



Original article

New record of *Scedosporium dehoogii* from Chile: Phylogeny and susceptibility profiles to classic and novel putative antifungal agents



Eduardo Alvarez^{a,*}, Camila Sanhueza^b

^a Mycology Unit/Filamentous Fungi, Biomedical Sciences Institute, Microbiology and Mycology Program, University of Chile, Chile

^b Medical Technology Department, University of Chile, Chile

ARTICLE INFO

Article history:

Received 20 August 2015

Accepted 10 March 2016

Available online 25 July 2016

Keywords:

Mycoses

Phylogeny

Scedosporiosis

Scedosporium dehoogii

Taxonomy

Antifungal agents

ABSTRACT

Background: *Scedosporium* species are considered emerging agents causing illness in immunocompromised patients. In Chile, only *Scedosporium apiospermum*, *Scedosporium boydii* and *Lomentospora prolificans* have been reported previously.

Aims: The study aimed to characterize genetically *Scedosporium dehoogii* strains from Chilean soil samples, and assessed the antifungal susceptibility profile to classic and novel putative antifungal molecules.

Methods: In 2014, several samples were obtained during a survey of soil fungi in urban areas from Chile. Morphological and phylogenetic analyses of the internal transcribed spacer region (ITS), tubulin (TUB), and calmodulin (CAL) sequences were performed. In addition, the susceptibility profiles to classic antifungal and new putative antifungal molecules were determined.

Results: Four strains of *Scedosporium dehoogii* were isolated from soil samples. The methodology confirmed the species (reported here as a new record for Chile). Antifungal susceptibility testing demonstrates the low activity of terpenes (α -pinene and geraniol) against this species. Voriconazole (VRC), posaconazole (PSC), and the hydroxyquinolines (clioquinol, and 5,7-dibromo-8-hydroxyquinoline) showed the best antifungal activity.

Conclusions: Our results demonstrate that *Scedosporium dehoogii* is present in soil samples from Chile. This study shows also that hydroxyquinolines have potential as putative antifungal molecules.

© 2016 Asociación Española de Micología. Published by Elsevier España, S.L.U. All rights reserved.

***Scedosporium dehoogii*, un nuevo reporte de Chile: filogenia y perfiles de sensibilidad a antifúngicos clásicos y a nuevas moléculas con potencial antifúngico**

RESUMEN

Antecedentes: Las especies de *Scedosporium* se consideran agentes emergentes responsables de enfermedad en pacientes inmunodeficientes. En Chile, únicamente se había publicado con anterioridad la existencia de las especies *Scedosporium apiospermum*, *Scedosporium boydii* y *Lomentospora prolificans*.

Objetivos: Este estudio tuvo como objetivo clasificar genéticamente aislamientos de *Scedosporium dehoogii* obtenidos de muestras del suelo de Chile. Asimismo, se evaluó el perfil de sensibilidad de las cepas a los antifúngicos clásicos y a nuevas moléculas con potencial antifúngico.

Métodos: En el año 2014, durante un estudio de evaluación de la biodiversidad fúngica en Chile, se tomaron diversas muestras del suelo de zonas urbanas del país. Se llevaron a cabo análisis morfológicos y filogenéticos de las secuencias pertenecientes a la región del espaciador transcrita interna (ITS), de la tubulina (TUB) y de la calmodulina (CAL). Además, se determinaron los perfiles de sensibilidad a los antifúngicos clásicos y a nuevas moléculas con potencial antifúngico.

Resultados: Se aislaron cuatro cepas de *Scedosporium dehoogii* de las muestras del suelo. Las pruebas morfológicas y moleculares confirmaron la especie (el presente estudio representa un nuevo reporte para Chile). Las pruebas de sensibilidad antifúngica mostraron baja actividad de los terpenos (α -pineno y geraniol). El voriconazol (VRC), el posaconazol (PSC) y las hidroxiquinolinas (clioquinol y 5,7-dibromo-8-hidroxiquinolina) presentaron la mejor actividad antifúngica.

Palabras clave:

Micosis

Filogenia

Escedosporiosis

Scedosporium dehoogii

Taxonomía

Antifúngicos

* Corresponding author at: Biomedical Sciences Institute, University of Chile, Independencia 1027, Santiago, Chile. Tel.: +56 229786639.

E-mail addresses: ead78@yahoo.com, ealvarezd@med.uchile.cl (E. Alvarez).

Conclusiones: Nuestros estudios demuestran que *Scedosporium dehoogii* está presente en los suelos de Chile. Asimismo, este estudio sugiere que las hidroxiquinolinas desempeñan una potencial actividad antifúngica.

© 2016 Asociación Española de Micología. Publicado por Elsevier España, S.L.U. Todos los derechos reservados.

Species of *Scedosporium* are ubiquitous fungi found in several substrates, such as compost, soil, sewage, and water. Some species can cause a wide spectrum of infections in humans and animals.^{17,24} Up to date, only five species have been related to human infections: *Scedosporium apiospermum*, *Scedosporium aurantiacum*, *Scedosporium dehoogii*, *Scedosporium boydii*, and *Lomentospora prolificans* (formerly *Scedosporium prolificans*).^{13,15,22,34} *Scedosporiosis* (mycosis caused by *Scedosporium* species)^{7,17} mostly affects immunocompromised patients, causing a wide range of infections as brain abscesses, lungs disease, meningitis, mycetoma, septic arthritis, osteomyelitis, etc.^{6,20,25–27} In Chile, only three clinical cases have been reported: a rhinosinusitis by *S. boydii*,³¹ a surgical site abscess by *L. prolificans*¹⁰ and a rhinosinusul infection by *S. apiospermum*.³

The treatment of scedosporiosis is still a challenge due to the high mortality rates and the increasing frequency.³⁷ In addition, several studies have reported that *Scedosporium* species show multiple *in vitro* resistance patterns to most antifungals, with high intraspecific variability.^{14,17,23,30} On the other hand, voriconazole (VRC) remains as the most effective antifungal agent. However, several species show high minimum inhibitory concentrations (MICs) values to VRC, as *S. dehoogii* (up to >16 µg/ml) or *L. prolificans* (geometric mean MIC of 15.4 µg/ml).²³

Currently, the number of antifungal drugs available for the treatment of fungal infections is still limited. Likewise, the increasing frequency of resistance makes the development of new therapeutic strategies necessary. In this way, several authors have reported the potential of natural compounds isolated from plants as antifungal agents.^{18,29,35} In this regard, some compounds such as terpenes (geraniol, carvacrol, α and β pinene) and quinolines (hydroxyquinolines) have been proposed as good candidates with biocide activity.^{1,9,12,33}

In the present work, we report the isolation of four strains of *S. dehoogii* from Chilean soil samples. Our searching in the literature showed no previous findings of *S. dehoogii* in Chile. In addition, we have performed antifungal assays in order to determine the susceptibility patterns to classic antifungal agents and new putative antifungal molecules.

Materials and methods

Fungal strains

A total of 84 soil samples were collected from 14 administrative regions. At each sampling site, triplicates were obtained and placed in sterilized polyethylene bags. Then, 1 ml of each soil suspension (1:10 w/v dilutions) was cultured on the selective medium Dichloran Rose-Bengal Chloramphenicol Agar (DRBC) (Becton & Dickinson). The DRBC plates were stored at 25 °C and 37 °C, and observed every two days. When typical features of *Scedosporium* species were observed, the colonies were subcultured in 2% malt extract agar (MEA), potato dextrose agar (PDA) and Sabouraud agar (SA), at 25 °C and 37 ± 1 °C in the dark. The strains studied in the present work were isolated from soil collected in urban areas from several Chilean cities: one from Arica (North of Chile); two from Valparaíso (central region of Chile) and the last from Puerto Varas (South of Chile).

Morphological and physiological study

Morphological features were determined on PDA at 25 ± 1 °C (7 days). Cultures were deposited at the Chilean Fungal Collection (ChFC), Mycology Unit, University of Chile as *S. dehoogii* ChFC 148, 151, 153, and 161. Microscopic features were studied in lactophenol stained with cotton blue (Merck) using Optika B1000Ph microscope (Optika, Italy). Microphotographs were taken with Optikam B9 camera (Optika, Italy).

DNA extraction, amplification, and sequencing

Fungal DNA was extracted directly from pure 7 day-old cultures by means of the E.Z.N.A® Fungal DNA Mini Kit (Omega Biotek Store, USA). PCR was conducted with a 3'Prime Thermal Cycler (Techne, UK) using ITS1 and ITS4 primers for ITS1-5.8 S-ITS2 region³⁶; Cal1 and Cal2 primers for the calmodulin gene,²⁸ and Tub2F and Tub2R primers for one region within the β-tubulin gene.⁸ A 25 µl-mixture containing 17 µl of BioMix™ Red (New England Biolabs), 10 pM of the primers, and 10 ng DNA was amplified following a protocol of initial denaturation at 94 °C for 5 min, 35 cycles of denaturation at 94 °C for 1 min, annealing at 52 °C for 1 min for ITS region, and 55 °C for CAL and TUB; extension at 72 °C for 2 min, and final extension at 72 °C for 7 min. The PCR product was checked on 1% agarose gel. Purification was performed with FavorPrep™ Gel/PCR purification Mini Kit (Favorgene, Taiwan) following the manufacturer guidelines. Sequencing was done on Macrogen (Macrogen, Korea). An NCBI Blastn search for each locus region was performed.² Sequences from type and reference strains were gotten from GenBank database. The sequences were aligned in MUSCLE software¹¹ followed by manual adjustments with a text editor.

Phylogenetic analyses

Phylogenetic analyses were performed using Maximum Parsimony (MP) method in PAUP 4.0b10 software. For MP inference the most-parsimonious tree (MPT) was obtained after one hundred heuristic searches with random sequence addition and tree bisection-reconnection branch-swapping algorithms, collapsing zero-length branches and saving all minimal-length trees (Multi-Trees). Support for internal branches was assessed using a heuristic parsimony search of 1000 bootstrapped data sets. The combined datasets of the ITS, CAL, and TUB were tested for incongruence with the partition homogeneity test and implemented in PAUP 4.0b10 software. *Lomentospora prolificans* FMR 7294 and *Petriellopsis africana* CBS 311.72 were used as outgroup.

Antifungal susceptibility testing

In vitro susceptibility testing was performed using broth microdilution for filamentous fungi according to CLSI document M38-A2.⁵ The MICs and MECs (minimal effective concentration) were read at 48 and 72 h. Although the clinical breakpoints for *Scedosporium* species have not been developed, the following suggested cut-off breakpoints were used: susceptible (S), ≤1 µg/ml; intermediate (I), 2 µg/ml; and resistant (R), ≥4 µg/ml (CLSI, M38 A-2). Pure active powders, of known potency, of amphotericin B (AMB) (Sigma-Aldrich), voriconazole (VRC)

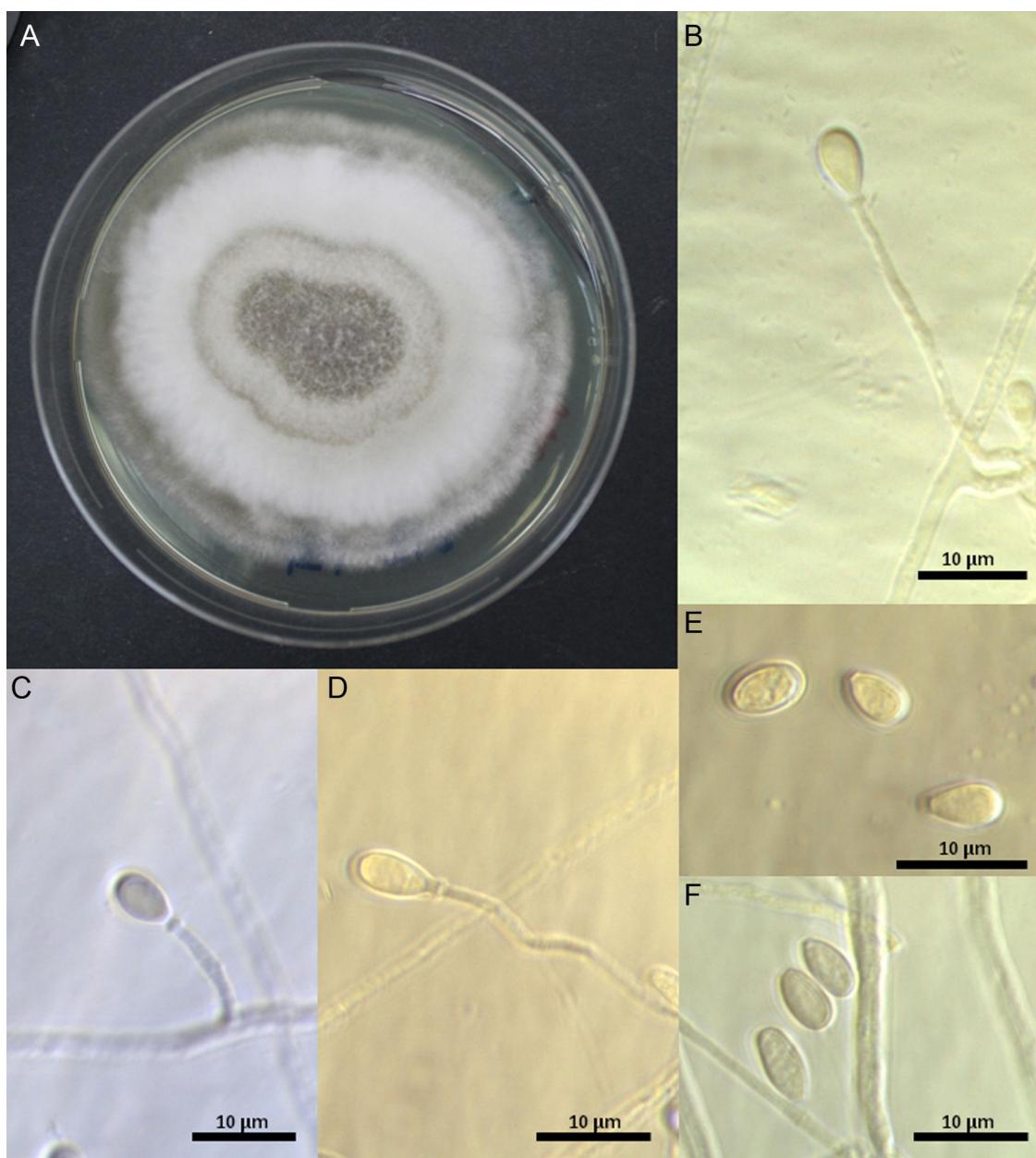


Fig. 1. (A–D) *Scedosporium dehoogii*, ChFC 151. (A) Colony growing on PDA after 14 days at 25 °C. (B–D) Conidiogenous cells and conidia. (E, F) Conidia.

(Sigma–Aldrich), itraconazole (ITC) (Sigma–Aldrich), posaconazole (PSC) (Sigma–Aldrich), caspofungin (CSF) (Sigma–Aldrich), micafungin (MCF) (Astellas Pharma), anidulafungin (ANF) (Pfizer), α -pinene (α -P) (partner laboratory), geraniol (GER) (partner laboratory), clioquinol (CLIO) (Sigma–Aldrich), and 5,7-dibromo-8-hydroxyquinoline (5–7C) (Sigma–Aldrich) were used. All tests were performed in duplicates. *Candida parapsilosis* ATCC 22019 and *Paecilomyces variotii* ATCC MYA 3630 were included as quality controls.

Results

Morphology and physiology

Several genera and species were isolated from soil samples (*i.e.*, *Aspergillus*, *Cunninghamella*). In the same way, other *Scedosporium* and related species such as *S. boydii*, *S. apiospermum*, and *L. prolificans* were isolated on DRBC plates (unpublished data).

In the first approach, all *Scedosporium* isolates were identified based on the morphological and physiological features. The final identification for each species recovered from DRBC plates was obtained by molecular analyses of tubulin gene sequences.

In general, the four strains studied displayed the typical features described by Gilgado et al.¹⁵ The colonies on PDA reached 28 mm diameter after 7 days at 25 ± 0.2 °C, they were dense, cottony to lanose, grayish-white to white, and with a whitish irregular margin; the reverse was yellowish or light-brown. Solitary conidiogenous cells were subhyaline smooth-walled, mainly cylindrical, 10–54 μm length by 1–1.4 μm width, and produced pale brown, obovoid or ellipsoidal 5.9–6.2 $\mu\text{m} \times$ 3.6–4.4 μm conidia. Sessile conidia were subhyaline, thin-walled, mostly obovate (5.9–)7.3(–7.8) \times (3.5–)3.8(–4.0) μm . Hyphae, 1.2–2.5 μm , were hyaline to subhyaline (Fig. 1). A teleomorph was not observed even after 60 days of incubation.

Complete dataset
1698 bp
Tree length = 773
CI = 0.8124
HI = 0.1876
RI = 0.8945

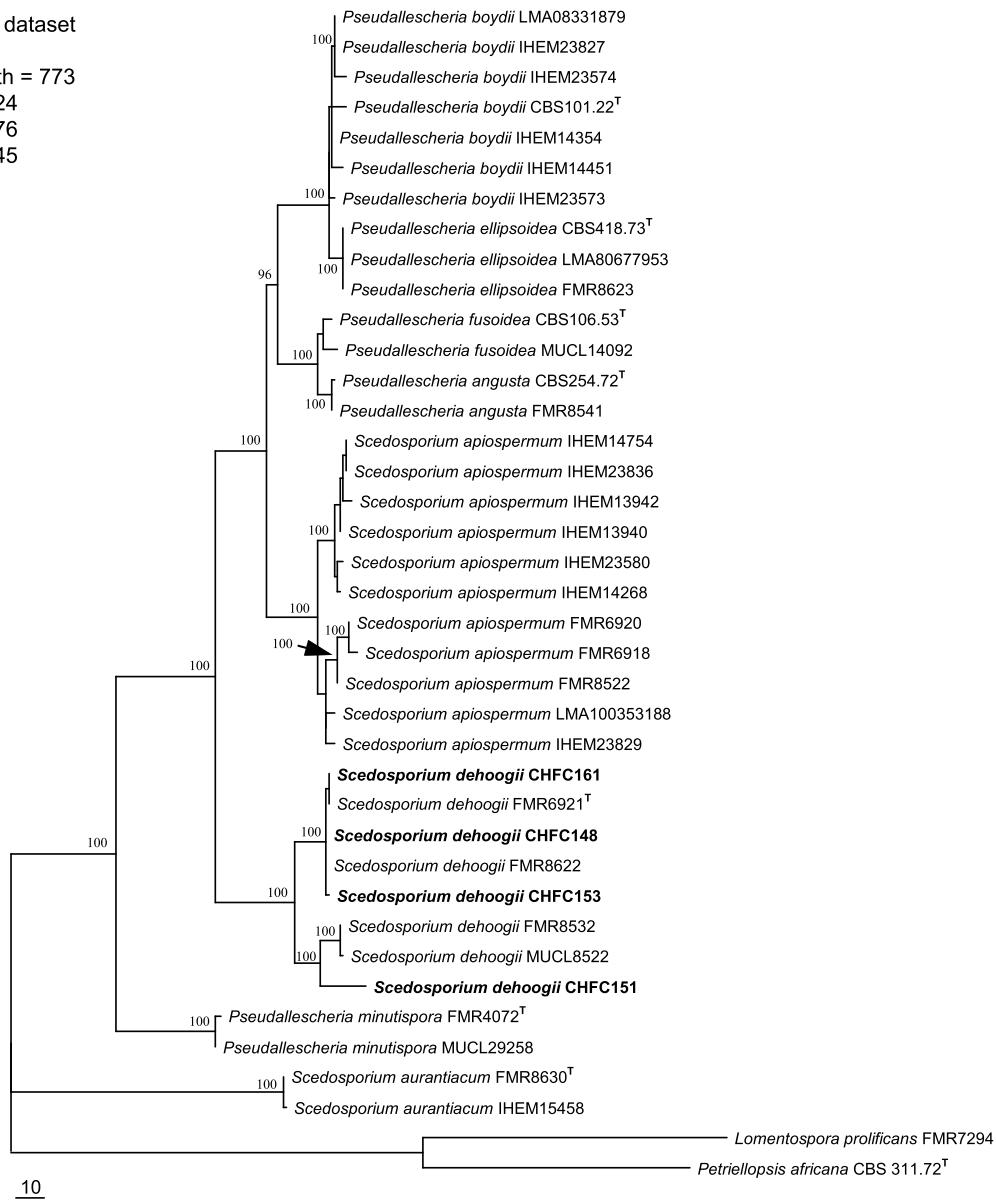


Fig. 2. One of the 90 most-parsimonious trees obtained from heuristic searches based on the analysis of the combined data set (ITS + CAL + TUB). Bootstrap support values of 100% are indicated at the nodes. CI, consistency index; RI, retention index; HI, homoplasy index. Chilean strains are indicated with boldface type. ^T = type strains.

The growth rates were similar in all media tested. The minimum, maximum and optimal temperatures were 15 °C, 40 °C and 36 ± 1 °C respectively.

Phylogeny

The identification of all the strains studied as *S. dehoogii* was feasible by means of the three loci used in the present study. The partition homogeneity test demonstrated that the three loci sequence data sets were congruent ($p = 0.05$) and could therefore be combined. A fragment of 1698 bp was obtained in the concatenated analysis. The phylogeny inference was analyzed as separate (data not shown) and combined dataset.

The tree topologies of concatenated loci by MP were highly similar to those observed in the individual tree for each gene tested. High clade support values were obtained in the concatenated analysis, being mainly over 90%. A total of 90 MPT were obtained from heuristic search from concatenated dataset (Fig. 2). The trees had a consistency index of 0.81, a retention index of

0.89, and a homoplasy index of 0.18. Analyses of the combined partitions allowed the identification and recognition of 8 species of *Scedosporium* (Fig. 2). In the same way, the separation of *L. prolificans* from the *Scedosporium* clade was observed. The four strains isolated from Chilean soil samples were placed at the *S. dehoogii* clade, with the reference and type strains sequences.

The sequences generated in this study were deposited in the GenBank database under the following accession numbers: KT600015 (ChFC148 ITS), KT600016 (ChFC151 ITS), KT600017 (ChFC153 ITS), KT600018 (ChFC161 ITS), KT820715 (ChFC148 TUB), KT820716 (ChFC151 TUB), KT820717 (ChFC153 TUB), KT820718 (ChFC161 TUB), KT820719 (ChFC148 CAL), KT820720 (ChFC151 CAL), KT820721 (ChFC153 CAL), KT820722 (ChFC161 CAL).

Antifungal susceptibility testing

The results of the antifungal susceptibility tests for the *Scedosporium* strains are shown in Table 1. Among the classical antifungal agents, voriconazole, itraconazole and posaconazole were the most

Table 1

In vitro activities of classical antifungal agents and novel putative antifungal compounds against four isolates of *Scedosporium dehoogii*. MIC ($\mu\text{g/ml}$) values MEC ($\mu\text{g/ml}$) values.

Strain	α -P	GER	CLIO	5–7C	AMB	PSC	VRC	ITC	ANF	CSF	MCF
ChFC 148	>128	>128	1	1	2	1	1	1	4	2	4
ChFC 151	64	>128	1	1	2	1	0.5	1	2	1	2
ChFC 153	>128	>128	1	1	2	1	0.5	2	4	4	4
ChFC 161	64	>128	1	1	4	1	0.5	2	4	4	4
ATCC 22019	64	64	0.5	0.25	0.25	0.12	0.25	0.5	1	1	0.5
ATCC MYA 3630	64	>128	0.5	0.5	0.5	0.25	0.25	0.5	0.12	0.12	0.12

AMB, amphotericin B; VRC, voriconazole; ITC, itraconazole; PSC, posaconazole; CSF, caspofungin; MCF, micafungin; ANF, anidulafungin; α -P, α -pinene; GER, geraniol; CLIO, clioquinol; 5–7C, 5,7-dibromo-8-hydroxyquinoline; ChFC, Chilean Fungal Collection; ATCC, American type culture collection.

active. Echinocandins were the less active against all the strains. Among the putative new antifungal molecules, geraniol and α -pinene were not active, obtaining high MIC values. On the other hand, clioquinol, and 5,7-dibromo-8-hydroxyquinoline showed promising results with low MICs values.

Discussion

Scedosporium species are opportunistic pathogenic fungi. Several species have been reported as etiological agents, especially in immunocompromised patients.

S. dehoogii is considered an environmental species. In recent studies, several strains were isolated from locations associated with high human activity: playgrounds, riverbanks or agricultural areas (except vineyards).^{19,21,32} Thus, *S. dehoogii* is not commonly associated with human infections, despite the high virulence observed in murine models.¹⁶ However, some clinical isolates have been recovered: one from a sympathetic ophthalmia in a 10 year-old boy,⁴ one from cystic fibrosis sputum, and two from cutaneous infections.²³

In Chile, few cases of scedosporiosis have been reported.^{3,10,31} These three cases described previously, with the identification based on the morphological features of the isolates, were infections by *S. apiospermum*, *S. boydii* and *L. prolificans*, the most prevalent agents described on the literature. In the present study, we report for the first time the presence of *S. dehoogii* in soil samples of our country. The preliminary morphological identification was confirmed using the molecular tools, agreeing with the information previously reported by other authors.¹⁶ In this way, four strains isolated from Chile were located in the *S. dehoogii* clade. However, some variability was observed in their nucleotide sequences, confirming the observations of Gilgado et al.¹³

Although data of the *in vitro* susceptibility of this fungus is limited, high MECs for echinocandins have been described in previous studies,^{14,23} which is in agreement with our results. In the present study VRC and PSC were the most active drugs, being within the range of susceptibility. This finding agrees with the results reported by Gilgado et al.¹⁴ However, other authors have found that *S. dehoogii* is associated with high VRC MICs.²³ This could be explained by the fact that these authors tested a greater number of isolates.

Clioquinol and 5,7-dibromo-8-hydroxyquinoline seem to be putative antifungal agents as they showed a good activity against the four strains, having similar MIC values to those obtained against some azoles tested here.

In summary, we have reported for the first time the presence of *S. dehoogii* in Chile, and its susceptibility against classical and novel putative antifungal drugs. More studies are necessary to evaluate the spectrum of all the *Scedosporium* species present in Chile. In addition, more *in vitro* and *in vivo* analyses with a large number of strains are necessary to elucidate the potential activity of clioquinol, 5,7-dibromo-8-hydroxyquinoline and other hydroxyquinolines as antifungal agents.

Conflict of interest

The authors declare no conflict of interest.

Acknowledgments

This work was supported by the FONDECYT project 11130381 and U-INICIA 560786.

The authors thank Dr. Valentina Salas for the reviewing of the manuscript and constructive comments. Authors also thank Felipe Salas for some soil samples collecting and geraniol and α -pinene samples. We thank Armin Araya for the Arica soil samples.

References

- Abbaszadeh S, Sharifzadeh A, Shokri H, Khosravi AR, Abbaszadeh A. Antifungal efficacy of thymol, carvacrol, eugenol and menthol as alternative agents to control the growth of food-relevant fungi. *J Mycol Med.* 2014;24: 51–6.
- Altschul SF, Gish W, Miller W, Myers EW, Lipman DJ. Basic local alignment search tool. *J Mol Biol.* 1990;215:403–10.
- Chanqueo L, Gutiérrez C, Tapia C, Silva V, Razeto L, Misad C. *Scedosporium apiospermum* rhinosinusal infection in an immunocompetent host. *Rev Chilena Infectol.* 2009;26:453–6.
- Chen S, Aronow ME, Wang C, Shen D, Chan C. Classical pathology of sympathetic ophthalmia presented in a unique case. *Open Ophthalmol J.* 2014;8: 32–8.
- Clinical and Laboratory Standards Institute. Reference method for broth dilution antifungal susceptibility testing of filamentous fungi: approved standard M38-A2. Wayne, PA, USA: CLSI; 2008.
- Cooley L, Spelman D, Thursky K, Slavin M. Infection with *Scedosporium apiospermum* and *S. prolificans*, Australia. *Emerg Infect Dis.* 2007;13:1170–7.
- Cortez KJ, Roilides E, Quiroz-Telles F, Meletiadis J, Antachopoulos C, Knudsen T, et al. Infections caused by *Scedosporium* spp. *Clin Microbiol Rev.* 2008;21:157–97.
- Cruse M, Telerant R, Gallagher T, Lee T, Taylor JW. Cryptic species in *Stachybotrys chartarum*. *Mycologia.* 2002;94:814–22.
- Dardari Z, Lemrani M, Bahloul A, Sebban A, Hassar M, Kitane S, et al. Antileishmanial Activity of a new 8-hydroxyquinoline derivative designed 7-[5'-(3'-phenylisoaxazolino)methyl]-8-hydroxyquinoline: preliminary study. *Farmaco.* 2004;59:195–9.
- Díaz M, Juliet C, Monzón A, Rodríguez-Tudela J. Absceso de herida operatoria por *Scedosporium prolificans*: Primer aislamiento en Chile. Revisión de la literatura. *Rev Chil Infect.* 2004;21:65–9.
- Edgar RC. MUSCLE: multiple sequence alignment with high accuracy and high throughput. *Nucleic Acids Res.* 2004;32:1792–7.
- Enquist PA, Gylfe A, Hägglund U, Lindström P, Norberg-Scherman H, Sundin C, et al. Derivatives of 8-hydroxyquinoline-antibacterial agents that target intra- and extracellular Gram-negative pathogens. *Bioorg Med Chem Lett.* 2012;22:3550–3.
- Gilgado F, Cano J, Gene J, Guarro J. Molecular phylogeny of the *Pseudallescheria boydii* species complex: proposal of two new species. *J Clin Microbiol.* 2005;43:4930–42.
- Gilgado F, Serena C, Cano J, Gene J, Guarro J. Antifungal susceptibilities of the species of the *Pseudallescheria boydii* complex. *Antimicrob Agents Chemother.* 2006;50:4211–3.
- Gilgado F, Cano J, Gene J, Sutton DA, Guarro J. Molecular and phenotypic data supporting distinct species statuses for *Scedosporium apiospermum* and *Pseudallescheria boydii* and the proposed new species *Scedosporium dehoogii*. *J Clin Microbiol.* 2008;46:766–71.
- Gilgado F, Cano J, Gene J, Serena C, Guarro J. Different virulence of the species of the *Pseudallescheria boydii* complex. *Med Mycol.* 2009;47:371–4.
- Guarro J, Kantarcıoglu AS, Horre R, Rodriguez-Tudela JL, Cuenca-Estrella M, Berenguer J, et al. *Scedosporium apiospermum*: changing clinical spectrum of a therapy-refractory opportunist. *Med Mycol.* 2006;44:295–327.

18. Grayer RJ, Kokubun T. Plant-fungal interactions: the search for phytoalexins and other antifungal compounds from higher plants. *Phytochemistry*. 2001;56:253–63.
19. Harun A, Gilgado F, Chen SC, Meyer W. Abundance of *Pseudallescheria*/*Scedosporium* species in the Australian urban environment suggests a possible source for scedosporiosis including the colonization of airways in cystic fibrosis. *Med Mycol*. 2010;48:70–6.
20. Horré R, Jovanić B, Marklein G, Schumacher G, Friedrichs N, Neuhaus T, et al. Fatal pulmonary scedosporiosis. *Mycoses*. 2003;46:418–21.
21. Kaltseis J, Rainer J, De Hoog G. Ecology of *Pseudallescheria* and *Scedosporium* species in human-dominated and natural environments and their distribution in clinical samples. *Med Mycol*. 2009;47:398–405.
22. Kim JS, Choi M, Nam CH, Kim JY, Hong SP, Kim MH, et al. Co-infection of *Scedosporium apiospermum* and *Mycobacterium chelonae* in an immunocompetent host. *J Dermatol*. 2014;41:922–5.
23. Lackner M, de Hoog GS, Verweij PE, Najafzadeh MJ, Curfs-Breuker I, Klaassen CH, et al. Species-specific antifungal susceptibility patterns of *Scedosporium* and *Pseudallescheria* species. *Antimicrob Agents Chemother*. 2012;56:2635–42.
24. Maslen M, Peel M. Human and animal isolates of *Pseudallescheria boydii* and *Scedosporium* species, from Melbourne, Australia, 1977–1995. *Mycoses*. 2011;54:442–9.
25. Mathew S, Rao RM, Raghavendra S, Chandramouli BA, Kamble R, Athmanathan S. Rare case of multiple brain abscess – *Scedosporium apiospermum*. *BMC Infect Dis*. 2012;12 Suppl. 1:59.
26. Meshram SH, Mishra GP. Pulmonary scedosporiosis – a rare entity. *Asian Pac J Trop Dis*. 2011;1:330–2.
27. Nath R, Gogoi RN, Saikia L. Keratomycosis due to *Scedosporium apiospermum*. *Indian J Med Microbiol*. 2010;28:414–5.
28. O'Donnell K, Nirenberg HI, Aoki T, Cigelnik E. A Multigene phylogeny of the *Gibberella fujikuroi* species complex: detection of additional phylogenetically distinct species. *Mycoscience*. 2000;41:61–78.
29. Oliveira MH, Innocente AM, Pereira AG, Dias DO, Oliveira EG, Barreto F, et al. Semi-synthesis: a solution to problems of pharmacological natural products. *Electronic J Pharm*. 2012;9:62–8.
30. Pfaller MA, Messer SA, Hollis RJ, Jones RN. Antifungal activities of posaconazole, ravuconazole, and voriconazole compared to those of itraconazole and amphotericin B against 239 clinical isolates of *Aspergillus* spp. and other filamentous fungi: report from SENTRY Antimicrobial Surveillance Program, 2000. *Antimicrob Agents Chemother*. 2002;46:1032–7.
31. Pruzzo E, Labarca J, Gúzman A, León P, Emmerich M, Bustos Rico B. Rinosinusitis por *Pseudallescheria boydii*: caso clínico. *Rev Otorrinolaringol Cir Cabeza Cuello*. 2000;60:109–16.
32. Rougeron A, Schulier G, Leto J, Sitterlé E, Landry D, Bougnoux ME, et al. Human impacted areas of France are environmental reservoirs of the *Pseudallescheria boydii*/*Scedosporium apiospermum* species complex. *Environ Microbiol*. 2015;17:1039–48.
33. Silva ACR, Lopes PM, Azevedo MMB, Costa DCM, Alviano CS, Alviano DS. Biological activities of α-pinene and β-pinene enantiomers. *Molecules*. 2012;17:6305–16.
34. Taylor A, Talbot J, Bennett P, Martin P, Makara M, Barrs VR. Disseminated *Scedosporium prolificans* infection in a Labrador retriever with immune mediated haemolytic anaemia. *Med Mycol Case Rep*. 2014;6:66–9.
35. Vengurlekar S, Sharma R, Trivedi P. Efficacy of some natural compounds as antifungal agents. *Pharmacogn Rev*. 2012;6:91–9.
36. White TJ, Bruns S, Lee S, Taylor J. Amplification and direct sequencing of fungal ribosomal RNA genes for phylogenetics. *PCR Protoc A Guid to Methods Appl*. 1990;31:5–22.
37. Wiederhold NP, Lewis RE. Antifungal activity against *Scedosporium* species and novel assays to assess antifungal pharmacodynamics against filamentous fungi. *Med Mycol*. 2009;47:422–32.