

un aislado de SARM *mecC* como SASM^{2,9}. Por ello, realizar antibiograma y utilizar técnicas específicas es imprescindible². El SARM *mecC* representa un porcentaje menor al 1% de los SARM y la mayoría pertenecen al complejo clonal (CC) CC130. En España, dentro de este CC, el secuenciotipo y tipo de *spa* más frecuentes son ST1945 y t843, respectivamente^{5,8}. En nuestro paciente, la PCR inicial y la técnica inmunocromatográfica que detecta la presencia de PBP2a identificaron al aislado como SASM. Pero tras la realización del antibiograma, la resistencia se confirmó mediante PCR específica y secuenciación, demostrándose también la presencia del gen *mecC* integrado en el cassette cromosómico *SCCmec XI*. Además, mediante *multilocus sequence typing* (MLST) se catalogó dentro del CC130, y mediante el tipado molecular por PCR y posterior secuenciación del *spa* se detectó un nuevo tipo de *spa* registrado como t20888, dentro del ST1945.

En conclusión, este es un caso de infección invasiva por SARM *mecC* con un nuevo tipo de *spa* y con fatal desenlace en el que el paciente reunía los factores de riesgo para padecer infección por este microorganismo: contacto estrecho con ganado, enfermedad subyacente, edad avanzada y colonización nasal. En los laboratorios de Microbiología es importante la realización de antibiograma manual junto a la utilización de técnicas moleculares rápidas, ya que se pueden identificar mecanismos no incluidos en las dianas moleculares de las PCR comerciales. Esto es esencial para elección de un tratamiento antibiótico adecuado.

Agradecimientos

Agradecemos la participación de Carmen Torres y su grupo de trabajo de La Universidad de La Rioja en la caracterización de la cepa de *S. aureus* portadora del gen *mecC*.

Bibliografía

- Lakhundi S, Zhang K. Methicillin-resistant *Staphylococcus aureus*: molecular characterization, evolution, and epidemiology. Clin Microbiol Rev. 2018;31:e00020-18, <http://dx.doi.org/10.1128/CMR.00020-18>.
- García-Álvarez L, Holden MTG, Lindsay H, Webb CR, Brown DFJ, Curran MD, et al. Methicillin-resistant *Staphylococcus aureus* with a novel *mecA* homologue in human and bovine populations in the UK and Denmark: a descriptive study. Lancet Infect Dis. 2011;11:595–603, [http://dx.doi.org/10.1016/S1473-3099\(11\)70126-8](http://dx.doi.org/10.1016/S1473-3099(11)70126-8).
- García-Garrote F, Cercenado E, Marín M, Bal M, Trincado P, Corredoira J, et al. Methicillin-resistant *Staphylococcus aureus* carrying the *mecC* gene: emergence in Spain and report of a fatal case of bacteraemia. J Antimicrob Chemother. 2014;69:45–50, <http://dx.doi.org/10.1093/jac/dkt327>.
- The European Committee on Antimicrobial Susceptibility Testing. Clinical breakpoints-bacteria version 11.0.2022. [consultado 2 Nov 2022]. Disponible en: www.eucast.org.
- Larsen J, Raisen CL, Ba X, Sadgrove NJ, Padilla-Gonzalez GF, Simmonds MSJ, et al. Emergence of methicillin resistance predates the clinical use of antibiotics. Nature. 2022;602:135–41, <http://dx.doi.org/10.1038/s41586-021-04265-w>.
- Harrison EM, Paterson GK, Holden MTG, Larsen J, Stegger M, Larson AR, et al. Whole genome sequencing identifies zoonotic transmission of MRSA isolates with the novel *mecA* homologue *mecC*. EMBO Mol Med. 2013;5:504–15, <http://dx.doi.org/10.1002/emmm.201202413>.
- Angen O, Stegger M, Larsen J, Lilje B, Kaya H, Pedersen KS, et al. Report of *mecC*-carrying MRSA in domestic swine. J Antimicrob Chemother. 2017;72:60–3, <http://dx.doi.org/10.1093/jac/dkw389>.
- Lozano C, Fernández-Fernández R, Ruiz-Ripa L, Gómez P, Zarazaga M, Torres C. Human *mecC*-carrying MRSA: clinical implications and risk factors. Microorganisms. 2020;8:1615, <http://dx.doi.org/10.3390/microorganisms8101615>.
- Ba X, Harrison EM, Lovering AL, Gleadall N, Zadiks R, Parkhill J, et al. Old drug to treat resistant bugs: methicillin-resistant *Staphylococcus aureus* isolates with *mecC* are susceptible to a combination of penicillin and clavulanic acid. Antimicrob Agents Chemother. 2015;59:7396–404, <http://dx.doi.org/10.1128/AAC.01469-15>.

Iratí Arregui García ^{a,*}, M. Eugenia Portillo ^{a,b}, Luis Torroba Álvarez ^{a,b} y Carmen Ezpeleta Baquedano ^{a,b}

^a Servicio de Microbiología Clínica, Hospital Universitario de Navarra, Pamplona, España

^b Instituto de Investigación Sanitaria de Navarra (IdisNA), Pamplona, España

* Autor para correspondencia.

Correo electrónico: iratiarreguig@gmail.com (I. Arregui García).

<https://doi.org/10.1016/j.eimc.2023.01.008>

0213-005X/ © 2023 Sociedad Española de Enfermedades Infecciosas y Microbiología Clínica. Publicado por Elsevier España, S.L.U. Todos los derechos reservados.

High frequency of *Streptococcus pneumoniae* serotype 3 in negative pleural fluid cultures from paediatric samples obtained in the Madrid region from 2018 to 2022, detected by direct identification using PCR-reverse-hybridization strip-based assay



Elevada frecuencia del serotipo 3 de *Streptococcus pneumoniae* detectado mediante una técnica de PCR e hibridación reversa, en muestras pediátricas de líquido pleural con cultivo negativo, obtenidas en la Comunidad de Madrid entre 2018 y 2022

The first pneumococcal conjugate vaccine introduced in 2006 in the paediatric immunization calendar of the Madrid region was the 7-valent (PCV7), which was substituted in 2010 by the 13-valent pneumococcal conjugate vaccine (PCV13). In 2012, PCV13 was withdrawn from the Madrid paediatric public immunization programme, being prescribed privately. In 2015, the PCV13 was re-introduced at the Spanish national paediatric public vaccine scheme.¹ The Quellung reaction is the gold standard for pneumococcal serotyping. This method requires strains isolated in culture. However, in samples as parapneumonic effusion (PPE) and pleural

empyema (PE), commonly grouped as PPE/PE,² the bacterial growing is fastidious and the availability of colonies may be difficult. The objective of this work was to identify the not growing *Streptococcus pneumoniae* serotypes (SPNGST) causing pleural infection in children, in which the standard microbiological culture shows negative results.

Thirty-five PFS from paediatric patients (aged six months to 8 years; mean 3.4 years, standard deviation 2.1) with negative culture results, obtained between September 2018 and December 2022, were processed for the detection of the α -fucosidase gene using a real time PCR method (*Streptococcus pneumoniae* alpha-fucosidase gene Genesig® Advanced Kit; Primerdesign Ltd, United Kingdom). Positive α -fucosidase samples were subsequently tested by a PCR reverse-hybridization strip-based assay (S. PneumoStrip test; Operon S.A., Zaragoza, Spain) that allows serotype identification.

The α -fucosidase real-time PCR method was positive in 30 of the 35 samples studied (85.7%). The PCR reverse-hybridization assay showed positive results for the *lytA*, *cpsA* and *ply* genes in 28 of the 30 samples (93.3%). The serotypes detected by this technique in these 28 samples were twenty-one serotype 3 (75%), two

Table 1

Distribution of not-growing *Streptococcus pneumoniae* serotypes detected by PCR in pleural fluid samples according to the year the sample was obtained.

Year	Number of samples	Positive real-time PCR for the α -fucosidase-gene	Positive PCR hybridization for <i>lytA</i> , <i>cpsA</i> , and <i>ply</i> genes	Serotypes detected by PCR hybridization						
				3	19A	8	10A	1	9A/9V	33F/33A
2018	4	3	3		1	1				1
2019	9	6	4		2		1			1
2020	3	3	3		1		1			
2021	1	1	1					1		
2022	18	17	17		17					
2018–2022	35	30	28		21	1	2	1	1	1

serotype 8 (7.1%), one serotype 19A (3.6%), one serotype 1 (3.6%), one serotype 10A (3.6%), one 9A/9V (3.6%) and one 33F/33A (3.6%).

Table 1 shows the distribution of PCR SPNGST detected in PFS according to time. Most serotype 3 cases occurred in 2022 (17/21). The only one PCV13 covered serotype 19A was identified in 2018. The only one PCV13 serotype 1 case was detected in 2020. The number of serotype 3 cases detected in PFS after 2020 was significantly higher (Fisher exact test $p < 0.01$) than the rest of the serotypes. Among the twenty-one children infected by serotype 3, one had received one dose of PCV13, another one two doses, fifteen children received three doses, one child four doses, and in three this data was unknown. The two cases caused by serotype 19A received two PCV13 doses and the case due to serotype 1 received four.

Although the number of cases included is small and this is not a systematic epidemiological study, the elevate frequency of detected SPNGST 3 in PFS is worrying. Serotype 3 is a well-identified cause of pleural PPE/PE in children and these cases are increasing in the PCV13 era.^{2–4} We do not have a definitive explanation for the high number of cases of serotype 3 in 2022. The synergy between Influenza virus and *S. pneumoniae*⁵ would to have influenced in this fact. During 2022, the number of influenza cases in the region of Madrid was higher than in the previous season and affected children to a great extent.⁶

Despite the fact that the use of PCV13 in children has led to a substantial reduction of IPD in Spanish children⁷ and pneumococcal PPE/PE (especially associated to the drop of serotype 1),⁸ the immunogenicity and the effectiveness against each of the 13 serotypes of the vaccine seems to be non-homogeneous.^{3,4} The immunogenic activity has been described lower for serotype 3 than for other PCV13 serotypes,⁹ and the individual vaccine protection against this serotype is also lower.¹⁰ Moreover, the sensitivity of culture seems to be particularly very low for this serotype. Thus, serotype 3 prevalence data in pleural infection may be underestimated when surveillance is only based in culture and conventional phenotypic serotyping techniques. The routine implementation of PCR procedures in order to identify the serotype involved in culture-negative specimens provides additional information and can improve substantially the microbiological investigation of *Streptococcus pneumoniae* pleural infections. This approach can support epidemiological surveillance and monitoring of immunization plans.⁴

Acknowledgements

To the Microbiological Laboratory Services of the Hospital Universitario 12 de Octubre, Hospital Clínico San Carlos, Hospital Universitario de Getafe, Hospital General Universitario Gregorio Marañón and Hospital Infantil Universitario Niño Jesús from Madrid for sending the pleural fluid samples to Laboratorio Regional de Salud Pública de la Comunidad de Madrid during the study period.

Bibliografía

- Latasa Zamalloa P, Sanz Moreno JC, Ordobás Gavín M, Barranco Ordoñez MD, Insúa Marquerena E, Gil de Miguel Á, et al. Trends of invasive pneumococcal disease and its serotypes in the Autonomous Community of Madrid. Enferm Infect Microbiol Clin. 2018;36:612–20, <http://dx.doi.org/10.1016/j.eimc.2017.10.026>.
- Goettler D, Streng A, Kemmling D, Schoen C, von Kries R, Rose MA, et al. Increase in *Streptococcus pneumoniae* serotype 3 associated parapneumonic pleural effusion/empyema after the introduction of PCV13 in Germany. Vaccine. 2020;38:570–7, <http://dx.doi.org/10.1016/j.vaccine.2019.10.056>.
- Silva-Costa C, Gomes-Silva J, Pinho MD, Friões A, Ramirez M, Melo-Cristino J. Continued vaccine breakthrough cases of serotype 3 complicated pneumonia in vaccinated children, Portugal (2016–2019). Microbiol Spectr. 2022;10:e0107722, <http://dx.doi.org/10.1128/spectrum.01077-22>.
- Martínez-Osorio J, García-García JJ, Moraga-Llop F, Díaz A, Hernández S, Solé-Ribalta A, et al. Enfermedad neumocócica invasiva en niños menores de 60 meses, antes y después de la introducción de la vacuna conjugada 13-valente. An Pediatr. 2022;96:501–10, <http://dx.doi.org/10.1016/j.anpede.2021.06.005>.
- McCullers JA. Insights into the interaction between influenza virus and pneumococcus. Clin Microbiol Rev. 2006;19:571–82, <http://dx.doi.org/10.1128/CMR.00058-05>.
- Subdirección General de Vigilancia en Salud Pública. Dirección General de Salud Pública. Consejería de Sanidad. Comunidad de Madrid. Informe epidemiológico Semanal Comunidad de Madrid. Semana 50. Red de Vigilancia Epidemiológica. Martes, 20 de diciembre de 2022 (datos provisionales). Available from: <https://www.comunidad.madrid/sites/default/files/doc/sanidad/epid/informe.epidemiologico.semanal.pdf> [accessed 9.2.23].
- De Miguel S, Domenech M, González-Camacho F, Sempere J, Vicioso D, Sanz JC, et al. Nationwide trends of invasive pneumococcal disease in Spain from 2009 through 2019 in children and adults during the pneumococcal conjugate vaccine era. Clin Infect Dis. 2021;73:e3778–87, <http://dx.doi.org/10.1093/cid/cia1483>.
- Díaz-Conradi A, Hernández S, García-García JJ, Muñoz-Almagro C, Moraga-Llop F, Ciruela P, et al. Complicated pneumococcal pneumonia with pleural effusion or empyema in the 13-valent pneumococcal conjugate vaccine era. Pediatr Pulmonol. 2019;54:517–24, <http://dx.doi.org/10.1002/ppul.24279>.
- Stacey HL, Rosen J, Peterson JT, Williams-Diaz A, Gakhar V, Sterling TM, et al. Safety and immunogenicity of 15-valent pneumococcal conjugate vaccine (PCV-15) compared to PCV-13 in healthy older adults. Hum Vaccin Immunother. 2019;15:530.
- Savulescu C, Krizova P, Valentiner-Branth P, Ladhani S, Rinta-Kokko H, Levy C, et al. Effectiveness of 10 and 13-valent pneumococcal conjugate vaccines against invasive pneumococcal disease in European children: SplDnet observational multicentre study. Vaccine. 2022;40:3963–74, <http://dx.doi.org/10.1016/j.vaccine.2022.05.011>.

Marta Pérez-Abeledo^a, Gloria Zaragoza^b, Belén Ramos^a, Juan Carlos Sanz^{a,c,*}

^a Unidad de Microbiología Clínica, Laboratorio Regional de Salud Pública de la Comunidad de Madrid, Dirección General de Salud Pública de la Comunidad de Madrid, Spain

^b Servicio de Microbiología y Parasitología del Hospital Universitario de Getafe, Spain

^c CIBER de Epidemiología y Salud Pública (CIBERESP), Madrid, Spain

* Corresponding author.

E-mail address: juan.sanz@salud.madrid.org (J.C. Sanz).

<https://doi.org/10.1016/j.eimc.2023.02.001>

0213-005X/ © 2023 Sociedad Española de Enfermedades Infecciosas y Microbiología Clínica. Published by Elsevier España, S.L.U. All rights reserved.