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Molecular (ticks) and serological (humans) study of Crimean-Congo hemorrhagic fever virus in the Iberian Peninsula, 2013–2015



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ABSTRACT

Introduction: Crimean-Congo hemorrhagic fever (CCHF) is a viral disease, mainly transmitted through tick bite, of great importance in Public Health. In Spain, Crimean-Congo hemorrhagic fever virus (CCHFV) was detected for the first time in 2010 in *Hyalomma lusitanicum* ticks collected from deer in Cáceres. The aim of this study was to investigate the presence of CCHFV in ticks from Cáceres, and from other Spanish areas, and to evaluate the presence of antibodies against the virus in individuals exposed to tick bites.

Methods: A total of 2053 ticks (1333 *Hyalomma marginatum*, 680 *H. lusitanicum* and 40 *Rhipicephalus bursa*) were analyzed using molecular biology techniques (PCR) for CCHFV detection. The determination of specific IgG antibodies against CCHFV in 228 serum samples from humans with regular contact with ticks (at risk of acquiring the infection) was performed by indirect immunofluorescence assay.

Results: The CCHFV was not amplified in ticks, nor were antibodies against the virus found in the serum samples analyzed.

Conclusion: The absence of the CCHFV in the ticks studied and the lack of antibodies against the virus in individuals exposed to tick bites would seem to suggest a low risk of acquisition of human infection by CCHFV in Spain.

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Estudio molecular (garrapatas) y serológico (humanos) del virus de la fiebre hemorrágica de Crimea-Congo en la Península Ibérica, 2013–2015

RESUMEN

Introducción: La fiebre hemorrágica de Crimea-Congo (FHCC) es una enfermedad vírica, transmitida fundamentalmente por garrapatas, con gran importancia en salud pública. La primera detección del virus de la fiebre hemorrágica de Crimea-Congo (vFHCC) en España se realizó en 2010 en garrapatas de la especie *Hyalomma lusitanicum* recogidas sobre ciervos en Cáceres. El objetivo de este estudio fue investigar la presencia del vFHCC en garrapatas de esta y otras zonas de España y evaluar la presencia de anticuerpos frente al virus en personas expuestas a picaduras de garrapatas.

Métodos: Se analizaron un total de 2.053 garrapatas (1.333 *Hyalomma marginatum*, 680 *Hyalomma lusitanicum* y 40 *Rhipicephalus bursa*) mediante técnicas de biología molecular (PCR) para la detección del

Palabras clave:

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vFHCC. Se realizó la determinación de anticuerpos IgG específicos frente al vFHCC mediante inmunofluorescencia indirecta a 228 sueros de personas en contacto habitual con garrapatas, y por tanto, en riesgo de adquirir la infección.

Resultados: No se amplificó el vFHCC en garrapatas ni se encontraron anticuerpos frente al mismo en los sueros analizados.

Conclusión: La ausencia del vFHCC en las garrapatas estudiadas y de anticuerpos frente al mismo en personas expuestas a picaduras indica un riesgo bajo de adquisición de la infección por el vFHCC en humanos en España.

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Introduction

Crimean-Congo hemorrhagic fever virus (CCHFV) is responsible for a human severe hemorrhagic disease, Crimean-Congo hemorrhagic fever (CCHF), transmitted via tick bite or by contact with infected body fluids.^{1,2} After an incubation period (2–7 days), the disease courses with non-specific symptoms (headache, high fever, myalgia, stomach pain, diarrhea and vomiting) in a period known as prehemorrhagic. This phase is followed by a hemorrhagic period characterized by ecchymosis and petechiae of skin and mucous membranes, epistaxis, gastrointestinal bleeding (haematemesis, melenas, etc.) and severe thrombocytopenia, leukopenia and the elevation of liver enzymes.^{1–3} The documented fatality rates can reach more than 50%.⁴ CCHF has a broad world distribution including Africa, Asia and Europe, and it is emerging and reemerging in several countries.^{1–5} The disease only occurs in humans, but the zoonotic life cycle of the virus includes ticks as vectors and reservoirs (mainly within *Hyalomma* genus), and animals as amplifiers of the infectious agent or dispersers of the vectors.^{1,6}

The occurrence of the CCHFV was not known in Southern Europe before 2010 when the virus was amplified in *Hyalomma lusitanicum* ticks captured from deer in Spain (Cáceres, 39.63° N; 7.33° W) by our team.⁷ The genetic sequence of this virus showed high similarity with strains from Senegal and Mauritania, and it was different from European or Asian strains. Therefore, the potential arrival of infected ticks transported by migratory birds from Africa was hypothesized,⁷ and our hypothesis was supported by the detection of a similar strain of CCHFV in Morocco, in *Hyalomma marginatum* ticks removed from migratory birds able to reach the Iberian Peninsula.⁶

The detection of CCHFV in ticks from Spanish deer and from Moroccan migratory birds, in addition to the presence of *H. marginatum* established populations (an anthropophilic tick species) and humans susceptible to develop FHCC infections, made necessary to establish a surveillance program of the CCHF in Spain. The present study, supported by the Instituto de Salud Carlos III (Fondo de Investigación Sanitaria, Ministerio de Ciencia e Innovación, Spain; PI12/02579), aimed to investigate the presence of CCHFV in ticks and to screen human antibodies against the virus in Spain.

Material and methods

Tick collection

From 2013 to 2015, ticks were collected from vegetation using dragging methods and from hosts (cattle and sheep) by direct capture. These techniques aimed to obtain the maximum number of *Hyalomma* spp. specimens, the known vector of the CCHFV. Therefore, samplings were carried out in the optimal season and areas, always taking into account the distribution and the biology of this genus.⁸ Some *Rhipicephalus bursa* specimens captured close to the

area where the virus had been previously detected⁷ were also added to the tick collection. A total of 22 Spanish provinces were screened for ticks with the invaluable help of multidisciplinary colleagues (Heads of Public Health Organizations, veterinarians, biologists, forest workers and ranchers).

The morphological classification of ticks was performed using conventional keys and reprints.^{8,9} This classification was genetically confirmed in doubtful specimens using a PCR targeting a fragment gene of the mitochondrial 16S rRNA,¹⁰ as it has been described in previous studies.¹¹ Ticks were kept frozen at –80 °C.

Sera collection

During 2014 sera samples from volunteers in risk for tick bites or animal contact were collected in Cáceres (7–40 km far away from the area where there was evidence of the virus) through the Health Centers from Santiago de Alcántara (31.61° N; 7.2° W), Cedillo (39.65° N; 7.5° W), Carbajo (39.60° N; 7.20° W), Membrío (39.53° N; 7.06° W), Salorino (39.48° N; 7.01° W), and Herrerueta (39.46° N; 6.9° W). Sera samples from humans routinely exposed to tick-bites (foresters and forest rangers) in La Rioja (Spain), and from patients with microbiological confirmation of tick-borne disease that attended the Infectious Diseases Department from Hospital San Pedro (which attends all 300,000 inhabitants of La Rioja region) from 2010 to 2014 were also included. All participants were informed of the purpose of the study and a consent was obtained from all of them. This study was performed with the ethical approval of the Committee for Ethics in Clinical Research in La Rioja (Ref. CEICLAR 109 Bis).

CCHFV detection

Ticks were washed in 70% ethanol and rinsed twice with sterilized MQ water. A half of each tick was homogenized using the Bullet Blender™ system and 6 mm glass balls (maximum speed during 2 min). The RNA was extracted from the homogenates using the RNeasy Mini Kit (Qiagen, Hilden, Germany), following the manufacturer's instructions. A negative control without tick was extracted in each batch of extractions. The RNA was retrotranscribed using the Omniscript RT kit (Qiagen, Hilden, Germany). The RNA and cDNA were frozen at –80 °C and –20 °C, respectively, until their use. The cDNAs were pooled (from 6 to 11 samples per pool, except for 3 ticks from sheep that were processed in one pool) by tick species, gender, source of capture (vegetation or type of host) and area.

The pooled cDNAs were used as templates for PCR assays targeting the 16S mitochondrial RNA¹⁰ as internal control to validate the extraction and retrotranscription of the RNA. The CCHFV detection was performed using 4 conventional PCRs (3 nested and 1 single reaction) and a real time assay (qPCR) that amplify fragment genes of the S segment.^{12–15} A positive control of the virus (AP92 strain)

Table 1
Tick samples analyzed in this study.

Spanish province	<i>Hyalomma lusitanicum</i>		<i>Hyalomma marginatum</i>			<i>Rhipicephalus bursa</i> ^a
	Vegetation	Cattle	Vegetation	Cattle	Sheep	Vegetation
Albacete		36				
Ávila		7		14		
Badajoz		3		12		
Burgos				148		
Cáceres	41			44		34
Castellón				33	3	
Ciudad Real	61			5		
Córdoba	257	8		2		4
Jaén	232					2
La Rioja			10	788		
Navarra			1			
Palencia				11		
Salamanca		34		52		
Segovia			2			
Soria				22		
Teruel				98		
Valladolid				5		
Zamora	1			83		
Total	592	88	13	1317	3	40

^a The detection of the CCHFV in this tick species has been reported, although its role as vector has not been demonstrated.¹

as well as negative controls of extraction and negative controls of PCR were included.

CCHF antibodies detection

The determination of IgG antibodies against CCHFV was carried out by indirect immunofluorescence assay with the commercial kit “Crimean-Congo Fever Virus Mosaic 2” (Euroimmun AG, Luebeck, Germany), following the manufacturer’s instructions.

Results

A total of 2053 adult ticks from 18 Spanish provinces were screened for the CCHFV detection; 1333 *H. marginatum*, 680 *H. lusitanicum* and 40 *R. bursa* specimens (Table 1 and Fig. 1). The cDNAs from tick specimens were grouped in 74 pools from *H. lusitanicum*, 151 from *H. marginatum*, and 4 from *R. bursa*. No samples yielded positive results for the PCRs used for the CCHFV detection.

One hundred and fourteen human sera specimens collected in Cáceres (54 from Santiago de Alcántara, 16 from Cedillo, 15 from Carbajo, 12 from Membrío, 10 from Salorino and 7 from Herrerueta)

were analyzed. From them, 97% volunteers had usual contact with animals, 61% were aware of having suffered a tick-bite, and 5% were conscious of having suffered a tick-borne disease. The same number of human sera ($n = 114$) from La Rioja region were also processed: 42 from foresters and forest rangers and 72 from patients with a confirmed tick-borne disease. A total of 70% lived and/or worked in rural areas and they had regular contact with animals, and 67% recalled tick-bites. IgG antibodies against CCHFV were not found in any of the sera samples analyzed.

Discussion

Previous studies carried out in the Center of Rickettsiosis and Arthropod-borne Diseases in La Rioja demonstrated the circulation of the CCHFV in *H. lusitanicum* ticks from Cáceres province and the possible arrival of this virus via migratory birds to Spain.^{6,7} These facts, in addition to the presence of *H. marginatum* populations that bite people, made us to be aware of the possible occurrence of autochthonous CCHF human cases in Spain. In fact, when we were writing this paper, the first CCHF case has been diagnosed in our country.^{16–18} Moreover, a second case of a nurse who treated

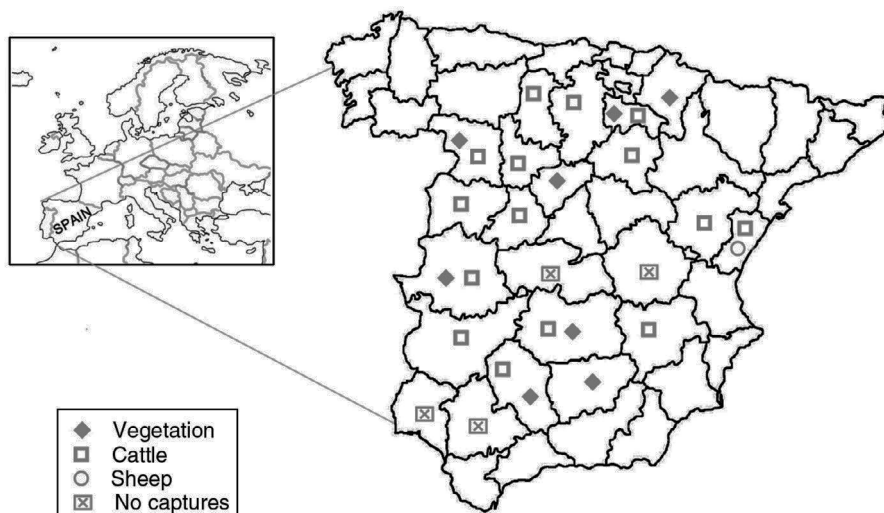


Fig. 1. Sampling areas.

the first patient has been reported.^{16–18} In the present work, the absence of CCHFV in the ticks analyzed and the lack of antibodies against CCHFV in human populations with high risk of exposition to tick-bites, suggest that, to date, the virus is not widespread and the risk of acquiring the infection and the disease in Spain is low.

Our negative results in ticks suggest a low prevalence of the FHCCV in the vectors, supported also by previous studies carried out in ticks removed from patients, vegetation and hosts (cattle and birds) in the North of Spain.^{11,19} Moreover, the virus has been repeatedly detected in the same tick species (*H. lusitanicum*) and from the same area where it was found for the first time (Cáceres).¹⁶ The reported rates of the virus infection in ticks from endemic European areas are highly variable (0 to $\geq 50\%$).^{20,21} These differences support that the screening of CCHFV in ticks should continue throughout Spain.

Despite the recent autochthonous CCHF case in Spain, our negative seroepidemiological results were not unexpected since the circulation of the virus in this area seems very recent and not widely distributed. Moreover, the reported seroprevalences of IgG antibodies against CCHFV in populations from the endemic areas showed significant differences, even in different regions of the same country.^{20,22} The occurrence of the virus in Spain and the recent appearance of the first human case in our country make necessary to continue monitoring the presence of antibodies against CCHFV in more sera samples and areas of the Iberian Peninsula. In addition, a seroepidemiological survey of CCHF in domestic and wild animals is also recommended.

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Conflict of interest

The authors declare that they have no conflict of interest.

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References

- Ergonul O. Crimean-Congo haemorrhagic fever. *Lancet Infect Dis.* 2006;20066:203–14.
- Bente DA, Forrester NL, Watts DM, McAuley AJ, Whitehouse CA, Bray M. Crimean-Congo hemorrhagic fever: history, epidemiology, pathogenesis, clinical syndrome and genetic diversity. *Antiviral Res.* 2013;100:159–89.
- Oteo JA. Conocimientos básicos sobre la fiebre hemorrágica de Crimea-Congo. Recomendaciones Sociedad Española de Enfermedades Infecciosas y Microbiología Clínica (SEIMC); 2016. Available from: http://www.seimc.org/contenidos/documentoscientificos/recomendaciones/seimc-rc-2016-conocimientos_basicos_fiebre_hemorragica_Crimea-Congo.pdf [accessed November 2016].
- Centers for Disease Control and Prevention (CDC). Crimean Congo hemorrhagic fever; 2013. Available from: <https://www.cdc.gov/vhf/crimean-congo/> [last updated 09.05.14, accessed November 2016].
- Messina JP, Pigott DM, Golding N, Duda KA, Brownstein JS, Weiss DJ, et al. The global distribution of Crimean-Congo hemorrhagic fever. *Trans R Soc Trop Med Hyg.* 2015;109:503–13.
- Palomar AM, Portillo A, Santibáñez P, Mazuelas D, Arizaga J, Crespo A, et al. Crimean-Congo hemorrhagic fever virus in ticks from migratory birds, Morocco. *Emerg Infect Dis.* 2013;19:260–3.
- Estrada-Peña A, Palomar AM, Santibáñez P, Sánchez N, Habela MA, Portillo A, et al. Crimean-Congo hemorrhagic fever virus in ticks, southwestern Europe, 2010. *Emerg Infect Dis.* 2012;18:179–80.
- Estrada-Peña A, Bouattour A, Camicas JL, Walker AR. Ticks of domestic animals in the Mediterranean region. A guide of identification of species. 1st ed. Zaragoza, Spain: University of Zaragoza Press; 2004.
- Apanaskevich DA, Santos-Silva MM, Horak IG. The genus *Hyalomma* Koch, 1844. IV. Redescription of all parasitic stages of *H. (Euhyalomma) lusitanicum* Koch, 1844 and the adults of *H. (E.) franchinii* Tonelli Rondelli, 1932 (Acari: Ixodidae) with a first description of its immature stages. *Folia Parasitol (Praha).* 2008;55:61–74.
- Black WC, Piesman J. Phylogeny of hard- and soft-tick taxa (Acari: Ixodida) based on mitochondrial 16S rDNA sequences. *Proc Natl Acad Sci U S A.* 1994;91:10034–8.
- Palomar AM, Portillo A, Mazuelas D, Roncero L, Arizaga J, Crespo A, et al. Molecular analysis of Crimean-Congo hemorrhagic fever virus and Rickettsia in *Hyalomma marginatum* ticks removed from patients (Spain) and birds (Spain and Morocco), 2009–2015. *Ticks Tick Borne Dis.* 2016;7:983–7.
- Burt FJ, Swanepoel R. Molecular epidemiology of African and Asian Crimean-Congo haemorrhagic fever isolates. *Epidemiol Infect.* 2005;133:659–66.
- Midilli K, Gargili A, Ergonul O, Sengöz G, Ozturk R, Bakar M, et al. Imported Crimean-Congo hemorrhagic fever cases in Istanbul. *BMC Infect Dis.* 2007;7:54.
- Midilli K, Gargili A, Ergonul O, Eleveli M, Ergin S, Turan N, et al. The first clinical case due to AP92 like strain of Crimean-Congo hemorrhagic fever virus and a field survey. *BMC Infect Dis.* 2009;9:90.
- Atkinson B, Chamberlain J, Logue CH, Cook N, Bruce C, Dowall SD, et al. Development of a real-time RT-PCR assay for the detection of Crimean-Congo hemorrhagic fever virus. *Vector Borne Zoonotic Dis.* 2012;12:786–93.
- Ministerio de Sanidad, Servicios Sociales e Igualdad. Informe de situación y evaluación del riesgo de transmisión de Fiebre Hemorrágica de Crimea-Congo (FHCC) en España, septiembre 2016; 2016. Available from: http://www.mssi.gob.es/profesionales/saludPublica/ccayes/alertasActual/Crimea_Congo/docs/Informe_situacion_evaluacion_CrimeaCongo_sep2016.pdf [accessed November 2016].
- European Centre for Disease Prevention and Control. Crimean-Congo haemorrhagic fever in Spain. Stockholm: ECDC; 2016. Available from: <http://ecdc.europa.eu/en/publications/Publications/crimean-congo-haemorrhagic-fever-spain-risk-assessment.pdf> [accessed November 2016].
- García Rada A. First outbreak of Crimean-Congo haemorrhagic fever in Western Europe kills one man in Spain. *BMJ.* 2016;354:i4891.
- Palomar AM, Portillo A, Eiros JM, Oteo JA. The risk of introducing tick-borne encephalitis and Crimean-Congo hemorrhagic fever into southwestern Europe (Iberian Peninsula). In: *Virology II – advanced issues 2013*. ISBN: 978-1-922227-45-4. Available from: <https://www.icconceptpress.com/book/virology-ii-advanced-issues/11000086/1206000594/> [accessed November 2016].
- Fajš L, Humolli I, Saksida A, Knap N, Jelovšek M, Korva M, et al. Prevalence of Crimean-Congo hemorrhagic fever virus in healthy population, livestock and ticks in Kosovo. *PLOS ONE.* 2014;9:e110982.
- Panayotova E, Papa A, Trifonova I, Christova I. Crimean-Congo hemorrhagic fever virus lineages Europe 1 and Europe 2 in Bulgarian ticks. *Ticks Tick Borne Dis.* 2016;7:1024–8.
- Sidira P, Maltezou HC, Haidich AB, Papa A. Seroepidemiological study of Crimean-Congo haemorrhagic fever in Greece, 2009–2010. *Clin Microbiol Infect.* 2012;18:E16–9.