) were diagnosed among other parasitic infections. Diabetes was registered in 26 cases and other non-infectious diseases in 41.

Table 6
Susceptibility test results of *C. neoformans* isolates to different antifungal drugs.

Antifungal	Genotype	Nr. isolates	Range μg/ml	MIC$_{50}$ μg/ml	MIC$_{90}$ μg/ml	GM[b] μg/ml
Amphotericin	Global	667	≤0.03–2	0.25	0.5	0.252
B	VNI	75	≤0.03–2	0.5	1.0	0.348
	VNII[a]	2	0.12			0.120
	VNIII[a]	4	0.06–0.5			0.173
Fluconazole	Global	691	0.12 to ≥64	4	8	2.72
	VNI	75	0.12–32	4	8	2.971
	VNII[a]	2	1–2			1.414
	VNIII[a]	4	2–4			2.378
Voriconazole	Global	243	≤0.03–0.25	0.06	0.06	0.046
Posaconazole		272	≤0.03–0.25	0.06	0.12	0.068
Albaconazole		271	≤0.03–0.12	0.03	0.06	0.036
Itraconazole		8[a]	≤0.03–0.5			0.112

[a] MIC$_{50}$, MIC$_{90}$ could not be determined due to the low number of isolates.
[b] GM: geometric mean.

Table 7
Survival time, and number of deceased and alive patients until the last clinical control of 2000 HIV-positive individuals diagnosed along the 6 quinquennia (*n* = 1798 patients).

Quinquennium	Dead patients Nr. (%)	Survival time mean/median (days)	Patients in follow up Nr. (%)	Follow up mean/median (days)
1	24 (88.9)	237.6/165.5	3 (11.1)	367.7/214
2	233 (65.3)	100.7/31	115 (32.2)	208.1/107
3	260 (49.3)	116.4/33.5	241 (46.0)	467.7/86
4	133 (35.6)	82.5/29	212 (56.7)	577.3/126.5
5	100 (25.4)	71.2/25.5	205 (52.1)	443.4/166
6	42 (11.6)	36.7/18.5	230 (63.4)	189.7/71.5
Total	792 (39.6)	99.8/30	1006 (50.3)	392.3/100

Fig. 1. CrAg* average titer in serum and CSF at diagnosis and during follow up.

Treatment

At the beginning of AIDS pandemic, 5-fluorocytosine (FC) was available in Argentina, and the standard treatment scheme during the first 2 quinquennia was the association of amphotericin B (AMB) with FC. As a result of FC discontinuity, treatment consisted of AMB alone for 2–3 weeks, followed by fluconazole (FCZ) by oral route. At the F. J. Muñiz Hospital, the combination of intravenous AMB (0.7 mg/kg daily) and FCZ (800 mg/day orally) at the beginning of the treatment, followed by FCZ alone for maintenance, has been the therapeutic scheme since 2010.[49]

Antifungal susceptibility of the isolates

MICs to the following antifungal drugs were determined: FCZ, AMB, ITZ, PCZ, ABZ and VCZ. The number of *C. neoformans* isolates studied for each drug along with MICs values are presented in Table 6. AMB was tested on 667 *C. neoformans*, and nearly all showed a MIC value ≤1 μg/ml, except for 4 isolates which had a value of 2 μg/ml. Susceptibility to FCZ was determined in 691 isolates. Only in 22 MIC was 16 μg/ml, in 5 cases the MIC value was 32 μg/ml and in another 6 isolates it was ≥64 μg/ml. Therefore only 1.6% of the isolates presented MICs above the ECV (95% ECV 16 μg/ml; 99% ECV 32 μg/ml).[32]

Patients follow-up parameters

A total of 1798 patients were followed at the Mycology Unit for pretty variable periods that varied between a week and more than 17 years. One hundred and four individuals passed away in less than one week after the diagnosis and some of them the same day they were hospitalized; in another 202 cases no control was done after the diagnosis. The maximum survival time of those patients who died (792 cases) was 3410 days (more than 9 years) and for those who did not die during the control period (1006 cases) was about 17.2 years. During the follow up, between 1 and 20 mycological
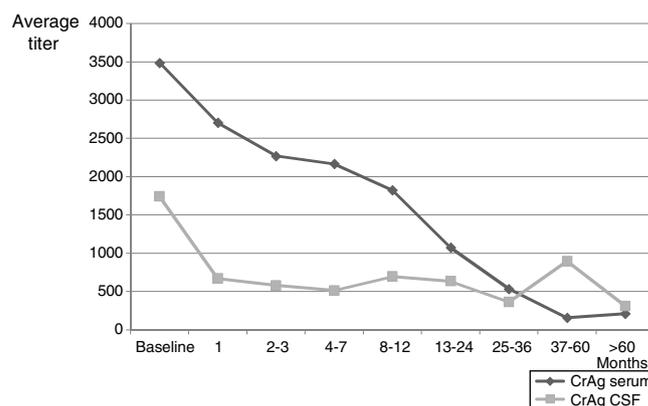
exams per patient were carried out on different samples, and *C. neoformans* grew in 292 blood cultures (from 240 patients), 1301 CSF (669 patients), 23/31 urine specimens, 6 sputa, 15 bronchoalveolar lavages, 3 lymph node punctures, 2 skin lesions, 2 hepatic biopsies, 2 pleural fluids and 4 bone marrow aspirations. Negative CSF cultures were obtained between the first and second month of treatment in many cases, even though few patients needed 4–5 months for their CSF cultures to turn negative. One of the parameters used in the follow up period was CrAg, both in serum and in CSF (when lumbar puncture was done). Mean data of CrAg through control period are shown in Fig. 1. The first antigen determination was carried out 3–4 weeks after the diagnosis and then it was repeated monthly or bimonthly during the first year and, after that, either every 3–4 months or when the patient was attended at the Mycology Unit for a clinical control. A total of 2464 serum samples and 2047 CSF samples were evaluated. CrAg serum titers were habitually equal or higher to the ones obtained in CSF. The values obtained with this last test decrease after 2–3 months, when most CSF cultures became negative (even when encapsulated yeasts were seen in Indian ink preparations).

Survival time through the six analyzed quinquennia

The mean survival time since the diagnosis of cryptococcosis in all the studied patients is presented in Table 7. The number of deceased patients in each quinquennium decreased from 88.9% in the first period to 11.6% in the last one. The survival time of those patients became shorter through the 6 periods as many deaths took place in the first weeks of the last quinquennia as shown in Fig. 2. All through the years the percentage of deaths during the first week diminished from 13.2% (2nd period) to 3.8% (6th period)
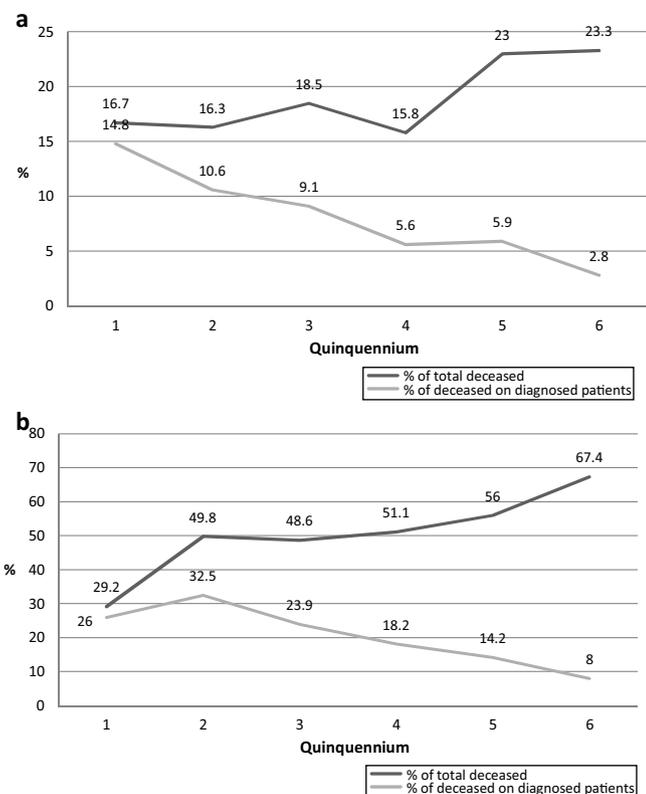
Fig. 2. Percentage of deceased patients during the first week (a) and during the first month (b) (black line). Percentage of deaths in the first week (a) and in the first month (b) among the diagnosed patients in each quinquennium (gray line).

($p = 0.0001$). However, this was not observed in the first quinquennium. This decrease was similar in the deaths of the first month (35% to 10.4%; $p < 0.0001$). Nevertheless, when the number of deaths during the first week or month was compared with the total number of dead patients of each quinquennium, an increasing proportion was observed in the first weeks (from 12.5 to 24.4%; $p = 0.3417$ in the first week and from 33.3 to 65.9% during first month; $p = 0.0195$).

b. Characteristics of HIV-negative patients

The principal characteristics of the 41 HIV-negative cases of cryptococcosis are presented in Table 8.

Among the risk factors the most frequent was the renal transplant (9 cases); two patients were under corticosteroids treatment, and there was one patient with each of the following conditions: a solitary kidney, idiopathic CD_{4+} lymphopenia with a previous disseminated histoplasmosis, a chronic pulmonary aspergillosis, diabetes, a lupus nephritis, concomitant paracoccidioidomycosis and strongyloidiasis (a badly nourished-patient infected with *C. gattii*), bullous pemphigoid, Herpes Virus infection, arterial hypertension, and contact with wood dust.

Discussion

Cryptococcosis is a systemic mycosis that continues causing a high number of infections in AIDS patients, mainly in not industrialized countries. The disease produces a great concern not only because of the clinical manifestations but also because of the high mortality associated with it. In 2009, Park estimated a million new cases yearly, mostly in sub-Saharan Africa (more than 700,000) with a mortality of about 600,000.[62,71] Cryptococcosis in Latin-American countries account for a great number of cases, but there are only estimations of the prevalence of this mycosis based on

Table 8
Characteristics of the 41 HIV-negative patients with cryptococcosis.

Demographic data		
Sex		
Male	20	Rate 0.95:1
Female	21	
Age		
Mean	44.8 years old	Range: 20–84 years old
Median	48 years old	
Clinical data		
Fever		
n = 39	35	89.7%
Neurological involvement		
Meningeal cryptococcosis	18	46.3%
Brain cryptococcoma	1	
Headache	16	
Nuchal rigidity	10	
Respiratory compromise		
Lung cryptococcoma	5	29.3%
Interstitial infiltrates	4	
Pleural effusion	1	
Nodular lesions	2	
Other lesions		
Cryptococcemia	4	9.8%
Vertebral bones	2	4.9%
Mediastinum tumor	2	4.9%
Skin lesions	2	4.9%
Diagnosis		
CSF (n = 28)		
Indian ink	17	60.7%
Culture	18	64.3%
Blood culture (n = 12)	4	33.3%
Bone (vertebrae) (n = 2)	2	
Respiratory samples (n = 12)		
Respiratory secretions	4	33.3%
Pleural fluid	1	8.3%
Lung biopsy	1	8.3%
Histopathology (n = 7)		
Lung	6	
Brain	1	
Cryptococcus species		
C. neoformans	25	
C. gattii	4	VGI 1, VGII 1, VGIII 1, not genotyped 1
C. albidus	1	
C. laurentii	2	
Not identified	9 (only histopathology in 7 samples; 2 samples from other institutions)	

Sample	Nr.+/total	CrAg LA[a] titers at diagnosis		
Serum	35/39	≤1:32 = 11	1:128 to 1:1024 = 14	≥1:4096 = 10
CSF	25/29	≤1:64 = 11	1:128 to 1:1024 = 12	≥1:4096 = 2

[a] CrAg: cryptococcal capsular polysaccharide antigen; LA: latex agglutination test.

information from research groups and not from national statistical data. In a recent epidemiological surveillance in Colombia, which covered the 1997–2014 period, a total of 1837 cases from different regions were reported, 76.9% (1413) in HIV-positive-individuals.[30] Cryptococcosis is not a notifiable disease in our country and no reliable data about the real incidence of this pathology in Argentina is available. An Argentine publication accounted for a total of 105 cases widespread in the country in 1981–1990.[6] A recent research done by the "Laboratorio Central de Redes y Programas" of Corrientes province informed about 26 detected cases between 2008 and 2013 in four general hospitals and one pediatric hospital. On the outskirts of Buenos Aires, 106 HIV-positive-patients (128 episodes) were attended between 1996 and 2007 at the Paroissien Hospital.[53]

Cryptococcosis is the most prevalent systemic mycosis in HIV+positive patients in the F.J. Muñiz Hospital. In the last years an average of 60 new cases have been diagnosed yearly, similar to those corresponding to *P. jirovecii* pneumonia and almost two fold the disseminated histoplasmosis. Almost half of the cryptococcosis diagnosis in Buenos Aires city and 15% of those cases from the whole country are made in this Infectious Diseases Hospital. These data show that this casuistic is very high compared to those of other institutions in Argentina and Colombia. The main risk factor in this cohort was the HIV infection (98%); this percentage was lower (76.9%) among the patients of a Colombian study in an 18 year-period.[30] The high percentage of HIV-positive-patients could be explained by the fact that the F.J. Muñiz Hospital is an institution devoted to infectious diseases that assists most of the aids individuals requiring hospitalization in Buenos Aires city; on the contrary, patients with other type of immunodeficiency are habitually attended in other institutions.

Male/female rate of the presented cases was 3:1, quite similar to the findings in Paroissien Hospital (70.1% men), lower than the rate obtained in Colombia on 526 cases (83.5% males), lower than in the surveillance carried out in 1997–2014 (where rates varied from 4.7:1 to 3.1:1), and higher than the cases diagnosed in Corrientes province (2.6:1) among HIV+positive patients.[15,53] The average age of the cohort studied increased through the 6 analyzed quinquennia as it happened with the HIV+positive population attending the Hospital. The mean age in the whole period was 34.1 years, with 26.7 years in the first quinquennium and reaching 38.5 years of age in the last one. In the Colombian population, 74.9% of the cases were between 21 and 50 years, and in Corrientes the median age was 38.5, the same as ours in the last quinquennium. In the cohort of Paroissien Hospital the median age was 34 years old.[15,30,31,53] No statistical difference in age was found between both sexes ($p = 0.45$). Most of the patients were in the late stages of HIV infection with a CD_{4+} lymphocyte median count of 35 cells/μl (interquartile range 15–70 cells/μl), being a population at high risk of suffering cryptococcosis. As it was expected, CSF was the sample rendering positive results in first place, followed by blood cultures, in consistence with previous findings in other investigations.[6,46,53] The increase in the proportion of positive blood cultures and decrease in CSF positive cultures seen during the 6th quinquenium is probably due to an earlier diagnosis. Direct culture of all the samples (except blood and bone marrow aspiration) onto sunflower agar favored and accelerated the identification of this microorganism (presence of melanin pigment) and allowed the differentiation from other yeasts unable to produce melanin in materials like respiratory secretions, urine, biopsies, etc. Moreover, the quick urease detection method on filter paper brought about fast information and contributed to the early diagnosis of this mycosis.[8,48]

The Mycology Unit does not have proteomic equipment (MALDI-TOF) for species identification, thus yeasts belonging to the species *C. neoformans* and *C. gattii* were phenotypically differentiated by conventional methods using GCB and Salkin medium.[43,64] All the isolates from HIV+positive patients were *C. neoformans* except for two *C. gattii* (0.1%). The last species was the causing agent in four HIV-negative patients (9.7%). Recently, the molecular identification by PCR-RFLP of URA5 gene in isolates from several periods included in this study was carried out. In this cohort 91.7% of *C. neoformans* isolates belonged to VNI genotype as it was found in other investigations. In 2003, the research performed with Spanish and Latin American environmental and clinical isolates, showed that 29/33 (87.9%) samples from HIV-positive patients from the F.J. Muñiz Hospital were VNI, 2 VNII and the remaining 2 VNIII, Considering all the Argentinean isolates, 57 (82.5%) were VNI, 8.9% VNII, and 3.6% VNIII.[51] Another investigation on *C. neoformans* isolates in Corrientes province found that 15/18 cases belonged to VNI genotype,

1 to VNII, and 2 to VNII-VNIV hybrid.[15] In the Perrando Hospital (Chaco province) 15/16 isolates from HIV-positive patients were VNI and 1 *C. gattii* VGI.[16] A global research carried out with clinical and environmental isolates in different world regions showed similar results: VNI genotype was the most prevalent all over the world and 60/87 isolates from Argentina were VNI, as 71% of Latin-American isolates.[21,33] On the other hand, a recent publication from Seville, Spain, showed that 64% of 28 isolates from 12 HIV-positive patients were *C. deneoformans* or *C. deneoformans* × *C. neoformans* hybrids.[35]

Another useful tool in the diagnose of the disease and the progress of the patients was the detection of CrAg in serum and CSF. In this study LA method was always used with the same kit (IMMY), with diagnostic and evolution control purposes. In order to compare the results it is very important using the same equipment since it has been shown that there is a marked difference in sensitivity among different commercial kits, which makes the comparison of data impossible.[29,39,67] Lateral flow chromatography (LFA) kit (IMMY) began to be commercialized in Argentina three to four years ago. There are many publications that demonstrate its great sensitivity (superior to LA) and specificity.[4,41,42,45,66,73] Currently, LFA is used in the Mycology Unit as a diagnostic tool in HIV-positive patients at risk of suffering cryptococcosis. If this test brings a positive result, the titer is determined by LA and different samples (especially CSF and blood) are further studied to microbiologically confirm the mycosis. Twenty years ago a research was carried out in order to make an early diagnosis of cryptococcosis in patients at risk (CD_{4+} counts <300 cells/μl, fever, without meningeal symptoms). It was done using LA and EIA tests on 193 patients. Seroprevalence of CrAg was 6.7% (13/193 patients) and only in three of them the cryptococcosis was microbiologically confirmed.[58] CrAg titer in serum and CSF is useful as an indirect measure of fungal load; during the follow up period the titer decrease is consistent with the patient improvement. In our study, this fact was more evident with CSF samples because low titers corresponded to negative cultures (67.4% of negative CSF showed titers ≤1:10 in the second month after diagnosis) even when Indian ink still showed encapsulated yeasts. Serum titers diminished more slowly and more time was required to obtain negative results.[3,13,29,36] This long follow up of CrAg titer in serum and CSF allowed us to observe negative results of these tests in patients after 18 or more months of secondary prophylaxis and HAART.[59]

The clinical symptoms and the location of the lesions were similar to the data presented in other publications.[34,47,54,55,64] Meningoencephalitis was the most frequent clinical form (90% of patients), and measuring opening pressure at diagnosis is a key parameter to assess the progress. Among the patients in whom this information was available, 78.9% had a high intracranial pressure, which is a sign of poor prognosis if not controlled adequately in the first days of hospitalization, as demonstrated in previous research.[11,24,27] Cerebral CT scan without contrast rarely shows pathologic findings; nevertheless brain magnetic resonance without gadolinium seems to be more sensitive and will show CNS images compatible with meningeal cryptococcosis. In a reduce number of cases they are observed as images with high signal intensities (at T2 and FLAIR), and usually correspond to enlargement of Virchow-Robin spaces, which are occupied by mucoid-protein material, affecting basal ganglia, perivascular parenchyma and, occasionally, the protuberance. Just exceptionally solid occupying lesions can be observed.[26]

Lung lesions appeared in 40% of the patients, but a great number of them also suffered other respiratory diseases (tuberculosis, atypical mycobacterial infection, *P. jirovecii* pneumonia, bacterial pneumonitis). The high frequency of respiratory involvement is due to the severe immunological compromise of this group of patients (the majority of them presented CD_{4+} cell counts ≤50/μl).[40]

Cryptococcus was isolated from 82/108 respiratory samples as not always those samples were referred to a mycological study. The sunflower medium for the primary isolation of *Cryptococcus* in this kind of samples is notably useful since it allows brown colonies to be seen even when other yeasts are present in the same sample.[48,61]

Like in most Latin-American countries, 5FC has not been available for more than 2 decades in Argentina so induction therapy consisted of AMB alone. From 2010 onwards, induction treatment has been the association of AMB (0.7 mg/kg daily, I.V.) with oral FCZ (800 mg/day), as recommended by IDSA guides. This antifungal combination was more effective than the use of AMB alone, and cultures became negative in 3–4 weeks, as it was found in a previous research. Consolidation therapy continued with FCZ (800 mg/day) and the results were very promising.[49] Another investigation associated AMB with a lower dose of FCZ (400 mg) during 2 weeks; the patients were followed up for only two months, and the results were not successful enough.[12,69] In a meta-analysis performed by Campbell et al. the authors pointed out that in 35 investigations no evidence of a decrease in the mortality rate was observed when AMB was administered together with 5FC; moreover, they did not find any additional benefit in the AMB + FCZ combination.[14] On the other hand, studies considering time and culture negativization rate found that AMB + 5FC is a more effective treatment.[63] In an attempt to elucidate if glucocorticosteroids could reduce mortality as in other types of meningitis, dexamethasone was combined with AMB + FCZ. The results were discouraging as there were more side effects, higher mortality rates and slower CSF clearance when compared with the control group.[7] The change from induction therapy to consolidation therapy was based on CSF culture results (conversion from positive to negative). Interestingly, the cryptococcal antigen concentration in CSF was coherent with the result in the CSF culture, and in many occasions Indian ink showed encapsulated yeasts despite the low titer CrAg in CSF, thus cultures resulted negative. In this study the count of colony forming units (CFU) of CSF cultures were not carried out as a parameter to monitor fungal load and early fungicidal activity like in other investigations.[63]

High intracranial pressure should be controlled, especially within the 5 first days. This parameter has been taken into account considering it was previously demonstrated that when pressure continues above normal values during the mentioned period, mortality risk increases (OR 7.23, 95% CI 2.53–20.14).[26] Serial lumbar punctures, even more than one a day, were useful to normalize the opening pressure in many patients.[28] In those cases in which this method was unable to control the opening pressure, a ventricle-peritoneal shunt was performed with good clinical results.[24]

In this study, the decrease in the mortality rate (especially evident in the last quinquennium) was probably due to the control over the intracranial pressure, the combined therapy and the early diagnosis. On the other hand, the number of resistant isolates (MIC above ECV) was scarce and the MIC values obtained were similar to those found in VNI strains from Brazil.[68] Therapeutic response and outcome were independent from MIC values, as it was seen in Colombian patients[1] but different from findings that showed a correlation between the treatment failure and MICs ≥ 16 μg/ml in Spain.[2] Nevertheless, heteroresistance should be considered in those patients with delayed negative CSF cultures.[18]

In this cohort the frequency of cryptococcosis in HIV-negative patients was very low (2%). In countries at other latitudes, like China or Iran, this condition is more frequent in HIV-negative individuals, probably due to genetic factors related to immunity (Han population in China). *C. neoformans* VNI and *C. gattii* VGI genotypes are prevalent there, in the manner of Australian isolates.[33] Most cases corresponded to renal transplanted patients, and according to some publications it is important to consider the possibility of cryptococcosis in of organ donor transmission. Transplant candidates with cirrhosis should also be considered at risk. Early diagnosis with LFC

in this group is highly recommended.[38] Another risk factor associated with female cryptococcosis is systemic lupus erythematous or other autoimmune diseases, as it was seen in 2 of our patients.[75] Other risk factors to be considered are diabetes and corticosteroids, as it was observed in this group and also in 5/6 cases in an intensive care unit in Arkansas.[44,70] This seems to be also a predisposing factor in HIV-positive individuals according to a previous study.[50]

Based on the data obtained through 30 years, it can be concluded that meningoencephalitis has been the predominant clinical form, frequently with few symptoms (headache and fever as the most important ones) in cryptococcosis. *C. neoformans* (genotype VNI) has been isolated in most of the cases. On numerous occasions the blood culture, as well as skin or respiratory cultures, made the diagnosis possible.[10,46,57,58] The use of sunflower agar in primary cultures has been of great help in these cases. Those HIV-positive individuals with CD$_{4+}$ counts <100 cells/μl are at risk for this mycosis, and CrAg detection using the FLC technique in serum can be of great help for an early diagnosis.[72] In Argentina, as in many countries, 5FC is not available and in the Muñiz Hospital the initial treatment with the AMB + FCZ combination has proved to be efficient. Along with a correct management of high intracranial pressure, especially in the first days, mortality rate diminished and results are encouraging. The failure in the treatment is probably due to a combination of causes more than an increased resistance to the antifungal drugs.[1,74]

Conflict of interest

The authors declare no conflict of interest.

References

1. Agudelo CA, Muñoz C, Ramírez A, Tobón AM, Bedout Bact C, et al. Response to therapy in patients with cryptococcosis and AIDS. Association with in vitro susceptibility to fluconazole. Rev Iberoam Micol. 2015;32:214–20.
2. Aller AI, Martin-Mazuelos E, Lozano F, Gomez-Mateos J, Steele-Moore L, Holloway WJ, et al. Correlation of fluconazole MICs with clinical outcome in cryptococcal infection. Antimicrob Agents Chemother. 2000;44:1544–8.
3. Antinori S, Radice A, Galimberti L. The role of cryptococcal antigen assay in diagnosis and monitoring of cryptococcal meningitis. J Clin Microbiol. 2005;43:5828–9.
4. Arechavala AI, Gianecini RA, Santiso GM. Evaluación de un equipo de inmunocromatografía para detección de antígeno polisacárido capsular de *Cryptococcus* en muestras clínicas. Rev Argent Infectol Dr. Francisco J. Muñiz. 2015;18:52–5.
5. Arechavala A, Robles AM, Negroni R, Bianchi M, Taborda A. Valor de los métodos directos e indirectos de diagnóstico en las micosis sistémicas asociadas al sida. Rev Inst Met trop Sao Paulo. 1993;35:163–9.
6. Bava AJ, Negroni R. Características epidemiológicas de 105 casos de criptococosis diagnosticados en la República Argentina entre 1981-1990. Rev Inst Med trop Sao Paulo. 1992;34:335–40.
7. Beardsley J, Wolkers M, Kibengo FM, Ggayi ABM, Kamali A, Cuc NTK, et al. Adjunctive dexamethasone in HIV-associated cryptococcal meningitis. N Engl J Med. 2016;374:542–54.
8. Bianchi MH, Bava A. Método rápido para la determinación de la actividad ureásica de las levaduras. Acta Bioquim Clin Latinoam. 1995;39:527–9.
9. Bianchi M, Robles AM, Vitale R, Helou S, Arechavala A, Negroni R. The usefulness of blood culture in diagnosing HIV-related systemic mycoses: evaluation of a manual lysis centrifugation method. Med Mycol. 2000;38:77–80.
10. Bianchi MH, Santiso G, Lehmann E, Walker L, Arechavala A, Maiolo E, et al. Utilidad del citodiagnóstico de Tzanck en un hospital de enfermedades infecciosas de la ciudad de Buenos Aires. Dermatol Argent. 2012;18:42–6.
11. Bicanic T, Brouwer AE, Meintjes G, Rebe K, Limmathurotsakul D, Chierakul W, et al. Relationship of cerebrospinal fluid pressure, fungal burden and outcome in patients with cryptococcal meningitis undergoing serial lumbar punctures. AIDS. 2009;23:701–6.
12. Brouwer AE, Rajanuwong A, Chierakul W, Griffin GE, Larsen RA, White NJ, et al. Combination antifungal therapies for HIV-associated cryptococcal meningitis: a randomised trial. Lancet. 2004;363:1764–7.
13. Brouwer AE, Teparrukkul P, Pinpraphaporn S, Larsen R, Chierakul W, Peacock S, et al. Baseline correlation and comparative kinetics of cerebrospinal fluid colony-forming unit counts and antigen titers in cryptococcal meningitis. J Infect Dis. 2005;192:681–4.
14. Campbell JI, Kanters S, Bennett JE, Thorlund K, Tsai AC, Mills EJ, et al. Comparative effectiveness of induction therapy for human immunodeficiency virus-associated cryptococcal meningitis: a network meta-analysis. Open Forum Infect Dis. 2015:1–11.

15. Cattana ME, Fernández MS, Rojas FD, Sosa MA, Giusiano G. Genotipos y epidemiología de aislamientos clínicos de *Cryptococcus neoformans* en Corrientes Argentina. Rev Argent Microbiol. 2015, http://dx.doi.org/10.1016/j.ram.2014.09.001.

16. Cattana ME, Tracogna MF, Fernández MS, Carol Rey MC, Sosa MA, Giusiano GE. Genotipificación de aislamientos clínicos del complejo *Cryptococcus neoformans/Cryptococcus gattii* obtenidos en el Hospital Dr. Julio C. Perrando, de la ciudad de Resistencia (Chaco, Argentina). Rev Argent Microbiol. 2013;45:89–92.

17. Chander FW, Kaplan W, Ajello L. Color atlas and text of the histopathology of mycotic diseases. Chicago: Wolfe Medical Publications Ltd; 1980. p. 54–8.

18. Cheong JWS, McCormack J. Fluconazole resistance in cryptococcal disease: emerging or intrinsic. Med Mycol. 2013;51:261–9.

19. CLSI. Clinical and Laboratory Standards Institute. Reference method for broth dilution antifungal susceptibility testing of yeasts. Approved standard. CLSI documentM27-A3. Wayne, PA: Clinical and Laboratory Standards Institute; 2008.

20. CLSI. Clinical and Laboratory Standards Institute. Reference method for broth dilution antifungal susceptibility testing of yeasts. Fourth Informational Supplement. CLSI document M27-S4. Wayne, PA: Clinical and Laboratory Standards Institute; 2012.

21. Cogliati M. Global molecular epidemiology of *Cryptococcus neoformans* and *Cryptococcus gattii*: an atlas of the molecular types. Scientifica. 2013, http://dx.doi.org/10.1155/2013/675213. Article ID 615213.

22. Colombo AC, Rodrigues ML. Fungal colonization of the brain: anatomopathological aspects of neurological cryptococcosis. An Acad Bras Cienc. 2015;87:1293–309, http://dx.doi.org/10.1590/0001-3765201520140704. PMID: 26247147.

23. Corti M, Boschi A, Villafañe MF, Messina F, Negroni R, Arechavala A, et al. Criptococosis e histoplasmosis diseminadas y simultáneas como primera manifestación de sida. Rev Patol Trop. 2013;42:459–67.

24. Corti M, Priarone M, Negroni R, Gilardi L, Castrelo J, Arechavala A, et al. Ventriculoperitoneal shunts for treating increased intracranial pressure in cryptococcal meningitis with or without ventriculomegaly. Rev Soc Bras Med Trop. 2014, http://dx.doi.org/10.1590/0037-8682-0176-2013.

25. Corti M, Solari R, Cangelosi D, Dominguez C, Yampolsky C, Negroni R, et al. Sudden blindness due to bilateral optic neuropathy associated with cryptococcal meningitis in an AIDS patient. Rev Iberoam Micol. 2010;27:207–9.

26. Corti M, Villafañe MF, Negroni R, Arechavala A, Maiolo E. Magnetic resonance imaging findings in AIDS patients with central nervous system cryptococcosis. Rev Iberoam Micol. 2008;25:211–4.

27. de Vedia L, Arechavala A, Calderón MI, Maiolo E, Rodríguez A, Lista N, et al. Relevance of intracranial hypertension control in the management of *Cryptococcus neoformans* meningitis related to AIDS. Infection. 2013;41:1013–77.

28. de Vedia L, Calderón MI, Lista N, Messina F, De Grazia N, Rodríguez A, et al. Meningitis por *Cryptococcus* asociada a sida: el control de la hipertensión endocraneana y la terapia antifúngica combinada mejoraron la sobrevida. In: Comunicación oral presentada en el XVI Congreso Argentino de Infectología, SADI 2016. 2016. Resumen No. 586. Available from: http://www.sadi.org.ar/

29. Diaz MR, Nguyen MH. Diagnostic approach based on capsular antigen, capsule detection, β-glucan, and DNA analysis. In: Heitman J, Kozel TR, Kwon-Chung KJ, Perfect JR, Casadevall A, editors. *Cryptococcus* from human pathogen to model yeast. Washington: ASM Press; 2011. p. 547–64. Cap 41.

30. Escandón P, Agudelo CI, Castañeda E. Criptococosis en Colombia. Datos de la encuesta epidemiológica sobre la criptococosis en Colombia 1997-2014. Bogotá Colombia: Instituto Nacional de Salud; 2015.

31. Escandón P, Bedout C, Lizarazo J, Agudelo CI, Tobón A, Bello S, et al. Cryptococcosis in Colombia: results of the national surveillance program for the years 2006–2010. Biomédica. 2012;32:386–98.

32. Espinel-Ingroff A, Aller A, Cantón E, Castañón-Olivares LR, Chowdhary A, Córdoba S, et al. *Cryptococcus neoformans–Cryptococcus gattii* Species Complex: an international study of wild-type susceptibility endpoint distributions and epidemiological cutoff values for fluconazole, itraconazole, posaconazole and voriconazole. Antimicrob Agents Chemother. 2012;56:5898–906.

33. Fang W, Fa Z, Liao W. Epidemiology of *Cryptococcus* and cryptococcosis in China. Fungal Genet Biol. 2014, http://dx.doi.org/10.1016/jfgb2014.10.017.

34. Fries B, Cox GM. Cryptococcosis in AIDS. In: Heitman J, Kozel TR, Kwon-Chung KJ, Perfect J, Casadevall A, editors. *Cryptococcus* from human pathogen to model yeast. Washington: ASM Press; 2011. p. 515–25. Cap 38.

35. Gago S, Serrano C, Alastruey-Izquierdo A, Cuesta I, Martín Mazuelos E, Aller AI, et al. Molecular identification, antifungal resistance and virulence of *Cryptococcus neoformans* and *Cryptococcus deneoformans* isolated in Seville, Spain. Mycoses. 2016, http://dx.doi.org/10.1111/myc.12543.

36. Grinsell M, Weinhold LC, Cutler JE, Han Y, Kozel TR. In vivo clearance of glucuronoxylomannan, the major capsular polysaccharide of *Cryptococcus neoformans*: a critical role for tissue macrophages. J Infect Dis. 2001;184:479–87.

37. Guelfand L, Cataldi S, Arechavala A, Perrone M. Manual práctico de micología médica. Acta Bioquim Clin Latinoam. 2015; Suppl. 1.

38. Haidar G, Singh N. *Cryptococcus* shedding new light on an inveterate yeast. J Fungi. 2015;1:115–29.

39. Hansen J, Slechta S, Gates-Hollingsworth MA, Neary B, Barker AP, Bauman S, et al. Large-scale evaluation of the immuno-mycologics lateral flow and enzyme-linked immunoassays for detection of cryptococcal antigen in serum and cerebrospinal fluid. Clin Vaccine Immunol. 2013;20:52–5.

40. Helou S, Robles AM, Arechavala A, Bianchi M, Negroni R. Criptococcosis respiratoria en pacientes VIH positivos. Rev Iberoam Micol. 1999;16:126–9.

41. Huang HR, Fan LC, Rajbanshi B, Xu JF. Evaluation of a new cryptococcal antigen ateral flow immunoassay in serum, cerebrospinal fluid and urine for the diagnosis of cryptococcosis: a meta-analysis and systematic review. PLoS ONE. 2015;10:e0127117, http://dx.doi.org/10.1371/journal.pone0127117.

42. Kabanda T, Siedner MJ, Klausner JD, Muzoora C, Boulware DR. Point-of-care diagnosis and prognostication of cryptococcal meningitis with the cryptococcal antigen flow assay on cerebrospinal fluid. Clin Infect Dis. 2014;58:113–6.

43. Kwon-Chung KJ, Polacheck I, Bennett JE. Improved diagnostic medium for separation of *Cryptococcus neoformans* var. *neoformans* (serotypes A and D) and *Cryptococcus neoformans* var. *gattii* (serotypes B and C). J Clin Microbiol. 1982;15:535–7.

44. Lin KH, Chen CM, Chen LT, Kuo SC, Kao CC, Jeng YC, et al. Diabetes mellitus is associated with acquisition and increased mortality in HIV-uninfected patients with cryptococcosis: a population-based study. J Infect. 2016;72: 608–14.

45. Lindsley MD, Mekha N, Baggett HC, Surinthong Y, Autthateinchai R, Sawatwong P, et al. Evaluation of a newly developed lateral flow immunoassay for the diagnosis of cryptococcosis. Clin Infect Dis. 2011;53:321–5.

46. López Moral L, Tiraboschi IN, Schijman M, Bianchi M, Guelfand L, Cataldi S. Fungemias en hospitales de la ciudad de Buenos Aires, Argentina. Rev Iberoam Micol. 2012;29:144–9.

47. Maziars EK, Perfect JR. Cryptococcosis. Infect Dis Clin N Am. 2016;30:179–206.

48. McTaggart L, Richardson SE, Seah C, Hoang L, Fothergill A, Zhang SX. Rapid identification of *Cryptococcus neoformans* var. *grubii C. neoformans* var. *neoformans*, and *C. gattii* by use of rapid biochemical tests, differential media and DNA sequencing. J Clin Microbiol. 2011;49:2522–7.

49. Messina F, Maiolo E, Negroni R, Arechavala A, Santiso G, Bianchi M. Alternativas terapéuticas de la criptococosis meníngea. Actual Infectol. 2015;23:25–32.

50. Messina FA, Negroni R, Maiolo EI, Arechavala A, Villafañe MF, Santiso G, et al. Criptococosis meníngea en pacientes con diabetes y sida. Enferm Infecc Microbiol Clin. 2014;32:643–6.

51. Meyer W, Castañeda A, Jackson S, Huynh M, Castañeda E, Iberoamerican Cryptococcal Study Group. Molecular typing of IberoAmerican *Cryptococcus neoformans* isolates. Em Infect Dis. 2003;9:189–95.

52. Meyer W, Trilles L. Genotyping of the *Cryptococcus neoformans/Cryptococcus gattii* species complex. Aust Biochem. 2010;41:11–5.

53. Mónaco LS, Tamayo Antabak N. Cryptococosis en pacientes con SIDA: estudio de casos en el Hospital Paoissien en el período 1996-2007. Rev Argent Microbiol. 2008;40:218–21.

54. Negroni R. Criptococosis. In: Benetucci J, editor. Sida y Enfermedades Asociadas. Diagnóstico, Clínica y Tratamiento. 3rd ed. Buenos Aires: FUNDAI; 2008. p. 332–6.

55. Negroni R. Cryptococcosis. Clin Dermatol. 2012;30:599–609.

56. Negroni R, Arechavala A, Maiolo E. Histoplasmosis clásica en pacientes inmunocomprometidos. Med Cutan Ibero Lat Am. 2010;38:59–69.

57. Negroni R, Arechavala A, Santiso G, Bonvehí P. Problemas clínicos en micología médica: problema número 46. Rev Iberoam Micol. 2014;31:207–9.

58. Negroni R, Cendoya C, Arechavala A, Robles AM, Bianchi M, Bava AJ, et al. Detection of *Cryptococcus neoformans* capsular polysaccharide antigen in asymptomatic HIV-infected patients. Rev Inst Med Trop Sao Paulo. 1995;37:385–9.

59. Negroni R, Helou SH, López Daneri G, Robles AM, Arechavala A, Bianchi MH. Interrupción exitosa de la profilaxis secundaria antifúngica en la criptococosis asociada al sida. Rev Argent Microbiol. 2004;36:113–7.

60. Negroni R, Helou SH, López Daneri G, Robles AM, Arechavala AI, Bianchi MH. Interrupción de la profilaxis secundaria antifúngica en la histoplasmosis asociada al sida. Rev Iberoam Micol. 2004;21:75–8.

61. Negroni R, Lloveras S, Arechavala A, Maiolo E, Bianchi M, Santiso G, et al. Problemas clínicos en micología médica: problema número 43. Rev Iberoam Micol. 2012;29:178–80.

62. Park BJ, Wannemuehler KA, Martson BJ, Govender N, Pappas PG, Chiller TM. Estimation of the current global burden of cryptococcal meningitis among persons living with HIV/AIDS. AIDS. 2009;23:525–30.

63. Perfect JR, Bicanic T. Cryptococcosis diagnosis and treatment: what do we know now. Fungal Genet Biol. 2015;78:49–54.

64. Quindós Andres G, Colom Valiente MF, Abarca Salat ML, Arechavala Silva A, Arévalo Morales MP, Calderón Sandubete EJ, et al. In: Quindós Andrés, editor. Criptococosis y otras micosis causadas por levaduras. Micología clínica. Barcelona: Elsevier España; 2015. p. 109–28.

65. Salkin IF, Hurd N. New medium for differentiation of *Cryptococcus neoformans* serotype pairs. J Clin Microbiol. 1982;15:169–71.

66. Tang MW, Clemons KV, Katzenstein DA, Stevens DA. The cryptococcal antigen lateral flow assay: a point-of-care diagnostic at an opportune time. Crit Rev Microbiol. 2016;42:634–42.

67. Tiltenot L, Hagen F, Han CK, Seibold M, Rickerts V, Boekhout T. Pitfalls in serological diagnosis of *Cryptococcus gattii* infections. Med Mycol. 2015;53:874–9.

68. Trilles L, Meyer W, Wanke B, Guarro J, Lázéra M. Correlation of antifungal susceptibility and molecular type within the *Cryptococcus neoformans/C. gattii* species complex. Med Mycol. 2012;50:328–32.

69. Vaidhya SA, Gupta BB, Jha RK, Kumar R. Combination versus monotherapy for the treatment of HIV associated cryptococcal meningitis. J Clin Diagn Res. 2015;9:OC14–6.

70. Vallabhaneni S, Haselow D, Lloyd S, Lockhart S, Moulton-Meissner H, Lester L, et al. Cluster of *Cryptococcus neoformans* infections in intensive care unit, Arkansas, USA, 2013. Emerg Infect Dis. 2015;21:1719–24.

71. Vallabhaneni S, Mody RK, Walker T, Chiller T. The global burden of fungal diseases. Infect Clin Dis N Am. 2016;30:1–11.

72. Vidal JE, Boulware DR. Lateral flow assay for cryptococcal antigen: an important advance to improve the continuum of HIV care and reduce cryptococcal meningitis-related mortality. Rev Inst Med Trop Sao Paulo. 2015;57 Suppl. 19:38–45.

73. Williams DA, Kiiza T, Kwizera R, Kiggundu R, Velamakanni S, Meya DB, et al. Evaluation of fingerstick cryptococcal antigen lateral flow assay in HIV-infected persons: a diagnostic accuracy study. Clin Infect Dis. 2015;61:464–7.

74. Zhang M, Sun D, Shi M. Dancing cheek to cheek: *Cryptococcus neoformans* and phagocytes. SpringerPlus. 2015;4:410–8, http://dx.doi.org/10.1186/s40064-015-1192-3.

75. Zheng H, Li M, Wang D, Yang JI, Chen Q, Zhang X, et al. Gender-specific contributing risk factors and outcome of female cryptococcal meningoencephalitis patients. BMC Infect Dis. 2016;16:22, http://dx.doi.org/10.1186/s12879-016-1363-z.