

Bibliografía

1. Kabeerdoss J, Pilania RK, Karkhele R, Kumar TS, Danda D, Singh S. Severe COVID-19, multisystem inflammatory syndrome in children, and Kawasaki disease: Immunological mechanisms, clinical manifestations and management [Internet]. *Rheumatol Int*. 2021;41:19–32 [consultado 2021 Abr 17]. Disponible en: <https://pubmed.ncbi.nlm.nih.gov/33219837/>.
2. Bastug A, Aslaner H, Aybar Bilir Y, Kemirtlek N, Gursoy FM, Bastug S, et al. Multiple system inflammatory syndrome associated with SARS-CoV-2 infection in an adult and an adolescent. *Rheumatol Int* [Internet]. 2021;41(5.) [consultado 17 Abr 2021]. Disponible en: <https://pubmed.ncbi.nlm.nih.gov/33742229/>.
3. Vogel TP, Top KA, Karatzios C, Hilmers DC, Tapia LI, Moceri P, et al. Multisystem inflammatory syndrome in children and adults (MIS-C/A): Case definition & guidelines for data collection, analysis, and presentation of immunization safety data. *Vaccine*. 2021.
4. La Rosée P, Horne AC, Hines M, Greenwood TVB, Machowicz R, Berliner N, et al. Recommendations for the management of hemophagocytic lymphohistiocytosis in adults. *Blood* [Internet]. 2019;133:2465–77 [consultado 17 Abr 2021]. Disponible en: <http://ashpublications.org/blood/article-pdf/133/23/2465/1553600/blood894618.pdf>.
5. Morris SB, Schwartz NG, Patel P, Abbo L, Beauchamps L, Balan S, et al. Case series of multisystem inflammatory syndrome in adults associated with SARS-CoV-2 infection – United Kingdom and United States March–August 2020. *MMWR Morb Mortal Wkly Rep* [Internet]. 2020;69:1450–6 [consultado 17 Abr 2021]. Disponible en: <https://pubmed.ncbi.nlm.nih.gov/33031361/>.
6. Davogustto GE, Clark DE, Hardison E, Yanis AH, Lowery BD, Halasa NB, et al. Characteristics associated with multisystem inflammatory syndrome among adults with SARS-CoV-2 infection [Internet]. *JAMA Network Open*. 2021;4 [consultado 10 Jul 2021]. Disponible en: <https://pubmed.ncbi.nlm.nih.gov/34009351/>.
7. Salzman MB, Huang C-W, O'Brien CM, Castillo RD. Multisystem inflammatory syndrome after SARS-CoV-2 infection and COVID-19 vaccination. *Emerg Infect Dis* [Internet]. 2021;27:1944–8 [consultado 10 Jul 2021]. Disponible en: <https://pubmed.ncbi.nlm.nih.gov/34034858/>.
8. García Salido A, Antón J, Marínez-Pajares JD, Giralt García G, Gómez-Cortés B, Tagarro A, Grupo de trabajo de la Asociación Española de Pediatría para el síndrome inflamatorio multisistémico pediátrico vinculado a SARS-CoV-2. Consenso nacional sobre diagnóstico, estabilización y tratamiento del síndrome inflamatorio multisistémico pediátrico vinculado a SARS-CoV-2. *Enfermedades Infecciosas y Microbiología Clínica*. 2021;40:399–410.

Rapid diagnosis of pulmonary tuberculosis using Xpert MTB/RIF assay in gastric aspirate samples from adult patients with sputum-absent disease: A first-step alternative to bronchoscopy?

Diagnóstico precoz de la tuberculosis pulmonar mediante Xpert MTB/RIF en muestras obtenidas mediante aspirado gástrico en pacientes adultos que no expectoran: ¿una alternativa previa a la broncoscopia?

Dear Editor,

Sputum-absent pulmonary tuberculosis (PTB) is common in adult patients. It leads to misdiagnoses and delays of PTB and forces to rely on alternative diagnostic approaches such as bronchoscopy (BC). XpertMTB/RIF on gastric aspirate (GA) sampling is an option in this scenario,¹ mainly in the pediatric population,² but evidence of its usefulness on the adult population is scarce and no previous work has studied its diagnostic performance compared with a reference standard such as tuberculosis culture (TC) from BC samples. TC from GA samples has been previously compared with TC from BC samples showing a positive culture yield of 21% vs. 34% respectively.³

In the present study we compared the diagnostic yield of the Xpert MTB/RIF assay in GA samples with regard to TC obtained through BC in adult patients with suspected PTB and no sputum production. The secondary aim was to compare the diagnostic performance of the Xpert MTB/RIF assay versus that of the TC in the same GA sample. Overall, confirmed PTB diagnosis was made if either Xpert MTB/RIF assay or TC were positive.



inflamatorio multisistémico pediátrico vinculado a SARS-CoV-2. (SIM-PedS). *Anales Pediatr*. 2021;94, 116.e1-11-.

9. Son MB, Murray N, Friedman K, Young CC, Newhams MM, Feldstein LR, et al. Multisystem inflammatory syndrome in children: Initial therapy and outcomes. *N Engl J Med* [Internet]. 2021;385:23–34 [consultado 10 Jul 2021]. Disponible en: <https://www.nejm.org/doi/full/10.1056/NEJMoa2102605>.
10. McArdle AJ, Vito O, Patel H, Seaby EG, Shah P, Wilson C, et al. Treatment of multisystem inflammatory syndrome in children. *N Engl J Med* [Internet]. 2021;385:11–22 [consultado 10 Jul 2021]. Disponible en: <https://pubmed.ncbi.nlm.nih.gov/34133854/>.

Jara Llenas-García ^{a,b,*}, Mari Luz Paredes-Martínez ^c, Pedro Luis Boils-Arroyo ^d e Isabel María Pérez-Gómez ^e

^a Servicio de Medicina Interna-Infecciosas, Hospital Vega Baja, FISABIO, Orihuela (Alicante), España

^b Departamento de Medicina Clínica, Universidad Miguel Hernández, Elche (Alicante), España

^c Servicio de Radiodiagnóstico, Hospital Vega Baja, FISABIO, Orihuela (Alicante), España

^d Servicio de Anatomía Patológica, Hospital Vega Baja, FISABIO, Orihuela (Alicante), España

^e Unidad de Medicina Intensiva, Hospital Vega Baja, FISABIO, Orihuela (Alicante), España

* Autor para correspondencia.

Correo electrónico: jarallenas@gmail.com (J. Llenas-García).

<https://doi.org/10.1016/j.eimc.2021.10.009>

0213-005X/ © 2021 Sociedad Española de Enfermedades Infecciosas y Microbiología Clínica. Publicado por Elsevier España, S.L.U. Todos los derechos reservados.

We retrospectively reviewed all the GA samples obtained between January 2015 and May 2018 from adult patients with clinical or radiological suspicion of PTB, no sputum production and which fulfilled the inclusion criteria: (a) GA samples were processed for TC and Xpert MTB/RIF; (b) when the GA Xpert MTB/RIF assay was negative a TC was obtained through BC. GA samples were obtained through nasogastric tube in the morning after overnight fasting, and then processed and decontaminated within less than 2 h from sampling.⁴ Since no bronchoscopic sampling was performed in patients with a positive Xpert MTB/RIF assay in GA, it was assumed that all GA-positive patients would have been diagnosed with TC obtained through BC.

Forty-three GA samples from 43 patients were reviewed and 31 were finally included in the analysis. The Xpert MTB/RIF was positive in 9 patients (29.0% [9/31]). Eight of them had a positive TC in the same GA sample, whereas the GA culture of the remaining patient was contaminated. Among the 22 patients with negative Xpert MTB/RIF assay in GA (70.9% [22/31]), two (9.1% [2/22]) had a positive TC only in the BC sample and one additional patient (4.5% [1/22]) had a positive TC only in the GA sample. Overall, the diagnosis of PTB was confirmed on the basis of TC performed in GA and/or BC samples in 12 patients (38.7% [12/31] of the overall study cohort).

The sensitivity and specificity of the Xpert MTB/RIF assay in GA samples by using TC in BC samples as reference method were 81.9% (95% CI: 48.2–97.7) and 100.0% (95% CI: 83.2–100), respectively. The NPV was estimated at 90.9% (95% CI: 74.1–97.2) (Table 1). On the other hand, the Xpert MTB/RIF assay exhibited a sensitivity of 88.9% (95% CI: 51.8–99.7) and a specificity of 95.5%

Table 1

Diagnostic performance of the Xpert MTB/RIF assay performed in GA samples.

| Comparison of diagnostic methods | Sensitivity, % (95% CI) | Specificity, % (95% CI) | PPV, % (95% CI) | NPV, % (95% CI) | Positive likelihood ratio (95% CI) | Negative likelihood ratio (95% CI) |
|--|-------------------------|-------------------------|------------------|--------------------|------------------------------------|------------------------------------|
| Xpert MTB/RIF in GA (vs. positive mycobacterial culture in BC sample) ^a | 81.9 (48.2–97.7) | 100 (83.2–100) | 100.0 (NA) | 90.9 (74.1–97.2) | NA | 0.18 (0.05–0.64) |
| Xpert MTB/RIF in GA (vs. positive mycobacterial culture in GA) | 88.9 (51.8–99.7) | 95.5 (77.2–99.9) | 88.9 (53.8–98.2) | 95.5 (76.8.0–99.3) | 19.56 (2.84–134.6) | 0.12 (0.02–0.74) |

BC: bronchoscopy; CI: confidence interval; GA: gastric aspirate; NA: not applicable; NPV: negative predictive value; PPV: positive predictive value.

^a It was assumed that all the patients diagnosed on the basis of a positive Xpert MTB/RIF assay in the GA sample would have been also diagnosed if a bronchoscopic sample had been processed.

(95% CI: 77.2–99.9) when compared with TC in the same GA sample.

The diagnostic performance showed of the Xpert MTB/RIF assay in GA consistent with previous studies when it is compared with TC performed on the same sample,^{1,5} although this evidence is mainly limited to the pediatric population. Interestingly, we observed that one out of 12 patients (8.3%) with a culture-proven diagnosis of PTB had a positive TC in GA but not in the BC sample, a complimentary diagnostic yield that has been shown in other studies.^{3,6} Altogether, our study shows that Xpert MTB/RIF and TC from GA sampling can avoid up to 32% (10/31) of the BC in this scenario.

Our study has some limitations. Firstly, it lacks of simultaneous BC sampling in patients with a positive Xpert MTB/RIF assay in GA. This invasive procedure was spared by the attending clinicians due to the high specificity of the Xpert MTB/RIF assay in GA samples when there is a suspicion of PTB. We addressed this issue by assuming that all these patients would have been diagnosed by TC in BC samples. Secondly, we used an assay that has been replaced by the more sensitive Xpert MTB/RIF ultra assay.⁷ Thus, it is likely that the actual sensitivity of the molecular diagnosis of PTB in GA samples was underestimated.

Our study supports the use of the Xpert MTB/RIF assay as a first line diagnostic approach in adult patients with PTB suspicion and who are unable to expectorate.

Conflicts of interest and source of funding

Mario Fernández-Ruiz holds a research contract "Miguel Servet" (CP 18/00073) from the Spanish Ministry of Science and Innovation, Instituto de Salud Carlos III.

The rest of the authors have no conflicts of interest nor funding sources to declare.

Bibliografía

- Maynard-Smith L, Larke N, Peters JA, Lawn SD. Diagnostic accuracy of the Xpert MTB/RIF assay for extrapulmonary and pulmonary tuberculosis when testing non-respiratory samples: a systematic review. *BMC Infect Dis*. 2014;14:709.

- Lewinsohn DM, Leonard MK, LoBue PA, Cohn DL, Daley CL, Desmond E, et al. Official American Thoracic Society/Infectious Diseases Society of America/Centers for Disease Control and Prevention Clinical Practice Guidelines: diagnosis of tuberculosis in adults and children. *Clin Infect Dis*. 2017;64:e1–33.
- Dickson SJ, Brent A, Davidson RN, Wall R. Comparison of bronchoscopy and gastric washings in the investigation of smear-negative pulmonary tuberculosis. *Clin Infect Dis*. 2003;37:1649–53.
- Fradejas I, Ontañón B, Muñoz-Gallego I, Ramírez-Vela MJ, López-Roa P. The value of xpert MTB/RIF-generated CT values for predicting the smear status of patients with pulmonary tuberculosis. *J Clin Tuberc Mycobact Dis*. 2018;13:9–12.
- Tan HK, Fan SJ, Xu YC, Zhou JJ, Chen YZ, Xie TA, et al. The clinical diagnostic value of Xpert MTB/RIF for the detection of *Mycobacterium tuberculosis* in gastric aspirates. *Biosci Rep*. 2020;40. BSR 20200138.
- Uskul BT, Turker H, Kant A, Partal M. Comparison of bronchoscopic washing and gastric lavage in the diagnosis of smear-negative pulmonary tuberculosis. *South Med J*. 2009;102:154–8.
- Kohli M, Schiller I, Dendukuri N, Yao M, Dheda K, Denkinger CM, et al. Xpert MTB/RIF Ultra and Xpert MTB/RIF assays for extrapulmonary tuberculosis and rifampicin resistance in adults. *Cochrane Database Syst Rev*. 2021;1:CD012768.

Guillermo Maestro-de la Calle^{a,*}, Mario Fernández-Ruiz^b, Paula López-Roa^c, José María Aguado^b

^a Department of Internal Medicine – Antibiotic Stewardship Program, Hospital Universitario “12 de Octubre”, Instituto de Investigación Hospital “12 de Octubre” (imas12), “Universidad Complutense de Madrid” School of Medicine, Madrid, Spain

^b Unit of Infectious Diseases, Hospital Universitario “12 de Octubre”, Instituto de Investigación Hospital “12 de Octubre” (imas12), “Universidad Complutense de Madrid” School of Medicine, Madrid, Spain

^c Department of Microbiology, Hospital Universitario “12 de Octubre”, Instituto de Investigación Hospital “12 de Octubre” (imas12), “Universidad Complutense de Madrid” School of Medicine, Madrid, Spain

* Corresponding author.

E-mail address: guillermo.maestro@salud.madrid.org (G. Maestro-de la Calle).

<https://doi.org/10.1016/j.eimc.2021.12.012>

0213-005X/ © 2022 Sociedad Española de Enfermedades Infecciosas y Microbiología Clínica. Published by Elsevier España, S.L.U. All rights reserved.