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REVIEW

Probiotics function and modulation of the immune system in allergic diseases

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Abstract Allergic diseases have been a global problem over the past few decades. The effect of allergic diseases on healthcare systems and society is generally remarkable and is considered as one of the most common causes of chronic and hospitalized disease. The functional ability of probiotics to modulate the innate/acquired immune system leads to the initiation of mucosal/systemic immune responses. Gut microbiota plays a beneficial role in food digestion, development of the immune system, control/growth of the intestinal epithelial cells and their differentiation. Prescribing probiotics causes a significant change in the intestinal microflora and modulates cytokine secretion, including networks of genes, TLRs, signaling molecules and increased intestinal IgA responses. The modulation of the Th1/Th2 balance is done by probiotics, which suppress Th2 responses with shifts to Th1 and thereby prevent allergies. In general, probiotics are associated with a decrease in inflammation by increasing butyrate production and induction of tolerance with an increase in the ratio of cytokines such as IL-4, IL-10/IFN- γ , Treg/TGF- β , reducing serum eosinophil levels and the expression of metalloproteinase-9 which contribute to the improvement of the allergic disease's symptoms. Finally, it can be said that the therapeutic approach to immunotherapy and the reduction of the risk of side effects in the treatment of allergic diseases is the first priority of treatment and the final approach that completes the first priority in maintaining the condition and sustainability of the tolerance along with the recovery of the individual

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Introduction

Allergic diseases have been a global problem over the past few decades.¹ The prevalence of eczema (atopic dermatitis), food allergies and asthma have increased dramatically during this period, especially in Western societies. It is believed that between 20–30% of people living in Western countries are suffering from at least one type of allergic disease. The effect of allergic diseases on health care systems and society is generally remarkable and is one of the most common causes of chronic and hospitalized illness.^{2,3} Allergic diseases are described with an inadequate immune response of T helper (Th2) cell lymphocytes to environmental or food antigens. Activating this response results in the secretion of interleukins (IL)-4, IL-5, IL-13, and the production of IgE specifically for allergens. The induction of Th2 cytokine responses also inhibits Th1 activity primarily via interferon (IFN)- γ , which helps maintain an allergic phenotype. The stability of the Th1/Th2 balance is governed by the expression of the transcription factors GATA-3 (Th2) and T-bet (Th1).^{4,5}

Several studies have shown that in patients with allergies, the number and function of regulatory T cells (Tregs) is associated with a decrease in the immune response, while the mutation in the FoxP3 Treg transcription factor leads to severe immunosuppression. The mechanisms that cause allergic diseases in early life are not yet fully understood.⁶ One of the well-known and very prominent ideas is the impact of intestinal microbiota, where the composition and structure of commensal bacteria are associated with the developing immune system. Such interactions can affect the maturity of the immune system, which potentially results in Th2-type allergic responses. Similarly, preventive or therapeutic strategies targeting intestinal microbiota are the subject of increasing scientific research.⁷ In this review, the beneficial effects of probiotics on the prevention and treatment of some allergic diseases are critically discussed.

The prevalence of allergic diseases and asthma are increasing worldwide, particularly in low and middle-income countries. Allergic diseases are common in children and adolescents and result in high costs to the health care system, as well as having an adverse impact on the quality of life.⁸ Complexity and severity of allergic diseases include life-threatening anaphylaxis, food allergies, certain forms of asthma, rhinitis, conjunctivitis, angioedema, urticaria, eczema, eosinophilic disorders, include eosinophilic esophagitis, and drug and insect allergies continue to escalate particularly in children and young adults, who are bearing the highest burden of these trends.^{8,9} Globally, 300 million people suffer from asthma and about 200–250 million from food allergies.⁹ Moreover, one-tenth of the population are affected by drug allergies and 400 million from rhinitis.¹⁰ In the United States, the prevalence of asthma is approximately 7%. Allergic rhinitis (AR) is more prevalent than asthma, ranging from 10% to 20% (500 million) worldwide and in the United States ranging from 10% to 40%.¹¹

The prevalence of atopic disease has reportedly been on the rise in the United States and around the world, although rates may have plateaued recently. The International Study of Asthma and Allergies in Childhood reported that the life-

time prevalence rates of atopic disease stabilized at 20%.¹² This plateau is a mixture of a substantial increase in the prevalence of the atopic disease in Mexico, Chile, Kenya, Algeria, and Southeast Asia, with decreases in New Zealand and the United Kingdom, which previously had high rates.¹³

The increase of urbanization in the developing world may contribute to the higher occurrence of aeroallergic disease in an urban area as compared with rural locations. The hygiene hypothesis by industrialization/urbanization led to reduced microbial exposure and infectious agents (biasing the immune system away from a Th1 immune-mediated response toward to Th2, in early life in contrast to rural societies. In addition, another risk factor of the aeroallergic disease includes prolonged and high exposure to dust mite populations, cockroach, or rodent allergens, lifestyle and obesity, elevated concentrations of air pollution^{14,15} (Fig. 1).

In the allergic response, the immune system must identify the allergens and induce a dynamic immune response. Sensitization to a specific antigen is an essential part of allergic immune responses mediated by naive T and B lymphocytes. Primary, recognized and processed allergens presented by antigen-presenting cells (APC) to naive T cells (Th0).¹⁶ Th2 cell differentiation is linked to the local cytokine environment provided by interactions among the epithelium, resident dendritic cells (DCs), and regional lymph nodes. The crucial cytokines responsible for the allergic response include IL-4, IL-5, and IL-13, as well as innate lymphoid (ILC-2) cells which may intensify and preserve local Th2-driven allergic inflammation. These interleukins switched immunoglobulin production by heavy chain gene rearrangement to class IgE in B cells¹⁷ (Fig. 1).

Allergy treatment and allergen immunotherapy

The primary and best way to prevent allergy symptoms is to avoid allergens as much as possible. This includes removing the source of allergens from your home and other places where you spend time. There are many harmless prescription medicines to relieve allergy symptoms. Non-pharmacological treatment of these patients may cause allergy and manifestation of painful problems such as sinus or ear infections. Allergy medications include nasal corticosteroids, antihistamines, mast cell stabilizers, corticosteroid, epinephrine.^{18,19}

The third treatment is immunotherapy as an appropriate therapeutic strategy chosen for some allergy patients. The most common types of immunotherapy are allergy shots and sublingual immunotherapy (SLIT).²⁰ Allergy shots involve giving injections of allergens in an increasing dose over time. The person becomes progressively less sensitive to that allergen. Allergy shots can work well for some people with allergies to pollen, pets, dust, bees or other stinging insects, as well as for asthma. Allergy shots do not usually work well for allergies to food, medicines, feathers, or for hives or eczema. SLIT is another way to treat certain allergies without injections.²¹ Allergists give patients small doses of an allergen under the tongue. This exposure improves tolerance to the substance and reduces symptoms. SLIT is fairly safe and effective for the treatment of nasal allergies and asthma. SLIT tablets are currently available for dust mites,

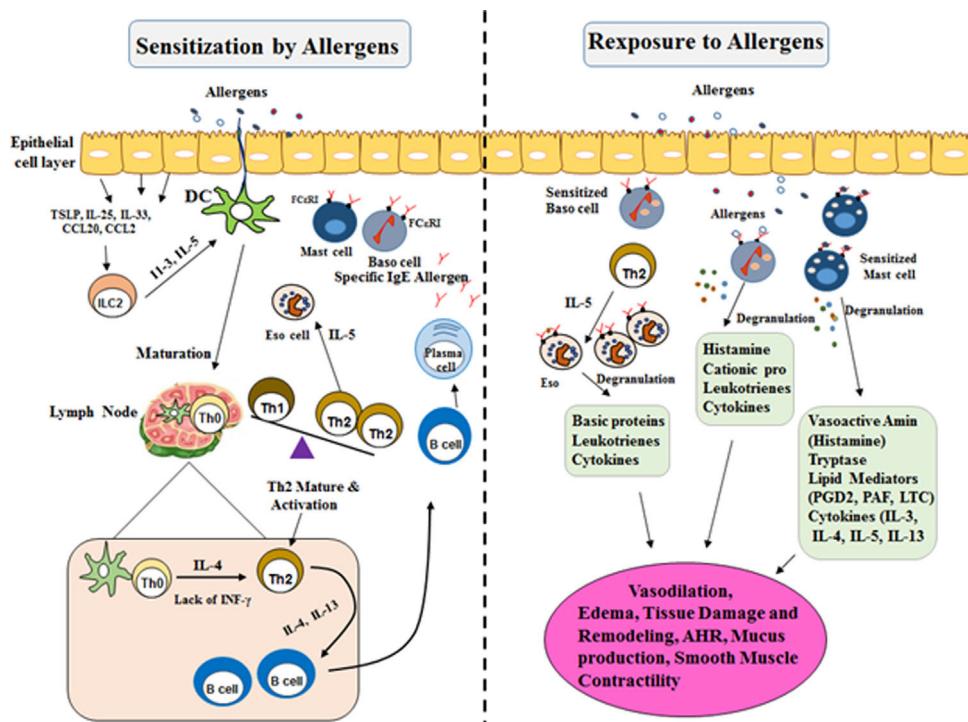


Figure 1 Mechanisms involved in allergic reactions. Sensitization to a specific antigen is a prerequisite for the onset of allergic diseases. Differentiation and expansion to Th2 cell subtypes lead to the production of inflammatory cytokines (IL-4, IL-5, and IL-13). They drive immunoglobulin E (IgE) class-switch in B cells and the recruitment and activation of pro-inflammatory cells in mucosal target organs. Allergic reactions are triggered when allergens cross-link preformed IgE bound to the high-affinity receptor Fc ϵ RI on Mast cells and Basophil cells. In the early phase of allergic reactions they alert the local immune cells and induce inflammatory reactions by secreting chemical mediators, leukotrienes, and cytokines. Mast cell and Basophil cell degranulation followed by a more constant inflammation, recognized as the late-phase response. This late response includes the employment of other effector cells, particularly Th2 lymphocytes, eosinophils, and basophils, which contribute significantly to the immunopathology of an allergic response. These activations contribute to the development of the inflammation and the symptoms of allergic disease.

TLR: Toll-like Receptor; TNF- α : Tumor Necrosis Factor A; IFN- γ : Interferon γ ; TGF- β : Transforming Growth Factor B; IL-10: Interleukin 10; IL-3: Interleukin 3; IL-4: Interleukin 4; IL-5: Interleukin 5; IL-13: Interleukin 13; IL-23: Interleukin 23; IL-25: Interleukin 25; Th0: T Cell Naive; Th2: Type 2 Helper T Cell; Th1: Type 1 Helper T Cell; IgE: Immunoglobulin E; DC: Dendritic Cells.

grass, and ragweed. While allergy shots are fairly safe, there is a chance of a severe, life-threatening allergic reaction to the injections, so they must always be given in an allergist's office under observation from a medical professional.²²

Merits and demerits of immunotherapy and new strategies for treating allergic diseases

When oral drugs and allergies are not able to control allergic reactions, allergy shots known as immunotherapy treatment may be the solution to this problem. There are many benefits to patients who receive an allergy shot, including some people who do not tolerate oral medication. While the cost of allergy shots is higher than oral medications, ultimately, the longer the cost of this treatment is less than the anti-allergic drug and should be spent by these patients who do not choose this treatment.²³ Oral allergy medications treat allergic symptoms, but allergy shots treat the cause of allergic reactions. When immunotherapy is successful, patients are recovered, because shots instead of symptoms cure the underlying causes of allergies, but like other types of medical treatments, the desensitization of allergy also

has disadvantages. Some of the common disadvantages of this type of allergy include: Immunotherapy does not affect all types of allergy. Shots do not have a good effect on common allergies such as pet, pollen, dust. Shots are not effective in treating allergies to food and hives. In addition, allergy to insect bites or spider bites cannot be prevented by allergic shots. In some patients, the reactions are reddening, swelling and hypersensitivity of the place of the injection. Losing these opportunities can delay or eliminate the treatment of allergies. Allergy shots will make allergic symptoms worse when the infusion regimen starts. Since allergic shots rarely cause anaphylaxis, patients should wait 30 min at the treatment faculty to reduce the risk of anaphylaxis.²⁴

One of the newest therapeutic approaches is allergenic changes that include the use of recombinant proteins, the combination of antigens with infectious carrier proteins, peptide immunotherapy and genetic vaccines containing code for allergic proteins, these methods include are oral and fecal immunotherapy, intralympathic immunotherapy, and epicutaneous immunotherapy.^{25,26} The first priority in the treatment of allergic diseases is the therapeutic approach to immunotherapy and the reduction of the risk

of side effects, and in the second stage, the maintenance of the tolerance and recovery of the individual.²⁷

The effect of microbiota and development of the immune system on allergic diseases

Reducing exposure to microorganisms in early life is one of the main mechanisms for increasing the prevalence of allergic diseases over the past few decades. This is typically referred to as the Health Hypothesis; which was originally used by Strachan (1989). Today, reducing exposure to microorganisms (increasing allergic conditions) has been pointed out to factors such as diet, antibiotic use, vaccination, households, and health improvements.²⁸ The digestive microbiota provide a significant amount of microbial contamination for infant growth. There are more than 10^{15} microorganisms which contain 1000 different species that are colonized in the gastrointestinal tract. Under normal conditions, these bacteria play a beneficial role in food digestion, development, and growth of the immune system, control, and growth of the intestinal epithelial cells and their differentiation.²⁹ Commensal bacteria also play an important role in the fermentation of dietary fiber, in addition to essential vitamins, a significant amount of short chain fatty acids (SCFAs) are also released. The colonization of commensal bacteria occurs immediately after birth and during the first years of life. The gastrointestinal microbiota acquisition affects the type of parturition, maternal microbiota, as well as genetic factors, breastfeeding, and other environmental factors.³⁰

One of the important functions of the intestinal microbiota is to contribute to the development of the immune system. The extensive antigenic stimulation after microbiota acquisition helps to organize the immune system through the development of gut-associated lymphoid tissue (GALT) and tolerogenic responses to harmless antigens, including food. These responses are performed by sampling and initial detection of antigens by intestinal epithelial cells and intrinsic immune cells through pattern recognition receptors (PRRs).³¹ These receptors are bonded to pathogen-associated molecular patterns (PAMPs), expressed by microorganisms. PAMPs include lipoteichoic acid on gram-positive bacteria and lipopolysaccharide on gram-negative bacteria. Identification of PAMPs by PRRs such as toll-like receptor (TLR)-2 and TLR4, triggers signals that determine the type of immune response, leads to the formation of a combination of regulatory and executive functions by DCs, Treg cells, chemokines, and cytokines.³²

The study of germ-free (GF) mice confirmed the key role of microbiota and their antigens in the development of GALT and inadequate immune responses. Several studies have shown that GF mice have dramatically reduced the number of intra-epithelial lymphocytes, payer's patches with incomplete germinal centers, and produce fewer IgA antibodies by plasma cells in the gastrointestinal lamina propria regions (which is actually the GALT executive region). Restoring different types of microbial species can help rebuild the GALT function in these mice, as well as the development and growth of the immune system. Additionally, in GF mice, there is no tolerance to prescriptive antigens due to the

deficiency of Treg cells. However, oral administration of microorganisms leads to stabilization of tolerance.³³⁻³⁵

Probiotics

Probiotics are defined as live microorganisms that confer a health benefit on the host when administered in adequate amounts.³⁶⁻³⁸ The history of the use of probiotics with extensive access to them reflects their high degree of safety.³⁹ These findings highlight the important role of microbiota in immune system development and its potential role for immune disorders, such as allergies and autoimmune disease, resulting in intestinal dysbiosis.^{40,41} The identification and use of probiotics date back to the early 20th century. Generally, probiotic bacteria need to have a set of criteria for inducing their beneficial effects, including: (1) resistance to PH, bile and digestive enzymes; (2) preventing the binding of pathogens and oral antigens to epithelial cells; (3) increasing the probability of biological efficacy in humans; (4) direct effect on toxicity of bacteria, viruses, fungi and parasites; and (5) proper tolerance and their importance in clinical use for their safety.^{42,43}

Safety of probiotics

The human body is colonized (mainly gastrointestinal tract) by 10–100 trillion microbes, including bacteria, archaea, viruses, and eukaryotic microbes. The indigenous intestinal microbiota is known to be a principal contributor in maintaining usual physiology, immune homeostasis, and energy production during life.⁴⁴ Protein products and metabolites of these microbes play an important role in the development and homeostasis of a number of body functions. The communication between these microorganisms and hosts has grown throughout the entire mammalian evolution, resulting in a distinctive gastrointestinal microbial community in humans.⁴⁵ The microbial community remains stable over time, moreover, diet and other environmental factors can affect the microbial community. Commensal microorganisms, in addition to inhibiting invasion of other pathogenic microorganisms; donate fermentation of non-digestible fibers; help to absorption of minerals and produce vitamins, SCFAs, and carbohydrates and also regulate inflammatory responses and the immune system.⁴⁶ A defect to maintain the balance among them (commensal microbiota and the host) interrupting the complex homeostasis (defined as dysbiosis) may cause a variety of syndromes (e.g. intestinal, metabolic, and immune disorders).⁴⁷

Probiotics are live microorganisms that are regarded as safe because of their long history of use in foods and dairy products for over a hundred years. Common probiotics are *Lactobacillus* and *Bifidobacterium*, some strains of *Enterococcus* and generally *Saccharomyces* species.⁴⁸ Because probiotics have been shown to affect both the innate and adaptive immune systems, a theoretical concern about the potential of probiotics to stimulate the immune response in some individuals, possibly leading to autoimmune or inflammation has not been reported in any human subjects.³⁴

The gut acts as an extremely discerning barrier and communication organ between the luminal bacterial environment and the host. Therefore, failure of this commu-

nication due to the loss of barrier function or failure of tolerance mechanisms have been associated with diseases, usually as opportunistic infections in some predisposing patients and development of inflammation. Additionally, the presence of transferable antibiotic resistance genes, which comprises a theoretical risk of transfer to a less harmless member of the gut microbial community, must also be considered.⁴⁹

Therefore, the safety of probiotics is related to the nature of the specific microbe proposed use, dose, and duration.⁵⁰ Overall, probiotics when administered in adequate amounts constitute a health benefit on the host. Agency for Healthcare Research and Quality (AHRQ) in support with National Institutes of Health and the FDA released a report which analyzed 622 studies and clinical trials that used organisms from six species (*Lactobacillus*, *Bifidobacterium*, *Saccharomyces*, *Streptococcus*, *Enterococcus*, and *Bacillus*) and revealed no evidence of increased risk in those studies.^{51,52}

Overall, the evaluation of probiotics safety includes consideration of a variety of factors: isolation history and taxonomic classification of the applicant probiotic; monitoring of contamination; lack of transferable antibiotic resistance genes; physiological status of the consuming population; dose and method administration (oral or otherwise); lack of allergenic material (for example, dairy proteins) for allergic populations.⁵³

Functional mechanisms of probiotics

Probiotics cause beneficial effects in the host via several mechanisms, which are divided into physiological and immunological sections.⁴⁰

Physiological functions

Probiotics are capable of modulating the intestinal microbiota. In this regard, studies have shown that probiotic supplements, such as *Lactobacillus rhamnosus* GG (LGG) or *Lactobacillus casei*, are capable of modulating the intestinal microbiota that involves the reduction of pathogenic bacteria, including cholesterol, and, conversely, an increase in the level of beneficial *Bifidobacteria* in the neonate with allergies.⁵⁴ These limiting physiological changes include the appropriate pH for the growth and reproduction of these pathological bacteria. Probiotics can also prevent invasion by competing with other microorganisms to bind to the host epithelial cells.⁵⁵ Treatment with *Bifidobacteria* and *Lactobacilli* in newborns results in modulation of the intestinal microbiota in early years of newborn life. It also stimulates the growth of other beneficial bacteria and their products in humans and other species. Modifying the colonization by probiotic bacteria can affect the prolonged presence of harmful pathogens in the intestine lumen, and thus facilitate their clearing by the immune system.⁵⁶ In addition, some of these strains prevent the growth of bacterial pathogens by producing bacteriocin. For example, the bacteriocin produced by *Lactobacillus acidophilus* La-14 inhibits the growth of *Listeria monocytogenes* bacteria⁵⁷ (Fig. 2).

Another of the probiotic functional mechanisms is their effect on epithelial barrier integrity and regulation of

the expression of the proteins which are involved in tight junctions and secretion of mucus.⁵⁸ Treatment with *Streptococcus thermophilus* and *Lactobacillus acidophilus* increased transepithelial resistance as well as the phosphorylation of proteins involved in cellular tight junctions, such as actinin and occluding.⁵⁹ Probiotics also release a significant amount of SCFAs during the fermentation of food fibers and the induction of strong anti-inflammatory activity. Butyrate is one of these SCFAs that is involved in modulating the expression of tight junction proteins, including cingulin, Zo and occludin, which improve the integrity of the epithelial barrier.⁶⁰ Another SCFA is acetate, which has beneficial effects in animal models by reducing inflammation in asthma and colitis. Neutrophils interfere with their anti-inflammatory effects by acting via G protein-coupled receptors (GPR 41, 43) through which they can attach to SCFAs.⁶¹ These effects of SCFAs are accomplished by modulating the NF-κB signaling and the activity of their cytokines network. It also appears that SCFAs have the potential to inhibit histone-deacetylase and alter the structure and function of chromatin, and so the downstream signal affects the gene expression⁶² Fig. 2.

Immunological functions

The activation of TLRs by microorganisms can lead to immune responses that have mucosal and systemic effects. *Lactobacillus* reduces pro-inflammatory responses by regulating NF-κB signaling. Probiotic bacteria also modulate the maturation of DCs towards anti-inflammatory cytokines such as IL-10. Although DCs derived from human monocytes treated with probiotics in culture can release IL-10 that cause differentiation and survival of Treg cells.^{43,63} *B. animals* and *B. longum* have been shown to induce the release of IFN-γ and TNF-α by DCs, while only *B. bifidum* can activate Th17 cells through the release of IL-17.⁶⁴ Sufficient evidence indicates that the Th1/Th2 balance is modulated by probiotics, thereby preventing inflammatory diseases such as allergies. Peripheral blood mononuclear cell (PBMCs) isolated from allergic patients in vitro with several lactic acid bacteria, including *L. plantarum*, *L. lactis*, *L. casei*, and *Lactobacillus* GG, before induction with house dust mite, reduced the Th2 responses, which is characterized by the reduction of IL-4 and IL-5 production. LGG and *B. lactis* reduce the allergic symptoms of asthma mouse models by inducing TGF-β secreting^{65,66} (Fig. 2).

Probiotics function in allergic disorders

Other effects of probiotics that make them suitable for modulation of allergic diseases include stimulating the levels of mucosal IgA, as well as allergen-specific T cells and B cells. These interactions are very complex, and include a network of genes, TLRs, signaling molecules and an increase in IgA intestinal responses.^{67,68} Many animal and in vitro studies, as well as several human experiments, show the positive effects of probiotics in allergic diseases. Several randomized studies have showed that when *Lactobacillus* GG or placebo was given to pregnant mothers with a serious family history of eczema, allergic rhinitis, or asthma and after to newborns during the first six months from delivery, the incidence of

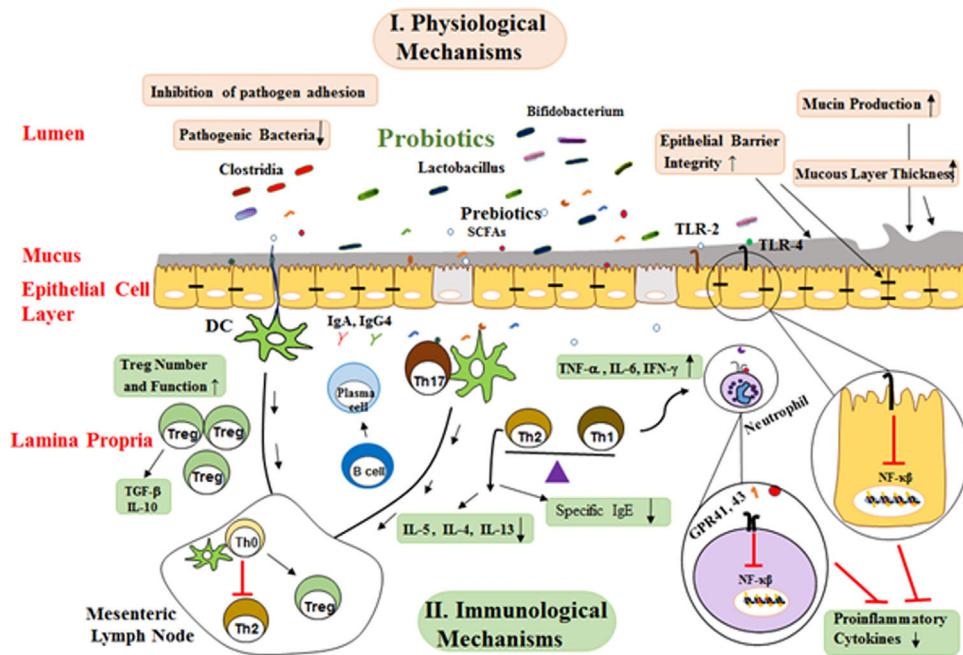


Figure 2 Possible mechanisms whereby probiotics affect allergic diseases. These mechanisms are generally divided into two groups of mechanisms of physiological and immunological mechanisms. SCFAs binding to various GPRs and TLR-2/TLR-4 activate several signaling pathways I. **physiological mechanisms include:** a) Probiotics create competitive conditions and inhibit bacterial adherence to mucosal layer, b) Enhancement epithelial barrier integrity and improved barrier function, c) Mucus production can also be increased by probiotics that stimulate goblet cells leading to activation of mucin gene expression and therefore altering colonization and persistence condition. II. **Immunological mechanisms:** a) Probiotics, directly and indirectly, affect the epithelial cells and modulate signaling pathways that lead to a reduced expression of inflammatory cytokines by suppressing NF- κ B signaling, b) Primary mode of action of probiotics includes restoration of Th1/Th2 cytokine balance and enhancement of Th2 cytokines (IL-4, IL-5, IL-13), c) Probiotics with their products stimulate dendritic cells might be leading to Treg differentiation induction of CD4+ Foxp3+ Treg cells and production of TGF- β and IL-10, d) Probiotics changes the cytokine profiles through effects on dendritic cells and so increase the production of secretory IgA and IgG4 by B cell and reduction in allergen-specific IgE by B cells. (SCFAs: short chain fatty acids; TLR: Toll-like Receptor; TNF- α : Tumor Necrosis Factor A; IFN- γ : Interferon γ ; TGF- β : Transforming Growth Factor B; IL-3: Interleukin 3; IL-4: Interleukin 4; IL-5: Interleukin 5; IL-6: Interleukin 6; IL-10: Interleukin 10; Th0: T Helper Cell Naive; Th2: Type 2 Helper T Cell; Th1: Type 1 Helper T Cell; Treg: Regulatory T Cell; IgE: Immunoglobulin E; DC: Dendritic Cells).

atopic dermatitis in children decreased by 50%, 44% and 36% at two years, four years and seven years, respectively.^{69,70}

Animal studies

Studies on probiotic effects in allergic diseases in animal models have promising results. In the dermatologic atopic model induced by house dust mite and dinitrochlorobenzene, treatment with probiotic mixture *L. acidophilus*, *L. casei*, *L. reuteri*, *B. bifidum* and *Streptococcus thermophilus*, was associated with an increase in the number of CD4+FOXP3+Treg cells parallel with the regression of clinical symptoms, including the specific and total IgE level, and the amount IL-4, IL-5, IL-13.⁷¹ In mouse allergic airway disease (AAD), which is a human asthma model, the probiotic efficacy has been studied. Hougee et al. demonstrated that six probiotic strains, when administered to an ovalbumin (OVA)-induced animal model of AAD led to improvement of lung function and the number of eosinophils, specific IgE levels, IL-4, IL-5 and IL-10 in the bronchoalveolar lavage fluid (BALF). Specifically, treating rats with *L. casei* and

L. plantarum before sensitization with OVA prevented the development of airway hypersensitivity (AHR) and increased eosinophil count and total IgE levels. *L. reuteri* can significantly reduce AAD and eosinophilia of the respiratory tract, AHR, and level of TNF- α , IL-5, and IL-13 in the BALF. It has also been shown that this bacterium can induce Treg CD4+ CD25+ FOXP3+ cells in the spleen and prevent the development of AAD in OVA-sensitive mice.⁷²

In animal models, *L. lactis* and *L. plantarum* cause a shift in response to Th1 and reduce the clinical symptoms associated with allergic diseases.⁷³ *B. bifidum* and *L. casei* reduces the incidence of OVA-induced allergy symptoms.⁷⁴ LGG can reduce the expression of metalloproteinase 9 (MP-9) and cell infiltration and in combination with *L. gasseri*, *L. reuteri* and *L. salivarius* reduce the symptoms of allergy.^{75,76} A mixture of LGG with *B. lactis* induces Treg responses along with increased TGF- β and leads to inhibition of allergic responses and decreased IgE levels.⁷⁷

B. longum and *L. casei* have a role in preventing Th2 responses and allergic diseases.^{78,79} *Streptococcus thermophilus*, *L. acidophilus*, *L. bulgaricus* and *B. Longum* cocktail had an anti-allergic effect which is manifested

Table 1 Probiotics used to prevent and treat allergic diseases in animal models.

Animal Model	Probiotic Strain	Product dose, Treatment Period	Treatment Outcome	Country	Year	Ref.
Female, BALB/c mice	<i>L. lactis</i>	10^{11} cfu/mL	Could be promising candidates for mucosal vaccination against type I allergy.	Austria	2003	73
7 weeks Murine model of birch pollen allergy	<i>L. plantarum</i>	10 day				
Female C3H/HeJ mice	<i>B. bifidum</i> BGN4	diet pellet (0.2% of bacteria)	Decreased OVA-induced allergy response. Useful for the prevention of allergy.	Korea	2005	74
3 weeks OVA-immunized mice food allergy	<i>L. casei</i> 911	7 weeks				
Male Brown Norway rats	LGG	1×10^{10} cfu/mL	Alleviate nasal allergic symptoms by suppressing the increase in nasal vascular permeability caused by local inflammation associated with allergic rhinitis.	Japan	2006	75
4 weeks BALB/c mice	<i>L. gasseri</i> TMC0356 <i>B. longum</i>	4 weeks 1 µg/mouse	Prevents antigen-induced Th2 immune responses	Japan	2006	79
6 weeks NC/Nga mice		2 weeks <i>L. casei</i> subsp. <i>casei</i>	Useful in preventing allergic disease. Prevention and treatment of allergic reactions.	Japan	2006	78
Six-week-old male Atopic dermatitis-like skin lesions in model Newborn BALB/c mice	LGG	8 weeks 10^9 cfu/mL	Inhibits subsequent allergic sensitization and airway disease by induction of Treg cells with increased TGF-β.	Germany	2007	77
Murine model of asthma	<i>B. lactis</i> (Bb-12)	8 weeks				
Male BALB/c or (TLR9-/-) mice	<i>L. reuteri</i>	1×10^9 cfu/mL	Therapeutic potential in the treatment of allergic airway disease.	Canada	2007	76
Mouse model of allergic airway inflammation BALB/c mice	<i>L. salivarius</i> <i>Streptococcus thermophilus</i> MC, <i>L. acidophilus</i> B, <i>L. bulgaricus</i> Lb, <i>L. bulgaricus</i> 448, <i>B. longum</i> B6.	9 days $1-2 \times 10^9$ cfu/mL	Antiallergic effects on OVA/CFA immunized mice via the increasing ratio of IFN-γ/IL-4 and decreasing the serum level of OVA-specific IgE.	Taiwan	2007	80

Table 1 (Continued)

Animal Model	Probiotic Strain	Product dose, Treatment Period	Treatment Outcome	Country	Year	Ref.
5-week-old female Female C3H/HeJ mice OVA-induced allergy mouse model	<i>L. acidophilus</i> AD031, <i>B. lactis</i> AD011	6 weeks 1.5×10^{10} cfu/g, 1×10^{10} cfu/g	Decreased levels of degranulated mast cells, eosinophil granules, and tail scabs. Useful for the prevention of allergy.	Korea	2008	108
Female and male BALB/c mice 6–8 weeks	<i>L. casei</i> Shirota (LcS)	$1-2 \times 10^9$ cfu/mL	Do not necessarily induce beneficial effects.	Netherlands	2008	81
a mouse model for respiratory allergy; A rat model for EAE.		7 weeks				
Pregnant BALB/c mice, Pregnant- Lewis rats (LEW/HanHsD)	<i>B. animalis</i>	1×10^9 cfu/mL	Reduced several immune parameters in the allergy as well as in the autoimmunity model.	Netherlands	2008	81
Rat model for EAE C57BL/6 mice	<i>L. casei</i> Shirota	6 weeks 5×10^8 cfu/mL	Intrinsic adjuvant and immunomodulatory properties that could potentially be exploited for secondary prevention /treatment of allergic respiratory diseases.	Singapore	2009	82
3–4 weeks Allergy mouse model		5 weeks				
Male BALB/c mice 6 weeks OVA-immunized mice	<i>B. breve</i> M-16V	5×10^8 cfu/mL	Antiallergic activity.	Japan	2009	83
Allergic hypersensitivity to cow's milk and atopic dermatitis.		3 weeks				
Male BALB/c ByJlco-specified, 5–8 weeks	<i>B. breve</i> M-16 V, <i>B. infantis</i> NumRes251, <i>B. animalis</i> Num- Res252/253, <i>L. plantarum</i>	10^9 cfu/mL	M-16 V as the potent anti-allergic strain. M-16 V and NumRes8 inhibited, response to methacholine, reduced the number of eosinophils in the bronchoalveolar lavage fluid, reduced both OVA-specific IgE/IgG1.	Netherlands	2010	72
OVA-sensitized	NumRes8, <i>L. rhamnosus</i> NumRes6.	3 weeks				
BALB/c mice	<i>L. casei</i> <i>rhamnosus</i>	1×10^9 cfu/mL	Lcr35 prior to sensitization can attenuate airway inflammation and hyperreactivity in this model.	Korea	2010	109

Table 1 (Continued)

Animal Model	Probiotic Strain	Product dose, Treatment Period	Treatment Outcome	Country	Year	Ref.
6 weeks Airway inflammation in an OVA-induced model of asthma.	Lcr35	8 weeks				
Allergic lung disease in a pig model	<i>L. rhamnosus</i> HN001 (HN001)	1×10^{10} cfu/mL	Decreased the severity of allergic skin and lung responses with an increase of IFN- γ expression.	USA	2011	86
36 male Wistar rats	<i>L. helveticus</i> R0052	6 weeks 10^9 cfu/mL	Anxiolytic-like activity.	France	2011	84
	<i>B. longum</i> R0175 (PF)	2 weeks				
Female BALB/c mice	<i>L. rhamnosus</i> (Lcr35)	1×10^9 cfu/mL	Attenuated the features of allergic asthma model and induced immune regulation by a CD4+CD25+Foxp3+Treg.	Korea	2012	87
4–6 weeks mouse model of asthma		7 days				
Canine model of atopic dermatitis	LGG	1×10^9 cfu/mL	Decreases allergen-specific IgE and prevents AD.	USA	2012	110
		3 weeks - 6 months				
Female mouse model (SKH-1/hr)	<i>L. rhamnosus</i> (Lcr35)	1×10^9 cfu/mL	Prevented the development of AM by suppressing Th2, Th17, and TSLP responses via a mechanism that may involve CD4+CD25+Foxp3+ Tregs in MLNs.	Korea	2014	111
4 weeks with AM		2 weeks				
Female BALB/c mice 5–8 weeks Allergy/asthma mouse model	LGG	Eighteen micrograms of LGG powder were dissolved in 10 mL ddH ₂ O	Reduced OVA-specific IgE levels in serum, decreased matrix metalloproteinase 9 expression in lung tissue and inhibited inflammatory cell infiltration.	Taiwan	2014	112
41 dogs with atopic dermatitis (cAD).	<i>L. paracasei</i> K71	4 weeks 1 mg/kg	Useful in dogs with cAD as a complementary therapy, by providing a steroid-sparing effect.	Japan	2015	85
Male BALB/c mice OVA-induced allergic asthma	<i>B. infantis</i>	12 weeks 5×10^{10} cfu/mL	Inhibits the secretion of allergen-induced IgE, IL-4 and IL-13, and attenuates allergic inflammation	China	2017	113

Table 1 (Continued)

Animal Model	Probiotic Strain	Product dose, Treatment Period	Treatment Outcome	Country	Year	Ref.
6–8 week Female BALB/c mice with asthma	<i>L. paracasei</i>	4 weeks 4×10^9 cfu/mL	Exert the anti-allergic benefit, possibly through rebalancing Th1/Th2 immune response and modulating IL-17 pro-inflammatory immune response	China	2017	114
6 weeks 18 hairless SKH-1 female mice with chronic skin inflammation	<i>B. longum</i> , <i>L. helveticus</i> , <i>L. lactis</i> , <i>Streptococcus thermophilus</i> , <i>L. rhamnosus</i>	3 weeks 1×10^9 cfu/day (or vehicle)	Help in preserving skin integrity and homeostasis	France	2018	115
Male BALB/c mice of OVA-induced allergic airway inflammation	<i>L. rhamnosus</i>	3 weeks 5×10^8 cfu/mL	Promote the expression of Tregs and MLN CD103+DCs to suppress Th2 inflammation	China	2018	116
6–8 weeks Rat and mouse	<i>L. plantarum</i>	46 day 2×10^{12} cells/g	Decreased the mRNA levels of Th2 and Th17 cell transcription factors, increased the transcription factors of Th1 and Treg cells, galactin-9, filaggrin	Republic of Korea	2019	117
6 weeks with AD BALB/c-mice 6 weeks with allergic asthma	<i>L. bulgaricus</i>	4 weeks 1×10^9 cfu/0.2 mL PBS/mouse	Inhibits the lung inflammation and airway remodeling by interfering on Th1/Th2 cytokines and STAT6/T-bet transcription factors.	Brazil	2019	118
40 male Balb/c mice with allergic asthma	<i>B. infantis</i>	4 weeks 5×10^{10} cfu/mL	Promoting Th1 and inhibiting Th2 immune responses	China	2020	119
		4 weeks				

Ova: Ovalbumin; LAB: Lactic Acid Bacteria; L: *Lactobacillus*; B: *Bifidobacterium*; EAE: Experimental Autoimmune Encephalomyelitis; LGG: *L. rhamnosus* GG; AM: Allergic March.

mainly by increasing the IFN- γ /IL-4 ratio and lowering the serum IgE level.⁸⁰ In the use of bacteria, *L. casei Shirota* results were variable and it was not found to be beneficial in a BALB-c animal model, while in cases of C57BL6, there was a reduction of allergic disease symptoms.^{81,82} Simultaneous use of *B. animalis*, *B. breve*, *L. helveticus* and *L. paracasei* improves allergic responses and alleviates the clinical symptoms of allergic disease.^{72,83–85} *B. infantis* and *L. rhamnosus* also reduce the level of eosinophil and specific IgE antibodies leading to an anti-allergic effect.⁸⁶ Moreover, by increasing the level of IFN- γ , it reduces the symptoms of

the disease and, by increasing Treg cells, reduces allergic asthma in the BALB-c model.^{72,87} (Probiotics used to prevent and treat allergic diseases in animal models are shown in Table 1).

Human studies

Early study demonstrates that administration for eight weeks of LGG, *Bifidobacterium lactis* Bb12, or *B. breve* M-16V strains led to the improvement of eczema symptoms in

infants and children compared to a control group.⁸⁸ In spite of the studies that were later carried out in this field, the results were poor in patients with eczema. Viljanen in 2005 and Sistek in 2006 studies showed that LGG, *L. rhamnosus* and *B. lactis* had a better Scorad in children with atopic dermatitis. Viljanen et al. observed that LGG decreased the IL-6 and C-reactive protein and thereby inflammation. On the other hand, it stimulated IL-10 and IgA production which indicated the beginning of repressive responses.^{89,90} The complementary formulation containing *L. acidophilus*, *B. lactis* and fructooligosaccharides (FOS) after eight weeks of treatment significantly reduced Scorad as compared to the control.⁹¹ Wu et al. demonstrated that treatment of children with *L. casei* for 12 weeks significant decreased Scorad scar as compared to placebo. In 2012, Yesilova studied the effect of *B. bifidum*, *L. acidophilus*, *L. casei*, and *L. salivarius* combination for eight weeks in 1–3 years aged children with eczema. A decrease in Scorad and IL-5, IL-6, IFN- γ , and IgE total compared to the control group was observed. Although the use of a combination of probiotics, prebiotics, and synbiotics, including *B. breve* M-16 and a mixture of galacto-fructooligosaccharides, was not able to improve the severity of eczema and reduce Scorad.⁹² In Toh et al., a study in children with moderate to severe eczema, the use of the combination of *L. salivarius* and FOS for eight weeks resulted in a significant reduction in Scorad severity compared to FOS alone.⁶⁸ In another similar adult study, treatment with *L. salivarius* for 16 months significantly decreased with Scorad in comparison with the control group, as well as decreasing the level of IFN- γ , IL-2, and Th1/Th2 cytokines.⁹³

Preventing allergies is one of the major clinical challenges. While there is a lack of evidence, there is a great gap in the use of probiotics in the treatment of allergic diseases.⁹⁴ Several randomized clinical trials (RCTs) have been conducted to evaluate the effects of various probiotic strains in children and adults with a history of allergic diseases. The duration of treatment with probiotics seems to be one of the key factors determining their beneficial effects.

Among the human studies conducted between 2014 and 2018, the strains that can affect AD (atopic dermatitis) could be *L. acidophilus*, LGG, *B. animalis*, *L. paracasei*, *L. fermentum*, and *L. plantarum*. The combination was generally accompanied by suppressing Th2 responses and improving AD clinical symptoms.^{95–98} In the Cow's Milk Allergy (CMA) studies, the use of LGG, which caused butyrate production, increased the induction of tolerance in CMA children, but no significant effect was observed in the presence of *B. lactis* Bb-12 and *L. casei* on CMA.^{99,100} The improvement of AR symptoms was reported for several strains such as LGG, *L. reuteri*, *L. casei*, *L. salivarius*, *B. Longum*, *B. infantis*, *B. breve*, *B. gasseri*, *B. bifidum* G9-1, and *Lactococcus lactis*, all of which were effective in reducing clinical symptoms.^{101–105} In patients with asthma, the effect of *L. acidophilus*, *L. reuteri*, and *Clostridium butyrate* were evaluated. The *L. reuteri* increased IL-10 and decreased inflammation, leading to symptom improvement in these patients.^{102,106,107} The probiotics used to prevent and treat allergic diseases in human studies shown in Table 2.

Conclusion

Currently, the prevention and treatment of allergic diseases is one of the major clinical challenges. The rapid rise in immune system disorders, such as allergic disease, is strongly associated with reduced exposure to early microorganisms. Intestinal microbiota partially stimulates the immune system, and the particular composition of the microbiota of the intestine may affect the risk of allergic disease. Therefore, these findings suggest a therapeutic approach for probiotics and prebiotics used in allergic diseases. Probiotics are selectively stimulating a number of beneficial bacteria that are evaluated in allergy treatment studies. In general, probiotics use major mechanisms to improve clinical symptoms in patients with allergic diseases and prevent them, including: (1) the suppression of Th2 responses and shift response to Th1; (2) butyrate production and increased induction of tolerance; (3) increase of IL-10 and decreased inflammation; (4) decreased eosinophil level and serum specific IgE levels; (5) increasing the IFN- γ /IL-4 ratio; (6) increasing Treg cells and inducing their responses; (7) increasing TGF- β responses and inhibiting allergic responses; and (8) reducing the expression of metalloproteinase 9 and cell infiltration. Finally, it can be said that the therapeutic approach to immunotherapy and the reduction of the risk of side effects in the treatment of allergic diseases is the first priority of treatment. The final approach that completes the first priority is maintaining the condition and sustainability of the tolerance along with the recovery of the individual.

There are several advantages and health profits associated with probiotics or probiotic food products. The primary results of evaluated studies in our review included improvement in the clinical score, influence on the immune system and allergy modulation, also effects on the fecal and plasma biomarkers of inflammation. But these studies also have limitations and disadvantages that will be discussed in detail. The major limitations of some of the studies is the reduced sample size, the randomization process and the small number of bacterial groups used. In addition, some studies do not clearly describe the conditions and goals of the selected probiotic species, inclusion and exclusion criteria and methods of working. However, there are several other risks associated with probiotics therapy in the clinical field, especially in the treatment of allergic diseases.

These risks are mainly concerned with safety in vulnerable target groups such as immune compromised subjects (high risk populations such as pregnant women, babies and the elderly individuals) and some immunodeficiency patients. Probiotics can affect the commensal bacteria and can also have a direct influence on the host in the long term and for these reasons the evolution of these interactions is one of the key concerns for future research. Therefore, immediate clinical and molecular examination is required to improve our knowledge about interaction among candidate probiotics and host microbiome, cells, mucus and immune defenses, in order to create efficacious interventions.

One theoretical concern with the safety of probiotics is that some have been designed or chosen to have good

Table 2 Probiotics used to prevent and treat allergic diseases in human studies.

Patient group	Study design	Probiotic strain	Product dose, Treatment Period	Treatment Outcome	Country	Year	Ref.
49 AD patients aged \geq 16 years	RDPC	<i>L. acidophilus</i> strain L-92 (L-92)	20.7 mg/day 4–8 weeks	Effective for AD symptoms. L-92 contributes to the suppression of Th2-dominant inflammation.	Japan	2014	95
415 pregnant women	RDPC	LGG, <i>L. acidophilus</i> La-5 <i>B. animalis</i> subsp. <i>lactis</i> Bb-12	5×10^{10} cfu/mL	Reduction in the cumulative incidence of AD, but not other allergy-related diseases.	Norway	2015	97
6 years children were re-assessed for AD, atopic sensitization, asthma, and ARC.			Bb12 5×10^9 cfu/mL				
62 children (1–10 years) with peanut allergy	RDPC	<i>L. rhamnosus</i>	6 year 2×10^{10} cfu/mL	Elimination in patients with peanut allergy.	Australia	2015	88
331 children eczema and atopic sensitization in infants	RDPC	CGMCC 1.3724 <i>L. rhamnosus</i> HN001 (HN001)	18 months. 10^9 cfu/d	Positively affected by probiotic. HN001 exhibits a stronger effect than HN019 in this respect.	New Zealand	2015	120
119 Children with CMPA	RDPC	<i>B. animalis</i> subsp. <i>lactis</i> HN019 (HN019) <i>L. casei</i> CRL431	2 years 10^7 cfu/g	Did not show any impact on these parameters.	France	2015	100
220 children 1–18 years moderate/severe AD	RDPC	<i>B. lactis</i> Bb-12 <i>L. paracasei</i> (LP)	6 months 2×10^9 cfu/mL	Improvement in children with AD.	Taiwan	2015	98
Infants with CMA	RDPC	<i>L. fermentum</i> (LF) LGG	3 months 4.5×10^7 – 8.5×10^7 cfu/g	Promotes tolerance in CMA infants, enrichment of specific strains of bacteria that are associated with butyrate production.	Italy	2016	99
1–12 months 100 children,	RDPC	LGG	4 weeks 3 billion live bacteria in 1 dose	Better clinical and immunologic response in children.	USA Poland	2016	104

Table 2 (Continued)

Patient group	Study design	Probiotic strain	Product dose, Treatment Period	Treatment Outcome	Country	Year	Ref.
5–12 years sensitive to grass pollen with allergic rhinitis 220 children	RDPC	LGG	Increasing doses (0.1, 0.3, 1, 3, 10, 30, and 100 mL) of fresh pasteurized cow's milk	EHC1LGG reduces the incidence of other AMs and hastens the development of oral tolerance in children with IgE-mediated CMA. 110 were placed in the EHC group, 110 were placed in the EHC1LGG group	Italy	2016	99
Allergic children with asthma	RDPC	<i>L. reuteri</i> DSM 17,938	10^8 cfu/mL	Reduced bronchial inflammation and significantly increased interleukin 10 in children with asthma.	Italy	2016	102
32 children (age range, 6–14 years) Asthma patients with mild to moderate clinical symptoms	RDPC	<i>Clostridium butyrate</i> (CB)	8 weeks	Potential therapeutic remedy in the treatment of allergic diseases.	China	2016	106
participants aged 5 years and older with asthma	RDPC	<i>L. aci- dophilus</i> CUL60 (NCIMB 30,157), CUL21 (NCIMB 30,156), <i>B. bifidum</i> CUL20 (NCIMB 30,153), <i>B. animalis</i> (var <i>lactis</i>) CUL34 (NCIMB 30,172)	3-month 2.5×10^{10} cfu/capsule.	No evidence of an effect on respiratory tract infections or asthma exacerbations.	United Kingdom	2016	107
1302 participants Children	RDPC	LGG	2 weeks 10^6 cfu/g	Normal growth and development, long-term safety through 5 years of age.	USA	2017	121
289 participants CMA			5-year				

Table 2 (Continued)

Patient group	Study design	Probiotic strain	Product dose, Treatment Period	Treatment Outcome	Country	Year	Ref.
Children with seasonal allergic rhinitis and intermittent asthma (40 children)	RDPC	<i>B. longum</i> BB536, <i>B. infantis</i> M-63, <i>B. breve</i> M-16V	3 × 10 ⁹ cfu/mL, 1 × 10 ⁹ cfu/mL, 1 × 10 ⁹ cfu/mL	Improving AR symptoms in children with pollen-induced AR and intermittent asthma.	Italy	2017	101
92 childhood with asthma and eczema	RDPC	LGG	4 weeks 10 billion cfu/day	Prevent the development of eczema or asthma.	USA	2017	122
173 participants who self-identified as having seasonal allergies	RDPC	<i>L. gasseri</i> KS-13, <i>B. bifidum</i> G9-1, <i>B. longum</i> MM-2	6 months 2 capsules/d, 1.5 billion cfu/capsule	Improved ARC specific quality of life during allergy season.	USA	2017	103
JCP	RDPC	LGG and TMC0356 in the fermented milk	8 weeks 1.4 × 10 ⁸ cfu/mL	Protection against JCP, Beneficial effects on blood lipid levels.	Japan	2017	123
Children with mild and moderate AD	RDPC	<i>L. plantarum</i> IS-10,506	1 × 10 ⁷ cfu/mL 10 weeks 10 ¹⁰ cfu/day	Reduce clinical symptoms.	Indonesia	2017	96
415 pregnant women assessed for AD during the first 2 years of life	RDPC	LGG, <i>B. animalis</i> subsp. <i>lactis</i> Bb-12 (Bb-12), <i>L. acidophilus</i> La-5 (La-5)	12 weeks 5 × 10 ¹⁰ cfu/mL LGG /Bb-12 and 5 × 10 ⁹ cfu/mL La-5	Preventive effect on AD.	Norway	2017	124
≥10 patients	ProPACT study Phase III placebo-controlled trial	<i>B. bifidum</i> W23, <i>B. lactis</i> W51, <i>L. acidophilus</i> W55, <i>L. casei</i> W56, <i>. salivarius</i> W57, <i>Lactococcus lactis</i> W58	1 year 1 × 10 ⁹ cfu/g	Improvement in quality-of-life metrics.	Australia	2018	105
40 patients AR			8 weeks				

ARC: Allergic Rhinoconjunctivitis; AD: Atopic Dermatitis; AR: Allergic Rhinitis; RDPC: Randomized, Double-blind, Placebo-Controlled trial; CMPA: Cow's Milk Protein Allergy; JCP: Japanese Cedar Pollinosis; LGG: *L. rhamnosus* GG.

adherence to the intestinal mucosa, and this is considered important for their mechanism of action. Adherence to the intestinal mucosa may also increase bacterial translocation and virulence. The most potent probiotics, therefore, may have increased pathogenicity. The relation between mucosal adhesion and pathogenicity in *Lactobacillus* spp. is supported by the finding that blood culture isolates of *Lactobacillus* spp. adhere to intestinal mucus in greater

numbers than isolates from human feces or dairy products do. Emerging insights from microbiome research allow an assessment of gut colonization by probiotics, species and strain, interactions with the indigenous microbiome, safety and impacts on the host, physiological effects and potentially beneficial medical indications such as allergic diseases.

Author contribution

Majid Eslami: Investigated and supervised the findings of this work, wrote the article. Processed the experimental data. Supervised the project.

Bahman Yousefi: Designed the study, helped supervise the project, conceived the original idea. Processed the experimental data. Developed the theoretical framework.

Mohsen Karbalaei: Provided critical feedback and helped the analysis of the manuscript. Contributed to the interpretation of the results.

Masoud Keikha: Contributed to the final version of the manuscript.

Aisa Bahar: Contributed to the final version of the manuscript.

Nazarii M. Kobyliaik: Contributed to the final version of the manuscript.

All authors: Designed the model and the computational framework and analyzed the data.

Conflict of interest

The authors have no conflict of interest to declare.

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