



MICROBIOLOGICAL IMAGE

Argentinean *Bacillus thuringiensis* strains exhibiting distinct morphology of their parasporal crystals



Cepas argentinas de *Bacillus thuringiensis* con distinta morfología en sus cristales paraesporales

Cecilia Peralta^a, Diego Herman Sauka^b, Antonela Marozzi^c,
Eleodoro E. Del Valle^d, Leopoldo Palma^{a,*}

^a Centro de Investigaciones y Transferencia CIT-VM-CONICET, Universidad Nacional de Villa María, Villa María, Córdoba, Argentina

^b Instituto Nacional de Tecnología Agropecuaria (INTA), Instituto de Microbiología y Zoología Agrícola (IMYZA), Hurlingham, Buenos Aires, Argentina

^c Instituto de Investigaciones en Biodiversidad y Medioambiente (INIBIOMA) – CONICET – UNCo, San Martín de los Andes, Neuquén, Argentina

^d ICiagro Litoral, Universidad Nacional del Litoral, CONICET Facultad de Ciencias Agrarias, Esperanza, Santa Fe, Argentina

Received 30 April 2020; accepted 1 September 2020

Available online 12 November 2020

Bacillus thuringiensis is a Gram-positive and sporulated bacterium exhibiting insecticidal activity against a wide range of insects.³ During sporulation, this bacterium produces a number of different proteins forming crystalline inclusions adjacent to the spores (parasporal crystals). Among these insecticidal proteins, the most abundant are those commonly known as Cry (Crystal) proteins, which are responsible for exerting a toxic activity (upon ingestion) against insects of different species.⁵ For this reason, *B. thuringiensis* has proved to be the most efficient and used bioinsecticide to date.² However, *Spodoptera cosmioides*, *Spodoptera eridania* and *Agrotis* sp. (Lepidoptera) are species that are not yet controlled by some transgenic crops (e.g. Intacta RR2Pro

soybean). Thus, in an attempt to enlarge the host spectrum of this bacterium it is necessary to search for novel strains. In this work we show a sporulated *B. thuringiensis* Bt-UNVM_84 strain exhibiting a number of rare amorphous to spherical crystal combinations, whereas sporulated *B. thuringiensis* strain Bt-UNVM_94 showed quasi symmetric bipyramidal parasporal crystals, by using scanning electron microscopy (SEM) (Fig. 1). Strains Bt-UNVM_84 and Bt-UNVM_94 were isolated from Oncativo (Córdoba, Argentina) and Cululú (Santa Fe, Argentina), respectively. The insecticidal activity of these different *B. thuringiensis* strains is currently under investigation. Each strain was grown in liquid CCY sporulation medium⁶ for ~48 h (150 rpm) until no vegetative cells were observed under a light microscope. The presence of parasporal crystals was first determined using Coomassie blue stained slides¹ (1000×) under a Nikon E100 light microscope and confirmed later by a Nikon Ti-Eclipse phase

* Corresponding author.

E-mail address: palma.leopoldo@gmail.com (L. Palma).

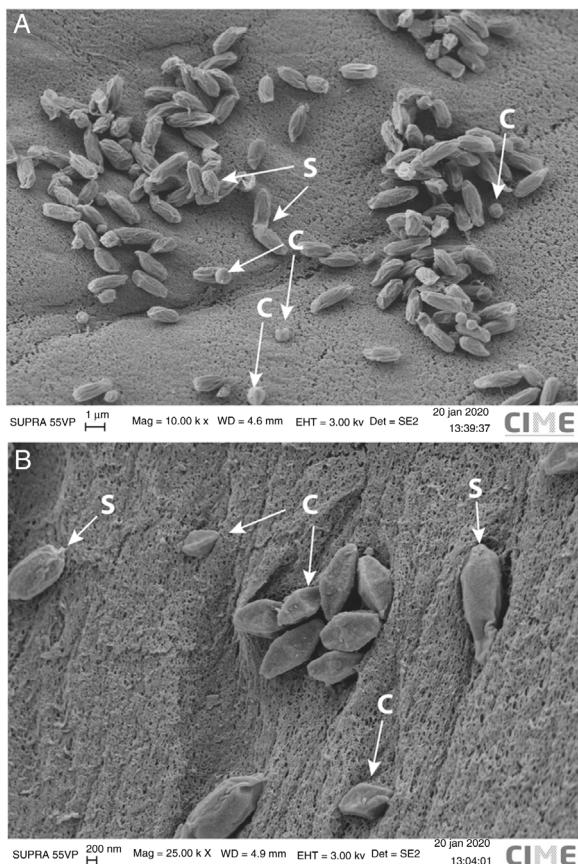


Figure 1 Scanning electron microscopy of parasporal crystals from *Bacillus thuringiensis* strains Bt-UNVM.84 and Bt-UNVM.94 (C=crystal, S=spore). (A) *B. thuringiensis* strain Bt-UNVM.84 showed combinations of amorphous to spherical parasporal crystals of ~0.7–0.9 μm from two points along the diametral axis. (B) *B. thuringiensis* strain Bt-UNVM.94 exhibited quasi symmetric bipyramidal parasporal crystals of ~1.0–1.2 μm from two points along the longitudinal axis. Crystal size was measured using ImageJ.⁴ Parameters used for image acquisition are shown: Mag=magnification (K \times =1000 \times), WD=work distance, Eht=energy high tension, Det=detector type and SE2=secondary electron.

contrast microscope (1000 \times) (data not shown). For the SEM analysis, aliquots of 1 ml were centrifuged for 5 minutes (16,000 g) at room temperature. Each pellet was washed three times with sterile distilled water and fixed with 100 μl 4% formaldehyde. Each fixed preparation was then sent to Centro Integral de Microscopía Electrónica (CIME – CONICET – UNT) for SEM examination.

Funding

This work was supported by Universidad Nacional de Villa María research grant PIC UNVM 2018-2019.

Acknowledgments

We thank to Hernán Esquivel, Luciano Martínez from CIME-CONICET-UNT and Vanessa Areco for their technical assistance. We also thank to the Instituto de Investigación of Universidad Nacional de Villa María for contributing to this ongoing research.

References

1. Ammons D, Rampersad J, Khan A. Usefulness of staining parasporal bodies when screening for *Bacillus thuringiensis*. *J Invertebr Pathol.* 2002;79:203–4.
2. Jouzani GS, Valijanian E, Sharifi R. *Bacillus thuringiensis*: a successful insecticide with new environmental features and tidings. *Appl Microbiol Biotechnol.* 2017;101:2691–711.
3. Palma L. *Bacillus thuringiensis*-based biopesticides, are they as effective as they should be? *Rev Argent Microbiol.* 2017;49:119.
4. Rueden CT, Schindelin J, Hiner MC, DeZonia BE, Walter AE, Arena ET, Eliceiri KW. ImageJ2: ImageJ for the next generation of scientific image data. *BMC Bioinformatics.* 2017;18:529.
5. Sauka DH, Benintende GB. Diversity and distribution of lepidopteran-specific toxin genes in *Bacillus thuringiensis* strains from Argentina. *Rev Argent Microbiol.* 2017;49:273–81.
6. Stewart GS, Johnstone K, Hagelberg E, Ellar DJ. Commitment of bacterial spores to germinate. A measure of the trigger reaction. *Biochem J.* 1981;198:101–6.