



ORIGINAL ARTICLE

# Comprehensive evaluation of abattoirs with no Hazard Analysis Critical Control Point plan in Tucumán, Argentina



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**Abstract** This work focused on the comprehensive study of two provincial transit abattoirs in Tucumán, Argentina, with no Hazard Analysis Critical Control Point (HACCP) plan. Visits ( $n=20$ ) were conducted between 2016 and 2018 during the operational and post-operational processes. Risk was estimated and the bacteriological analysis of carcass and environmental samples was performed. Risk estimation showed the predominance of high risk in both abattoirs. The main deviations from the HACCP plan were: deficient building conditions, deficient workflow, lack of sectorization of changing rooms and bathrooms, lack of implementation of Standardized Sanitary Operational Procedures, and no food safety training of workers. The counts of indicator microorganisms from both abattoirs were not significant. *Salmonella* spp. was isolated from 7.5% carcass and 7.3% environmental samples. The *Salmonella* serovars identified were Cerro, Corvallis, Havana and Agona. Shiga toxin (*stx*) genes were detected in 24.4% carcass and 30.9% environmental samples. The isolates were characterized as *Escherichia coli* O8:H7/*stx*<sub>1</sub>, O116:H49/*stx*<sub>2</sub> and O136:H40/*stx*<sub>2</sub>. Based on these results, it would be possible to implement

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an improvement plan in Tucumán abattoirs together with the local health authorities. Still, the need to work jointly with the sanitary authority in search of a unique sanitary standard for Argentina remains unaddressed.

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## PALABRAS CLAVE

Estimación de riesgo;  
*Salmonella* spp.;  
*Escherichia coli*;  
Frigoríficos;  
Carne

## Evaluación integral de frigoríficos sin un plan de análisis de peligros y puntos críticos de control en Tucumán, Argentina

**Resumen** Este trabajo se centró en el estudio integral de dos frigoríficos de tránsito provincial en Tucumán, Argentina, carentes de un plan de análisis de peligros y puntos críticos de control (HACCP, por sus siglas en inglés). Las visitas (n = 20) se realizaron entre 2016 y 2018 durante los procesos operativos y posoperativos. Se realizó la estimación del riesgo y el análisis bacteriológico de medias reses y muestras ambientales. La estimación del riesgo demostró un predominio de riesgo alto en ambos frigoríficos. Las principales desviaciones del plan HACCP fueron las deficientes condiciones edilicias, un inadecuado flujo de trabajo, la falta de sectorización de vestuarios y baños, una implementación nula de procedimientos operativos estandarizados de saneamiento y una insuficiente capacitación en seguridad alimentaria de los operarios. Los recuentos de microorganismos indicadores de ambos frigoríficos no presentaron diferencias significativas. *Salmonella* spp. se aisló del 7,5% de muestras de medias reses y del 7,3% de muestras ambientales. Se identificaron las siguientes serovariiedades de *Salmonella*: Cerro, Corvallis, Havana y Agona. Se detectaron genes de toxina Shiga (stx) en el 24,4% de las muestras de medias reses y en el 30,9% de las muestras ambientales. Los aislamientos se caracterizaron como *Escherichia coli* O8:H7/stx<sub>1</sub>, O116:H49/stx<sub>2</sub> y O136:H40/stx<sub>2</sub>. Teniendo en cuenta estos resultados, sería posible implementar un plan de mejoramiento en frigoríficos de Tucumán conjuntamente con las autoridades locales de salud. Aun así, sigue sin abordarse la necesidad de trabajar en vinculación con las autoridades sanitarias en la búsqueda de una norma integrada única para Argentina.

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## Introduction

Meat is a valuable source of protein in the human diet, which makes it prone to high microbial contamination that can cause spoilage and foodborne infections in human beings<sup>2</sup>. The microorganisms most commonly present in meat and meat products are *Salmonella* spp., *Escherichia coli*, *Bacillus* spp., *Clostridium* spp., and *Shigella* spp.<sup>22</sup>. For this reason, the safety and quality of meat must be ensured.

Salmonellosis is among the main foodborne diseases affecting human beings globally. Since animals are the principal reservoirs of *Salmonella* spp., beef has been implicated as a vehicle of human infection in several outbreaks<sup>27</sup>. Shiga toxin-producing *Escherichia coli* (STEC) is a known enteric pathogen associated with foodborne illnesses, representing an important human health problem<sup>40</sup>. Ruminants are one of the main STEC reservoirs, and it has been recently shown that around 18.2% of all STEC reported cases worldwide were associated with beef<sup>40</sup>.

In abattoirs, carcass contamination with pathogens can occur through contact with the hide and contents of the digestive tract of slaughtered animals as well as through

contact with contaminated workers' hands, equipment or utensils<sup>10</sup>.

Argentine abattoirs are classified into exporter, federal transit and provincial transit according to their slaughter capacity, the marketing area for meat and viscera, and the sanitary authority responsible for their control<sup>16</sup>. Hygiene and sanitation standards differ among the abattoir categories described above<sup>34</sup>. For instance, most provincial transit abattoirs do not apply Hazard Analysis Critical Control Point (HACCP), Standardized Sanitary Operational Procedures (SSOP) or any intervention procedure at the end of meat processing<sup>8</sup>.

In Argentina, Costa et al.<sup>16</sup> reported the presence of *E. coli* O157:H7 (3.3%), non-O157 STEC (18.8%) and *Salmonella* spp. (5.5%) in 60 carcasses from three abattoirs with no HACCP plan in Buenos Aires province, before making improvements. In addition, in Tucumán, Perez Terrazzino<sup>33</sup> et al. reported *E. coli* O157:H7 (0.46%) and non-O157 STEC (1.9%) in 274 carcasses from six abattoirs without HACCP plan.

This work focused on the comprehensive study of two provincial transit abattoirs without an HACCP plan from

Tucumán (Argentina) by evaluating their sanitary status, including risk estimation and determination of the bacteriological quality of carcass and environmental samples.

## Materials and methods

The province of Tucumán (22 524 km<sup>2</sup>; 1 448 188 inhabitants) has eight provincial transit abattoirs controlled by the Subsecretaría de Asuntos Agrarios y Alimentos. This study was conducted in two of these abattoirs with no HACCP plan, identified as abattoir I (26°49'26" S 65°13'21" W) and abattoir II (27°3'15" S 65°24'12" W). Each abattoir was visited once a week for 20 consecutive weeks, during the operational and post-operational processes, in the period 2016–2018. Risk was estimated using two checklists. In addition, the bacteriological analysis of carcass and environmental samples (platforms, saw machine, worker's hands, knives, trays, sinks) was performed.

## Risk estimation

Risk was estimated using the two checklists described by Costa et al.<sup>16</sup> with minimal modifications. The pre-operational checklist of this work was used as post-operational checklist (once work was finished and after complete cleaning of the abattoir) in the present work. Ten operational and 10 post-operational checklists were completed alternately for 20 weeks in each abattoir. They were used to evaluate different variables associated with the building conditions of the establishment, the production line and good manufacturing practices (GMP), good hygiene practices (GHP) and SSOP. Each variable was assigned a numerical value and grouped into six blocks. Each block was assigned a score depending on the importance of the sector in the risk of contamination. The four possible qualifications of abattoir conditions were acceptable (perfect), marginal (not ideal), unacceptable (not corresponding) or not applicable (could not be evaluated but did not influence process outcomes).

The final block score (BS) was obtained with a formula that included the sum of the total acceptable and marginal grades obtained (TAM) multiplied by the importance (I) assigned to each block, divided by the sum of all acceptable scores (AA) minus total grades referred to as "does not apply" (TDNA)<sup>16</sup>.

$$BS = \frac{TAM \times I}{AA - TDNA}$$

The sum of all BS gave a final score of 100. Accordingly, risk was estimated on a 1–100 scale as high (1–40), moderate (41–70) or low (71–100).

The operational checklist included the following blocks and scores: (1) pens (15.0); (2) slaughter area (25.0); (3) head and viscera area (10.0); (4) control points (15.0); (5) cool chambers (20.0); (6) offal area (15.0). The post-operational checklist included the following blocks and scores: (1) pens (15.0); (2) slaughter area (35.0); (3) cool chambers (10.0); (4) quartering (10.0); (5) offal area (20.0); (6) outdoors (10.0).

## Sample collection

Carcass (n=160) and environmental (n=136) samples were obtained using a sterile sponge (Whirl-Pak Speci-Sponge, Nasco, USA) soaked in 10ml of buffered peptone water (BPW) (Britania S.A., CABA, Argentina).

At each visit, carcass samples (n=4) were taken during the operational (airing chambers) and post-operational (cool chambers) processes. Two samples from each carcass were taken. One was a combination of four carcass areas of 100 cm<sup>2</sup> each (chest, neck, buttock and posterior lateral hock), which were swabbed for the count of indicator microorganisms<sup>35</sup>: *E. coli*/coliform and mesophilic. The other sample was obtained by swabbing the carcass entire surface (external and internal side) with another sterile sponge (Whirl-Pak) and used for the detection of *Salmonella* spp., *E. coli* O157:H7 and non-O157 STEC<sup>16</sup>.

In the case of environmental samples, different pools were obtained during each operational visit from platforms (n=4), saw machine (chest and half, n=2), workers' hands (n=4) and knives (n=4). During post-operational visits, pools of platform (n=4), saw machine (chest and half, n=2), tray (n=4) and sink (n=4) samples were used for the detection of *Salmonella* spp., *E. coli* O157:H7 and non-O157 STEC. All samples were stored at 4°C and immediately sent to the laboratory for analysis.

## Bacteriological analysis

Carcass samples were analyzed for indicator microorganisms with 3M Petrifilm™ *E. coli*/coliform count plates and mesophilic count plates (3M™, Minnesota, USA). After the addition of 10ml BPW (Britania S.A.), the bags with the carcass sampling sponges were homogenized and squeezed. Then, 1 ml of the sample was placed into each 3M Petrifilm™ plate, incubated and counted according to the manufacturer's specifications. Results were expressed as log CFU/cm<sup>2</sup>.

*Escherichia coli* O157:H7, non-O157 STEC and *Salmonella* spp. were detected and isolated from carcass and environment samples. Sponges were aseptically cut in half and 500 ml BPW was added. They were then aseptically divided into two portions of 250 ml each to analyze the different bacteria.

The detection and isolation of *E. coli* O157:H7 and non-O157 STEC was performed according to USDA MLG 5C.00<sup>31</sup>, with some modifications. The *rfbO157* gene and Shiga-toxin (*stx*) genes *stx*<sub>1</sub> and *stx*<sub>2</sub> were screened by multiplex-polymerase chain reaction (PCR)<sup>28</sup>. *Escherichia coli* serotyping was carried out using agglutination tests on a microplate, slide and/or tube containing specific antisera (Instituto Nacional de Producción de Biológicos [INPB] – ANLIS "Dr. Carlos G. Malbrán", Argentina). Cultures that could not be resolved by serotyping were subjected to multiplex PCR to determine the gene encoding somatic antigen (O)<sup>1</sup>.

Isolation of *Salmonella* spp. was carried out according to ISO 6579-1:2017<sup>24</sup> and serotyping was performed according to the White-Kauffmann-Le Minor scheme by slide (O antigen) and tube (H antigen) agglutination, using specific antisera (INPB – ANLIS).

**Table 1** Risk estimation obtained in the operational and post-operational visit of abattoirs I and II.

Abattoir	Operational			Post-operational		
	Block	Average scores <sup>a</sup>	Risk estimation <sup>b</sup>	Block	Average scores <sup>a</sup>	Risk estimation <sup>b</sup>
I	Pens	4.7	H	Pens	5.6	H
	Slaughter area	11	H	Slaughter area	17.6	H
	Head and viscera	2.9	H	Cool chambers	2.9	H
	Control points	11.6	M	Quartering	-	-
	Cool chambers	3.5	H	Offal area	6.9	H
	Offal area	7	H	Outdoors	3.0	H
	Total	39.7	H	Total	38.7	H
II	Pens	7	H	Pens	5.0	H
	Slaughter area	10.7	H	Slaughter area	17.1	H
	Head and viscera	6.3	H	Cool chambers	6.2	M
	Control points	6.4	H	Quartering	-	-
	Cool chambers	10.2	M	Offal area	9.5	M
	Offal area	8.9	M	Outdoors	2.7	H
	Total	49.6	M	Total	41.9	M

<sup>a</sup> Average scores: average final score obtained in total visits (10 operational and 10 post-operational checklists for each abattoir).

<sup>b</sup> Risk estimation: H: high; M: moderate; L: low.

## Statistical analyses

The microbiological quality of carcasses (mesophilic microorganisms, coliforms and *E. coli* counts) from both abattoirs was evaluated using the Student's *t*-test for independent variables. Enumeration data were log-transformed before performing the analysis of variance. All statistical analyses were performed using IBM® SPSS® version 24. Significance was set at *p*<0.05.

## Results

### Risk estimation

Risk was high to moderate in the two abattoirs and in both operational and post-operational processes, with predominance of high risk.

In abattoir I, the results obtained with the operational checklist showed that risk was high and moderate (range, 33–49; 10 visits), with an average of 39.7 (high) in the pens, slaughter, head and viscera, cool chamber and offal sectors. Only control points were rated as having moderate risk. Likewise, the risk values obtained with the post-operational checklist were also classified as high and moderate (range, 26–51; 10 visits), and 38.7 in average (high risk). Such high risk was detected in all sectors (pens, slaughter area, cool chambers, offal area and outdoors) during at least one visit. The average scores of each block are shown in Table 1.

In abattoir II, the operational checklist rendered moderate risk results in the cool chambers and offal area (range, 46–53; 10 visits), with an average of 49.6. However, the risk was high in the pens, slaughter area, head and viscera area and control points. Concerning the post-operational checklist, the values obtained were in the range of 33–48 (10 visits) and also classified as high (pens, slaughter area and outdoors in at least one visit) and moderate (cool chambers

and offal area) risk. In this case, the average corresponded to moderate risk (41.9). The average scores of each block are shown in Table 1.

The main deviations from HACCP observed in both abattoirs were (a) deficient building conditions: pen infrastructure, slaughter area, carcass and offal cool chambers (walls, floors and ceilings, platforms, lighting and ventilation, among others); (b) inadequate workflow, including lack of workers; (c) lack of sectorization of changing rooms and bathrooms; (d) lack of implementation of SSOP, particularly during the post-operational process, and (e) no food safety training of workers.

### Bacteriological analysis

The counts of mesophilic microorganisms, coliforms and *E. coli* from abattoirs I and II did not differ significantly. In Argentina, only *E. coli* is included as microbiological criteria for microorganism counts on carcass surfaces (limit,  $\leq 0.7 \log \text{CFU/cm}^2$ )<sup>35</sup>. In abattoir I, counts were as follows ( $\log \text{CFU/cm}^2$ ): mesophilic,  $3.1 \pm 0.14$ ; coliform,  $0.56 \pm 0.01$ ; *E. coli*,  $0.49 \pm 0.03$ . According to local regulations (SENASA, 2008), 32 samples (40%) did not comply with the microbiological criteria for *E. coli*. Likewise, abattoir II showed the following counts ( $\log \text{CFU/cm}^2$ ): mesophilic,  $2.6 \pm 0.42$ ; coliform,  $0.47 \pm 0.05$ ; *E. coli*,  $0.41 \pm 0.07$ . Accordingly, 13 samples (16.2%) did not meet the microbiological criteria for *E. coli*.

The *stx* genes were detected in 39 (24.4%) carcass and 42 (30.9%) environmental samples. While all samples (carcass and environmental) and sampling times (operational and post-operational) from both abattoirs had at least one *stx*-positive sample, *E. coli* O157:H7 was not isolated from any of them. On the other hand, STEC was isolated from three (1.9%) carcass and five (3.7%) environmental samples, as follows: in abattoir I, it was

**Table 2** STEC detection and isolation and *Salmonella* spp. isolation from carcass and environmental samples from each abattoir during operational and post-operational processes.

Microorganism	Abattoir I				Abattoir II			
	Carcass		Environment		Carcass		Environment	
	Operational n = 40 (%)	Post-operative- n = 40 (%)	Operational n = 40 (%)	Post-operative- n = 30 (%)	Operational n = 40 (%)	Post-operative- n = 40 (%)	Operational n = 36 (%)	Post-operative- n = 30 (%)
<b>STEC</b>								
Detection	13 (32.5)	16 (40.0)	14 (35.0)	8 (26.7)	7 (17.5)	3 (7.5)	6 (16.6)	14 (46.7)
Isolation	0	2 (5.0)	1 (2.5)	0	0	1 (2.5)	1 (2.8)	3 (10.0)
<i>Salmonella</i> spp. isolation	0	2 (5.0)	0	1 (3.3)	2 (5.0)	2 (5.0)	6 (16.6)	3 (10.0)

**Table 3** Sources of STEC detection and isolation and *Salmonella* spp. isolation.

Microorganism	Platforms		Knives		Workers' hands		Saw machines		Trays/sinks	
	I n = 19 (%)	II n = 20 (%)	I n = 10 (%)	II n = 8 (%)	I n = 10 (%)	II n = 8 (%)	I n = 20 (%)	II n = 20 (%)	I n = 10 (%)	II n = 10 (%)
<b>STEC</b>										
Detection	6 (31.6)	7 (35.0)	3 (30.0)	1 (12.5)	4 (40.0)	1 (12.5)	7 (35.0)	8 (40.0)	2 (20.0)	3 (30.0)
Isolation	1 (5.3)	1 (5.0)	0	0	0	1 (100.0)	0	0	0	2 (20.0)
<i>Salmonella</i> spp. isolation	0	1 (5.0)	0	2 (25.0)	0	3 (37.5)	0	2 (10.0)	1 (10.0)	1 (10.0)

isolated from two carcasses (post-operational) and one environmental (operational) sample. The STEC isolates were characterized as *E. coli* O8:H7/stx<sub>1</sub> and O116:H49/stx<sub>2</sub> in carcass and O136:H40/stx<sub>2</sub> in platform samples. In abattoir II, STEC was isolated from one carcass (post-operational) and four environmental (operational and post-operational) samples, and the isolates were characterized as *E. coli* O113:H21/stx<sub>2</sub> (carcass), O116:H49/stx<sub>2</sub> and O7:H7/stx<sub>2</sub> (trays), O8:H16/stx<sub>2</sub> (platforms) and O178:H19/stx<sub>2</sub> (workers' hands).

*Salmonella* spp. was isolated from six (7.5%) carcass and 10 (7.3%) environmental samples. In abattoir I, *Salmonella* isolates (n = 3) came from the post-operational process. The serovars identified were *S. Cerro* in two carcass samples and *S. Corvallis* in one tray sample. In abattoir II, *Salmonella* was isolated from carcass and environmental samples (n = 13) in both the operational and post-operational processes. Serovars were identified as *S. Cerro* (n = 4, platform, saw machine, knives and worker's hands), *S. Corvallis* (n = 3, carcass), *S. Havana* (n = 3, two from workers' hands and one from knife) and *S. Agona* (n = 3, carcass, saw machine and tray).

Results of STEC detection and isolation and *Salmonella* spp. isolation in carcass and environmental samples are shown in Table 2. Sources of *Salmonella* spp. isolation and STEC detection and isolation from environmental samples are detailed in Table 3.

## Discussion

In this work, the comprehensive study of two provincial transit abattoirs with no HACCP plan from Tucumán, Argentina, was carried out through risk estimation and sampling of carcasses and the environment.

Around 30.0% of the beef produced in Argentina is for export, and the remaining 70.0% is destined for domestic consumption<sup>8</sup>. Exporting companies produce premium quality products which comply with the high levels of demand and the required high quality standards. In the domestic market, however, these standards depend on local demands and local-state control. Like in other countries, this may occasionally result in multiple abattoir hygiene and sanitation standards<sup>17,25,34</sup>. It has been shown that consuming meat from abattoirs without HACCP increases the risk of contracting diseases such as hemolytic uremic syndrome<sup>8</sup>. In this sense, 16% of the Argentinian bovine slaughter comes from provincial abattoirs with no HACCP plan<sup>16</sup>.

In a previous study conducted in Argentinian abattoirs, risk estimation checklists and microbiological analyses were useful to identify relevant deviations from HACCP<sup>16</sup>. In this study, the same model was applied in the two abattoirs evaluated during the operational and post-operational processes. The results obtained indicated the predominance of high risk of contamination in both abattoirs and highlighted the lack of control along the production line and

after the sanitation process, two important points where the microbiological quality of meat should be ensured<sup>13</sup>. In addition, the existence of major failures was evidenced, not only in buildings (lack of sanitary filters, divisions between dirty and clean areas, enclosures) and processes (lack of GMP, GHP, SSOP, HACCP), but also in the lack of safety training of workers. Similar problems have been recently reported in abattoirs from Ethiopia and Namibia<sup>6,21,32</sup>. In this regard, three Argentinian abattoirs without HACCP managed to reduce the risk of contamination from high to moderate through the implementation of improvement actions<sup>16</sup>. Based on the results obtained in the current study, it would be possible to implement an improvement plan in each abattoir from Tucumán jointly with the local health authorities, including the training of workers.

The mesophilic counts reported here were similar to those obtained in Buenos Aires (BA) abattoirs without HACCP<sup>16</sup>, but lower by more than 1 log CFU/cm<sup>2</sup> in abattoir II compared with BA results. Coliform counts were in the same order as those obtained in two abattoirs from BA after applying improvement actions<sup>16</sup>, while *E. coli* counts were higher. However, counts in abattoirs without HACCP from Argentina were lower (1–4 log CFU/cm<sup>2</sup>) than those from Pakistan and Ethiopia<sup>2,6</sup>. On the other hand, mesophilic, coliform and *E. coli* counts were almost 2 log CFU/cm<sup>2</sup> higher in Argentine abattoirs without HACCP than in exporting abattoirs applying an HACCP plan<sup>36</sup>, supporting the importance of applying a quality control system.

High counts can be indicative of the possible presence of pathogens<sup>29</sup>, although low counts do not ensure the total absence of potentially pathogenic bacteria such as STEC. In this study, the detection of *stx* from carcass and environmental samples was low (24.4 and 30.9%, respectively). In BA abattoirs, *stx* was detected in 97.2% of carcass and 76.1% of environmental samples<sup>15</sup>. Differences could be due to the *stx* screening technique used, i.e., real-time PCR in Costa<sup>15</sup> and conventional multiplex PCR in this work, whose detection limit is 1000 times higher<sup>28</sup>. In addition, the PCR currently used did not include an internal amplification control, so that the detection of *stx* could have been underestimated by false negative results<sup>23</sup>. On the other hand, the limited success to isolate non-O157 STEC should be highlighted. In BA abattoirs, STEC were isolated from 18.8% carcass and 6.1% environmental samples in the initial stage<sup>16</sup>, while in the current work this was only possible in 1.9% carcass and 3.7% environmental samples. Factors such as high levels of background bacteria, volume of samples plated, amount of plates necessary to achieve STEC isolates, number of colonies selected per plate, and laboratory personnel experience probably impaired the accurate isolation of STEC<sup>4,9,17,26</sup>.

The STEC serotypes O8:H16, O113:H21 and O178:H19 isolated in this study have been previously associated with unique cases of hemolytic uremic syndrome<sup>20</sup>. However, there are no reports of disease cases caused by these serotypes in Tucumán. Serotypes O113:H21 and O178:H19 were also reported in carcass, viscera and environmental samples from BA provincial abattoirs<sup>16</sup>. The other STEC serotypes isolated in this study were previously isolated in cattle from Argentina, reaffirming that cattle are a reservoir for a wide variety of STEC strains<sup>11,18,19,38</sup>. Meanwhile, *E. coli* O157:H7 was not isolated, probably due to the high

levels of background flora detected in the samples and the low capacity to compete for this *E. coli* serotype<sup>37,39</sup>.

A wide variety of reports account for the presence of *Salmonella* spp. both in carcasses and abattoir environments, highlighting the possibility of cross-contamination<sup>1,26,31</sup>. The prevalence of *Salmonella* spp. in carcasses differs among countries, ranging from 0.1% in Mid-western USA<sup>5</sup> to 18.0% in Mexico<sup>30</sup> and 9.9% in Ethiopia<sup>6</sup>. In the present study, the prevalence of *Salmonella* spp. was 3.8% in carcass and 7.4% in environmental samples from both abattoirs. Such prevalence was higher than that described by Costa et al.<sup>16</sup> in abattoirs from BA (Argentina), where *Salmonella* spp. was not detected in carcasses, being its prevalence 4.1% in environmental samples after the implementation of improvement actions.

The *Salmonella* serovars isolated here were *S. Cerro*, *S. Corvallis*, *S. Agona* and *S. Havana*. Even though all of them had already been reported in Argentina in different types of samples<sup>14,16</sup>, none of them corresponded to the most prevalent serovars in human diseases in Argentina<sup>12</sup>. Considering that *Salmonella* can be transmitted directly and indirectly<sup>7</sup>, its presence in any food compromises food safety and is indicative of poor hygiene practices such as regular and effective handwashing after visiting the toilet or mixing meat with intestinal contents during carcass dressing.

Non-compliance of companies with regulations and lack of adequate state-control has been defined as "organized irresponsibility"<sup>3</sup>. In Argentina, with the exception of exporting companies, provincial bovine abattoirs do not receive economic stimulus for applying high hygiene standards. This does not promote fair competition, considering that abattoirs applying good hygiene practices and selling their production in the local market do not receive an appropriate economic reward either.

## Conclusions

Based on the results of this work, it was possible to corroborate that checklists together with sampling to validate their application were useful tools to know the sanitary condition of two abattoirs, mainly classified as having high risk. Carcasses had high counts of indicators (mesophilic bacteria, coliforms and *E. coli*) as well as STEC and *Salmonella*, which were found on the products and the environment at two specific times of the production process, operational and post-operational. These results will allow to plan improvements in Tucumán abattoirs aimed at solving the problems identified through risk estimation and microbiological analysis. Still, the need to work jointly with the sanitary authority in search of a single sanitary standard for Argentina based on the implementation of HACCP in all abattoirs, regardless of category, remains unaddressed.

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## Conflict of interest

The authors declare that they have no conflicts of interest.

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## References

1. Acuña P, Florentín M, Rojas N, Rodríguez F, Guillén R. Estandarización de una técnica de PCR múltiple para la detección de los serogrupos O157, O104 y "big six" de *Escherichia coli* productora de la toxina Shiga (STEC). Mem Inst Investig Cienc Salud. 2019;17:71–6, <http://dx.doi.org/10.18004/mem.iics/1812-9528/2019.017.02.71-076>.
2. Ahmad MUD, Sarwar A, Najeeb MI, Nawaz M, Anjum AA, Ali MA, Mansur N. Assessment of microbial load of raw meat at abattoirs and retail outlets. J Anim Plant Sci. 2013;23:745–8.
3. Albersmeier F, Schulze H, Jahn G, Spiller A. The reliability of third-party certification in the food chain: from checklists to risk-oriented auditing. Food Control. 2009;20:927–35.
4. Auvray F, Lecureuil C, Dilasser F, Tache J, Derzelle S. Development of a real-time PCR assay with an internal amplification control for the screening of Shiga toxin-producing *Escherichia coli* in foods. Lett Appl Microbiol. 2009;48:554–9.
5. Barkocy-Gallagher GA, Arthur TM, Rivera-Betancourt M, Nou X, Shackelford SD, Wheeler TL, Koochmarie M. Seasonal prevalence of Shiga toxin-producing *Escherichia coli*, including O157:H7 and non-O157 serotypes, and *Salmonella* in commercial beef processing plants. J Food Prot. 2003;66:1978–86, <http://dx.doi.org/10.4315/0362-028x-66.11.1978>.
6. Bersisa A, Tulu D, Negera C. Investigation of bacteriological quality of meat from abattoir and butcher shops in Bishoftu, Central Ethiopia. Int J Microbiol. 2019;1–8.
7. Bogere P, Baluka SA. Microbiological quality of meat at the abattoir and butchery levels in Kampala City, Uganda. Internet J Food Saf. 2014;16:29–35.
8. Brusa V, Costa M, Padola NL, Etcheverría A, Sampedro F, Fernandez PS, Leotta GA, Signorini ML. Quantitative risk assessment of haemolytic uremic syndrome associated with beef consumption in Argentina. PLOS ONE. 2020;15:e0242317, <http://dx.doi.org/10.1371/journal.pone.0242317>.
9. Brusa V, Piñeyro PE, Galli L, Linares LH, Ortega EE, Padola NL, Leotta GA. Isolation of Shiga toxin-producing *Escherichia coli* from ground beef using multiple combinations of enrichment broths and selective agars. Foodborne Pathog Dis. 2016;13:163–70, <http://dx.doi.org/10.1089/fpd.2015.2034>.
10. Brusa V, Restovich V, Signorini M, Pugin D, Galli L, Diaz VR, Arias R, Leotta GA. Evaluation of intervention measures at different stages of the production chain in Argentinian exporting abattoirs. Food Sci Technol Int. 2019;25:491–6, <http://dx.doi.org/10.1177/1082013219836326>.
11. Cáceres ME, Etcheverría AI, Fernández D, Rodríguez EM, Padola NL. Variation in the distribution of putative virulence and colonization factors in Shiga toxin-producing *Escherichia coli* isolated from different categories of cattle. Front Cell Infect Microbiol. 2017;7:147, <http://dx.doi.org/10.3389/fcimb.2017.00147>.
12. Caffer MI, Alcain A, Panagopulo M, Moroni M, Brengi S, Terragno R. *Salmonella* serovars in Argentina, 2007–2009. Rev Argent Microbiol. 2010;42:80.
13. Cevallos-Almeida M, Burgos-Mayorga A, Gómez CA, Lema-Hurtado JL, Lema L, Calvache I, Jaramillo C, Ruilova IC, Martínez EP, Estupiñán P. Association between animal welfare indicators and microbiological quality of beef carcasses, including *Salmonella* spp., from a slaughterhouse in Ecuador. Vet World. 2021;14:918–25, <http://dx.doi.org/10.14202/vetworld.2021.918-925>.
14. Colello R, Ruiz MJ, Padín VM, Rogé AD, Leotta G, Padola NL, Etcheverría AI. Detection and characterization of *Salmonella* serotypes in the production chain of two pig farms in Buenos Aires Province, Argentina. Front Microbiol. 2018;9:1370, <http://dx.doi.org/10.3389/fmicb.2018.01370>.
15. Costa M [Doctoral dissertation] *Calidad microbiológica de la carne bovina y cuantificación del riesgo en plantas de faena de la provincia de Buenos Aires*. Universidad Nacional de La Plata; 2019.
16. Costa M, Pracca G, Sucari A, Galli L, Ibargoyen J, Gentiliuomo J, Brusa V, Martinez Zugazua M, Figueroa Y, Londero A, Roge A, Silva H, Van Der Ploeg C, Signorini M, Oteiza JM, Leotta GA. Comprehensive evaluation and implementation of improvement actions in bovine abattoirs to reduce pathogens exposure. Prev Vet Med. 2020;176:104933, <http://dx.doi.org/10.1016/j.prevetmed.2020.104933>.
17. Essendoubi S, Stashko N, So I, Gensler G, Rolheiser D, Mainali C. Prevalence of Shiga toxin-producing *Escherichia coli* (STEC) O157: H7, six non-O157 STECs, and on beef carcasses in provincially licensed abattoirs in Alberta, Canada. Food Control. 2019;105:226–32.
18. Fernández D, Krüger A, Polifroni R, Bustamante AV, Sanso AM, Etcheverría AI, Lucchesi PM, Parma AE, Padola NL. Characterization of Shiga toxin-producing *Escherichia coli* O130:H11 and O178:H19 isolated from dairy cows. Front Cell Infect Microbiol. 2013;3:9, <http://dx.doi.org/10.3389/fcimb.2013.00009>.
19. Fernández D, Sanz ME, Parma AE, Padola NL. Short communication: characterization of Shiga toxin-producing *Escherichia coli* isolated from newborn, milk-fed, and growing calves in Argentina. J Dairy Sci. 2012;95:5340–3, <http://dx.doi.org/10.3168/jds.2011-5140>.
20. Galli L, Miliwebsky E, Irino K, Leotta G, Rivas M. Virulence profile comparison between LEE-negative Shiga toxin-producing *Escherichia coli* (STEC) strains isolated from cattle and humans. Vet Microbiol. 2010;143:307–13, <http://dx.doi.org/10.1016/j.vetmic.2009.11.028>.
21. Gutema FD, Agga GE, Abdi RD, Jufare A, Duchateau L, De Zutter L, Gabrél S. Assessment of hygienic practices in beef cattle slaughterhouses and retail shops in Bishoftu, Ethiopia: implications for public health. Int J Environ Res Public Health. 2021;18:2729, <http://dx.doi.org/10.3390/ijerph18052729>.
22. Hameed MAK, Muhammed HA, Al-Rashedi AAM. The bacterial load and contamination rate of raw meat in butcher shops in different areas of Karbala City. Pak J Med Health Sci. 2021;15: 960.
23. Hoofar J, Malony B, Abdulmawjood A, Cook N, Wagner M, Fach P. Practical considerations in design of internal amplification controls for diagnostic PCR assays. J Clin Microbiol. 2004;42:1863–8, <http://dx.doi.org/10.1128/JCM.42.5.1863-1868.2004>.
24. The International Organization for Standardization. *Microbiology of the food chain – horizontal method for the detection, enumeration and serotyping of Salmonella – Part 1: Detection of *Salmonella* spp.* 6579-1:2017. Geneva: International Organization for Standardization; 2017.
25. Jaja IF, Green E, Muchenje V. Aerobic mesophilic, coliform, *Escherichia coli*, and *Staphylococcus aureus* counts of raw meat from the formal and informal meat sectors in South Africa. Int J Environ Res Public Health. 2018;15:819, <http://dx.doi.org/10.3390/ijerph15040819>.
26. Ju W, Shen J, Li Y, Toro MA, Zhao S, Ayers S, Najjar MB, Meng J. Non-O157 Shiga toxin-producing *Escherichia coli* in retail ground

- beef and pork in the Washington D.C. area. *Food Microbiol.* 2012;32:371–7, <http://dx.doi.org/10.1016/j.fm.2012.07.017>.
27. Kore K, Asrade B, Demissie K, Aragaw K. Characterization of *Salmonella* isolated from apparently healthy slaughtered cattle and retail beef in Hawassa, southern Ethiopia. *Prev Vet Med.* 2017;147:11–6, <http://dx.doi.org/10.1016/j.prevetmed.2017.08.018>.
28. Leotta GA, Chinen I, Epszteyn S, Miliwebsky E, Melamed IC, Motter M, Ferrer M, Marey E, Rivas M. Validación de una técnica de PCR múltiple para la detección de *Escherichia coli* productor de toxina Shiga [Validation of a multiplex PCR for detection of Shiga toxin-producing *Escherichia coli*]. *Rev Argent Microbiol.* 2005;37:1–10.
29. Lues JFR, Van Tonder I. The occurrence of indicator bacteria on hands and aprons of food handlers in the delicatessen sections of a retail group. *Food Control.* 2007;18:326–32.
30. Martínez-Chávez L, Cabrera-Díaz E, Pérez-Montaño JA, Garay-Martínez LE, Varela-Hernández JJ, Castillo A, Lucia L, Ávila-Novoa MG, Cardona-López MA, Gutiérrez-González P, Martínez-González NE. Quantitative distribution of *Salmonella* spp. and *Escherichia coli* on beef carcasses and raw beef at retail establishments. *Int J Food Microbiol.* 2005;210:149–55, <http://dx.doi.org/10.1016/j.ijfoodmicro.2005.06.016>.
31. USDA-FSIS. Detection, isolation and identification of top seven Shiga toxin-producing *Escherichia coli* (STECs) from meat products and carcass and environmental sponges. *Microbiology Laboratory Guidebook.* Athens, GA, USA: United States Department of Agriculture, Food Safety and Inspection Service (USDA FSIS), Office of Public Health Science; 2019. Available at <https://www.fsis.usda.gov/wps/portal/fsis/>
32. Nehoya KN, Hamatui N, Shilangale RP, Onywera H, Kennedy J, Mwapaghla LM. Characterization of Shiga toxin-producing *Escherichia coli* in raw beef from informal and commercial abattoirs. *PLOS ONE.* 2020;15:e0243828, <http://dx.doi.org/10.1371/journal.pone.0243828>.
33. Pérez Terrazzino GB, Condorí MS, López Campo A, Vega S, Carbonari C, Chinen I, Rivas M, de Castillo MC, Jure MA. Calidad higiénico-sanitaria en plantas de faena de la provincia de Tucumán. Detección, aislamiento y caracterización de *Escherichia coli* productor de toxina Shiga [Hygienic-sanitary quality in abattoirs from Tucuman province, Argentina. Detection, isolation and characterization of Shiga toxin-producing *Escherichia coli*]. *Rev Argent Microbiol.* 2017;49:242–6, <http://dx.doi.org/10.1016/j.ram.2016.11.005>.
34. Santángelo F, Robert S. Análisis tecnológico sectorial, Cap. XXI, Complejo productivo frigorífico-bovino. October 2012. p. 603–34. <https://www.argentina.gob.ar/ciencia/publicaciones/estudios/publicaciones/analisis-tecnologico-sectorial> [online].
35. Servicio Nacional de Sanidad y Calidad Agroalimentaria. Circular SENASA No. 3834: Prevención y Control de *Escherichia coli* O157:H7; 2008.
36. Signorini M, Costa M, Teitelbaum D, Restovich V, Brasesco H, García D, Superno V, Petroli S, Bruzzone M, Arduini V, Vanzini M, Sucari A, Suberbé G, Maricel T, Rodríguez R, Leotta GA. Evaluation of decontamination efficacy of commonly used antimicrobial interventions for beef carcasses against Shiga toxin-producing *Escherichia coli*. *Meat Sci.* 2018;142:44–51, <http://dx.doi.org/10.1016/j.meatsci.2018.04.009>.
37. Tamplin ML. Growth of *Escherichia coli* O157:H7 in raw ground beef stored at 10°C and the influence of competitive bacterial flora, strain variation, and fat level. *J Food Prot.* 2002;65:1535–40, <http://dx.doi.org/10.4315/0362-028x-65.10.1535>.
38. Tanaro JD, Galli L, Lound LH, Leotta GA, Piaggio MC, Carbonari CC, Irino K, Rivas M. Non-O157:H7 Shiga toxin-producing *Escherichia coli* in bovine rectums and surface water streams on a beef cattle farm in Argentina. *Foodborne Pathog Dis.* 2012;9:878–84, <http://dx.doi.org/10.1089/fpd.2012.1182>.
39. Vold L, Holck A, Wasteson Y, Nissen H. High levels of background flora inhibits growth of *Escherichia coli* O157:H7 in ground beef. *Int J Food Microbiol.* 2000;56:219–25, [http://dx.doi.org/10.1016/s0168-1605\(00\)00215-4](http://dx.doi.org/10.1016/s0168-1605(00)00215-4).
40. World Health Organization. Shiga toxin-producing *Escherichia coli* (STEC) and food: attribution, characterization and monitoring, vol. 19. Rome: Food and Agriculture Organization of the United Nations, World Health Organization; 2019.