# West Nile virus past infections in the general population of Southern Spain

Máximo Bernabeu-Wittela, Maite Ruiz-Pérezb, María Dolores del Toroc, Javier Aznarb, Ángel Muniainc, Fernando de Oryd, Cristina Domingo<sup>d</sup> and Jerónimo Pachón<sup>e</sup>

<sup>a</sup>Department of Internal Medicine, <sup>b</sup>Department of Microbiology and <sup>e</sup>Department of Infectious Diseases. Hospitales Universitarios Virgen del Rocío. Sevilla. <sup>e</sup>Department of Infectious Diseases. Hospital Universitario Virgen Macarena. Sevilla. <sup>d</sup>Centro Nacional de Microbiología. Instituto de Salud Carlos III. Madrid. España.

OBJECTIVE. To analyze the prevalence of past and recent infections by West Nile virus (WNV) and the risk factors associated with WNV exposure in a representative population from southern Spain.

METHODS. Sample size was established for an estimated prevalence of past WNV infections of 5 ± 2.5% in 504 subjects. A pre-stratification was performed according to age distribution and place of residence. After random telephone solicitation and acquisition of informed consent, a serum sample was collected and an epidemiologic survey performed on all participating subjects. Samples were tested with ELISA-IgG and MAC-ELISA to detect specific IgG and IgM antibodies; results were confirmed by the plaque reduction neutralization test (PRNT). Multivariate analysis using a forward stepwise logistic regression model was performed to assess potential risk factors associated with WNV exposure.

RESULTS. Prevalence of past WNV infections confirmed by PRNT in the 504 participants was 0.6%, affecting mainly older persons (mean age 65  $\pm$  23 vs. 34  $\pm$  22 years; P = 0.018), those living in rural areas (5.4% vs. 0% in urban areas; P = 0.01), and individuals with risk professions (prevalence 2.8% vs. 0%; P = 0.048). None of the five recent infections detected by MAC-ELISA was confirmed by PRNT. CONCLUSIONS. These results strongly suggest past circulation and exposure of the human population to WNV in southern Spain.

Key words: Flavivirus. West Nile Virus. Seroprevalence studies. Spain.

Seroprevalencia de infecciones por el virus del Nilo Occidental en la población general del sur de España

OBJETIVO. Analizar la prevalencia de infecciones pasadas y recientes por el virus del Nilo Occidental (VNO), así como los factores de riesgo asociados con la exposición al

Correspondence: Dr. M. Bernabeu-Wittel. Servicio de Medicina Interna. Hospitales Universitarios Virgen del Rocío. Avda. Manuel Siurot, s/n. 41013 Seville. Spain. E-mail: MAXBW@telefonica.net

Manuscript received on October 9, 2006; accepted for publication on January 2, 2007. mismo, en una muestra representativa de la población del sur de España.

MÉTODOS. El tamaño de la muestra se estableció para una seroprevalencia de infección pasada del 5 ± 2,5% en 504 sujetos, preestratificándose ésta por edad y lugar de residencia. Los voluntarios se incluyeron tras solicitud telefónica y consentimiento, y a todos ellos se les realizó una extracción de suero y una encuesta epidemiológica. Las muestras se analizaron para detectar IgG e IgM específicas, mediante enzimoinmunoensayo (ELISA-IgG) y enzimoinmunoensayo por captura (MAC-ELISA), confirmando ulteriormente los resultados positivos mediantereducción-neutralización en placa (PRNT. Finalmente se realizó un análisis de regresión logística multivariante paso a paso hacia delante para determinar los posibles factores asociados con la exposición al VNO.

RESULTADOS. La prevalencia de infecciones por VNO confirmadas por PRNT en los 504 sujetos incluidos fue del 0,6%, y se vieron principalmente afectadas las personas de mayor edad (edad media 65 ± 23 frente a 34 ± 22 años; p = 0,018), las que vivían en áreas rurales (prevalencia del 5,4% frente al 0% en áreas urbanas; p = 0,01), y las personas con profesiones de riesgo (prevalencia del 2,8% frente al 0%; p = 0,048). Por el contrario, ninguna de las infecciones recientes detectadas por MAC-ELISA fue confirmada ulteriormente por PRNT. CONCLUSIONES. Estos resultados apoyan firmemente la circulación en el pasado del VNO, así como la exposición humana del mismo, en áreas del sur de España.

Palabras clave: Flavivirus. Virus del Nilo Occidental. Estudios de seroprevalencia. España.

## Introduction

The global ecology of West Nile virus (WNV) has experienced noteworthy changes in recent years. From an endemic pattern of infection in specific areas dotted with sporadic local outbreaks every 25-30 years<sup>1,2</sup>, this flavivirus has spread to non-endemic areas over the past 10 years, causing a significant burden of human and animal disease<sup>3-8</sup>. This expansion, the causes of which are not yet totally understood, has been featured by the forthcoming

of human outbreaks preceded by severe and highly mortal epizootia affecting mainly indigenous birds<sup>9,10</sup>. An increase in WNV virulence has also been noted in these outbreaks, confirmed by human mortality ranging between 4.3% in Romania in 1996 and 16.7% in Israel in 2000<sup>4,6</sup>. Because of these issues, WNV is now considered a paradigm of emerging pathogen<sup>1,3</sup>.

The presence of circulating WNV strains has also increased in the Mediterranean basin in the past years, in countries geographically and climatologically closely related to Spain, such as Italy, France, Tunisia, Algeria, and Morocco<sup>11,12</sup>. Many factors concur in our environment to enable the introduction and expansion of WNV in Spain: a) the ubiquitous presence of wetlands (particularly the Doñana National Park in southwestern Spain, one of the most extensive in Europe), and stable populations of various ornitophilic Culex spp. mosquitoes; b) the strategic geographical location for numerous migratory birds coming from endemic areas of Africa (e.g., storks and flamingos), which annually share these natural areas and wetlands with indigenous birds; and c) recent animal and human outbreaks occurring in many bordering countries, configuring a virtual "ring" of WNV circulation around

Despite these issues, there is little data assessing the ecobiology and possible circulation of WNV in sylvatic cycles occurring in our geography. With respect to humans, no cases of confirmed acute infection have been declared, and the few existing seroepidemiologic studies were performed with techniques having poor specificity (indirect hemagglutination)<sup>13,14</sup>. Thus, the current impact of human exposure to WNV in Spain remains uncertain. The aim of the present study was to analyze the prevalence of past and recent WNV infections and the possible risk factors associated with exposure to WNV in a representative population from southern Spain.

#### Material and methods

## Geographical area and study population

Individuals from the city of Seville and the northern, western, and southern parts of Seville province (1,194,570 inhabitants). The province of Seville (southwestern Spain) covers an area of 14,000 square kilometers and has 1,758,720 inhabitants (65% of whom live in urban areas). The capital of the province, Seville, has 705,000 inhabitants. This area is located close to numerous wetlands, in particular Doñana National Park, one of the most extensive wetlands in Europe, and the delta of the Guadalquivir river, both of which shelter a large variety of indigenous and migratory avian species. Numerous livestock exploitations (mainly of equine cattle) are established in the vicinity of the park. The area has a mild Mediterranean climate, with a mean temperature of 19.6 °C (12.9-26.3), and mean rainfall of 327 L/m²/year. The topography is divided into farms and crops (66%), woods (19%), pastures (7.5%), and settled areas (7.5%).

#### Sample size/stratification

Sample size was established for an estimated prevalence of past WNV infections of 5  $\pm$  2.5% (confidence level 99%) in 504 subjects. A pre-stratification was performed using the Annual Statistics Report of Andalusia (2001 census)^{15}, according to age distribution (0-14  $[n=110],\ 15\text{-}29\ [n=135],\ 30\text{-}44\ [n=98],\ 45\text{-}64\ [n=101],\ and > 64 [n=60]\ years),$  and place of residence of the population (> 50,000 = urban  $[n=310],\ 5000\text{-}50,000$  = suburban  $[n=157],\ <5000$  = rural [n=37]).

### Inclusion criteria and sample collection

Subjects of all age groups were selected by random telephone solicitation, with the exception of the youngest age group (0-14 years). Subjects from this last group were pre-selected in our two central laboratories, which receive blood samples from the entire area (primary care, outpatient clinics, and hospital). Based on this pre-selection, we ultimately collected samples originating from primary care centers and pre-anesthetic assessment in outpatient clinics (to avoid selection of hospitalized, chronically ill, or immunosuppressed children). Subsequently, legal guardians were asked by phone for informed consent to use the remaining serum for our study, and were questioned to detect possible exclusion criteria. Once informed consent had been granted, the serum sample was stored at -80 °C and an epidemiologic survey was performed on all participating subjects. The data collected included gender, age, profession, place of residence, outdoor activities, travel over the past 12 months to other parts of Spain and Europe, contact with animals, and insect bites in the past month. Risk professions were defined as those involving close contact with animals (farmers, stockbreeders, veterinarians), nature and wetlands (rangers, foresters), or mosquitoes (extermination and pest control workers). The following persons were excluded: those unable to answer the epidemiologic survey, those who had traveled to endemic areas in the previous 12 months, and persons previously vaccinated against yellow fever, hospitalized in the last 30 days, with febrile disease in the last 30 days, or with any type of immunosuppression.

#### Serological techniques

Samples were processed and tested for WNV IgG by enzyme-linked immunosorbent assay (ELISA) and for WNV IgM by antibody-capture ELISA (MAC-ELISA) (Focus Technologies, Rendon, VA, USA) according to the manufacturer's guidelines. Assays were performed in duplicate. All the positive samples were retested and the final result was the mean value of all determinations. When available, IgM-positive samples were tested with a confirmatory ELISA procedure recommended by the manufacturer (background subtract procedure).

Positive or indeterminate sera in both the IgG and IgM ELISA tests were assayed by a plaque reduction microneutralization test against WNV (PRNT). For this purpose, samples were tested in duplicate and assayed twice. Two-fold dilutions (25  $\mu L$ ) of the samples ranging from 1:16 to 1:256 were placed in a 96-well tissue culture microplate (Nunc A/S, Roskilde, Denmark) and mixed with 25  $\mu L$  containing 100 tissue culture infectivity doses (100 TCID $_{50}$ ) $^{16}$  of a reference E101 WNV preparation. After 1 hour of incubation at 37 °C, 50  $\mu L$  of a Vero E6 cell suspension containing  $4x10^5$  cells/mL was added to each well. Cultures were maintained for 7 days at 37 °C and 5% CO $_2$ , and then fixed with a solution containing 10% formaldehyde and 1% naphtol blue-black dye. The endpoint titer was defined as the highest serum dilution that showed no cytopathic effect in 50% of cell cultures. All these procedures were performed in a level-3 biosafety facility.

Sera showing an IgG/IgM titer against WNV underwent further commercial serological analyses against other flaviviruses (dengue and tick-borne encephalitis [TBE]). The assays for dengue IgG (Panbio Diagnostics, Queensland, Australia) and for TBE IgG and IgM (Dade Behring, Deerfield, IL, USA) were based on indirect ELISA; the method for dengue IgM was a  $\mu$ -chain capture ELISA (Panbio Diagnostics).

### **Definition of WNV infection**

Subjects with any ELISA IgG titer were considered as having possible past WNV infection; subjects with any MAC-ELISA IgM titer were considered as having possible recent WNV infection. Those with any ELISA IgG or MAC-ELISA IgM titer, subsequently confirmed by PRNT assay, were considered as having definitive past or recent WNV infection, respectively.

#### Statistical analysis

Seroprevalence was determined in the overall sample, and by place of residence and age group. Continuous variables are expressed as

TABLE 1. Epidemiological features of subjects with past West Nile virus infection in the population of southern Spain

	Subject (gender, age, place of residence, risk-profession, outdoor activities)	ELISA-IgG*	MAC-ELISA-IgM*	PRNT	Classification	
	Cases with WNV IgG positive result					
1	Male, 77 years, rural, yes, no 217VM	3.75	Negative	> 256	WNV past infection	
2	Male, 80 years, suburban, yes, no 7VM	3.33	Negative	128	WNV past infection	
3	Male, 38 years, rural, yes, no 112VR	3,18	Negative	64	WNV past infection	
4	Female, 30 years, urban, no, no 234VR	1.79	Negative	< 16	Not confirmed	
5	Female, 30 years, urban, yes, no 117VM	1.53	Negative	< 16	Not confirmed	
	Cases with WNV IgM positive result					
6	Male, 38 years, urban, yes, yes 199VR	Negative	4.30/negative**	< 16	Not confirmed	
7	Female, 5 years, urban, no, no 210VM	Negative	2.23/negative**	< 16	Not confirmed	
8	Male, 20 years, suburban, no, no 76VM	Negative	1.86/na***	< 16	Not confirmed	
9	Female, 11 years, urban, no, no 228VR	Negative	1.66/negative**	< 16	Not confirmed	
10	Male, 19 years, urban, no, yes 284VR	Negative	1.31/negative**	< 16	Not confirmed	

<sup>\*</sup>Absorbance ratio.

mean ± standard deviation. Univariate analysis was performed to determine possible risk factors associated with past infection. Univariate group comparisons were performed using the chi-square, Fisher, and Student t tests and correlations were determined with Spearman's Rho. Multivariate analysis of factors showing significant differences on univariate analysis was performed using a forward stepwise logistic regression model. Group differences were determined with the odds-ratio (OR) and 95% confidence interval (CI). Significance was set at a P-value of < 0.05.

## Results

Five subjects (nearly 1% of the 504 included) presented anti-WNV IgG titers with a mean ELISA absorbance ratio of 2.72 ± 1 (table 1); none of the remaining sera had indeterminate absorbance ratios. Prevalence of definitive past WNV infection confirmed by PRNT was 0.6% (3 positive samples, with PRNT titers of 64, 128, and > 256, respectively (table 1), and was significantly higher in rural areas (5.4%), than in suburban (0.6%; P = 0.09), or urban areas (0%; P = 0.01). Mean age of subjects with past WNV infections was higher than that of uninfected subjects (65  $\pm$  23 vs.  $34 \pm 22$  years, P = 0.01). Comparisons of demographic and epidemiological features between subjects with and without past WNV infection are shown in table 2. No differences were detected with the exception of activity in any risk profession (P = 0.048), which was associated with prior WNV exposure. Multivariate analysis detected no independent risk factors associated with past WNV infection.

Five subjects (nearly 1% of the 504 included) showed IgM titers against WNV with a mean MAC-ELISA absorbance ratio of 2.03 ± 1.15 (table 1); WNV IgG antibodies were not detected in any of these subjects, and none of the remaining sera had indeterminate absorbance ratios. None of these individuals presented specific anti-WNV neutralizing antibodies when PRNT assay was performed (table 1). Application of a background subtract procedure to four of the remaining sera also yielded negative results.

TABLE 2. Epidemiological features of subjects with definitive past WNV infection in the population of Southern Spain, compared with those without previous exposure

_	=	
Epidemiologic feature	Prevalence of past WNV infection	P
Gender (male/female)	1.2%/0	NS
Risk profession (yes/no)	2.8%/0	0.04
Travel to rural areas (yes/no)	1.2%/0.3%	NS
Outdoor activities (yes/no)	0/0.6%	NS
Contact with animals (yes/no)	0.5%/0.7%	NS
Insect bites in past month (yes/no)	0/0.7%	NS

NS: not significant; WNV: West Nile virus.

Analysis of the IgG immune response against dengue and TBE viruses in the 5 subjects with anti-WNV IgG titers by ELISA showed IgG titers against dengue in sera from 3 subjects (subjects 1, 2, and 3 in table 1); no IgG immune response was detected against TBE virus. Similarly, analysis of IgM immune response against dengue and TBE viruses in the 5 subjects with anti-WNV IgM titers by ELISA detected IgM titers against dengue in the serum of one subject (subject 6 in table 1). After application of a subtract background assay to this serum sample (in the presence and absence of specific dengue antigen), the specific IgM immune response was not confirmed, and therefore ruled out. No IgM immune response against TBE virus was detected in any of the 5 subjects.

## Discussion

The data from the present study show the presence of definite past WNV infection in the general population of southern Spain, confirming exposure of the population to this pathogen. These data complete previous preliminary studies carried out in Spain with less specific techniques that reported past infection rates of 8% in the human population of the northwestern part of the country and 3.1%

<sup>\*\*</sup>Results obtained after application of background subtract procedure.

<sup>\*\*\*</sup>Remaining serum not available.

ELISA-IgG: enzyme-linked immunosorbent assay; MAC-ELISA-IgM: antibody-capture enzyme-linked immunosorbent assay; PRNT: plaque reduction neutralization test; WNV: West Nile virus.

in small mammals collected from several parts  $^{13,14,17,19}$ . In addition, the findings are in accordance with the results of a similar recent study performed in northeastern areas of the country, which detected about 0.2% of confirmed WNV past infection in the general population<sup>20</sup>.

Definitive past WNV infection was characterized by affecting older persons, subjects living in rural areas, and those having risk professions. This epidemiological pattern probably reflects the means of acquisition, which may have been in the former past (older people are exposed during longer periods of time), and in subjects with closer contact to the wild cycle of WNV, since rural areas are located closer to wetlands, and therefore to reservoirs and potential vectors; the same is true for persons with risk professions.

As other authors have pointed out with respect to European countries, the present study raises questions as to the absence of human outbreaks of WNV disease in Spain, despite documented circulation of the virus, population exposure to WNV, and even declaration of sporadic outbreaks affecting equine cattle in surrounding countries<sup>8,21</sup>. This epidemiologic feature contrasts with the dynamics of WNV disease in North America, in which WNV has expanded considerably, causing more than 7,000 cases of human disease since 1999<sup>22</sup>. Several factors may be related to this difference. Natural immunization of the majority of the population due to early exposure to WNV is hardly admissible, because the prevalence rates found in the present study are far from those reported in endemic areas, which range from 8% to 15%3,23. Partial immunization conferring some kind of protection against WNV due to population exposure to antigenically-related flaviviruses is also unlikely, because no indigenous cases or evidence of circulation of the Japanese encephalitis virus group or dengue virus have been reported in Spain, and yellow fever was eradicated in the final years of the 19th century. Exposure to other flaviviruses (mainly TBE virus, which is endemic in vast territories of Europe) could highlight this argument explaining the discordance between seroprevalences determined by ELISA (1%) and the definitive PRNT assay (0.6%) in the present study, and could also explain the recent human cases imported by Irish tourists after a visit to southern Portugal (an area close to Andalusia), whereas there is no evidence of native human cases despite detection of WNV in indigenous mosquitoes<sup>24,25</sup>. This explanation is not supported by our data, which showed a lack of specific antibodies against TBE virus in the sera tested. We hypothesize that the main reason may lie in climatic factors. The relatively warm, dry summer weather, together with the migratory pattern of avian species originating from endemic areas, which arrive to our wetlands in winter when the activity of potential indigenous biologic vectors is lower (they emerge from aquatic stages in spring and bite until fall), and when female mosquitoes begin diapause and feed less frequently, establish unfavorable environmental conditions for significant amplification of the cycle to "jump" to bridging vectors (mosquitoes that bite both birds and humans), and to indigenous avian species. Nonetheless, this pattern can easily change in the future, due to the global climatic changes we are witnessing<sup>26,27</sup>.

Another issue our data raises is the value of ELISA-IgG and MAC-ELISA in the diagnosis of recent and past WNV

infection. The lack of specificity of these methods for accurate diagnosis of WNV infection has been discussed<sup>28,29</sup>, and discordance between the high specificity to detect immune response against WNV endowed by the technique's manufacturers and the well-recognized serological cross reactions against flaviviral antigens is a frequent and unsolved issue<sup>29-32</sup>. In fact, we detected IgG antibodies against dengue virus in the 3 subjects with specific neutralizing IgG titers against WNV, confirming cross-reactivity of the ELISA techniques with respect to closely related flaviviruses (dengue is not an endemic disease in Spain, and subjects with past WNV infections reported no travel to dengue endemic areas). Subjects with a single IgG against WNV, unconfirmed by neutralization are difficult to explain according to current knowledge. A plausible explanation might be prior exposure to an antigenic-related flavivirus unrecognized in our area. Clearly, future studies in this area are necessary, focusing on the impact of WNV (and other antigenic-related flavivirus) circulation in indigenous and migratory fauna, and on mosquitoes and other potential vectors (e.g., ixodidae and argasidae) to fully understand local epidemiology of WNV in humans33.

In conclusion, our data have documented the presence of definite past WNV infection in the general population of southern Spain. These infections were associated with older age, rural areas, and risk professions.

#### Acknowledgements

This study was financially supported by the Consejería de Salud, Junta de Andalucía (44/03), Spain.

#### References

- 1. Petersen LR, Roehrig JT. West Nile Virus: a reemerging global pathogen. Emerg Infect Dis. 2001;7:611-4.
- 2. McIntosh BM, Jupp PG, Dossantos I, Meenehan GM. Epidemics of West Nile and Sindbis viruses in South Africa with Culex (Culex) univittatus Theobald as vector. S Afr J Sci 1976;72:295.
- Marfin AA, Gubler DJ, West Nile encephalitis; an emerging disease in the United States. Clin Infect Dis. 2001;33:1713-9.
- Weinberger M, Pitlik SD, Gandacu D, Lang R, Nassar F, Ben David D, et al. West Nile Fever Outbreak, Israel, 2000: Epidemiologic aspects. Emerg Infect Dis. 2001;7:686-91.
- Weiss D, Carr D, Kellachan J. Clinical findings of West Nile Virus infection in hospitalized patients, New York, and New Jersey, 2000. Emerg Infect Dis. 2001:7:654-9.
- 6. Han LL, Popovici F, Alexander JP Jr, Laurentia V, Tengelsen LA, Cernescu C, et al. Risk factors for West Nile virus infection and meningoencephalitis, Romania, 1996. J Infect Dis. 1999;179:230-3.
- 7. L'vov DK, Butenko AM, Gaidamovich SIa, Larichev VF, Leshchinskaia EV, Zhukov AN, et al. Epidemic outbreak of meningitis and meningoencephalitis, caused by West Nile virus, in the Krasnodar territory and Volgograd region, Vopr Virusol, 2000:45:37-8.
- Durand B, Chevalier V, Pouillot R, Labie J, Marendat I, Murgue B, et al. West Nile virus outbreak in horses, Southern France, 2000: Results of a serosurvey. Emerg Infect Dis. 2002;8:777-82
- 9. Mostashari F, Kulldorff M, Hartman JJ, Miller JR, Kulasekera V. Dead bird clusters as an early warning system for West Nile virus activity. Emerg Infect Dis. 2003;9:641-6.
- 10. Guptill SC, Julian KG, Campbell GL, Price SD, Marfin AA, Early-season avian deaths from West Nile virus as warnings of human infection. Emerg Infect Dis. 2003;9:483-4
- 11. Hubalek Z, Halouzka J. West Nile Fever-a reemerging mosquito-borne viral disease in Europe. Emerg Infect Dis. 1999, 5:643-50
- 12. Murgue B, Murri S, Triki H, Deubel V, Zeller HG. West Nile in the Mediterranean basin. Ann NY Acad Sci. 2001;951:117-26.
- 13. Garea MT, Filipe Ar. Antibodies to arboviruses in Northwestern Spain. Am J Trop Med Hyg. 1977;26:792-7.

- 14. Lozano A, Filipe AR. Anticuerpos frente al virus West Nile y otros virus transmitidos por artrópodos en la población del delta del Ebro. Rev Esp Salud Pública. 1998;72:245-50.
- 15. Anuario Estadístico de Andalucía 2003. Instituto de Estadística de Andalucía. Available at: (http://www.juntadeandalucia.es/institutodeestadistica/ anuario/anuario03/index.htm).
- 16. Reed LJ, Muench HA. Simple method of estimating fifty per cent endpoints. Am J Hyg. 1938;27:493-7.
- 17. Chastel C, Launay H, Rogues G, Beaucournu JC. Arbovirus infections in Spain: serological survey on small mammals. Bull Soc Pathol Exot Filiales. 1980:73:384-90.
- 18. Filipe AR, De Andrade HR. Arboviruses in the Iberian Peninsula. Acta Virol. 1990;34:582-91.
- 19. Sánchez-Seco MP, Echevarría JM, Hernández L, Estévez D, Navarro-Mari JM. Tenorio A. Detection and identification of Toscana and other phleboviruses by RT-nested-PCR assays with degenerated primers. J Med Virol. 2003;71:140-9.
- 20. Bofill D, Domingo C, Cardeñosa N, Zaragoza J, De Ory F, Minguell S, et al. Human West Nile virus infection, Catalonia, Spain. Emerg Infect Dis. 2006;12:1163-4.
- 21. Cantile C, Di Guardo G, Eleni C, Arispici M. Clinical and neuropathological features of West Nile virus equine encephalomyelitis in Italy. Equine Vet J. 2000:32:31-5.
- 22. Hayes EB, Komar N, Nasci RS, Montgomery SP, O'Leary DR, Campbell GL. Epidemiology and transmission dynamics of West Nile virus disease. Emerg Infect Dis. 2005;11:1167-73.
- 23. Petersen LR, Marfin AA. West Nile virus: a primer for the clinician. Ann Intern Med. 2002;137:173-9.

- $24.\,$ Sánchez-Seco MP, Navarro JM. Infecciones por el virus Toscana, el virus del Nilo Occidental y otros arbovirus de interés en Europa. Enferm Infecc Microbiol Clin. 2005;23:560-8.
- Connell J, McKeown P, Garvey P, Cotter S, Conway A, O'Flannagan D, et al. Two linked cases of West Nile Virus acquired by an Irish tourist in Algarye. Portugal. Eurosurveillance weekly 8. 2004. Available at: http://www.eurosurveillance.org/ew/2004/040805.asp
- 26. Mellor PS, Leake CJ. Climatic and geographic influences on arboviral infections and vectors. Rev Sci Tech. 2000;19:41-54.
- 27. Patz JA, Epstein PR, Burke TA, Balbus JM. Global climate change and emerging infectious diseases. JAMA. 1996;275:217-23.
- 28. Tardei G, Ruta S, Chitu V, Rossi C, Tsai TF, Cernescu C. Evaluation of immunoglobulin M (IgM) and IgG enzyme immunoassays in serologic diagnosis of West Nile Virus infection. J Clin Microbiol. 2000;38:2232-9.
- 29. Holmes DA, Purdy DE, Chao DY, Noga AJ, Chang GJ. Comparative analysis of immunoglobulin M (IgM) capture enzyme-linked immunosorbent assay using virus-like particles or virus-infected mouse brain antigens to detect IgM antibody in sera from patients with evident flaviviral infections. J Clin Microbiol. 2005:43:3227-36.
- 30. Ebel GD, Dupuis AP 2nd, Nicholas D, Young D, Maffei J, Kramer LD. Detection by enzyme-linked immunosorbent assay of antibodies to West Nile virus in birds. Emerg Infect Dis. 2002;8:979-82.
- Ozkul A, Yildirim Y, Pinar D, Akcali A, Yilmaz V, Colak D. Serological evidence of West Nile Virus (WNV) in mammalian species in Turkey. Epidemiol Infect. 2005;29:1-4.
- 32. Kuno G. Serpodiagnosis of flaviviral infections and vaccinations in humans. Adv Virus Res. 2003;61:3-65.
- 33. Lawrie CH, Uzcategui NY, Gould EA, Nuttall PA. Ixodid and argasid tick species and West Nile virus. Emerg Infect Dis. 2004;10:653-7.