



Note

Candidaemia due to *Candida parapsilosis* species complex at a hospital in Brazil: Clinical characteristics and antifungal susceptibility profile



Débora de Souza Olartechea de Alencar^a, Rosianne Assis de Sousa Tsujisaki^a, Fernanda Luiza Espinosa Spositto^a, Maína de Oliveira Nunes^a, Adriana Araújo de Almeida^a, Marilena dos Anjos Martins^b, Marcia de Souza Carvalho Melhem^b, Marilene Rodrigues Chang^{a,*}

^a Laboratório de Pesquisas Microbiológicas, Universidade Federal de Mato Grosso do Sul, Campo Grande, MS, Brazil

^b Divisão de Micologia, Instituto Adolfo Lutz, São Paulo, SP, Brazil

ARTICLE INFO

Article history:

Received 11 July 2015

Accepted 24 June 2016

Available online 15 February 2017

Keywords:

Candidaemia

Candida parapsilosis

Candida orthopsilosis

Antifungal agents

ABSTRACT

Background: Recent decades have seen a global emergence of candidaemia caused by non-*Candida albicans* *Candida* species, particularly the *Candida parapsilosis* complex.

Aims: To evaluate the clinical features and antifungal susceptibility profiles of isolates belonging to the *C. parapsilosis* species complex in patients with candidaemia in a midwestern Brazilian tertiary-care teaching hospital.

Methods: Yeast identification was performed using an automated Vitek 2 Compact system. PCR-RFLP was employed for species differentiation.

Results: Five cases of infection by *C. parapsilosis sensu stricto* and two by *Candida orthopsilosis* were found. Of the seven cases, five were adult patients undergoing haemodialysis. The only isolate of *C. parapsilosis sensu stricto* resistant to fluconazole ($MIC = 8 \mu\text{g/ml}$) was obtained from a patient on a long-term regimen with this drug. This was the only patient who evolved to death.

Conclusions: Resistance to antifungal agents poses a therapeutic challenge, especially for non-*C. albicans* *Candida* species, and requires continuous monitoring using susceptibility tests because resistance *in vitro* can be predictive of treatment failure. In the present study, *in vitro* antifungal susceptibility proved consistent with clinical outcome.

© 2016 Asociación Española de Micología. Published by Elsevier España, S.L.U. All rights reserved.

Candidemia por especies del complejo *Candida parapsilosis* en un hospital de Brasil: características clínicas y perfil de sensibilidad a los antifúngicos

RESUMEN

Palabras clave:

Candidemia

Candida parapsilosis

Candida orthopsilosis

Agentes antifúngicos

Antecedentes: En las últimas décadas se ha visto un surgimiento mundial de la candidemia causada por especies de *Candida* no-*C. albicans*, en particular del complejo *Candida parapsilosis*.

Objetivos: Evaluar las características clínicas y los perfiles de sensibilidad antifúngica en aquellos aislamientos del complejo de especies *C. parapsilosis* responsables de candidemia en un hospital universitario de tercer nivel en la región centro-oeste de Brasil.

Métodos: La identificación se realizó en un sistema automatizado Vitek 2 compact. Se utilizó PCR-RFLP para la diferenciación de las especies.

Resultados: Se encontraron cinco casos de candidemia por *C. parapsilosis sensu stricto* y dos por *Candida orthopsilosis*. Cinco eran pacientes adultos sometidos a hemodiálisis. El único aislamiento de *Candida parapsilosis sensu stricto* resistente a fluconazol ($CIM, 8 \mu\text{g/ml}$) se obtuvo de un paciente en régimen largo de tratamiento con este antifúngico. Este fue el único paciente que murió.

* Corresponding author.

E-mail address: marichang@yahoo.com.br (M.R. Chang).

Conclusiones: La resistencia a los antifúngicos constituye un desafío terapéutico, en especial contra las especies de *Candida* no-*C. albicans*, que requieren la monitorización continua por medio de pruebas de sensibilidad en vista de que la resistencia *in vitro* puede ser predictiva de fracaso del tratamiento. En el presente estudio la sensibilidad antifúngica *in vitro* resultó consistente con el curso clínico.

© 2016 Asociación Española de Micología. Publicado por Elsevier España, S.L.U. Todos los derechos reservados.

Recent decades have seen a global emergence of candidaemia caused by non-*Candida albicans* *Candida* species, particularly of the *Candida parapsilosis* complex.^{2,6,15} Determination of antifungal susceptibility is important to ensure effective monitoring and detection of resistance, contributing to appropriate treatment selection.¹³

The purposes of this study were to evaluate the clinical features of patients with candidaemia caused by species of the *C. parapsilosis* complex and to determine the antifungal susceptibility profiles of these agents.

The yeasts were isolated from a tertiary-care teaching hospital located in Campo Grande, Mato Grosso do Sul State, Midwest Brazil, from March 2010 to March 2012. The isolates were detected in blood culture using an automated Bact/Alert system and identified employing an automated Vitek 2 Compact system (both from bioMérieux, France).

Species differentiation was performed by polymerase chain reaction – restriction fragment length polymorphism (PCR-RFLP). DNA extraction, conducted as described by Sambrook et al.,¹⁶ was carried out at the Mycology Laboratory of the Instituto Adolfo Lutz, in São Paulo City. PCR-RFLP was performed according to Tavanti et al.¹⁷ The results were interpreted by comparison against standard strains (*C. parapsilosis* ATCC 22019, *Candida orthopsilosis* ATCC 96139, and *Candida metapsilosis* ATCC 96144).

The minimum inhibitory concentrations of the antifungal agents amphotericin B (Sigma Aldrich, USA), fluconazole (Sigma Aldrich), itraconazole (Sigma Aldrich), and voriconazole (Pfizer, USA) were determined using a broth microdilution method according to the M27-A3 guidelines of the Clinical and Laboratory Standards Institute.³ Interpretation of the minimum inhibitory concentrations for amphotericin B and itraconazole was based on the M27-S3 document³; for fluconazole and voriconazole, the M27-S4 guidelines⁴ were followed.

Retrospective demographic and clinical data were obtained from medical records. The study was approved by the Universidade Federal de Mato Grosso do Sul Research Ethics Committee (permit 1591/2010).

In the two-year period investigated, 32 patients were laboratory-diagnosed with candidaemia, predominantly caused by non-*C. albicans* *Candida* species (65.6% of cases). The *C. parapsilosis*

complex predominated (7/32; 21.9%), comprising five isolates of *C. parapsilosis* *sensu stricto* and two of *C. orthopsilosis*. Except for one *C. parapsilosis* isolate resistant to fluconazole, all yeasts were susceptible to the antifungals tested. Table 1 shows the principal clinical and laboratory features of the seven patients.

C. parapsilosis *sensu stricto* and *C. orthopsilosis* jointly accounted for the majority of non-*C. albicans* *Candida* species isolated from the blood cultures – a finding that corroborates data collected from other Brazilian hospitals.^{5,10}

In a tertiary-care hospital in Spain,⁸ *C. parapsilosis* *sensu stricto* was the most frequent microorganism (90.3%) out of the 62 isolates pertaining to the *C. parapsilosis* complex isolated from blood, followed by *C. orthopsilosis* (9.7%).

Fever, tachycardia, and dyspnea were the most frequent clinical symptoms (3/7; 42.9%) at the time of candidaemia diagnosis. All patients had previously undergone procedures known for their associated high risk of nosocomial infection by *Candida* species – namely, haemodialysis or previous use of vancomycin (5 cases each); surgery within 30 days before the onset of candidaemia (3 cases); and use of nasogastric, orogastric, nasoenteral, or indwelling urinary catheter (2 cases each). These findings are similar to those of previous studies conducted in Brazil.^{1,2,12}

All patients with candidaemia had used central venous catheters. In the two pediatric patients included in the present study, the yeasts were also isolated from the catheter tips, corroborating studies that underscore the risks of invasive fungal infection associated with these devices, given the ability of these microorganisms to form biofilms.^{8,14} All five adult subjects were chronic renal failure patients undergoing haemodialysis with double-lumen catheters. The research literature reports cases of infection by species of the *C. parapsilosis* complex in patients with this profile.^{5,9} In the present study, four of the five adults with chronic renal failure had diabetes mellitus, which also constitutes a risk factor for invasive fungal infection.

A 33-year-old patient with diabetes mellitus and chronic renal failure on haemodialysis developed endocarditis. *C. orthopsilosis* was isolated not only from blood cultures but also from a post-surgical fragment of cardiac valve – a documented case of infective endocarditis caused by this microorganism, albeit rarely reported in the literature.¹¹ Endocarditis as a complication in patients

Table 1

Characteristics of seven patients with haematogenous candidaemia caused by species of the *Candida parapsilosis* complex and respective antifungal susceptibility profiles.

Patient	Age (years)	Ward	Underlying condition	Treatment	FLZ ^a (µg/ml)	ITZ ^b (µg/ml)	VCZ ^a (µg/ml)	AMB ^b (µg/ml)	Species (PCR-RFLP)
1	33	AICU	CRF, DM	FLZ	0.25	0.125	0.015	0.5	<i>C. orthopsilosis</i>
2	4	PICU	–	FLZ + AMB	0.5	0.06	0.015	1	<i>C. parapsilosis</i>
3	48	RCU	CRF, DM	FLZ	0.25	0.06	0.015	0.5	<i>C. parapsilosis</i>
4	72	RCU	CRF, DM	–	0.25	0.06	0.015	0.5	<i>C. parapsilosis</i>
5	<1	NNU	PB	FLZ	0.125	0.06	0.015	0.5	<i>C. orthopsilosis</i>
6	69	RCU	CRF	FLZ	0.125	0.015	0.015	0.5	<i>C. parapsilosis</i>
7	59	RCU	CRF, DM	FLZ	8	0.125	0.06	1	<i>C. parapsilosis</i>

AICU: adult intensive care unit; PICU: pediatric intensive care unit; RCU: renal care unit; NNU: newborn nursery unit; CRF: chronic renal failure, DM: diabetes mellitus; PB: preterm birth; FLZ: fluconazole; ITZ: itraconazole; VCZ: voriconazole; AMB: amphotericin B.

^a M27-S4 (CLSI 2012), 24 h reading.

^b M27-S3 (CLSI 2008), 48 h reading.

with candidaemia not previously submitted to heart surgery is an uncommon event.⁵

Of the seven patients with candidaemia, the majority (6/7; 85.7%) received fluconazole as a first-choice antifungal agent. The single untreated patient developed transient candidaemia, recognizable by an episode of shivering and chills following a haemodialysis session, although no subsequent manifestations of invasive fungal disease were reported.

Blood cultures from a patient with chronic renal failure on haemodialysis were positive for *C. parapsilosis sensu stricto* for nine consecutive months. This patient's clinical condition required haemodialysis via central venous catheter and proved too critical for catheter replacement. Antifungal therapy was therefore maintained, as was the catheter. Previous long-term use of fluconazole by this patient is thought to have facilitated the development of *in vitro* ($\text{MIC}=8\ \mu\text{g/ml}$) and *in vivo* resistance to this antifungal. This was the only patient who evolved to death. Extensive use of fluconazole has been associated with the emergence of resistant clinical isolates.⁷

Antifungal resistance is an increasing clinical issue, especially in non-*C. albicans* *Candida* species, and requires continuous monitoring using susceptibility tests because resistance *in vitro* can be predictive of treatment failure.¹³ In the present study, *in vitro* anti-fungal susceptibility proved consistent with clinical outcome.

Acknowledgment

Financial support was provided by the Fundação de Apoio ao Desenvolvimento do Ensino, Ciência e Tecnologia do Estado de Mato Grosso do Sul, Brazil.

References

1. Almirante B, Rodríguez D, Cuenca-Estrella M, Almela M, Sanchez F, Ayats J, et al. Epidemiology, risk factors, and prognosis of *Candida parapsilosis* bloodstream infections: case-control population-based surveillance study of patients in Barcelona, Spain, from 2002 to 2003. *J Clin Microbiol.* 2006;44:1681–5.
2. Chang MR, Correia FP, Costa LC, Xavier PCN, Palhares DB, Taira DL, et al. *Candida* bloodstream infection: data from a teaching hospital in Mato Grosso do Sul, Brazil. *Rev Inst Med Trop São Paulo.* 2008;50:265–8.
3. Clinical and Laboratory Standards Institute (CLSI). Reference method for broth dilution antifungal susceptibility testing of yeasts: approved standard M27-A3. 3rd ed. Wayne: Clinical and Laboratory Standards Institute; 2008.
4. Clinical and Laboratory Standards Institute (CLSI). Reference method for broth dilution antifungal susceptibility testing of yeasts: fourth informational supplement M27-S4. Wayne: Clinical and Laboratory Standards Institute; 2012.
5. Colombo AL, Guimarães T, Camargo LFA, Richtmann R, Queiroz-Telles F, Salles MJC. Brazilian guidelines for the management of candidiasis – a joint meeting report of three medical societies: Sociedade Brasileira de Infectologia, Sociedade Paulista de Infectologia and Sociedade Brasileira de Medicina Tropical. *Braz J Infect Dis.* 2013;17:283–312.
6. Colombo A, Garnica M, Camargo LFA, Cunha CA, Bandeira AC, Borghi D, et al. *Candida glabrata*: an emerging pathogen in Brazilian tertiary care hospitals. *Med Mycol.* 2013;51:38–44.
7. Déry M, Hasbun R. Fluconazole-resistant *Candida*: mechanisms and risk factor identification. *Curr Fungal Infect Rep.* 2011;5:23–8.
8. De Toro M, Torres MJ, Maite R, Aznar J. Characterization of *Candida parapsilosis* complex isolates. *Clin Microbiol Infect.* 2010;17:418–24.
9. Gauna TT, Oshiro E, Luzio YC, Paniago AMM, Pontes ERJC, Chang MR. Blood-stream infection in patients with end-stage renal disease in a teaching hospital in central-western Brazil. *Rev Soc Bras Med Trop.* 2013;46:426–32.
10. Herkert PF, Gomes RR, Muro MD, Pinheiro RL, Fornari G, Vicente VA, et al. In vitro susceptibility and molecular characterization of *Candida* spp. from candidemic patients. *Rev Iberoam Micol.* 2015;32:221–8.
11. Lefort A, Chartier L, Sendid B, Wolff M, Mainardi J-L, Podglajen I, et al. Diagnosis, management and outcome of *Candida* endocarditis. *Clin Microbiol Infect.* 2012;18:E99–109.
12. Pereira GH, Müller PR, Szesz MW, Levin ASC, Melhem MSC. Five-year evaluation of bloodstream yeast infections in a tertiary hospital: the predominance of non-*C. albicans* *Candida* species. *Med Mycol.* 2010;48:839–42.
13. Pfaller MA. Antifungal drug resistance: mechanisms, epidemiology, and consequences for treatment. *Am J Med.* 2012;125:S3–13.
14. Pires RH, Santos JM, Zaia JE, Martins CHG, Mendes-Giannini MJS. *Candida parapsilosis* complex water isolates from a haemodialysis unit: biofilm production and in vitro evaluation of the use of clinical antifungals. *Mem Inst Oswaldo Cruz.* 2011;106:646–54.
15. Quindós G. Epidemiology of candidaemia and invasive candidiasis. A changing face. *Rev Iberoam Micol.* 2014;31:42–8.
16. Sambrook J, Fritsch EF, Maniatis T. Commonly used techniques in molecular cloning. In: Sambrook J, Fritsch EF, Maniatis T, editors. Molecular cloning. A laboratory manual, appendix E. New York: Cold Spring Harbour Laboratory Press; 1989. p. 3–6.
17. Tavanti A, Davidson AD, Gow NAR, Maiden MCJ, Odds FC. *Candida orthopsilosis* and *Candida metapsilosis* spp. nov. to replace *Candida parapsilosis* groups II and III. *J Clin Microbiol.* 2005;43:284–92.