Revisión

Inmunología Vol. 28 / Núm 1/ Enero-Marzo 2009: 32-45

The Th17 lineage: Answers to some immunological questions

Coral González-García, Francisco M. Martín-Saavedra, Alicia Ballester, Sara Ballester

Unidad de Regulación Génica, Centro Nacional de Microbiología, Instituto de Salud Carlos III, Majadahonda, Madrid.

EL LINAJE TH17: RESPUESTAS A ALGUNAS CUESTIONES INMUNOLÓGICAS

Recibido: 1 Diciembre 2008 Aceptado: 26 Enero 2009

RESUMEN

En los últimos años se han estudiado exhaustivamente las funciones y las rutas de desarrollo del subtipo de células T helper especializado en la producción de IL-17 (Th17). Este linaje celular de células efectoras desempeña un papel decisivo tanto en la respuesta inmune a agentes infecciosos, como en inmunopatologías. Al igual que para los subtipos Th1 y Th2, la definición de Th17 está dirigida por citocinas y factores de transcripción específicos. La combinación de TGF-β e IL-6, y los factores de transcripción RORyt, RORα y Stat3 son esenciales para comprometer el subtipo Th17. IL-23 juega un papel clave en la estabilización del fenotipo y de la actividad patogénica de células productoras de IL-17. La citocina IL-21 producida por células Th17 participa en un mecanismo de retroalimentación para favorecer el desarrollo de células productoras de IL-17, mientras que las citocinas IL-27, IL-4, IFN-γ, IL-25 e IL-2 limitan el fenotipo Th17. Las células T reguladoras CD4+CD25+Foxp3+ (Treg) siguen una ruta de desarrollo divergente al establecimiento de las células IL-17, aunque ambas alternativas son gobernadas por TGF-β, el cual dirige el destino de células CD4+ naïve hacia uno u otro de estos subtipos celulares mutuamente excluyentes dependiendo de la presencia de IL-6. Además, datos recientes indican que células Treg ya establecidas pueden modificar su programa genético para convertirse en células Th17. En esta revisión se resumen y analizan los datos disponibles actualmente acerca de la biología de las células Th17.

PALABRAS CLAVE: Th17 / TGF-\u03b3 / IL-6 / IL-23 / RORyt.

ABSTRACT

In recent years the function and developmental pathway for the T helper subset specialized in IL-17 production (Th17) have been exhaustively studied. This lineage of effector cells plays a decisive role in the immune response to infectious agents, as well as in immunopathologies. Similar to the Th1 and Th2 subsets, the Th17 definition is orchestrated by specific cytokines and transcription factors. A combination of TGF-β plus IL-6, and the transcription factors RORyt, ROR α and Stat3 are essential for Th17 commitment. IL-23 plays a key role in the stabilization of the phenotype and in the promotion of the pathogenic activity of IL-17-producer cells. The IL-21 cytokine produced by Th17 cells participates in a feedback mechanism to favour this phenotype, while IL-27, IL-4, IFN-y, IL-25 and IL-2 cytokines limit the Th17 response. CD4+CD25+Foxp3+ regulator cells (Treg) follow a development pathway divergent to Th17 establishment, although both alternatives are governed by TGF- $\!\beta$ that directs the fate of naïve CD4+ cells to each of these mutually exclusive T cell subsets depending on the presence of IL-6. Furthermore, recent data indicate that preestablished Treg cells can switch its genetic program to become IL-17-producer cells. In this review we summarize and discuss the current available data about the biology of Th17 cells.

KEY WORDS: Th17 / TGF-β / IL-6 / IL-23 / RORyt.

THE IL-17 FAMILY

Since Mossman et al. proposed the model wherein CD4⁺ T helper cells were classified in Th1 and Th2 subtypes with different functions in immune responses according to the profile of cytokines produced⁽¹⁾, the Th1/Th2 paradigm provided a valuable tool to understand the interplay of innate and adaptive immunity and CD4⁺ T cell function. However, some discrepancies have arisen related to results that did not fit in the Th1/Th2 hypothesis. During the past years new studies have identified a distinct subset of CD4⁺ T cells that secrete IL-17 and the closely related cytokine IL-17F as well as other inflammatory cytokines such as IL-6 and IL-22^(2,3).

Murine IL-17 was described as CTLA-8⁽⁴⁾, and a 63% homologous human cytokine was soon found⁽⁵⁾. Currently, the IL-17 cytokines include a family of six members (IL-17A-F), with at least two of them having potent proinflammatory properties: IL-17A or CTLA-8 (the founder member of the family also named IL-17), and IL17-F. Both are produced by the recently described Th17 cell subset, are localized at the same chromosomal locus (1A4), share a 55% of homology at the protein level, and seem to have similar functions. IL-17A and IL-17F work mostly as homodimers, but IL-17A/F heterodimers have been recently described in several independent reports⁽⁶⁻⁸⁾, suggesting a role in inflammatory response regulation for such IL-17 complexes. IL-17D and IL-17E (alternative names: IL-27 and IL-25) are the two members of the IL-17 family with lowest homology (16% at protein level) to IL-17A. None of them is produced by Th17 cells, and both of them, as discussed later, exert a negative control on the Th17 subset development.

IL-17 receptors are a family of five members of membrane proteins, IL-17RA-F⁽⁹⁾. Except for IL-17RA, each of these receptors has alternative splicing variants, and, for IL-17RB and IL-17RC, result in secreted soluble proteins which could serve to antagonize their ligands⁽¹⁰⁾. IL-17A and IL-17F bind to IL-17RA, although IL-17A binds with much higher affinity⁽¹¹⁾, which correlates with its greater potency in functions like induction of chemokine expression⁽⁸⁾. In addition to IL-17RA, IL-17F can also bind to IL-17RC. By signalling through IL-17RA, which is ubiquitously expressed, IL-17 can induce the production of different kinds of proteins, many of them related to inflammation, including chemokines (CXCL-1, CXCL-2, CXCL-8-10, CCL-2, CCL-20), cytokines (IL-6, TNF α , G-CSF, GM-CSF), proteins of the acute phase response, tissue remodelling factors (MMP1, MMP3, MMP9, MMP13, TIMP2), and anti-microbial products (β-defensins, mucins, calgranulins)⁽¹²⁾. The signal transduction of IL-17 is mainly mediated by NFkB and C/EBP transcription factors, with the involvement of MAP kinase pathways⁽¹³⁻¹⁶⁾. Moreover,

TRAF6 was shown to be involved in the activation of NF κ B by IL-17⁽¹⁷⁾. On the other hand, IL-17 can increase expression of some of its target genes through mRNA stabilization⁽¹⁸⁾.

IL-17B and IL-17C are members of the family whose cellular sources are unknown yet, and whose biology seems unrelated to IL-17A. In this review, we will refer to IL-17A as IL-17.

CLUES TO THE IDENTIFICATION OF A NEW T HELPER CELL LINEAGE PRODUCING IL-17

The first suggestion of a new T helper subset, distinct from Th1 and Th2, was provided by the finding of T CD4+ cells producing high levels of IL-17 without expression of IFN-γ or IL-4, the respective prototype cytokines produced by Th1 and Th2⁽¹⁹⁾. Other important clues were supplied by studies on animal models of autoimmunity. Pathologies such as experimental autoimmune encephalomyelitis (EAE) and collagen-induced arthritis (CIA), or mouse models for human multiple sclerosis (MS) and rheumatoid arthritis (RA), have been traditionally considered as Th1-mediated diseases. However, over the years, a number of experimental results showed inconsistencies with the Th1/Th2 hypothesis. For example, depletion of the Th1 cytokine IFN- γ , its receptor IFN- γ R, or the IL-12 receptor (IL-12R β 2), the master inductor of the Th1 phenotype, increased the susceptibility to EAE(20-26).

With the discovery of IL-23 as an heterodimeric cytokine sharing its p40 subunit with IL-12(27), the relative contribution of IL-12 and IL-23 to chronic inflammation could be analyzed. The IL-12 heterodimer is composed by the p40 and p35 subunits, while IL-23 comprises p40 and a different p19 protein. By using mice lacking IL-23 (p19-/-), IL-12 (p35-/-) or both cytokines $(p40^{-1})$, it could be demonstrated that IL-23, and not IL-12, is the critical cytokine for autoimmune inflammation of the central nervous system (CNS) during EAE⁽²⁸⁾. This study was followed by many other showing the role of IL-23 in inflammation⁽²⁹⁻³¹⁾. In parallel with these findings, it was reported that IL-17-producer cells could be generated independently of the cytokines and transcription factors required for Th1 or Th2 differentiation. The development of Th17 from naïve cells was potently inhibited by IFN-y and IL-4, whereas memory Th17 cells were resistant to suppression by Th1 or Th2 cytokines, indicating that these cells had a permanent commitment. All these data together indicated that a T helper lineage distinct from Th1 and Th2 can differentiate from naïve CD4⁺ cells to produce IL-17^(2,3).

In 2006, three independent reports provided the basis for understanding the process of Th17 cell promotion⁽³²⁻³⁴⁾. All of them found that transforming growth factor β (TGF-



Figure 1. The homeostasis of Treg and Th17 cells can be balanced to each phenotype by the cytokine environment. TGF-β is involved in both pathways, and IL-6 is the decisive cytokine directing the cell subset fate. In the absence of IL-6, TGF-β induces the establishment of Treg cells that in turn produce TGF-β and control inflammation and autoimmunity by suppression of T effector cells. This process is favoured by IL-2 and retinoids. The Th17 pathway is triggered by inflammatory cytokines like IL-6. Once Th17 cells are committed, the IL-23R is expressed, allowing responsiveness to IL-23, which stabilizes the phenotype of IL-17 producer cells. IL-6 and IL-21 are also produced, and both of them take part in an autocrine loop that collaborates in Th17 development. Negative regulators of Th17 differentiation are IL-27, IL-4, IFN-γ, IL-25, IL-2 and retinoids. In addition to IL-17, IL-21 and IL-6, Th17 cells secrete other products, like chemokines, GM-CSF and IL-22, all of them involved in host defence and inflammation. In the absence of IL-23, Th17 cells can also produce IL-10, which may control the pathogenic phenotype of IL-17 producer cells.

β) is critical while IL-23 is not required for Th17 commitment. TGF-β is associated with immunosuppressive functions through the induction of Foxp3 expression and Treg cell activity with inhibitory properties on effector cells⁽³⁵⁾. Nevertheless, it is now clear that TGF-β may also facilitate proinflammatory responses through Th17 development, although it needs to act in concert with inflammatory cytokines such as IL-6. Thus, TGF-β induction of Treg or Th17 subsets are mutually exclusive depending on the presence of IL-6^(32, 34). The finding of these reciprocal pathways promoted by TGF-β could explain the apparent discrepancy that TGF-β is involved in both anti- and pro-inflammatory events in the immune system.

Therefore, when the immune system is not activated, in the absence of inflammatory cytokines, TGF- β favours the generation of adaptive Treg cells, which prevent inflammation and autoimmunity. After infection, cytokines like IL-6 derived from the innate response can inhibit such process and collaborate with Treg-produced TGF- β to induce proinflammatory Th17 cells (Fig. 1).

TGF- β ACTION ON HUMAN TH17 ESTABLISHMENT

Despite the well-known role for TGF- β in murine Th17 differentiation, many doubts arose about such a function on human Th17. Several important reports in 2007 refuted the requirement of TGF- β in human IL-17 production⁽³⁶⁻³⁹⁾. While some of these studies considered that the central cytokines needed for human Th17 function were IL-23 and IL-1 β , other found that IL-1, collaborates with IL-6; however, all of them emphasized that TGF- β was dispensable, suggesting important differences in the requirements for the differentiation of Th17 in humans and mice.

More recently, three simultaneously published reports assured that TGF- β was indeed required for the establishment of human Th17⁽⁴⁰⁻⁴²⁾. Several explanations for these divergent

conclusions are conceivable: i) the difficulty to obtain real naïve cells from human samples, a caution which has been ensured in the works of Manel et al.⁽⁴¹⁾ and Volpe et al.⁽⁴²⁾; ii) the importance of eliminating any residual contamination of platelets, a major store of TGF- β ; iii) or the culture media used in which serum may contain endogenous TGF- β . These issues are discussed at length by O'Garra⁽⁴³⁾. Thus, these new findings support that similar cytokine pathways are involved in Th17 development in mice and humans.

OTHER INFLUENTIAL CYTOKINES THAT FAVOUR TH17 ACTIVITY

As mentioned above, the early proposal that IL-23 directly drives differentiation to Th17 cells has been discontinued due to three main findings: i) IL-23-deficient mice produce Th17 cells which can be expanded *in vitro* by exogenous IL-23⁽³³⁾; ii) although IL-23 can induce memory T cells to produce IL-17, naïve T cells do not express the receptor for IL-23⁽⁴⁴⁾; and iii) it was clearly demonstrated that IL-23 is dispensable for initial IL-17 generation^(2,3,29). Nevertheless, there is no doubt about the essential role of IL-23 in the maintenance of Th17 cell activity. It has been proposed that IL-23 can have a function in promoting Th17 cell expansion or survival⁽⁴⁵⁾. A recent report suggests that IL-23 maintains the Th17 phenotype without affecting proliferation or survival⁽⁴⁶⁾. On the other hand, IL-23 has been demonstrated to maintain the pathogenic Th17 functions compared with culture under TGF- β and IL-6, depending on IL-10 production by Th17 cells⁽⁴⁷⁾.

Currently, IL-6 is the main partner of TGF- β in priming naïve T cells to IL-17 production^(32,33,36,45,48,49). It is able to inhibit the expression of Foxp3, which directs the differentiation of CD4+CD25⁺ Treg cells⁽⁵⁰⁾. This may constitute the beginning of the Th17 commitment and the decline of Treg activity during immune responses. Moreover, IL-6 orchestrates a series of downstream cytokine-dependent signalling pathways to amplify Th17 cell differentiation. In fact, IL-6 is able to induce the expression of IL-23R in T cells making them responsive to the stabilizer phenotype of IL-23⁽⁴⁸⁾. On the other hand, after induction of IL-17 expression, Th17 cells start to secrete IL-6 and IL-21, which in turn act as autocrine factors^(49,51).

Although IL-21 does not look like an essential factor for Th17 lineage commitment, it is able to induce IL-17 expression in collaboration with TGF- β even in the absence of IL-6^(48,51,52). Furthermore, generation of Th17 cells is attenuated by blocking IL-21⁽⁵³⁾, and loss of its expression, or its receptor, results in defective Th17 differentiation. Similar to IL-6, IL-21 inhibits Foxp3 expression induced by TGF- β ^(51,52). IL-21

is produced by Th17 cells under IL-6 induction and autocrinally induces its own synthesis and the expression of IL-23R to allow IL-23 responsiveness^(48,51).

Another positive regulator of Th17 development is IL-1. It has been reported that IL-1, can increase the effect of IL-6 and TGF- β on Th17 definition⁽⁴⁵⁾. Besides, the induction of antigen-specific Th17 cells is abrogated in IL-1R1-deficient mice, without effect on Th1 or Th2 cells⁽⁵⁴⁾. However, the mechanism through which IL-1 influences Th17 differentiation is not determined yet.

The IL-18 signalling pathway might be involved in Th17 cell definition as well. Remarkably, the receptor of IL-18 (IL-18R α), but not IL-18 itself, would be involved in this action. Gutcher et al.⁽⁵⁵⁾ showed that, while IL-18-deficient mice were susceptible to EAE, IL-18R α -deficient animals were resistant to the disease showing a deficient Th17 response, and proposed that IL-18 signalling is involved in Th17-mediated immunopathology through binding of an unknown alternative ligand distinct from IL-18.

NEGATIVE INFLUENTIAL CYTOKINES FOR THE PROMOTION OF THE TH17 SUBSET

As Th1 and Th2 cells inhibit the polarization of one another through IFN- γ and IL-4, respectively⁽⁵⁶⁾, there is evidence that these cytokines antagonize initial Th17 development^(2,3). However, after Th17 cells become established effectors, IFN- γ and IL-4 cannot suppress or revert their phenotype.

One of the main negative regulators of Th17 development is IL-27^(57,58), a cytokine structurally related to IL-6, but with many different actions. Definitive studies showing such damaging role of IL-27 on IL-17 producer cells were reported by Batten et al. and by Stumhofer et al.^(59,60). Both reports conclude that the absence of IL-27 signalling exacerbates chronic inflammation in correlation with increased number of Th17 cells. Moreover, IL-27 is able to promote IL-10 production⁽⁶¹⁾, another negative player in the network of Th17 activity regulation⁽⁶²⁾.

IL-10 was first described as a product of Th2 cells, but it is now well-known that it is also produced by other cell types like Th1 cells or regulatory Tr1 cells, with an important role in limiting T-cell mediated immunopathology. The involvement of IL-10 in regulating the pathogenic function of Th17 cells has been definitively demonstrated by McGeachy et al.⁽⁴⁷⁾, who have described a non-pathogenic Th17 subtype expressing IL-10 generated by IL-6 and TGF- β in the absence of IL-23. Such type of cells are not only non-pathogenic in EAE, but they also prevent the induction of the disease in an IL-10-dependent way. Another negative regulator of Th17 cells is IL-25 (IL-17E), identified by database searching for genes homologous to IL-17. Although IL-25 is included in the IL-17 family, it is not produced by Th17, but by Th2 and mast cells. This cytokine is involved in the expression of the Th2 products IL-5 and IL-13, and favours Th2 responses⁽⁶³⁾. On the other hand, IL-25 deficiency is involved in pathologic inflammation, associated with increased expression of IL-17 and IL-23^(64,65).

At least in mice, IL-2 also antagonizes Th17 activity. In spite of the essential function of IL-2 as growth factor of T effector cells, its deficiency leads to systemic autoimmune disease⁽⁶⁶⁾. This is justified in part by its involvement in differentiation and survival of Treg cells⁽⁶⁷⁾. Besides, a recent work has revealed that IL-2 constrains IL-17 production since IL-2 deficiency promotes differentiation of the Th17 cell subset in a Stat5-dependent mechanism⁽⁶⁸⁾.

ROLE OF TH17 CELLS IN HOST DEFENCE

The role of IL-17 in immune responses against infections has been widely demonstrated⁽⁶⁹⁾. IL-17 is a potent activator of neutrophils, both through cell expansion and through regulation of chemokine expression for cell recruitment. Signalling mediated by IL-17 induce target cells to express CXCL-8 (IL-8) and G-CSF, which results in the generation and accumulation of neutrophils. In addition, IL-17 induces expression of various anti-microbial genes such as β -defensins. IL-17 deficiencies are associated with neutrophil defects leading to disease⁽⁷⁰⁾. The lamina propria, constituent of mucosa located at the respiratory, gastrointestinal and urogenital tracts, is the only tissue where constitutive expression of IL-17 is detected. Due to the permanent interaction of this tissue with microbial flora, a defensive T cell population promptly responding to infection may be very useful⁽⁷¹⁾.

Both IL-17- and IL-17RA-deficient mice show enhanced susceptibility to experimental *Klebsiella pneumoniae* pulmonary infection and reduced lung G-CSF and CXCL-1 in response to this infection^(72,73). In addition to *K. pneumoniae* IL-17F has also demonstrated to be involved in pulmonary recruitment of neutrophils⁽⁷⁴⁾. Other bacteria preferentially inducing a Th17 response are *Borrelia burgdoferi*, *Bacteroides fragilis*, or *Mycobacterium tuberculosis*^(19,75).

IL-17 seems to have also a role in protective immunity against fungal infections. Deficiency in IL-17R leads to decreased survival and increased damage in kidney after infection with *Candida albicans*, with delayed mobilization of peripheral neutrophils⁽⁷⁶⁾. Another major human fungal pathogen which provokes a Th17 response is *Aspergillus* *fumigata*⁽⁷⁷⁾. Other fungal infections activating the IL-23/IL-17 axis are *Cryptococcus neoformans* and *Pneumocystis carinii*^(78,79).

IL-17 could be also defensive against some parasites, like infection with the protozoan *Toxoplasma gondii*^(60,80). However, and although an homologue of IL-17 is encoded by herpes virus Saimiri⁽⁸¹⁾, the role of Th17 in viral infections is unclear. Several studies suggest a pathogenic activity of IL-17 rather than protective function^(82,83). A protective role of Th17 cells has only been clearly demonstrated against rotavirus infection⁽⁸⁴⁾.

ROLE OF TH17 CELLS IN IMMUNOPATHOLOGY

The Th17 subset has a key role in both induction and progress of immunopathologies. IL-17 and other cytokines related to the development and function of Th17 cells are closely associated with several immune disorders, not only in animal models, but also in human diseases. As we have mentioned before, the importance of the Th17 cell activity in autoimmunity was first demonstrated in mice deficient in the p19 chain of IL-23, which showed an important impairment of IL-17 production and were highly resistant to EAE⁽²⁸⁾ and CIA⁽³¹⁾. Using passive transfer studies in EAE, it was demonstrated that IL-17 producing cells are highly pathogenic and essential for the establishment of organspecific inflammation⁽²⁹⁾. On the other hand, neutralization of IL-23 can decrease IL-17 expression in the central nervous system and prevent EAE(30). In addition, IL-17A deficient mice show reduced symptoms of EAE with delayed onset and early recovery⁽⁸⁵⁾, and are resistant to CIA⁽⁸⁶⁾. In several independent assays it could be demonstrated that vaccination against IL-17 prevents EAE and CIA⁽⁸⁷⁻⁸⁹⁾. All these experiments highlighted the importance of the IL-23/IL-17 axis in the pathogenesis of several disorders that was previously thought to be mediated by the Th1 subset. Many previous data, inconsistent with the Th1 hypothesis, could be then understood⁽²⁰⁻²⁶⁾. The role of IL-17 in autoimmunity has been later underlined by the relationship between therapeutic treatments and IL-17 production impairment. Thus, IFN- β , currently the main therapy used for MS⁽⁹⁰⁾, inhibits IL-17 expression in peripheral and CNS infiltrates of T lymphocytes during EAE^(91,92), and increases the expression of the IL-17 negative regulator IL-27(93). On the other hand, after transfer of ICOS+ cells to ICOS-deficient mice, that have enhanced susceptibility to EAE, lymph node cells showed decreased IL-17 production and could reduce the severity of the disease(94).

IL-17 seems to be also associated with the human diseases MS and RA. IL-17 and IL-23 are present in the sera, synovial fluids and synovial biopsies of RA patients⁽⁹⁵⁻⁹⁷⁾. In addition,

it is known that IL-17 mediates induction of IL-6 and IL-8 in RA synovial fibroblasts⁽⁹⁸⁾. Before the explosion of the recent advances in the knowledge of the murine Th17 subset, high IL-17 mRNA expression had been observed in mononuclear cells of the blood and CSF in human MS⁽⁹⁹⁾, and a gene-microarray analysis of MS lesions had suggested that IL-17 and IL-6 could be possible targets for MS therapy⁽¹⁰⁰⁾. Although human Th17 cells are less well characterized than the murine subset, some specific features in addition to IL-17 production have been proposed for them, mainly related with specific patterns of chemokine receptor expression, which suggest their involvement in migration of Th17 cells and recruitment of other inflammatory cells^(101,102). More recently, an elevated number of IL-23-expressing dendritic cells has been found in MS patients, concurrent with increased IL-17 production by T cells⁽¹⁰³⁾. Moreover, evidence has been provided that IL-17 and IL-22 induce a breach in the bloodbrain barrier and promote recruitment of additional CD4+ lymphocytes⁽¹⁰⁴⁾. Of particular interest has been the report of Tzartos et al., who found IL-17 mRNA and protein expression in active areas of MS lesions, where the cell sources of IL-17 were both infiltrating T cells and resident astrocytes and oligodendrocytes(105).

Teunissen et al.⁽¹⁰⁶⁾ described the upregulation of IL-17A in psoriasis, an autoimmune disorder affecting the skin. IL-17 cooperates with IL-22 in the induction of anti-microbial peptides as a function in host defence against pathogens. However, these defensive peptides can enhance the expression of factors related to psoriatic skin⁽¹⁰⁷⁾. Moreover, expression of different products of Th17 cells has been found in psoriatic skin⁽³⁹⁾. Another autoimmune disease that has been recently linked to inappropriate Th17 cell response is systemic lupus erythematosus (SLE). IL-17, IL-23 and the number of Th17 cells were higher in plasma from SLE patients than in control individuals, suggesting involvement of the IL-23/IL-17 axis in inflammatory immunity in SLE⁽¹⁰⁸⁾.

There are also data indicating that IL-17 and IL-23 are involved in animal models of induced colitis⁽¹⁰⁹⁻¹¹¹⁾. Patients with ulcerative colitis or Crohn's disease display an elevated expression of IL-17 in the intestinal mucosa, which is augmented during active exacerbations of inflammatory bowel disease^(112,113). All these data showing the contribution of IL-17 activity to autoimmune diseases justify clinical trials using a monoclonal antibody anti-IL-17 (AIN457) for Crohn's disease and psoriasis⁽¹¹⁴⁾.

Allergic asthma is considered to be a Th2-dominant chronic inflammatory disease with a major involvement of eosinophil activity. However, some asthmatic processes seem to be mediated by neutrophil infiltration rather than by eosinophil cells, and have been described as "noneosinophilic asthma"⁽¹¹⁵⁻¹¹⁹⁾. Some results in experimental models suggest that Th17 cells may be important for neutrophilic activity in acute airway inflammation⁽¹²⁰⁻¹²²⁾. These data lead some researchers to think that Th17 might be responsible for allergic processes mediated by neutrophil cells. However, the role of IL-17 in allergy is still largely unclear. Current results related to this issue have been discussed in length by Oboki et al.⁽¹²³⁾.

THE GENETIC PROGRAM GOVERNING TH17 COMMITMENT

In contrast to our wide understanding of the genetic and epigenetic control of Th1 and Th2 regulation, we are at the beginning of the discernment of the genetic mechanisms for the commitment of the Th17 subset. Th1 and Th2 development are initiated by TCR signalling in conjunction with master transcription factor regulators, T-bet for Th1 and GATA-3 for Th2, which trigger integrated signals with inheritable epigenetic changes that allow tissue-specific gene expression⁽¹²⁴⁾. Each of these processes requires the participation of factors that are activated by specific cytokines. IL-12 signalling through Stat4 is associated with Th1 differentiation, and IL-4 through Stat6 governs Th2 cell fate⁽⁵⁶⁾. The transcription factors RORyt and RORa seem to be hallmark regulators of Th17 development^(71,125). Both factors belong to the family of retinoic-acid-related orphan nuclear receptors, which in turn is included in the hormone nuclear receptor superfamily. The orphan label is due to the fact that the ligand for these receptors is unknown. While RORy is broadly expressed, the isoform RORyt is exclusively found in cells of the immune system⁽¹²⁶⁾. RORyt is coexpressed with IL-17 in the mucosa constituent lamina propria. Ivanov et al. found that RORyt is required for IL-17 expression in response to IL-6 and TGF- β , and RORyt-deficient mice have attenuated EAE and lack tissue-infiltrating Th17 cells⁽⁷¹⁾. Nevertheless, residual Th17 cells are still present in conditions of RORy deficiency and EAE was not completely abolished. This can be justified by the results of Yang et al., showing that another related nuclear receptor, ROR α , also induced by TGF- β and IL-6, can collaborate in Th17 promotion⁽¹²⁵⁾. In fact, double deficiencies in ROR α and RORy impair Th17 generation and confer more resistance to inflammatory disease than RORy deficiency alone. However, single RORa deficiency in T cells only resulted in modest decrease of IL-17 and IL-23R expression, and had a very moderate inhibition of EAE. Thus, these two ROR factors seem to have redundant functions acting in a synergistic way in promoting Th17 cells.

Upregulation of ROR γ t is Stat3-dependent⁽¹²⁷⁾. This is not the only level at which Stat3 regulates Th17 development

since it can also induce the expression of IL-23R⁽¹²⁸⁾. Moreover, the positive effect of IL-6, IL-21 and IL-23 on the Th17 subset is dependent on Stat3(48,51,129). On the other hand, negative regulation of Th17 generation by Socs3 was found to act mainly on IL-23-mediated Stat3 phosphorylation⁽¹³⁰⁾. A definitive result to ascribe a role for Stat3 in Th17 development was the finding that Stat3 deficiency resulted in defective Th17-cell differentiation in vivo and protection against EAE⁽¹²⁸⁾. Collecting these data together, Stat3 seems to act at several levels during the process of Th17 phenotype definition, taking part in sequential and feedback loops of cytokine functions: first, IL-6 induces IL-21 production in a Stat3-dependent fashion. Once produced, IL-21 autocrinally stimulates its own synthesis and IL-23R expression through Stat3 activity. This allows IL-23 signalling leading to RORyt induction, which in turn up-regulates IL-23R expression in collaboration with Stat3. In addition, IL-21 cooperates with TGF- β to promote IL-17 expression with the involvement of Stat3 and RORyt (and/or ROR α).

Interferon-regulatory factor 4 (IRF4), essential for the development of Th2 cells⁽¹³¹⁻¹³³⁾, is also critical for the generation of Th17 cells and for EAE induction⁽¹³⁴⁾. IRF4-deficient mice did not develop EAE and showed a fault of IL-17 expression by T helper cells, while transfer of wild-type T helper cells allowed EAE susceptibility. Another feature of Irf4^{-/-} T cells is the reduced expression of RORyt. However, overexpression of this factor in Irf4^{-/-} T cells only restored Th17-cell differentiation partially. Thus, whether IRF4 acts upstream or downstream of RORyt is not clear.

Recently, it has been reported that Ets-1 deficiency is associated with increased expression of IL-17, IL-22 and IL-23R in response to IL-6 and TGF- β , indicating an enhanced efficiency to Th17 cell differentiation⁽¹³⁴⁾. However, Ets-1 apparently does not affect directly the expression of these genes. Rather, Ets-1-deficient T cells produce less IL-2 and have impaired responsiveness in terms of IL-2-mediated inhibition of Th17 differentiation. The resistance to IL-2 suppression was caused by a defect downstream of Stat5 phosphorylation, but was not caused by a difference in the levels of RORyt.

Regarding direct regulation of IL-17 promoter, chromatin immunoprecipitation (Chip) assays determined that Stat3 directly binds to the murine IL-17A and IL-17F promoters⁽¹³⁰⁾. Ichiyama et al. reported that RORyt directly binds to the IL-17 promoter, and found two potential ROR binding sites⁽¹³⁵⁾. Besides, RORyt was sufficient for IL-17 promoter activation. Another recent work describes that the transcription factor Runx-1 collaborates with RORyt to activate this promoter, although in the absence of RORyt, Runx-1 was not able to induce IL-17 transcription⁽¹³⁶⁾. Fewer information is available about the activity of the human IL-17 gene promoter, and it is unknown if RORyt is able to bind to it. Nevertheless, the minimal promoter has been defined in a region between 232 and 159 nucleotides upstream from the start transcription point. This region contains two NFAT recognition sites which seem to be functional after TCR stimulation at least in Jurkat cells⁽¹³⁷⁾. However, no more data about the role of NFAT in the control of IL-17 expression have been reported up to now.

In spite of our poor understanding of the genetic programs directing Th17 cells development and activity, the current thought is that, as for Th1 and Th2 subsets, Th17 polarization obeys to an epigenetic control, most probably controlled by ROR γ t and ROR α transcription factors, with contribution of other players like Stat3, IRF4, or other still unidentified factors. In mice, the genes for IL-17A and IL-17F are linked in chromosome 1, and their expression also appears to be linked, similarly to the IL-4, IL-5 and IL-13 locus in Th2 cells. The possibility of chromatin remodelling events allowing epigenetic control of Th17-cell development is supported by the results obtained by Akimzhamov et al.⁽¹³⁸⁾, who showed that Il17a and Il17f genes undergo H3 acetylation in response to TGF- β and IL-6, implying increased accessibility of the locus.

REGULATION OF THE TH17/Treg BALANCE

Current data indicate that the reciprocal developmental pathways for the generation of effector Th17 and Treg cells are controlled by the cytokine environment through an strict transcription program. It is established that CD4+CD25+Foxp3+ Treg cells are generated in the thymus (natural Treg) or may be induced by TGF- β in the periphery from naïve CD4+CD25-cells (induced or adaptive Treg)^(67,139-142). The dichotomy between the generation of Th17 and regulatory Foxp3+ T cells was first demonstrated by Betelli et al. using IL-6, which completely inhibited the generation of Foxp3+ Treg cells induced by TGF- β , directing the phenotype fate toward IL-17-producer cells⁽³²⁾.

TGF- β alone is able to induce the expression of both master transcription factors involved in each genetic program: ROR γ t for Th17 cells and Foxp3 for Treg cells (Fig. 2). In spite of this, TGF- β does not initiate Th17 differentiation unless pro-inflammatory factors, such as IL-6 or IL-21, are also present. In the absence of these cytokines, Foxp3 interacts with ROR γ t suppressing IL-17 transcription⁽¹³⁵⁾. IL-6, IL-21, and IL-23 are able to relieve Foxp3 inhibition of ROR γ t, and to activate Stat3, allowing Th17 cell differentiation. Zhou et al. have proposed that this subtle decision can be influenced



Figure 2. Antagonic Treg and Th17 generations share an intrinsic genetic program. TGF- β activity on naïve CD4+ cells induces the expression of both Foxp3 and RORyt transcription factors. In the absence of IL-6, RORyt and Runx-1 are sequestered by Foxp3 binding; this interaction prevents contact with ROR elements (RORE) in the IL-17 promoter. Cascade signalling triggered by IL-6 leads in naïve, and probably in Treg defined cells, to Stat3 activation, RORyt expression, down-regulation of Foxp3 expression, and suppression of the binding of pre-existing Foxp3 to RORyt. In a Stat3-dependent mechanism, the activity of RORyt, ROR α and Runx-1 allows IL-17 gene expression. The subsequent expression of IL-23R, IL-6 and IL-21 guide a feedback of signals to amplify and stabilize the Th17 phenotype. IRF4 activation by antigen (Ag) presentation also collaborates to Th17 cell differentiation, whether upstream or downstream of RORyt remaining to be determined. Dark boxes correspond to Stat3 regulated genes.

by the environmental amount of TGF- $\beta^{(143)}$. According to these authors, at low concentrations, TGF- β would synergize with interleukin IL-6 and IL-21, favouring Th17 cell differentiation, whereas high concentrations of TGF- β would repress IL-23R expression and favour Foxp3⁺ Treg cells. Runx1 has just been proposed as a new player in this network of transcription factors regulating Th17/Treg balance⁽¹³⁶⁾. Results in this report show that Runx1, in addition to interact with ROR γ t on the IL-17 promoter, can be bound by Foxp3 and probably be neutralized in its ability to activate IL-17 transcription.

The function of IL-2, needed for Treg cell survival and negative regulator of Th17 cells, could constitute an additional control in the homeostasis of the Treg and Th17 subsets. On the other hand, retinoids appear to be other physiologic regulators of Th17/Treg differentiation⁽¹⁴⁴⁻¹⁴⁶⁾. Vitamin A derivatives are protective in animal models of autoimmune disease and current data suggest that retinoic acid produced by dendritic cells reciprocally regulates Th17 and Treg differentiation.

It is well-known that cAMP inhibits T cell proliferation^(147,148). This second messenger is involved in suppressor activity by Treg cells⁽¹⁴⁹⁾. In effector cells, cAMP is strongly increased upon coactivation with Treg cells, and it was proposed that cAMP can be transferred from Treg to effector targets via cell contact-dependent gap junctions. Although the role of cAMP concerning to Th17 cell function is largely unclear, there are some indications of increased IL-17 production mediated by this second messenger. Chizzolini et al.⁽¹⁵⁰⁾, have demonstrated that prostaglandin E2, whose signalling leads to elevated cAMP, synergizes with IL-23 to favour Th17 expansion. Nevertheless,

the authors did not analyze if this effect is cAMP-dependent or not. Besides, Yadav et al.⁽¹⁵¹⁾ found that the vasoactive intestinal peptide (VIP) induces Th17 differentiation, in a mechanism suppressed by inhibition of protein kinase A, a main target of cAMP signalling. On the other hand, our unpublished results suggest that some agents increasing cAMP favour *in vitro* Th17 development. Although more clarifying data about cAMP involvement in Th17 induction is needed, cAMP might be another mediator involved in the Th17/Treg balance, which, like intracellular signalling of TGF- β , could have a double function collaborating in Treg or Th17 differentiation depending on environmental signals.

A remarkable novel suggestion about the mechanisms of Th17/Treg balance has been made by Yang et al.⁽¹⁵²⁾, who suggest that proinflammatory cytokines produced in the inflamed tissue not only affect the generation of induced Treg cells, but also might inhibit the function of already existing Treg cells. In addition, the absence of Foxp3 expression in T cells leads to increased Th1 cell differentiation without enhancing Th17 development. These data could be interpreted as that the real opposite pathways are those followed by Th1 and Treg cells, while Th17 and Treg generation share intrinsic common programs which allow some plasticity to reprogramming the phenotype of Treg cells towards Th17 effectors. If confirmed, these data could have important implications in therapies against autoimmune diseases with Treg cells, and it should be required to complement them with suppression of IL-6 to avoid redifferentiation into pathogenic Th17 cells.

ACKNOWLEDGEMENTS

The authors thank M. J. Jerez for the careful grammar correction of this manuscript. This work was supported by grants from Instituto de Salud Carlos III (FIS-PI061012) and MM Foundation (MPY-1156/07).

DISCLOSURES

The author declares no financial conflicts of interest.

CORRESPONDENCIA:

Sara Ballester

Unidad de Regulación Génica, Centro Nacional de Microbiología, Instituto de Salud Carlos III Ctra. Majadahonda-Pozuelo Km 28220 Madrid Tel: +34 91 822 3922 E-mail: sballes@isciii.es

REFERENCES

- Mosmann TR, Cherwinski H, Bond MW, Giedlin MA, Coffman RL. Two types of murine helper T cell clone. I. Definition according to profiles of lymphokine activities and secreted proteins. J Immunol 1986;136:2348-2357.
- Harrington LE, Hatton RD, Mangan PR, Turner H, Murphy TL, Murphy KM, et al. Interleukin 17-producing CD4⁺ effector T cells develop via a lineage distinct from the T helper type 1 and 2 lineages. Nat Immunol 2005;6:1123-1132.
- 3. Park H, Li Z, Yang XO, Chang SH, Nurieva R, Wang YH, et al. A distinct lineage of CD4 T cells regulates tissue inflammation by producing interleukin 17. Nat Immunol 2005; 6: 1133-1141.
- Rouvier E, Luciani MF, Mattei MG, Denizot F, Golstein P. CTLA-8, cloned from an activated T cell, bearing AU-rich messenger RNA instability sequences, and homologous to a herpesvirus saimiri gene. J Immunol 1993; 150: 5445-5456.
- Yao Z, Painter SL, Fanslow WC, Ulrich D, Macduff BM, Spriggs MK, et al. Human IL-17: A novel cytokine derived from T cells. J Immunol 1995; 155: 5483-5486.
- Chang SH, Dong C. A novel heterodimeric cytokine consisting of IL-17 and IL-17F regulates inflammatory responses. Cell Res 2007; 17: 435-440.
- Wright JF, Guo Y, Quazi A, Luxenberg DP, Bennett F, Ross JF, et al. Identification of an interleukin 17F/17A heterodimer in activated human CD4⁺ T cells. J Biol Chem 2007; 282: 13447-13455.
- Liang SC, Long AJ, Bennett F, Whitters MJ, Karim R, Collins M, et al. An IL-17F/A heterodimer protein is produced by mouse Th17 cells and induces airway neutrophil recruitment. J Immunol 2007; 179: 7791-7799.
- 9. Moseley TA, Haudenschild DR, Rose L, Reddi AH. Interleukin-17 family and IL-17 receptors. Cytokine Growth Factor Rev 2003; 14: 155-174.
- 10. Haudenschild D, Moseley T, Rose L, Reddi AH. Soluble and transmembrane isoforms of novel interleukin-17 receptor-like protein by RNA splicing and expression in prostate cancer. J Biol Chem 2002; 277: 4309-4316.
- Hymowitz SG, Filvaroff EH, Yin JP, Lee J, Cai L, Risser P, et al. IL-17s adopt a cystine knot fold: Structure and activity of a novel cytokine, IL-17F, and implications for receptor binding. EMBO J 2001; 20: 5332-5341.
- 12. Gaffen SL. An overview of IL-17 function and signaling. Cytokine 2008; 43: 402-407.
- Shalom-Barak T, Quach J, Lotz M. Interleukin-17-induced gene expression in articular chondrocytes is associated with activation of mitogen-activated protein kinases and NF-κB. J Biol Chem 1998; 273: 27467-27473.
- Hata K, Andoh A, Shimada M, Fujino S, Bamba S, Araki Y, et al. IL-17 stimulates inflammatory responses via NF-κB and MAP kinase pathways in human colonic myofibroblasts. Am J Physiol Gastrointest Liver Physiol 2002; 282: G1035-1044.
- Shen F, Hu Z, Goswami J, Gaffen SL. Identification of common transcriptional regulatory elements in interleukin-17 target genes. J Biol Chem 2006; 281: 24138-24148.
- Patel DN, King CA, Bailey SR, Holt JW, Venkatachalam K, Agrawal A, et al. Interleukin-17 stimulates C-reactive protein expression in hepatocytes and smooth muscle cells via p38 MAPK and ERK1/2dependent NF-κB and C/EBP, activation. J Biol Chem 2007; 282: 27229-27238.

- 17. Schwandner R, Yamaguchi K, Cao Z. Requirement of tumor necrosis factor receptor-associated factor (TRAF)6 in interleukin 17 signal transduction. J Exp Med 2000; 191: 1233-1240.
- Hartupee J, Liu C, Novotny M, Li X, Hamilton T. IL-17 enhances chemokine gene expression through mRNA stabilization. J Immunol 2007; 179: 4135-4141.
- 19. Infante-Duarte C, Horton HF, Byrne MC, Kamradt T. Microbial lipopeptides induce the production of IL-17 in Th cells. J Immunol 2000; 165: 6107-6115.
- Bettelli E, Sullivan B, Szabo SJ, Sobel RA, Glimcher LH, Kuchroo VK. Loss of T-bet, but not STAT1, prevents the development of experimental autoimmune encephalomyelitis. J Exp Med 2004; 200: 79-87.
- Zhang GX, Gran B, Yu S, Li J, Siglienti I, Chen X, et al. Induction of experimental autoimmune encephalomyelitis in IL-12 receptorbeta 2-deficient mice: IL-12 responsiveness is not required in the pathogenesis of inflammatory demyelination in the central nervous system. J Immunol 2003; 170: 2153-2160.
- Billiau A, Heremans H, Vandekerckhove F, Dijkmans R, Sobis H, Meulepas E, et al. Enhancement of experimental allergic encephalomyelitis in mice by antibodies against IFN-γ. J Immunol 1988; 140: 1506-1510.
- Gran B, Chu N, Zhang GX, Yu S, Li Y, Chen XH, et al. Early administration of IL-12 suppresses EAE through induction of interferon-γ. J Neuroimmunol 2004; 156: 123-131.
- 24. Gran B, Zhang GX, Yu S, Li J, Chen XH, Ventura ES, et al. IL-12p35-deficient mice are susceptible to experimental autoimmune encephalomyelitis: Evidence for redundancy in the IL-12 system in the induction of central nervous system autoimmune demyelination. J Immunol 2002; 169: 7104-7110.
- 25. Voorthuis JA, Uitdehaag BM, De Groot CJ, Goede PH, van der Meide PH, Dijkstra CD. Suppression of experimental allergic encephalomyelitis by intraventricular administration of interferonγ in Lewis rats. Clin Exp Immunol 1990; 81: 183-188.
- 26. Willenborg DO, Fordham SA, Staykova MA, Ramshaw IA, Cowden WB. IFN-γ is critical to the control of murine autoimmune encephalomyelitis and regulates both in the periphery and in the target tissue: A possible role for nitric oxide. J Immunol 1999; 163: 5278-5286.
- 27. Oppmann B, Lesley R, Blom B, Timans JC, Xu Y, Hunte B, et al. Novel p19 protein engages IL-12p40 to form a cytokine, IL-23, with biological activities similar as well as distinct from IL-12. Immunity 2000; 13: 715-725.
- Cua DJ, Sherlock J, Chen Y, Murphy CA, Joyce B, Seymour B, et al. Interleukin-23 rather than interleukin-12 is the critical cytokine for autoimmune inflammation of the brain. Nature 2003; 421: 744-748.
- 29. Langrish CL, Chen Y, Blumenschein WM, Mattson J, Basham B, Sedgwick JD, et al. IL-23 drives a pathogenic T cell population that induces autoimmune inflammation. J Exp Med 2005; 201: 233-240.
- Chen Y, Langrish CL, McKenzie B, Joyce-Shaikh B, Stumhofer JS, McClanahan T, et al. Anti-IL-23 therapy inhibits multiple inflammatory pathways and ameliorates autoimmune encephalomyelitis. J Clin Invest 2006; 116: 1317-1326.
- Murphy CA, Langrish CL, Chen Y, Blumenschein W, McClanahan T, Kastelein RA, et al. Divergent pro- and antiinflammatory roles for IL-23 and IL-12 in joint autoimmune inflammation. J Exp Med 2003; 198: 1951-1957.

- Bettelli E, Carrier Y, Gao W, Korn T, Strom TB, Oukka M, et al. Reciprocal developmental pathways for the generation of pathogenic effector TH17 and regulatory T cells. Nature 2006; 441: 235-238.
- Mangan PR, Harrington LE, O'Quinn DB, Helms WS, Bullard DC, Elson CO, et al. Transforming growth factor-β induces development of the T(H)17 lineage. Nature 2006; 441: 231-234.
- 34. Veldhoen M, Hocking RJ, Flavell RA, Stockinger B. Signals mediated by transforming growth factor-β initiate autoimmune encephalomyelitis, but chronic inflammation is needed to sustain disease. Nat Immunol 2006; 7: 1151-1156.
- 35. Lohr J, Knoechel B, Abbas AK. Regulatory T cells in the periphery. Immunol Rev 2006; 212: 149-162.
- 36. Acosta-Rodriguez EV, Napolitani G, Lanzavecchia A, Sallusto F. Interleukins 1, and 6 but not transforming growth factor-β are essential for the differentiation of interleukin 17-producing human T helper cells. Nat Immunol 2007; 8: 942-949.
- Chen Z, Tato CM, Muul L, Laurence A, O'Shea JJ. Distinct regulation of interleukin-17 in human T helper lymphocytes. Arthritis Rheum 2007; 56: 2936-2946.
- van Beelen AJ, Zelinkova Z, Taanman-Kueter EW, Muller FJ, Hommes DW, Zaat SA, et al. Stimulation of the intracellular bacterial sensor NOD2 programs dendritic cells to promote interleukin-17 production in human memory T cells. Immunity 2007; 27: 660-669.
- Wilson NJ, Boniface K, Chan JR, McKenzie BS, Blumenschein WM, Mattson JD, et al. Development, cytokine profile and function of human interleukin 17-producing helper T cells. Nat Immunol 2007; 8: 950-957.
- 40. Yang L, Anderson DE, Baecher-Allan C, Hastings WD, Bettelli E, Oukka M, et al. IL-21 and TGF- β are required for differentiation of human T(H)17 cells. Nature 2008; 454: 350-352.
- Manel N, Unutmaz D, Littman DR. The differentiation of human T(H)-17 cells requires transforming growth factor-, and induction of the nuclear receptor RORyt. Nat Immunol 2008; 9: 641-649.
- Volpe E, Servant N, Zollinger R, Bogiatzi SI, Hupe P, Barillot E, et al. A critical function for transforming growth factor-β, interleukin 23 and proinflammatory cytokines in driving and modulating human T(H)-17 responses. Nat Immunol 2008; 9: 650-657.
- O'Garra A, Stockinger B, Veldhoen M. Differentiation of human T(H)-17 cells does require TGF-β! Nat Immunol 2008; 9: 588-590.
- 44. Aggarwal S, Ghilardi N, Xie MH, de Sauvage FJ, Gurney AL. Interleukin-23 promotes a distinct CD4 T cell activation state characterized by the production of interleukin-17. J Biol Chem 2003; 278: 1910-1914.
- Veldhoen M, Hocking RJ, Atkins CJ, Locksley RM, Stockinger B. TGF, in the context of an inflammatory cytokine milieu supports de novo differentiation of IL-17-producing T cells. Immunity 2006; 24: 179-189.
- Stritesky GL, Yeh N, Kaplan MH. IL-23 promotes maintenance but not commitment to the Th17 lineage. J Immunol 2008; 181: 5948-5955.
- 47. McGeachy MJ, Bak-Jensen KS, Chen Y, Tato CM, Blumenschein W, McClanahan T, et al. TGF-β and IL-6 drive the production of IL-17 and IL-10 by T cells and restrain T(H)-17 cell-mediated pathology. Nat Immunol 2007; 8: 1390-1397.
- Zhou L, Ivanov, II, Spolski R, Min R, Shenderov K, Egawa T, et al. IL-6 programs T(H)-17 cell differentiation by promoting sequential

engagement of the IL-21 and IL-23 pathways. Nat Immunol 2007; 8: 967-974.

- 49. Ogura H, Murakami M, Okuyama Y, Tsuruoka M, Kitabayashi C, Kanamoto M, et al. Interleukin-17 promotes autoimmunity by triggering a positive-feedback loop via interleukin-6 induction. Immunity 2008; 29: 628-636.
- 50. Kimura A, Naka T, Kishimoto T. IL-6-dependent and -independent pathways in the development of interleukin 17-producing T helper cells. Proc Natl Acad Sci USA 2007; 104: 12099-12104.
- 51. Nurieva R, Yang XO, Martinez G, Zhang Y, Panopoulos AD, Ma L, et al. Essential autocrine regulation by IL-21 in the generation of inflammatory T cells. Nature 2007; 448: 480-483.
- 52. Korn T, Bettelli E, Gao W, Awasthi A, Jager A, Strom TB, et al. IL-21 initiates an alternative pathway to induce proinflammatory T(H)17 cells. Nature 2007; 448: 484-487.
- Wei L, Laurence A, Elias KM, O'Shea JJ. IL-21 is produced by Th17 cells and drives IL-17 production in a STAT3-dependent manner. J Biol Chem 2007; 282: 34605-34610.
- Sutton C, Brereton C, Keogh B, Mills KH, Lavelle EC. A crucial role for interleukin (IL)-1 in the induction of IL-17-producing T cells that mediate autoimmune encephalomyelitis. J Exp Med 2006; 203: 1685-1691.
- 55. Gutcher I, Urich E, Wolter K, Prinz M, Becher B. Interleukin 18independent engagement of interleukin 18 receptor-α is required for autoimmune inflammation. Nat Immunol 2006; 7: 946-953.
- 56. Murphy KM, Reiner SL. The lineage decisions of helper T cells. Nat Rev Immunol 2002; 2: 933-944.
- 57. Stumhofer JS, Silver J, Hunter CA. Negative regulation of Th17 responses. Semin Immunol 2007; 19: 394-399.
- Colgan J, Rothman P. All in the family: IL-27 suppression of T(H)-17 cells. Nat Immunol 2006; 7: 899-901.
- Batten M, Li J, Yi S, Kljavin NM, Danilenko DM, Lucas S, et al. Interleukin 27 limits autoimmune encephalomyelitis by suppressing the development of interleukin 17-producing T cells. Nat Immunol 2006; 7: 929-936.
- 60. Stumhofer JS, Laurence A, Wilson EH, Huang E, Tato CM, Johnson LM, et al. Interleukin 27 negatively regulates the development of interleukin 17-producing T helper cells during chronic inflammation of the central nervous system. Nat Immunol 2006; 7: 937-945.
- Stumhofer JS, Silver JS, Laurence A, Porrett PM, Harris TH, Turka LA, et al. Interleukins 27 and 6 induce STAT3-mediated T cell production of interleukin 10. Nat Immunol 2007; 8: 1363-1371.
- 62. Jankovic D, Trinchieri G. IL-10 or not IL-10: That is the question. Nat Immunol 2007; 8: 1281-1283.
- 63. Fort MM, Cheung J, Yen D, Li J, Zurawski SM, Lo S, et al. IL-25 induces IL-4, IL-5, and IL-13 and Th2-associated pathologies in vivo. Immunity 2001; 15: 985-995.
- 64. Owyang AM, Zaph C, Wilson EH, Guild KJ, McClanahan T, Miller HR, et al. Interleukin 25 regulates type 2 cytokine-dependent immunity and limits chronic inflammation in the gastrointestinal tract. J Exp Med 2006; 203: 843-849.
- 65. Kleinschek MA, Owyang AM, Joyce-Shaikh B, Langrish CL, Chen Y, Gorman DM, et al. IL-25 regulates Th17 function in autoimmune inflammation. J Exp Med 2007; 204: 161-170.
- 66. Abbas AK. The control of T cell activation vs. tolerance. Autoimmun Rev 2003; 2: 115-118.

- 67. Shevach EM, DiPaolo RA, Andersson J, Zhao DM, Stephens GL, Thornton AM. The lifestyle of naturally occurring CD4⁺ CD25⁺ Foxp3⁺ regulatory T cells. Immunol Rev 2006; 212: 60-73.
- Laurence A, Tato CM, Davidson TS, Kanno Y, Chen Z, Yao Z, et al. Interleukin-2 signaling via STAT5 constrains T helper 17 cell generation. Immunity 2007; 26: 371-381.
- 69. Matsuzaki G, Umemura M. Interleukin-17 as an effector molecule of innate and acquired immunity against infections. Microbiol Immunol 2007; 51: 1139-1147.
- Linden A, Laan M, Anderson GP. Neutrophils, interleukin-17A and lung disease. Eur Respir J 2005; 25: 159-172.
- Ivanov, II, McKenzie BS, Zhou L, Tadokoro CE, Lepelley A, Lafaille JJ, et al. The orphan nuclear receptor RORγt directs the differentiation program of proinflammatory IL-17⁺ T helper cells. Cell 2006; 126: 1121-1133.
- Aujla SJ, Chan YR, Zheng M, Fei M, Askew DJ, Pociask DA, et al. IL-22 mediates mucosal host defense against Gram-negative bacterial pneumonia. Nat Med 2008; 14: 275-281.
- 73. Ye P, Rodriguez FH, Kanaly S, Stocking KL, Schurr J, Schwarzenberger P, et al. Requirement of interleukin 17 receptor signaling for lung CXC chemokine and granulocyte colony-stimulating factor expression, neutrophil recruitment, and host defense. J Exp Med 2001; 194: 519-527.
- Yang XO, Chang SH, Park H, Nurieva R, Shah B, Acero L, et al. Regulation of inflammatory responses by IL-17F. J Exp Med 2008; 205: 1063-1075.
- Chung DR, Kasper DL, Panzo RJ, Chitnis T, Grusby MJ, Sayegh MH, et al. CD4⁺ T cells mediate abscess formation in intra-abdominal sepsis by an IL-17-dependent mechanism. J Immunol 2003; 170: 1958-1963.
- Huang W, Na L, Fidel PL, Schwarzenberger P. Requirement of interleukin-17A for systemic anti-Candida albicans host defense in mice. J Infect Dis 2004; 190: 624-631.
- Zelante T, De Luca A, Bonifazi P, Montagnoli C, Bozza S, Moretti S, et al. IL-23 and the Th17 pathway promote inflammation and impair antifungal immune resistance. Eur J Immunol 2007; 37: 2695-2706.
- Kleinschek MA, Muller U, Brodie SJ, Stenzel W, Kohler G, Blumenschein WM, et al. IL-23 enhances the inflammatory cell response in Cryptococcus neoformans infection and induces a cytokine pattern distinct from IL-12. J Immunol 2006; 176: 1098-1106.
- Rudner XL, Happel KI, Young EA, Shellito JE. Interleukin-23 (IL-23)-IL-17 cytokine axis in murine Pneumocystis carinii infection. Infect Immun 2007; 75: 3055-3061.
- Kelly MN, Kolls JK, Happel K, Schwartzman JD, Schwarzenberger P, Combe C, et al. Interleukin-17/interleukin-17 receptor-mediated signaling is important for generation of an optimal polymorphonuclear response against Toxoplasma gondii infection. Infect Immun 2005; 73: 617-621.
- Yao Z, Fanslow WC, Seldin MF, Rousseau AM, Painter SL, Comeau MR, et al. Herpesvirus Saimiri encodes a new cytokine, IL-17, which binds to a novel cytokine receptor. Immunity 1995; 3: 811-821.
- Maertzdorf J, Osterhaus AD, Verjans GM. IL-17 expression in human herpetic stromal keratitis: Modulatory effects on chemokine production by corneal fibroblasts. J Immunol 2002; 169: 5897-5903.

- Patera AC, Pesnicak L, Bertin J, Cohen JI. Interleukin 17 modulates the immune response to vaccinia virus infection. Virology 2002; 299: 56-63.
- 84. Smiley KL, McNeal MM, Basu M, Choi AH, Clements JD, Ward RL. Association of γ interferon and interleukin-17 production in intestinal CD4⁺ T cells with protection against rotavirus shedding in mice intranasally immunized with VP6 and the adjuvant LT(R192G). J Virol 2007; 81: 3740-3748.
- Komiyama Y, Nakae S, Matsuki T, Nambu A, Ishigame H, Kakuta S, et al. IL-17 plays an important role in the development of experimental autoimmune encephalomyelitis. J Immunol 2006; 177: 566-573.
- Nakae S, Nambu A, Sudo K, Iwakura Y. Suppression of immune induction of collagen-induced arthritis in IL-17-deficient mice. J Immunol 2003; 171: 6173-6177.
- 87. Lubberts E, Koenders MI, Oppers-Walgreen B, van den Bersselaar L, Coenen-de Roo CJ, Joosten LA, et al. Treatment with a neutralizing anti-murine interleukin-17 antibody after the onset of collageninduced arthritis reduces joint inflammation, cartilage destruction, and bone erosion. Arthritis Rheum 2004; 50: 650-659.
- Rohn TA, Jennings GT, Hernandez M, Grest P, Beck M, Zou Y, et al. Vaccination against IL-17 suppresses autoimmune arthritis and encephalomyelitis. Eur J Immunol 2006; 36: 2857-2867.
- Uyttenhove C, Van Snick J. Development of an anti-IL-17A autovaccine that prevents experimental auto-immune encephalomyelitis. Eur J Immunol 2006; 36: 2868-2874.
- Weinstock-Guttman B, Ramanathan M, Zivadinov R. Interferonβ treatment for relapsing multiple sclerosis. Expert Opin Biol Ther 2008; 8: 1435-1447.
- 91. Martin-Saavedra FM, Flores N, Dorado B, Eguiluz C, Bravo B, Garcia-Merino A, et al. Beta-interferon unbalances the peripheral T cell proinflammatory response in experimental autoimmune encephalomyelitis. Mol Immunol 2007; 44: 3597-3607.
- 92. Martin-Saavedra FM, Gonzalez-Garcia C, Bravo B, Ballester S. Beta interferon restricts the inflammatory potential of CD4+ cells through the boost of the Th2 phenotype, the inhibition of Th17 response and the prevalence of naturally occurring T regulatory cells. Mol Immunol 2008; 45: 4008-4019.
- Guo B, Chang EY, Cheng G. The type I IFN induction pathway constrains Th17-mediated autoimmune inflammation in mice. J Clin Invest 2008; 118: 1680-1690.
- Rojo JM, Pini E, Ojeda G, Bello R, Dong C, Flavell RA, et al. CD4+ICOS+ T lymphocytes inhibit T cell activation 'in vitro' and attenuate autoimmune encephalitis 'in vivo'. Int Immunol 2008; 20: 577-589.
- Kotake S, Udagawa N, Takahashi N, Matsuzaki K, Itoh K, Ishiyama S, et al. IL-17 in synovial fluids from patients with rheumatoid arthritis is a potent stimulator of osteoclastogenesis. J Clin Invest 1999; 103: 1345-1352.
- 96. Kim HR, Cho ML, Kim KW, Juhn JY, Hwang SY, Yoon CH, et al. Up-regulation of IL-23p19 expression in rheumatoid arthritis synovial fibroblasts by IL-17 through PI3-kinase-, NF-κB- and p38 MAPK-dependent signalling pathways. Rheumatology (Oxford) 2007; 46: 57-64.
- 97. Cho ML, Yoon CH, Hwang SY, Park MK, Min SY, Lee SH, et al. Effector function of type II collagen-stimulated T cells from rheumatoid arthritis patients: Cross-talk between T cells and synovial fibroblasts. Arthritis Rheum 2004; 50: 776-784.

- Hwang SY, Kim JY, Kim KW, Park MK, Moon Y, Kim WU, et al. IL-17 induces production of IL-6 and IL-8 in rheumatoid arthritis synovial fibroblasts via NF-κB- and PI3-kinase/Akt-dependent pathways. Arthritis Res Ther 2004; 6: R120-128.
- Matusevicius D, Kivisakk P, He B, Kostulas N, Ozenci V, Fredrikson S, et al. Interleukin-17 mRNA expression in blood and CSF mononuclear cells is augmented in multiple sclerosis. Mult Scler 1999; 5: 101-104.
- 100. Lock C, Hermans G, Pedotti R, Brendolan A, Schadt E, Garren H, et al. Gene-microarray analysis of multiple sclerosis lesions yields new targets validated in autoimmune encephalomyelitis. Nat Med 2002; 8: 500-508.
- 101. Acosta-Rodriguez EV, Rivino L, Geginat J, Jarrossay D, Gattorno M, Lanzavecchia A, et al. Surface phenotype and antigenic specificity of human interleukin 17-producing T helper memory cells. Nat Immunol 2007; 8: 639-646.
- 102.Sato W, Aranami T, Yamamura T. Cutting edge: Human Th17 cells are identified as bearing CCR2+CCR5- phenotype. J Immunol 2007; 178: 7525-7529.
- 103. Vaknin-Dembinsky A, Balashov K, Weiner HL. IL-23 is increased in dendritic cells in multiple sclerosis and down-regulation of IL-23 by antisense oligos increases dendritic cell IL-10 production. J Immunol 2006; 176: 7768-7774.
- 104. Kebir H, Kreymborg K, Ifergan I, Dodelet-Devillers A, Cayrol R, Bernard M, et al. Human TH17 lymphocytes promote blood-brain barrier disruption and central nervous system inflammation. Nat Med 2007; 13: 1173-1175.
- 105. Tzartos JS, Friese MA, Craner MJ, Palace J, Newcombe J, Esiri MM, et al. Interleukin-17 production in central nervous systeminfiltrating T cells and glial cells is associated with active disease in multiple sclerosis. Am J Pathol 2008; 172: 146-155.
- 106. Teunissen MB, Koomen CW, de Waal Malefyt R, Wierenga EA, Bos JD. Interleukin-17 and interferon-γ synergize in the enhancement of proinflammatory cytokine production by human keratinocytes. J Invest Dermatol 1998; 111: 645-649.
- 107. Liang SC, Tan XY, Luxenberg DP, Karim R, Dunussi-Joannopoulos K, Collins M, et al. Interleukin (IL)-22 and IL-17 are coexpressed by Th17 cells and cooperatively enhance expression of antimicrobial peptides. J Exp Med 2006; 203: 2271-2279.
- 108. Wong CK, Lit LC, Tam LS, Li EK, Wong PT, Lam CW. Hyperproduction of IL-23 and IL-17 in patients with systemic lupus erythematosus: Implications for Th17-mediated inflammation in auto-immunity. Clin Immunol 2008; 127: 385-393.
- 109. Uhlig HH, McKenzie BS, Hue S, Thompson C, Joyce-Shaikh B, Stepankova R, et al. Differential activity of IL-12 and IL-23 in mucosal and systemic innate immune pathology. Immunity 2006; 25: 309-318.
- 110. Yen D, Cheung J, Scheerens H, Poulet F, McClanahan T, McKenzie B, et al. IL-23 is essential for T cell-mediated colitis and promotes inflammation via IL-17 and IL-6. J Clin Invest 2006; 116: 1310-1316.
- 111. Zhang Z, Zheng M, Bindas J, Schwarzenberger P, Kolls JK. Critical role of IL-17 receptor signaling in acute TNBS-induced colitis. Inflamm Bowel Dis 2006; 12: 382-388.
- 112. Fujino S, Andoh A, Bamba S, Ogawa A, Hata K, Araki Y, et al. Increased expression of interleukin 17 in inflammatory bowel disease. Gut 2003; 52: 65-70.

- 113. Annunziato F, Cosmi L, Santarlasci V, Maggi L, Liotta F, Mazzinghi B, et al. Phenotypic and functional features of human Th17 cells. J Exp Med 2007; 204: 1849-1861.
- 114. Ouyang W, Kolls JK, Zheng Y. The biological functions of T helper 17 cell effector cytokines in inflammation. Immunity 2008; 28: 454-467.
- 115.Fahy JV, Kim KW, Liu J, Boushey HA. Prominent neutrophilic inflammation in sputum from subjects with asthma exacerbation. J Allergy Clin Immunol 1995; 95: 843-852.
- 116. Wenzel SE, Schwartz LB, Langmack EL, Halliday JL, Trudeau JB, Gibbs RL, et al. Evidence that severe asthma can be divided pathologically into two inflammatory subtypes with distinct physiologic and clinical characteristics. Am J Respir Crit Care Med 1999; 160: 1001-1008.
- 117. Jatakanon A, Uasuf C, Maziak W, Lim S, Chung KF, Barnes PJ. Neutrophilic inflammation in severe persistent asthma. Am J Respir Crit Care Med 1999; 160: 1532-1539.
- 118. Ordonez CL, Shaughnessy TE, Matthay MA, Fahy JV. Increased neutrophil numbers and IL-8 levels in airway secretions in acute severe asthma: Clinical and biologic significance. Am J Respir Crit Care Med 2000; 161: 1185-1190.
- 119. Gibson PG, Simpson JL, Saltos N. Heterogeneity of airway inflammation in persistent asthma: Evidence of neutrophilic inflammation and increased sputum interleukin-8. Chest 2001; 119: 1329-1336.
- 120. Hellings PW, Kasran A, Liu Z, Vandekerckhove P, Wuyts A, Overbergh L, et al. Interleukin-17 orchestrates the granulocyte influx into airways after allergen inhalation in a mouse model of allergic asthma. Am J Respir Cell Mol Biol 2003; 28: 42-50.
- 121.Prause O, Bozinovski S, Anderson GP, Linden A. Increased matrix metalloproteinase-9 concentration and activity after stimulation with interleukin-17 in mouse airways. Thorax 2004; 59: 313-317.
- 122. Hoshino H, Laan M, Sjostrand M, Lotvall J, Skoogh BE, Linden A. Increased elastase and myeloperoxidase activity associated with neutrophil recruitment by IL-17 in airways in vivo. J Allergy Clin Immunol 2000; 105: 143-149.
- 123. Oboki K, Ohno T, Saito H, Nakae S. Th17 and allergy. Allergol Int 2008; 57: 121-134.
- 124. Lee GR, Kim ST, Spilianakis CG, Fields PE, Flavell RA. T helper cell differentiation: Regulation by cis elements and epigenetics. Immunity 2006; 24: 369-379.
- 125. Yang XO, Pappu BP, Nurieva R, Akimzhanov A, Kang HS, Chung Y, et al. T helper 17 lineage differentiation is programmed by orphan nuclear receptors ROR α and ROR γ . Immunity 2008; 28: 29-39.
- 126. Eberl G, Littman DR. The role of the nuclear hormone receptor RORγt in the development of lymph nodes and Peyer's patches. Immunol Rev 2003; 195: 81-90.
- 127. Yang XO, Panopoulos AD, Nurieva R, Chang SH, Wang D, Watowich SS, et al. STAT3 regulates cytokine-mediated generation of inflammatory helper T cells. J Biol Chem 2007; 282: 9358-9363.
- 128. Harris TJ, Grosso JF, Yen HR, Xin H, Kortylewski M, Albesiano E, et al. Cutting edge: An in vivo requirement for STAT3 signaling in TH17 development and TH17-dependent autoimmunity. J Immunol 2007; 179: 4313-4317.

- 129. Chen Z, Laurence A, O'Shea JJ. Signal transduction pathways and transcriptional regulation in the control of Th17 differentiation. Semin Immunol 2007; 19: 400-408.
- 130. Chen Z, Laurence A, Kanno Y, Pacher-Zavisin M, Zhu BM, Tato C, et al. Selective regulatory function of Socs3 in the formation of IL-17-secreting T cells. Proc Natl Acad Sci USA 2006; 103: 8137-8142.
- 131. Hu CM, Jang SY, Fanzo JC, Pernis AB. Modulation of T cell cytokine production by interferon regulatory factor-4. J Biol Chem 2002; 277: 49238-49246.
- 132. Lohoff M, Mittrucker HW, Prechtl S, Bischof S, Sommer F, Kock S, et al. Dysregulated T helper cell differentiation in the absence of interferon regulatory factor 4. Proc Natl Acad Sci USA 2002; 99: 11808-11812.
- 133. Rengarajan J, Mowen KA, McBride KD, Smith ED, Singh H, Glimcher LH. Interferon regulatory factor 4 (IRF4) interacts with NFATc2 to modulate interleukin 4 gene expression. J Exp Med 2002; 195: 1003-1012.
- 134. Brustle A, Heink S, Huber M, Rosenplanter C, Stadelmann C, Yu P, et al. The development of inflammatory T(H)-17 cells requires interferon-regulatory factor 4. Nat Immunol 2007; 8: 958-966.
- 135. Ichiyama K, Yoshida H, Wakabayashi Y, Chinen T, Saeki K, Nakaya M, et al. Foxp3 inhibits RORγt-mediated IL-17A mRNA transcription through direct interaction with RORγt. J Biol Chem 2008; 283: 17003-17008.
- 136. Zhang F, Meng G, Strober W. Interactions among the transcription factors Runx1, RORgammat and Foxp3 regulate the differentiation of interleukin 17-producing T cells. Nat Immunol 2008; 9: 1297-1306.
- 137. Liu XK, Lin X, Gaffen SL. Crucial role for nuclear factor of activated T cells in T cell receptor-mediated regulation of human interleukin-17. J Biol Chem 2004; 279: 52762-52771.
- 138. Akimzhanov AM, Yang XO, Dong C. Chromatin remodeling of interleukin-17 (IL-17)-IL-17F cytokine gene locus during inflammatory helper T cell differentiation. J Biol Chem 2007; 282: 5969-5972.
- 139. Sakaguchi S. Regulatory T cells: Key controllers of immunologic self-tolerance. Cell 2000; 101: 455-458.
- 140. Chen W, Jin W, Hardegen N, Lei KJ, Li L, Marinos N, et al. Conversion of peripheral CD4+CD25- naive T cells to CD4+CD25+ regulatory T cells by TGF-beta induction of transcription factor Foxp3. J Exp Med 2003; 198: 1875-1886.
- 141. Faria AM, Weiner HL. Oral tolerance and TGF-β-producing cells. Inflamm Allergy Drug Targets 2006; 5: 179-190.
- 142. Pini E, Ojeda G, Portolés P. The renaissance of T regulatory cells: Looking for markers in a haystack. Inmunología 2007; 26: 100-107.
- 143. Zhou L, Lopes JE, Chong MM, Ivanov, II, Min R, Victora GD, et al. TGF-β-induced Foxp3 inhibits T(H)17 cell differentiation by antagonizing RORγt function. Nature 2008; 453: 236-240.
- 144. Schambach F, Schupp M, Lazar MA, Reiner SL. Activation of retinoic acid receptor-α favours regulatory T cell induction at the expense of IL-17-secreting T helper cell differentiation. Eur J Immunol 2007; 37: 2396-2399.
- 145. Elias KM, Laurence A, Davidson TS, Stephens G, Kanno Y, Shevach EM, et al. Retinoic acid inhibits Th17 polarization and enhances FoxP3 expression through a Stat-3/Stat-5 independent signaling pathway. Blood 2008; 111: 1013-1020.

- 146. Mucida D, Park Y, Kim G, Turovskaya O, Scott I, Kronenberg M, et al. Reciprocal TH17 and regulatory T cell differentiation mediated by retinoic acid. Science 2007; 317: 256-260.
- 147. Molina CA, Foulkes NS, Lalli E, Sassone-Corsi P. Inducibility and negative autoregulation of CREM: An alternative promoter directs the expression of ICER, an early response repressor. Cell 1993; 75: 875-886.
- 148. Lalli E, Lee JS, Masquilier D, Schlotter F, Foulkes NS, Molina CA, et al. Nuclear response to cyclic AMP: Central role of transcription factor CREM (cyclic-AMP-responsive-element modulator). Biochem Soc Trans 1993; 21: 912-917.
- 149.Bopp T, Becker C, Klein M, Klein-Hessling S, Palmetshofer A, Serfling E, et al. Cyclic adenosine monophosphate is a key component

of regulatory T cell-mediated suppression. J Exp Med 2007; 204: 1303-1310.

- 150. Chizzolini C, Chicheportiche R, Alvarez M, de Rham C, Roux-Lombard P, Ferrari-Lacraz S, et al. Prostaglandin E2 synergistically with interleukin-23 favors human Th17 expansion. Blood 2008; 112: 3696-3703.
- 151.Yadav M, Rosenbaum J, Goetzl EJ. Cutting edge: Vasoactive intestinal peptide (VIP) induces differentiation of Th17 cells with a distinctive cytokine profile. J Immunol 2008; 180: 2772-2776.
- 152. Yang XO, Nurieva R, Martinez GJ, Kang HS, Chung Y, Pappu BP, et al. Molecular antagonism and plasticity of regulatory and inflammatory T cell programs. Immunity 2008; 29: 44-56.