



Contents lists available at ScienceDirect

Acta Colombiana de Cuidado Intensivo

journal homepage: www.elsevier.es/accii

Review article

Pathogenicity of sepsis and its effects on cell physiology

Patogenicidad de la sepsis y sus efectos sobre la fisiología celular

Qurban Ali^{a,*}, Mohammad Arshad Javed^a, Mohd Amir^b, Ajaz Ahmad^c^a Department of Plant Breeding and Genetics, Faculty of Agricultural Sciences, University of the Punjab, Lahore, Pakistan^b Department of Natural Products & Alternative Medicine, College of Pharmacy, Imam Abdulrahman Bin Faisal University, Dammam, Saudi Arabia^c Department of Clinical Pharmacy, College of Pharmacy, King Saud University, Riyadh, Saudi Arabia

ARTICLE INFO

Keywords:

Costimulatory molecules

Neutrophils

Coagulation

Palabras clave:

Moléculas coestimulantes

Neutrófilos

Coagulación

ABSTRACT

Sepsis is a serious medical condition with a high mortality rate where patients experience a severe infection. In a normal operation, the immune and physiological systems work together to eliminate dangerous infections. On the other hand, sepsis occurs when the body cannot control these normal physiological reactions. In an ideal situation, the first interaction between the immune system and a pathogen would result in the total elimination of the infection and a quick return to balance in the host's body. Increased macrophage and neutrophil activity can accelerate the septic response. Sepsis happens because of many things, such as more cell and tissue damage, faster lymphocyte cell death, longer neutrophil cell death, and too many lymphocytes costimulatory molecules being made. Various interactions between the coagulation system and the inflammatory response result in an imbalanced reaction from both systems. Identifying patients who could potentially benefit from immunomodulatory therapy and assisting in diagnosing sepsis are important applications of biomarkers.

RESUMEN

La sepsis es una enfermedad grave con una alta tasa de mortalidad, en la que los pacientes experimentan una infección grave. En condiciones normales, los sistemas inmunitario y fisiológico trabajan en conjunto para eliminar infecciones peligrosas. Por otro lado, la sepsis ocurre cuando el organismo no puede controlar estas reacciones fisiológicas normales. Idealmente, la primera interacción entre el sistema inmunitario y un patógeno resultaría en la eliminación total de la infección y un rápido restablecimiento del equilibrio en el organismo del huésped. El aumento de la actividad de macrófagos y neutrófilos puede acelerar la respuesta séptica. La sepsis se produce debido a diversos factores, como un mayor daño celular y tisular, una muerte celular más rápida de los linfocitos, una muerte celular más prolongada de los neutrófilos y una producción excesiva de moléculas coestimulantes de linfocitos. Diversas interacciones entre el sistema de coagulación y la respuesta inflamatoria resultan en una reacción desequilibrada de ambos sistemas. Identificar a los pacientes que podrían beneficiarse de la terapia inmunomoduladora, y ayudar en el diagnóstico de la sepsis son aplicaciones importantes de los biomarcadores.

Introduction

Sepsis is defined as life-threatening organ dysfunction caused by a dysregulated host response to infection. When two or more criteria including temperature ($>38^{\circ}\text{C}$ or $<36^{\circ}\text{C}$), heart rate (>90 beats per minute), respiratory rate (>20 or $\text{PaCO}_2 <32$ mm Hg) and white blood cell count ($>12 \times 10^9$ or $<4 \times 10^9 \text{ mm}^{-3}$, or $>10\%$ bands) are present, clinicians classify the condition as systemic inflammatory response syn-

drome (SIRS). SIRS can be diagnosed as sepsis when an infection is identified as the root cause.¹ It is not always necessary to have a positive pathogen culture to diagnose sepsis. A culture may not be required in cases with a strong indication of infection, such as the presence of neutrophils in a normally sterile area like the peritoneum.² A serious condition, severe sepsis, occurs when the septic process reaches a level of severity where the functioning of one or more organs is disrupted. Severe sepsis can cause low blood pressure, which is one indication of septic shock.³ Septic shock is a subset of sepsis in which underlying circulatory and cellular/metabolic abnormalities are profound enough to substantially increase mortality. It is clinically identified by the require-

* Corresponding author.

E-mail address: saim1692@gmail.com (Q. Ali).<https://doi.org/10.1016/j.acci.2025.08.010>

Received 27 November 2024; Accepted 25 August 2025

Available online xxx

0122-7262/© 2025 Asociación Colombiana de Medicina Crítica y Cuidado Intensivo. Published by Elsevier España, S.L.U. All rights are reserved, including those for text and data mining, AI training, and similar technologies.

ment of vasopressor therapy to maintain a mean arterial pressure (MAP) ≥ 65 mm Hg and serum lactate level > 2 mmol/L despite adequate fluid resuscitation. Ranked by severity: Sepsis is a more serious condition compared to septic shock, which is essentially a form of sepsis.⁴ A comprehensive analysis of individuals with severe sepsis from an international database provided valuable insights into the development of the condition, according to a 2009 assessment.⁵ Based on data collected from over 11,000 patients across 37 countries, the study has identified several key symptoms of sepsis.⁶ A significant number of patients (57%) presented with gram-negative infections, while 44% had gram-positive infections. Additionally, fungal infections were observed in many patients, affecting approximately 11% of the individuals.^{7–10} It is important to note that there were certain cases where the total percentage of illnesses exceeded 100%. In 47% of cases, the lungs had the highest infection rate among the patients. The abdomen was affected in 23% of cases, while the urinary tract was affected in 8% of cases.¹¹ A notable portion of patients, including the individual in this case study, also experienced diabetes (24%), chronic lung disease or malignancy (16%), congestive heart failure (14%), and renal insufficiency (11%).¹² The database revealed a significant number of fatalities, highlighting the severity of sepsis as a life-threatening condition. There was no noticeable decrease in sepsis mortality throughout the patient enrollment period. Having a thorough understanding of sepsis pathology is crucial for enhancing survival rates.

Sepsis and phagocytic cells

When a bacterial pathogen enters a sterile environment, the local cells that detect the infiltrator usually trigger an inflammatory response. In cases with only a few invading germs, the body's local immune responses can often eliminate the infections successfully.¹³ Macrophages play a crucial role in engulfing bacteria and releasing proinflammatory cytokines, which activate the innate immune system to fight against the bacterial pathogen.¹⁴ This probably occurred to patient during the initial stages of the infection, after the rupture of her colonic diverticulum. M1 cells are macrophages that produce chemokines like IL-8 (CXCL8) and cytokines such as IL-1 β , TNF, and IL-6¹⁵ (additional information regarding biomarkers will be provided below). Dendritic cells and macrophages are specialized cells that play a crucial role in the immune response by alerting the body to the presence of an infection.¹⁶ They achieve this by identifying pathogen-associated molecular patterns and common microbial molecules in a diverse range of bacteria, fungi, and viruses. Activated peripheral blood cells contain receptors that can detect various substances.^{17–21} As a result, cytokines are generated, contributing to the natural inflammatory response. One of the most widely recognized PRRs are toll-like receptors, which can detect various components of bacterial and fungal cell walls and viral and bacterial nucleic acids.²² During the innate response to bacterial contact, macrophages experience an increase in the levels of CD80 and CD86, which are costimulatory molecules. The molecules are crucial in the interactions between innate and adaptive immune responses (see the following section).²³ The initial discharge of pathogens from the perforated diverticulum was prevented by local peritoneal macrophages when bacteria invaded a sterile zone. The patient's condition is being closely monitored to ensure minimal complications, primarily focusing on providing effective local treatment.²⁴ It is expected that the newly recruited neutrophils, if they overcome this initial defense barrier, will be able to eliminate the pathogens effectively.²⁵

Inflammatory cells

In most cases, when inflammation occurs, more cells are needed to help clear the pathogens. Endothelial cell surface adhesion molecules are produced due to cytokine secretion from nearby inflammatory cells.²⁶ The transportation of white blood cells to the site of inflama-

tion occurs when they temporarily attach to the lining cells of the blood vessels and pass through the vessel wall. MicroRNAs have also been linked to the modulation of microbial adhesion molecules.²⁷ Peripheral blood contains a variety of cell categories, such as monocytes, lymphocytes, and neutrophils. Neutrophils make up more than half of the blood cells in individuals who are in good health.²⁸ Neutrophils, also known as polymorphonuclear leukocytes, have a variety of nuclear structures. Phagocytic cells are commonly referred to as macrophages and neutrophils.²⁹ Neutrophils and macrophages have different ways of eliminating pathogens. The body utilizes energy for various functions, some of which are necessary while others are not. Phagocytosis is an important intracellular mechanism crucial in eradicating pathogens by phagocytic cells. It serves as the initial stage in the eradication process.³⁰ Upon entering the body, bacteria are often surrounded by host proteins, such as complement fragments and antibodies. Neutrophil surface receptors facilitate phagocytosis by recognizing opsonized proteins on bacterial surfaces. PRRs include complement receptors and the Fc subunit of immunoglobulins.³¹ The elimination of the bacterium is a direct outcome of the various processes taking place within the neutrophils. Vacuoles called pneumosomes contain microorganisms that have been ingested through phagocytosis. It forms phagosomes by interacting with specific intracellular granules.³² Neutrophils possess granules with antimicrobial properties. The earliest (azurophilic) granules contain α -defensins, antimicrobial proteases such as cathepsin G and elastase, myeloperoxidase, and a protein that enhances bacterial permeability. In addition, secondary granules contain antimicrobial peptides like lactoferrin, metalloproteases, and lysozyme.³³ The interaction between bacteria and neutrophil granules creates an unfavorable local environment. This environment has a lower pH and contains powerful proteases designed to eliminate pathogens. In addition, there are implications of mortality related to the dependence on oxygen and other mechanisms. Reactive oxygen intermediates produced by neutrophils, such as superoxide anion and hydroxyl radicals, can trigger a respiratory surge.³⁴ A desirable scenario entails the effective collaboration of mediators, ensuring they stay within the phagolysosome. This allows for the swift eradication of bacteria while maintaining the host's well-being. In certain situations, sepsis can occur when microbes can evade the body's defenses or when the body is harmed by its response.³⁵ Neutrophil extracellular traps (NETs) are formed in response to bacterial interactions. The neutrophil extracellular matrix (NET) comprises fragments of neutrophil DNA, antimicrobial peptides, and histones. This response helps in the fight against pathogens. Nets are made up of substances that help prevent the growth of microorganisms.³⁵

Sepsis occurs when the body's inflammatory response to an infection becomes severe enough to disrupt important physiological functions. The given description may not be entirely accurate, but this response can be classified as an overly enthusiastic or exaggerated inflammatory reaction.³⁶ In cases with a significant bacterial load, especially with highly virulent bacteria, it can trigger an inflammatory reaction corresponding to the level of bacterial stimulation. Nevertheless, for individuals with a susceptibility to sepsis, the resulting unintended consequences pose significant challenges in terms of effective management.³⁷ A careful examination of the medical case study reveals that an intervention aimed at reducing inflammation could have positive and negative effects in this scenario. This has the potential to reduce the harmful effects of inflammation. However, it can weaken the body's ability to fight off infections. It is widely acknowledged that individuals with weakened immune systems are at a higher risk of developing infections, including sepsis.³⁸

Lymphocytes

During bacterial infections, lymphocytes and antigen-presenting cells (APCs) have a strong and essential interaction. APCs play a crucial role in the development of an adaptive immune response.³⁹ Antigen-presenting cells (APCs) work with costimulatory factors and

CD3 proteins on the cell surface to present microbial antigens to T lymphocytes. After receiving the appropriate signals, effector CD4⁺ T cells release cytokines such as interferon (IFN)- γ . These cytokines activate phagocytic cells to eliminate pathogens within the cell.⁴⁰ Moreover, effector CD4⁺ T cells play a crucial role in stimulating the production of antibodies, which offer defense against microbial infection by interacting with B cells. Furthermore, T cells enhance the production of CD40 ligand, which combines with APC CD40 to form a complex. This method enhances the communication between innate and adaptive systems by maintaining the expression of costimulatory molecules and increasing the secretion of IL-12.⁴¹ Patients suffering from sepsis often experience a significant decrease in lymphocyte counts as a result of apoptosis. The decrease in question is expected to significantly impact the development of an immunosuppressive state, a common occurrence in the final stages of sepsis. Patients with compromised health are more susceptible to acquiring additional infections.⁴² Immunohistochemistry staining revealed a decrease in the number of splenic B cells and CD4⁺ T cells in the postmortem spleens of patients with sepsis. Patients with sepsis often experience decreased CD4⁺ T cells in their peripheral blood. Unlike individuals who are in a state of excellent health. It is worth mentioning that individuals who show improvement in sepsis also experience a decrease in T cell apoptosis.⁴³ Using a two-hit paradigm, an evaluation was conducted on T cells in the spleens of mice. In this specific case, a mixed microbial infection occurred due to cecal ligation and puncture (CLP). After five days, *Pseudomonas aeruginosa* was introduced intravenously as part of an experiment. Based on the findings, it is evident that the mice produced a lower amount of IFN- γ .⁴⁴ Nevertheless, in a controlled laboratory environment, the splenic T cells demonstrate similar levels of IFN- γ production when stimulated with IL-12, as observed in the control group. The findings suggest that the ability of T cells to respond to subsequent infections may be compromised due to a potential lack of effective stimulation by APCs following an initial infection.⁴⁵ A possible explanation for decreased T cell functionality in sepsis is modifying antigen-presenting cell (APC) signaling pathways through costimulatory molecules. This can result in the development of anergic cells and ultimately cell death. It has been noted that patients diagnosed with sepsis tend to have a higher prevalence of the inhibitory costimulatory ligand CTLA-4/CD152 on their T cells.⁴⁶ Alongside this rise, there is a decrease in the expression of the costimulatory molecule CD86 on monocytes. Based on research conducted on individuals who have recovered from sepsis, it has been observed that the decline in T cell apoptosis becomes less pronounced over time. This decline was associated with an increase in CD86 over-expression and a decrease in CTLA-4 expression.⁴⁷ One possible reason for the reduced lymphocyte activity in septic patients is the excessive presence of regulatory T cells (Tregs), specifically CD4⁺CD25⁺ cells. Whole-blood samples collected from septic patients showed a reduced ability of T cells to respond to antigens, as observed in studies conducted outside of the body. When examining splenocytes from septic rodents, it was observed that the proliferation response was restored when Foxp3, a crucial transcription factor for regulatory T cells (Tregs), was absent.⁴⁸

Immune responsible molecules

One well-known and well-researched aspect of developing sepsis is the increased generation of proinflammatory cytokines during the innate immune response (see the following biomarker section).⁴⁹ Recent studies have demonstrated that the host response to sepsis is significantly influenced by the interplay between antigen-presenting cells (APCs) and the adaptive immune system. Some interactions happened because of the septic patient's weakened immune system. The interaction between the two branches of the immune system is expanding, particularly how the first reaction affects the second response and possible consequences for the prognosis of sepsis patients in the long run. Significant lymphocyte apoptosis is frequently observed in sepsis patients, which reduces the capacity of septic mouse mono-

cytes to activate T cells.⁵⁰ The immunological synapse that links APC and T cells depends on cell surface proteins called CSMs. Antigen-presenting cells (APCs) must express these substances to regulate T cell activation properly. The fate of T cell growth is largely determined by the secondary signals, which can either stimulate or inhibit it. This can ultimately lead to anaerobic responses and cell death.⁵¹ A thorough analysis of the B7 family's cell surface molecules, CD80 (B7-1) and CD86 (B7-2) has been conducted. As ligands for CD28/CTLA-4 receptors on T cells, CSMs are found in APCs and proliferate in response to various microbiological stimuli. Like other signaling systems, CD80 and CD86 can interact with CD28 or CTLA-4 to produce stimulatory or inhibitory responses. T cells become more active and proliferative during transit when CD28 is activated.⁵² Conversely, CTLA-4 is only generated after T cell activation and functions to impede the T cell's reaction to the antigen. A monoclonal agonist antibody for CD28 was used in a clinical experiment that revealed the possible significance of the B7:CD28 pathway in the innate response. The study subjects treated with antibodies exhibited sepsis-like clinical symptoms and a strong inflammatory response.⁵³ Research findings demonstrate that CLP can lead to increased CD80 expression in various types of monocytes found in rodents' peritoneal, splenic, and peripheral blood. This phenomenon mimics the development of sepsis in humans and the occurrence of bowel perforation. In contrast, the expression of CD86 is reduced in the peritoneum, but increased in the spleen and peripheral circulation.⁵⁴ Although there is significant sequence and ligand overlap, the change in expression indicates that the roles of CD80 and CD86 expression may differ depending on the location of the antigen-presenting cells about the infection site. A comparable form of compartmentalization has been observed in the cytokine response to infection.⁵⁵ In addition, it may substantially impact the probability of a severe localized infection advancing to systemic immunosuppression, a condition frequently observed in the later stages of sepsis.⁵⁶ There are noticeable differences in CD80 and CD86 expression levels in sepsis cases, which can impact survival. Comparing the survival rates of CD86-/- mice and wild-type controls to CD80-/- mice or mice treated with anti-CD80 monoclonal antibody before CLP reveals a significant improvement. Severe patients in the intensive care unit (ICU) show increased CD80 expression on their circulating monocytes.⁵⁷ Elevated levels of CD80, associated with shock, indicate the negative impact of the protein. However, there is no observed connection with survival. The expression of CD86 in these cells is lower compared to the healthy control group. On the other hand, individuals who have survived septic patients show higher levels of expression than those who did not survive.⁵⁸

A medical case study highlights the connection between CD40 and its T cell ligand, CD154 or CD40L, shedding light on the immune response to infection. For macrophages to efficiently engulf bacteria, CD40 must be expressed. The patient needs to undergo this procedure to develop a response to the infection. T cells that are activated show higher levels of CD40L expression.⁵⁹ Due to the binding of CD80 and CD86 to CD40 on APCs, their expression levels are elevated. In addition, it enhances the secretion of IL-12, an important regulator of T cell differentiation and activation. Mortality rates in CD40-/- mice are lower than wild-type controls, even though septic mice have higher levels of CD40. In addition, the rodents' blood lacking CD40-/- showed a significant decrease in IL-6 levels.⁶⁰ Patients with sepsis showed increased levels of CD40 expression on mononuclear cells in their peripheral blood. There is a correlation between shock and higher levels of CD40 expression.²⁹ On the other hand, higher survival rates are linked to higher expression levels. Clearly, when mice were given an agonistic anti-CD40 antibody, it was observed that lymphocyte apoptosis decreased and overall survival improved after CLP.⁶¹

In addition, PD-L1 and PD-L2 are important members of the B7 family. Through the PD-1 T cell receptor, these individuals experience decreased energy and ultimately succumb. PD-L1 is widely expressed on various cells, including APCs, B cells, splenic T cells, and other non-hematopoietic cells. Some specific immune cells, such as peritoneal B

cells, dendritic cells, macrophages, and bone marrow-derived mast cells, can be activated to express PD-L2.^{31,32} T cells infected with the virus exhibit increased levels of PD-1.⁶² It has been observed that animals lacking the PD-1 signaling pathway show increased resistance to *Listeria monocytogenes* infection. This indicates that the signaling system may affect the host's immune response to pathogens.⁶² Although PD-1 has been studied in animal models involving bacterial infections and sepsis, there is limited research specifically focused on sepsis patients and their relationship with PD-L1 and PD-L2. When CLP is present, rodents that lack PD-1 show lower mortality rates and decreased production of inflammatory cytokines and bacterial burden in the body and abdomen.⁶³ The findings of this study come as a surprise to some extent, as the lack of a T cell-inhibitory pathway could have indicated a stronger inflammatory reaction. This is because activated T cells release IL-2, which triggers the activation of cytotoxic T cells. Despite the lack of extensive evidence, it is undeniable that CSMs play a crucial role in the body's immune response to pathogens. They greatly disturb the delicate balance between clearing infections and the excessive inflammatory response seen in sepsis.⁶⁴

Effects of sepsis on cell proliferation

In sepsis cases, mortality remains high, with approximately 40% of patients not surviving beyond 28 days, particularly when complications arise during anesthesia and critical care management. The pathophysiology of sepsis places great importance on the occurrence of individual cell death.⁶⁵ Two mechanisms contribute to the death of cells: apoptosis and necrosis. This section describes the responses of specific cell types to necrotic and apoptotic pathways. The patient's condition probably worsened due to the dysregulation of apoptosis and necrosis in inflammatory cells.⁶⁶

Cell apoptosis

Apoptosis, or "programmed cell death," is a complex process involving multiple synchronized stages. The plasma membrane's division occurs during the later stages of apoptosis. Rare instances occur where harmful substances are released from cells into the surrounding environment, as long as the plasma membranes remain undamaged.⁶⁷ Apoptotic cells display various changes in their appearance, such as the development of plasma membrane blebs, nuclear fragmentation, and chromatin condensation. Various techniques can be used to detect apoptotic cells, such as staining for terminal deoxynucleotidyl transferase, chromatin laddering, or activation of caspases.⁶⁸ Apoptosis plays a crucial role in maintaining proper development by regulating proliferation. Apoptotic mechanisms play a crucial role in removing unnecessary cells during the development of organs. Scientific research has found that over time, a significant amount of bone marrow and lymph nodes can accumulate in the body if cells are not eliminated through apoptosis.⁶⁹ In addition, apoptosis could potentially contribute to the elimination of cancer cells. As a result, any disruptions in this process might be linked to the formation of certain types of cancer.⁷⁰

Two fundamental pathways initiate apoptosis: extrinsic and intrinsic.⁷¹ Cell death occurs through the extrinsic pathway when external proteins attach to receptors on the cell surface. The process in question is called the death receptor pathway.⁷² Several molecules are involved in this particular case, including FAS, TRAIL, and TNF. Upon receptor binding, the FAS-associated death domain of the adaptor protein relocates to the inner surface of the cell membrane.⁷³ As a result, a cascade of internal mechanisms is set in motion, leading to the synthesis of more proteins that prevent cell death through a negative feedback loop. According to research, caspase-3 is commonly called the "master executioner" due to its effective performance.⁷⁴

When it comes to the intrinsic pathway of apoptosis, the mitochondria play a significant role. This method heavily depends on the presence

of γ radiation, oxygen radicals, or DNA damage. In a fragile balance, various proapoptotic proteins, such as Bim, Bax, and PUMA, exist alongside antiapoptotic proteins, like BCL-2, BCL-XL, and others.⁷⁵ Factors that influence specific proteins are linked to the onset of the apoptotic signal. For example, PUMA regulates apoptosis, a cellular response to DNA damage. When proapoptotic proteins are present, mitochondria release cytochrome c, triggering a series of events that activate caspase-9. Following the completion of the extrinsic pathway, caspase-3 is activated.⁷⁶

Cell necrosis

The second primary cause of cellular demise is necrosis. Ischemia-induced cellular ATP depletion has historically been associated with ischemia-induced necrosis. The compromised membranes of necrotic cells permit detrimental proteolytic enzymes to pass through. These enzymes are released by intracellular organelles like lysosomes into the cytoplasm or, if a plasma membrane rupture occurs, into the adjacent tissue.⁷⁷ Phagocytic cells are responsible for eliminating pathogens during the initial phases of an infection, as previously stated in the corresponding section. Tests on adoptive transfer subjects have demonstrated that apoptosis is more significant than necrosis in sepsis. Injecting necrotic cells decreased mortality rates in sick rodents, whereas administering apoptotic cells increased mortality.⁷⁸

Apoptosis role

Existing evidence suggests that apoptosis-associated cell death holds significant therapeutic significance in this particular situation, as necrosis has been observed in animal models of sepsis and endotoxemia and septic patients.⁷⁹ Apoptosis has been observed in various cell types, such as endothelial, muscle, and neuronal cells. However, sepsis causes the most severe apoptosis in lymphocytes and gastrointestinal epithelial cells.⁸⁰ The Hotchkiss group conducted complex and time-sensitive investigations on patients who had succumbed to sepsis, and they were the first to document this important discovery in human subjects. Increased cell death has been noted in lymphoid organs like the thymus and spleen and in lymphoid sections of other organs such as the large intestine. The occurrence of this condition in tissues outside of lymph nodes is relatively low.⁸¹ Furthermore, there was no conclusive evidence linking the organ injury observed in these patients to the low occurrence of apoptosis in kidney and lung epithelial cells and other organs such as the liver. Further research by the same group revealed that in the spleens of septic patients, macrophages and other T cell subsets showed very low susceptibility to apoptosis.⁸² Nevertheless, dendritic cells (including follicular and interdigitating cells), B cells, and CD4 + T lymphocyte subsets showed increased susceptibility. A similar pattern of cell death was later observed in newborn and young patients who died from sepsis.⁸² Septic patients often experience prolonged lymphopenia due to increased lymphocyte apoptosis, affecting various subpopulations of lymphocytes in circulation.⁸³ Like splenic lymphocytes, most circulating CD4 + T and B cells go through apoptosis. However, there have been findings of reduced levels of CD8 + T cells and natural killer cells.⁸⁴ It is important to note that the severity of symptoms and prognosis strongly correlated with the level of apoptosis in both studies.⁸⁵

Much data obtained from animal studies supports the previously mentioned results. Similar levels of lymphocyte apoptosis have been observed in the intestinal epithelium and lymphoid organs of the murine CLP model of polymicrobial sepsis, mirroring the findings in humans with sepsis.⁸⁶ Apoptosis can occur in various nonlymphoid sites and cell subsets, including kidney tubule cells, skeletal muscle cells, and lung alveolar, respiratory, and capillary endothelial cells, during CLP sepsis.⁸⁶ Interestingly, despite the decrease in lymphocyte levels caused by CLP during the initial stage of sepsis, several studies have found higher peripheral lymphocyte counts in mice that did not survive than those that did. However, these counts did not specify the exact ratio of cells undergoing apoptosis to those not undergoing.⁸⁷

The rate of lymphocyte apoptosis can harm neutrophils. Powerful antibacterial neutrophils are initially recruited to the infection site, where they play a vital role in containing the infectious attack. Furthermore, neutrophils can undergo apoptosis, ensuring strict control over the inflammation they generate. Extensive documentation exists on the prolonged suppression of neutrophil apoptosis in patients with sepsis.⁸⁸ It appears that extended neutrophil apoptosis may have a role in causing harm to organs and leading to death. Continuous neutrophil activation can cause significant damage to nearby cells and tissues by constantly producing harmful metabolites.⁸⁹

There are still many unanswered questions regarding apoptosis in septic patients. The range of cells or tissues that can induce apoptosis is likely much larger than currently understood. Examining the origin or qualities of septic damage may reveal new patterns in apoptotic depletion. For instance, *Listeria monocytogenes* triggers a rapid apoptosis in hepatocytes, whereas *Staphylococcus pyogenes* does not induce the same apoptosis in macrophages and neutrophils.⁹⁰ Further research in this area can be greatly improved by using a recently developed mouse model that includes hCD34 + hematopoietic cord blood stem cells. This model accurately represents the full functionality of the human innate and adaptive immune systems.⁹¹

In lymphocytes, accelerated apoptosis may significantly impact the development of sepsis. An advanced adoptive transfer experiment was conducted using septic rodents to examine the concept mentioned earlier.⁹² In addition, a significant decrease in immune function occurs during the later stages of sepsis. The primary cause of this illness is most likely the loss of lymphocytes due to apoptosis. The immunosuppressive mechanism is believed to encompass two key processes: the direct elimination of crucial effector cells or the induction of immune tolerance through apoptosis in macrophages and dendritic cells that survive.⁹³ Several antiapoptotic medications have been suggested as potential approaches to hinder or reverse these processes, as the long-term weakening of the immune system's defenses undoubtedly puts the individual at a disadvantage and increases susceptibility to future infections. Various therapeutic approaches have been extensively studied, including the inhibition of CD95, the overexpression of BCL-2, the inhibition of caspases (such as caspase-3 and caspase-8), and the use of no caspase protease inhibitors.⁹⁴ Unfortunately, trials for these experimental treatments have not yet started. The complexity of the signaling networks that regulate apoptosis may contribute to the unsuitability of these targets for therapeutic purposes.⁹⁵

Sepsis and blood coagulation

The association between sepsis-induced coagulation dysfunction and an atypical accumulation of fibrin in blood vessels is widely acknowledged. Beyond that, it remains exceedingly challenging to reach an agreement.⁹⁶ There is intense debate regarding the function of coagulopathy in sepsis, with divergent opinions regarding whether it is a causal factor or merely an observer. The inconsistent results of anticoagulant medications in clinical studies for sepsis add layer of complexity to the subject, especially when compared to their effect on 28-day all-cause mortality.⁹⁷ A week after contracting a bacterial infection, our patient developed severe sepsis and hypotension as a result of disseminated intravascular coagulation. When the body's typical coagulation process is disrupted, virtually impermeable blood clots may develop. These clots have the potential to obstruct the organ's minute blood vessels.⁹⁸ Those with protracted clotting times, low platelet counts, and low fibrinogen levels may experience a depletion of platelets and coagulation components that surpasses their restoration rate. Patients diagnosed with DIC pose a peculiar dilemma due to their propensity for thrombosis and increased risk of hemorrhaging in comparison to fit individuals. Skin petechiae are prominent indications of micro bleeding in various anatomical sites and are frequently observed in patients with DIC-induced consumption coagulopathy.⁹⁹

The resolution to this issue ought to be uncomplicated. The microvasculature was physically obstructed in this instance by the aggregates. Damage caused by ischemia and reperfusion can result in severe complications, such as failure of multiple organs and mortality.¹⁰⁰ To safeguard organ function, it is necessary to halt or prevent the formation of blood clots. Nevertheless, when confronted with practical situations, it is imperative to consider many intricate elements.¹⁰¹ Potential variables to consider include shifts in the population's demographic composition, the coexistence of supplementary medical conditions, and the justifications for the inappropriateness of certain frequently prescribed medications, such as heparin. The impact of heparin therapy on the effectiveness of an investigational medication is being investigated in two clinical trials involving patients with sepsis.¹⁰²

Anticoagulation and sepsis

Despite the difficulties, efforts are being made to prevent unnecessary blood clot formation in patients with sepsis. Various points in the coagulation cascade are frequently targeted for pharmaceutical intervention.¹⁰³ Various pharmaceutical or natural methods can be employed to achieve this goal. Certain molecules play a crucial role in the successful synthesis of fibrin. These molecules, such as tissue factor, cofactors Factor Va and VIIIa, and Factor Xa, act as checkpoints in the process. The extrinsic pathway is activated by tissue factor and is sped up by factors VIIIa and Va. Factor Xa is the coagulation enzyme required for both the extrinsic and intrinsic routes.¹⁰⁴

While studying sepsis and its effects on patients, it has been observed that using anticoagulants can lead to different outcomes. However, by conducting correlation tests, it is possible to distinguish between trauma-induced coagulopathy and infection in patients. These tests can also help predict the prognosis of the patients.¹⁰⁵ The clinic routinely conducts two clotting tests. The measurement of molecules in the tissue factor pathway is done using the prothrombin time (PT). Tissue factor (thromboplastin) from an external source is introduced into the patient's plasma to compensate for any inconsistencies in the laboratory reagents.¹⁰⁶ The coagulation time is calculated as a ratio, known as the international normalized ratio, to a standard reagent value. It is common practice to use physical therapists to monitor individuals who are taking oral anticoagulants such as warfarin. The contact activation pathway that leads to the activation of Factor Xa can be assessed through the activated partial thromboplastin time, commonly known as APTT.¹⁰⁷ Clotting time is evaluated by administering an exogenous phospholipid in the patient's plasma. APTT tests are commonly utilized to evaluate patients receiving intravenous anticoagulants, like heparin or low-molecular-weight heparin. Various assays can be utilized to measure clot disintegration, clot formation regulation, and clot creation. Only a few tests have practical value in treatment, even though numerous tests are used in research. As an example, markers such as D-dimer and fibrin degradation products (FDPs) can provide valuable insights into fibrinolysis and help assess the extent of coagulopathy.¹⁰⁸

Thrombin is the final serine protease generated by the cascade. The primary role of this protein in the coagulation process is to break down fibrinogen into two smaller peptides known as fibrinopeptides A and B, resulting in the formation of oligomerized fibrin monomers. As mentioned, thrombin has various effects including fibrinolysis, inflammation, cell proliferation, angiogenesis, and tumor metastasis. While there is potential for thrombin inhibitory drugs in sepsis, it is important to consider their significant role in inflammation through protease-activated receptors (PARs).¹⁰⁹ Therefore, it may be premature and unproductive to utilize these drugs during severe coagulopathy of sepsis. A study examined the effectiveness of hirudin, a direct thrombin inhibitor derived from the medical leech *Hirudo medicinalis*, in patients experiencing acute thrombotic events. The study involved Phase II and III clinical trials. However, the trials found that hirudin had a significant bleeding rate, no clear advantage over low-molecular-weight heparin or conventional heparin, and no noticeable impact on mortality outcomes.¹¹⁰

Researchers discovered that hirudin has been shown to decrease fibrin deposition in animal models of mixed microbial sepsis. However, it does not appear to impact organ perfusion significantly.¹¹¹ In the KyberSept trial, a significant international Phase III clinical trial aimed at assessing antithrombin therapy for patients with severe sepsis, it was found that despite the natural ability of antithrombin to slow thrombin, the use of antithrombin medication did not improve survival rates. It is worth noting that antithrombin therapy has shown potential benefits for patient subpopulations with a significant risk of mortality (30–60%), as indicated by a meta-analysis of the KyberSept data.¹¹² This discovery shows potential for future clinical research using patient classification standards.

Tissue factors

Tissue factor, also known as thromboplastin, CD142, and Factor III, is a versatile transmembrane cofactor and receptor. The body initiates the clotting process.¹¹³ Tissue factor is typically not readily available to blood components as it is absent in the bloodstream. However, it can also manifest mysteriously and become procoagulant through an unidentified process. The mechanism by which the tissue factor transitions between high and low activity levels is fascinating. This concept highlights the redox-induced disulfide bonding between two specific cysteine residues in tissue factor (Cys186 and Cys209).¹¹⁴ Nevertheless, the limited conclusive evidence regarding free thiols in tissue factor, combined with insights from crystal structure data, indicates the potential presence of other factors contributing to the identification of tissue factor activity.¹¹⁵ Recent research has uncovered a soluble tissue factor that has prothrombotic properties. Due to alternative splicing, the transmembrane domain is absent and exon 5 is not produced in this factor. During coagulation, tissue factor binds to Factor VII on cell surfaces, activating it. Consequently, an enzyme-cofactor complex is formed, leading to an increase in the production of Factor Xa. Endothelial cells and platelets produce tissue factor pathway inhibitors (TFPI), effectively suppressing this checkpoint.¹¹⁶ TFPI inhibits the tissue factor-Factor VIIa complex in the presence of Factor Xa, leading to a dysfunctional quaternary structure. Heparin can displace TFPI, which has a relatively loose connection with endothelial cells and binds to their glycocalyx layer. In animal models of bacterial sepsis and human endotoxemia challenge tests, TFPI has demonstrated potential as a treatment for reducing. Nevertheless, the effectiveness of recombinant TFPI (rTFPI) in reducing the 28-day all-cause mortality rate among patients diagnosed with severe sepsis was found to be insignificant in a coagulopathy.¹¹⁷ Phase III randomized clinical trial involving a substantial number of participants ($n = 1754$). Interestingly, individuals with a low international normalized ratio (< 1.2) experienced a notable improvement in their chances of survival. The survival rate showed a notable difference between the placebo group (22.9%) and the rTFPI group (12.0%), with the latter exhibiting a lower survival rate. The survival effect remained consistent despite accounting for therapy, baseline APACHE score, and log10 IL-6 levels. There may be some concerns about this anticoagulant's effectiveness in treating sepsis patients. There is a higher likelihood of bleeding in the central nervous, gastrointestinal, and respiratory systems in the rTFPI arm. In addition, there is a concerning interaction with heparin for medical purposes.

Va and VIIIa factors

During the coagulation process, two cofactors, factors VIIIa and Va, enhance the production of fibrin. Given the critical function these molecules play as checkpoints, nature has ingeniously devised the protein C anticoagulant pathway to inhibit both cofactors effectively. Within cellular contexts, the transportation of the protein C precursor to a preassembled complex is facilitated by the endothelium protein C receptor. This complex comprises thrombin and thrombomodulin (CD141), which act as cofactors on the membrane. This promotes the

cleavage of activated protein C's precursor, a serine proteinase that breaks down Factors Va and VIIIa through limited proteolysis. When these crucial elements are absent, the coagulation process experiences a significant slowdown, leading to a notable decrease in thrombin production. In addition, components of the protein C pathway play a beneficial role in safeguarding cells from the harmful effects of sepsis. An interesting example is the ability of activated protein C to degrade cytotoxic histones released by injured cells. An investigation in the late 1980s involving nonhuman primates showcased the efficacy of using activated protein C as a pretreatment to decrease mortality caused by high doses of the gram-negative bacteria *Escherichia coli*. Based on the PROWESS clinical study, the FDA approved recombinant human activated protein C (drotrecogin alfa, activated) in 2001 as an additional treatment for patients diagnosed with severe sepsis.¹¹⁸

The significant concern of bleeding associated with drotrecogin alfa in almost all trials dampened the initial excitement surrounding a hopeful clinical study that demonstrated a 6.1% decrease in mortality and improved long-term organ function in acutely septic patients.¹¹⁹ Furthermore, research has indicated a minimal risk of mortality associated with this treatment option. However, it is important to note that it is not effective for managing severe sepsis in adult patients. In addition, research has indicated that it does not provide positive outcomes for children.¹²⁰ Drotrecogin alfa is currently only prescribed to patients who meet specific criteria, and it is not considered a commonly used treatment option. Meeting certain criteria, such as a high APACHE score, being in a terminal condition, or having multiple organ failures are some examples of the necessary conditions. It suggests that administering anticoagulants early to critically ill patients provides a certain level of protection. A 30-nation volunteer initiative known as Surviving Sepsis aimed to provide comprehensive and evidence-based therapeutic care to patients suffering from severe sepsis and shock. The promotional material stated that individuals who were administered drotrecogin alfa within the initial 24 h of admission to the intensive care unit while in a state of shock had an increased likelihood of survival.¹²¹

Various studies, including the PROWESS experiment, have shown that patients with low levels of circulating protein C tend to experience severe illness and have a less favorable prognosis. A Phase II trial was conducted to assess the effectiveness of protein C concentrate in critically ill children with meningococemia, and the results were quite promising. The survival time of 12.3 h is similar to what has been observed in previous small-scale clinical investigations. This medication effectively increased the levels of activated protein C in the blood, partially restoring the coagulation balance without any adverse bleeding effects.¹²² The results of this investigation disproved the conventional wisdom that patients suffering from acute sepsis ought to get the active enzyme since they are incapable of activating the protein C precursor. The only available option for treating severe protein C deficiency in newborns, an uncommon autosomal recessive disorder, is CeprotinTMR, a recombinant human protein C. A recent study focused on four premature infants who were at high risk of mortality due to severe bacterial sepsis, shock, and other comorbidities. The newborns were administered antithrombin and protein C concentrate, not part of the standard critical care treatment. After 48-h treatment with protein C concentrate, the levels of circulating protein C returned to normal, the purpuric cutaneous lesions disappeared, and the newborn's multiple organ dysfunction score became stable. All four babies made a complete recovery without any complications related to bleeding, clotting, or disruption of blood flow. There is a high demand for clinical trials investigating the efficacy of human protein C concentrate in pediatric sepsis patients. This particular group of individuals has a higher likelihood of experiencing blood coagulation problems. Although recombinant activated protein C (drotrecogin alfa) was initially approved based on early trial data (PROWESS trial), subsequent larger studies failed to replicate the survival benefit. The PROWESS-SHOCK trial, a Phase III randomized controlled study, demonstrated no significant reduction in mortality, leading to the withdrawal of the drug from the market.^{123,124} Further-

more, the high risk of serious bleeding events raised safety concerns. These findings underscore the complexity of sepsis treatment and the limitations of targeting a single pathway in such a heterogeneous syndrome.

Factor Xa

Although leeches and hirudin have a significant historical background as anticoagulants, several contemporary alternatives have surfaced since the 1930s introduction of heparin. Despite the potential drawbacks, low-molecular-weight heparin (specific to Factor Xa; 1987) and vitamin K antagonists (warfarin and coumadin; early 1950s) remain widely used as anticoagulants. The mentioned side effects include the requirement for intravenous administration, medication-induced thrombocytopenia, and difficulties in determining the ideal dosage. As a result, the finding of new oral Factor Xa inhibitors is highly promising. Factor Xa plays a crucial role in coagulation, being involved in both phases. A crucial cascade element affects both the intrinsic and extrinsic coagulation pathways. By inhibiting this component, we can effectively prevent the formation of blood clots. Apixaban, a newly discovered Factor Xa inhibitor, has shown promising results in animal models and could help patients prevent complications after knee replacement surgery.¹²⁵ The anti-Factor Xa activity of BAY 59-7934 (rivaroxaban). This specific chemical belongs to a class of oxazolidinone derivatives discovered through high-throughput screening. These compounds are competitive inhibitors of Factor Xa by targeting its active site. Their oral bioavailability remains high despite their short half-life. Rivaroxaban has been extensively studied in various settings, including pre-operative evaluation in healthy individuals, animal models of thrombosis, and patients undergoing major orthopedic surgery.¹²⁶ Little is known about the effectiveness of these new medications in treating blood clotting disorders during a severe infection. A recent study has highlighted the important role of Factor Xa in signaling, particularly in the activation of PARs (PAR-1 and PAR-2). It is important to mention that this communication occurs when the coagulation activity of Factor Xa is not present. The findings of this study suggest that altering this molecule could have important implications for various physiological processes commonly seen in severe sepsis, such as acute lung injury, wound healing, and tissue regeneration.¹²⁷

Cellular biomarkers

Biomarkers are chemical entities that correlate with physiological or pathological changes. Although these compounds may not be the main cause of the illness, they suggest a biological basis. The plasma concentration of troponin, used for diagnosing acute myocardial infarction in patients, is a remarkable example of a biomarker.¹²⁸ Utilizing biomarkers can be instrumental in guiding treatment decisions and diagnosing various diseases. The discovery of biomarkers for accurate sepsis diagnosis and patient monitoring in critical illness is a topic of significant interest, given the complex nature of the septic response. In the past, sepsis management focused on evaluating physiological markers.¹²⁹

Physiology of septic patients

Currently, medical professionals in the intensive care unit (ICU) closely monitor various laboratory parameters such as platelet count and serum sodium levels. They also closely monitor physiological data like heart rate, blood pressure, and respiration rate. This is particularly important for septic patients, who comprise most ICU cases. Nevertheless, the standard metrics mentioned lack the required reliability to provide a definitive diagnosis of sepsis.¹³⁰ Furthermore, it is important to mention that patients with sepsis may show either heightened or reduced inflammation levels, even if they have noticeable physiological scores. The scoring systems commonly used in medical case studies, such as APACHE, SOFA, and PIRO, cannot fully capture patients' complex

immunoinflammatory status. As a result, several Phase III clinical trials have shown that the widespread use of therapeutic anti-inflammatory drugs can be dangerous or have no effect at all.¹³¹ Improved results can be achieved by identifying specific biomarkers, allowing for more precise targeting of therapy techniques that were previously ineffective and more selective patient selection. However, it is unlikely that the current set of physiological indicators will provide appropriate diagnostic and prognostic signals.

Immunology

Some claim that as compared to traditional clinical monitoring, immunomonitoring has no advantages. An immunoinflammatory biomarker is beneficial when it can provide information beyond what can be obtained from a typical physiological examination. This information guarantees that therapy can be beneficial before the end of the treatment time. Given the complex immunoinflammatory response associated with sepsis, it is critical to look at symptoms from a range of perspectives. Procalcitonin (PCT), C-reactive protein (CRP), and pro- and anti-inflammatory cytokines are a few examples of inflammatory biomarkers that have been studied in conjunction with human sepsis to learn more about their diagnostic utility and possible relationship to mortality.¹³² PCT has shown great promise as a diagnostic marker in distinguishing sepsis from noninfectious SIRS etiology. Because PCT has a comparable effect on septic animal survival, it has also been studied as a possible target for therapeutic therapies. However, higher death rates were the outcome of neutralizing calcitonin precursors. The use of PCT in critical care settings is the subject of ongoing research. Furthermore, several clinical trials are also being conducted to evaluate the effectiveness of PCT immunomonitoring in sepsis patients. The possible effects of daily PCT levels on patient outcomes are presently being studied. The goal of the PASS study, which is filed under the NCT00271752, is to obtain important information in this field. You can visit <http://www.clinicaltrials.gov> for additional information. Daily PCT levels may shorten the duration of antimicrobial therapy (Pro-SEPS trial; NCT01025180) and offer helpful information for treatment decisions in septic patients (SISPCT study; NCT00832039). The usefulness of IL-6 as a biomarker for sepsis is still debatable. Opinions on its function vary; some consider it to be little more than a disease marker, while others think it is essential in assessing the severity of the sickness.¹³³ Nevertheless, IL-6 strongly predicts septic complications and mortality in both animal and human models. CRP in sepsis has little predictive and diagnostic significance because of its low specificity.¹³⁴

Besides the extensive studies on humoral markers, scientists have also found that sepsis patients have significant changes in immunoinflammatory responses in cellular compartments, particularly leukocytes.¹³⁵ The goal of the research was to assess a variety of sepsis-related biochemical indicators without needless duplication. A relationship was discovered between the patient's immunological status and the monocytes' expression of the fractalkine receptor (CX3CR1) and HLA-DR. Patients' expression levels dropped when they experienced immunosuppression due to clinical sepsis.¹³⁶ Studies have demonstrated a strong association between a lower lymphocyte count in sepsis patients and a poor outcome. Studies have shown that preventing lymphocyte apoptosis in animal cases of sepsis significantly increased survival rates.¹³⁷ It has been found that sepsis outcome in hospitalized patients can be accurately predicted by measuring the proportion of neutrophils expressing CD64. Circulating endothelial progenitor cells in septic patients can give important information about the stage and severity of the illness.¹³⁸

Immunomonitoring

Biomarker-based immunomonitoring is a versatile method used in various preventive and disease scenarios. When examining sepsis, the main focus has been on prognosis and diagnosis. In addition, previ-

ous studies have been conducted to determine if biomarkers can help in the timely discontinuation of antibiotics. When evaluating the effectiveness of a biomarker for sepsis diagnosis, it is important to consider various performance indicators. These include reducing the time it takes to make a diagnosis, improving the ability to distinguish between bacterial and viral infections, determining whether inflammation is caused by an infectious or noninfectious source, and assessing the effectiveness of infection control measures.¹³⁹ The first two factors are extremely important because prompt intervention greatly enhances the outlook for patients suffering from sepsis. In the realm of medical research, there is a constant pursuit of innovative approaches. However, it is crucial to emphasize the importance of promptly administering antibiotics in the early stages of sepsis treatment. Patients suffering from septic shock experienced a significant decrease in survival rates when effective antibiotic treatment was delayed, with a decline of 8% for every additional hour of delay.¹⁴⁰ In comparison to the widely accepted blood culture and SIRS criterion, biomarkers are expected to reduce the waiting time in this critical period. While the overall goal is practical, there is a significant limitation: it only identifies sepsis, not infection. Various biomarker-based tests, both experimental and commercial, can assist in detecting bacterial infections in intensive care unit (ICU) patients. However, it is important to note that these tests do not provide information about the pathogen's susceptibility to antibiotics or identify the specific causative pathogen. Insufficient administration of antibiotics in the early stages of sepsis has been associated with a significant decrease in the chances of survival. Consequently, omitting this crucial treatment may jeopardize the well-being of patients.¹⁴¹ By integrating biomarker-based monitoring with advanced molecular techniques, like nucleic acid-based technologies that can detect specific diseases, we may be able to fill this diagnostic gap. It is possible that a biomarker could have provided early warning signs of sepsis before the patient experienced hypotension. Ideally, this would have helped with timely and suitable intervention, thus preventing the patient's deterioration into severe sepsis. It appears that biomarkers have a significantly greater ability to predict and guide treatment than diagnostic methods. In contrast to APACHE or SOFA scores, immunomonitoring effectively identified the group at high risk and the varying immune response, allowing for more personalized and targeted treatment approaches. Serial biomarker measurements have been thoroughly examined in animal models and septic patients,¹⁴² indicating they are the more cautious in this situation. Customizing treatment for septic patients based on their immunoinflammatory condition is a relatively new advancement. There is a notable connection between early sepsis mortality and an excessive hyperinflammatory response. Interestingly, the characteristic features of hyperinflammatory and hypo-inflammatory reactions in this condition are remarkably similar. A thorough analysis has found that providing certain immunomodulators to a specific group of critically ill septic patients can greatly improve their chances of survival. This approach has been utilized in clinical trials with varying levels of effectiveness: A study conducted on patients with a higher risk of mortality, as indicated by the biomarker IL-6, found that treatment with anti-TNF antibodies showed a slight improvement in survival rates. However, this advantage was observed in only one out of the two studies.¹⁴³ The lack of consensus on accurately characterising immune-inflammatory signatures in different septic patients remains a significant challenge in advancing this field. Nevertheless, sepsis in experimental animals demonstrated the successful utilization of a biomarker to guide general treatments, like glucocorticoids, and enhance survival.¹⁴⁴ Various subtypes of sepsis can result in different clinical outcomes. Sepsis is commonly diagnosed by assessing a localized infection, such as peritonitis, pneumonia, or abscesses. Regarding disease diagnosis and predicting clinical progression, it might be more beneficial to measure inflammatory markers at the site of infection rather than in plasma. According to research, local manifestations of infection occur before systemic changes. The intraperitoneal micro dialysis data noted significant elevation in lactate pyruvate ratios following an urgent laparotomy.¹⁴⁵ In this study, it was observed

that there was no correlation between peritoneal cytokine levels and plasma levels after major surgery, even though the former were higher than the latter. A highly accurate identification of pneumonia patients in the lungs was achieved by enhancing soluble triggering receptor on myeloid cells 1 expression during Mini bronchoalveolar lavage.¹⁴⁶ Although monitoring local biomarker levels can have advantages, it may not always be possible or practical. In most cases, peripheral blood is the easiest specimen to obtain for diagnostic purposes.¹⁴⁷

Prediction paradigms

Biomarkers have a great chance of being useful if they exceed current physiological scoring systems. Numerous investigations have directly compared the diagnostic accuracy of physiological scoring systems and biomarkers. The analysis entailed determining the receiver operator characteristic's area under the curve using the provided data. The biomarkers perform at a level comparable to, or even higher than, that of the scoring systems. Researchers discovered contradictory findings in a recent study¹⁴⁸ about the predictive efficacy of several biomarkers for sepsis. It was discovered that PCT performed noticeably better than CRP, and IL-6 even performed even better. However, in contrast to the previous study¹⁴⁹ there was no discernible variation in the blood plasma levels of IL-6 between patients who had SIRS following surgery and those who did not. It is interesting to note that the ratio of TNF to IL-10 was found to possess predictive significance. According to earlier studies, people with septic shock or severe sepsis had greater PCT levels than people with sepsis alone. Furthermore, research has demonstrated that IL-10 and TNF are more reliable death indicators.

Ninety-four individuals were included in a comprehensive study aimed at identifying the best reliable indicators for sepsis death prediction. To guarantee thorough results, a wide range of characteristics were explored in the study. Initial research showed that IL-6 and TNF-soluble receptor I levels were correlated with death. Subsequent analysis of several variables, including age and APACHE II, showed that the early plasma IL-6 concentrations are the sole consistent predictor of 28-day mortality.¹⁵⁰ Natriuretic peptide plasma levels and APACHE II scores in sepsis patients appear to be related, suggesting that these levels may have use as biomarkers.¹¹² A study was done to look at the relationship between hospital mortality, physiologic scores, and brain natriuretic protein (BNP) levels. Both BNP and SAPS II (the new simplified acute physiology score) were proven to be independent variables that potentially predict mortality in a hospital setting by a logistic regression analysis.¹⁵¹ The results of a study on PCT and SOFA score levels indicated that both variables had comparable predictive power. But by day three, the SOFA score correctly predicted the result, and by day six following admission, the PCT demonstrated a good predictive capacity.¹⁴⁴

Modern markers and approaches

Additional investigation is necessary to ascertain the optimal biomarkers and their efficacy in various scenarios. Recent research has put forward a range of potential new biomarkers. Pro-BNP is a reliable indicator of mortality¹⁵² and has shown a strong correlation with survival in individuals diagnosed with severe sepsis and septic shock. During the sepsis examination, two biomarkers attracted attention: high mobility group box 1 and an abnormal late rise in retin.¹⁵³ According to the analysis, high mobility group box 1 consistently showed higher plasma concentration levels than cytokines such as IL-6, IL-8, and TNF. A kinetic analysis was utilized to determine this.¹⁵⁴ This discovery highlights the significant clinical relevance of the biomarker. In addition, it has been noted that sepsis-related mortality is linked to higher plasma concentrations of tissue inhibitors, and individuals with sepsis show raised levels of matrix metalloproteinases.¹⁵⁵ There is an interesting correlation between the levels of gelsolin and inter-alpha inhibitor protein in the bloodstream and the severity of illness in septic patients. Based on a previous study, it was found that the indi-

Table 1
Sepsis registered traits.

Intervention	Trial number	Sponsor
Microcirculation guided therapy	NCT00484133	Onze Lieve Vrouwe Gasthuis
ART-123, recombinant soluble thrombomodulin	NCT00487656	Artisan Pharma, Inc.
Activated protein C with glucocorticoids	NCT00625209	University of Versailles
Methylene blue, inhaled nitric oxide, combination of both	NCT00159510	Northern State Medical University
Activated protein C	NCT00604214	Eli Lilly and Company
Rapid administration of glucose, insulin, and potassium	NCT00823108	Carolinas Healthcare System
Anti-tissue factor antibody	NCT00879606	Altos Bioscience Corporation
Thymosin α 1	NCT00711620	Sun Yat-sen University
Levosimendan versus dobutamine	NCT00093301	Wentworth Area Health Services
External cooling	NCT00527007	Assistance Publique, Hôpitaux de Paris
<i>n</i> -3 fatty acids for cognitive function	NCT00772096	University Hospital, Basel, Switzerland
Ready-to-use parenteral nutrition	NCT00798681	Fernandes Tavora Hospital
Inhaled nitric oxide	NCT00608322	National Institutes of Health
Protocol-driven hemodynamic support	NCT00335907	National Institute of General Medical Sciences
Eritoran, Toll-like receptor 4 antagonist	NCT00334828	Eisai, Inc.
Glutamine and antioxidants	NCT00133978	Kingston General Hospital
Sodium selenite guided by procalcitonin	NCT00832039	Biosyn Brahms AG
Vasoactive intestinal peptide	NCT00004494	U.S. Food and Drug Administration Office of Orphan Products
Factor V1a agonist	NCT01000649	Ferring Pharmaceuticals

Resuscitation studies

Trial numbers

Colloid (typically albumin) versus crystalloid	NCT00707122, NCT00318942, NCT00327704
Hydroxy-ethyl starch	NCT00962156, NCT00464204, NCT00273728
Antibiotic studies	Trial number
Adjusted antibiotic dosing in patients with renal failure	NCT00816790
Meropenem	NCT00534287
Renal dialysis and antibiotics	NCT00451373
Cotrimoxazole versus vancomycin for MRSA	NCT00427076
Duration of antibiotics for peritonitis	NCT00657566
Daptomycin versus vancomycin for MRSA	NCT00770341
Liposome encapsulated amphotericin B	NCT00697944
Azithromycin (macrolide antibiotic) for early therapy	NCT00708799
30-min versus 3-h infusion of meropenem (antibiotic)	NCT00891423

cator showed a high level of accuracy (area under the curve of 0.89) in differentiating between sepsis and noninfectious SIRS in critically ill patients. This presents a significant difference compared to a previous indicator. Plasma concentrations of selenium or selenoprotein P can be valuable prognostic markers in critically ill patients.¹⁵⁶ It suggests that physiological activity may not be necessary for biomarkers to provide valuable data. Considering the wide range of septic conditions, conducting medical case studies in a controlled setting can reveal new and promising indicators, enhancing the accuracy of sepsis diagnosis and prediction. There is a possibility of sepsis occurring in individuals who have undergone multiple traumas without any traumatic brain injury. It is important to consider the levels of Pro-C-type natriuretic peptide¹⁵⁷ in this case. When examining newborn sepsis, it was discovered that serum amyloid A proved to be more reliable and accurate in both diagnosis and monitoring, surpassing PCT and CRP. The timely identification of inflammation proved extremely beneficial as it led to a faster diagnosis of the condition.¹⁵⁸

The challenges in evaluating a patient's severity during the initial phases of goal-directed therapy are elucidated.¹⁵⁹ This study revealed a concerning trend where the severity of sepsis was often underestimated in emergency rooms. Using SIRS criteria alone is not enough to assess the progression of non-life-threatening illnesses, such as community-acquired pneumonia, to severe sepsis, septic shock, and mortality. To address these limitations, researchers¹⁶⁰ have utilized advanced data

analysis techniques, such as artificial neural networks, to accurately identify the specific group of septic patients facing the highest mortality risk shortly after admission to the ER. A team of researchers conducted a study using mathematical models to predict their patients' progress. A comprehensive study used a Monte Carlo microsimulation model to predict the sequential SOFA scores, in-hospital death rates, and hospital discharges for sepsis patients.¹⁶¹ These findings highlight the significance of the duration of the illness in forecasting the outcome of sepsis.

Modern therapeutic measures

Various pathogenic factors of sepsis have been discovered, resulting in the development of several treatment options to improve survival rates. In February 2010, the sepsis trials listed on <http://www.clinicaltrials.gov> are displayed in Table 1. Efforts were made to counteract substances that trigger an exaggerated inflammatory response. Monoclonal antibodies specifically targeted TNF, IL-1 receptors, and endotoxin antibodies. However, despite these efforts, there was no noticeable improvement in survival. These results highlight the intricate and interconnected nature of the immune system's innate defenses in cases of sepsis. Prior attempts to reduce inflammation with high-dose steroids have not improved survival.¹⁶² Decreasing the number of steroids administered, specifically 200 mg of hydrocortisone per day, did not have any impact on the mortality rate. It is used as an addi-

tional treatment for sepsis because it helps speed up the recovery from shock. A growing array of innovative treatments, such as a synthetic antagonist of Toll-like receptor 4, focus on addressing inflammation in its early stages. A recent analysis¹⁶³ found that this drug's Phase II clinical trial showed no significant decrease in mortality. It is worth mentioning that the group that received the highest dose had a lower fatality rate. Activated protein C is an FDA-approved treatment for sepsis. It is designed to target the microthrombosis that occurs during the disease specifically. Furthermore, its anti-inflammatory properties have been widely acknowledged.¹⁶⁴ However, currently, there is insufficient evidence to support this assertion. For more information about this treatment, please refer to the section on coagulation and peritonitis. EGDt does not focus on the microbiological and inflammatory pathophysiology of sepsis. Currently, EGDt is part of the Surviving Sepsis recommendations.¹⁶⁵ Extensive clinical trials have shown its effectiveness in reducing mortality rates in severe cases of sepsis and septic shock.¹⁶⁶ These recommendations emphasise the importance of providing timely and thorough supportive care to septic patients. These recommendations aim to effectively manage hyperglycemia, minimize the risk of barotrauma associated with mechanical ventilation, address infections, and optimize organ perfusion. Alternative therapeutic methods, like the use of low-dose steroids or activated protein C, may be considered if deemed necessary. The study found that higher adherence to the recommended criteria is associated with lower death rates. This correlation was observed across multiple medical centers.¹⁶⁷

CRedit authorship contribution statement

All authors contribute equally and approved final version of manuscript to publish.

Conflicts of interest

The authors declare no conflict of interest.

References

- Sikora JP, Karawani J, Sobczak J. Neutrophils and the systemic inflammatory response syndrome (SIRS). *Int J Mol Sci*. 2023;24:13469.
- Schertz AR, Lenoir KM, Bertoni AG, Levine BJ, Mongraw-Chaffin M, Thomas KW. Sepsis prediction model for determining sepsis vs SIRS, qSOFA, and SOFA. *JAMA Network Open*. 2023;6:e2329729.
- Coggins S, Harris MC, Grundmeier R, Kalb E, Nawab U, Srinivasan L. f pediatric systemic inflammatory response syndrome and organ dysfunction criteria in late-onset sepsis in a quaternary neonatal intensive care unit: a case-control study. *J Pediatr*. 2020;219, 133-9.e1.
- Chae BR, Kim YJ, Lee YS. Prognostic accuracy of the sequential organ failure assessment (SOFA) and quick SOFA for mortality in cancer patients with sepsis defined by systemic inflammatory response syndrome (SIRS). *Support Care Cancer*. 2020;28:653-659.
- Prasad PA, Fang MC, Abe-Jones Y, Calfee CS, Matthay MA, Kangelaris KN. Time to recognition of sepsis in the emergency department using electronic health record data: a comparative analysis of SIRS, SOFA, and Qsofa. *Crit Care Med*. 2020;48:200.
- Sparks R, Harada A, Chavada R, Trethewy C. Comparison of different sepsis scoring systems and pathways: qSOFA, SIRS, Shapiro criteria and CEC SEPSIS KILLS pathway in bacteraemic and non-bacteraemic patients presenting to the emergency department. *BMC Infect Dis*. 2022;22:76.
- Lind ML, Phipps AI, Mooney S, et al. Predictive value of 3 clinical criteria for sepsis (quick sequential organ failure assessment, systemic inflammatory response syndrome, and national early warning score) with respect to short-term mortality in allogeneic hematopoietic cell transplant recipients with suspected infections. *Clin Infect Dis*. 2021;72:1220-1229.
- Amin F, Hassan N, Bashir K, et al. Antimicrobial susceptibility profile of various bacteria isolated from respiratory tract infection. *Bull Biol All Sci Res*. 2023;2023:48.
- Din S, Fazal M, Ishtiaque A, Ullah A. Antimicrobial activity of Lantana camara against *Pseudomonas aeruginosa*, *Serratia marcescens* and *Staphylococcus aureus* to develop ointment based therapy. *Bull Biol All Sci Res*. 2023;2023:33.
- Hassan N, Amin F, Bashir K, et al. Antiviral response of drugs used against hbv patients of Khyber Pakhtunkhwa, Pakistan. *Bull Biol All Sci Res*. 2023;2023:49.
- Oduncu AF, Kiyan GS, Yalcinli S. Comparison of Qsofa, SIRS, and NEWS scoring systems for diagnosis, mortality, and morbidity of sepsis in emergency department. *Am J Emerg Med*. 2021;48:54-59.
- Mignot-Evers L, Raaijmakers V, Buunk G, et al. Comparison of SIRS criteria and qSOFA score for identifying culture-positive sepsis in the emergency department: a prospective cross-sectional multicentre study. *BMJ Open*. 2021;11, e041024.
- Arina P, Singer M. Pathophysiology of sepsis. *Curr Opin Anesthesiol*. 2021;34:77-84.
- Wasyluk W, Zwolak A. Metabolic alterations in sepsis. *J Clin Med*. 2021;10:2412.
- Jarczszak D, Kluge S, Nierhaus A. Sepsis – pathophysiology and therapeutic concepts. *Front Med*. 2021;8:609.
- Habimana R, Choi I, Cho HJ, Kim D, Lee K, Jeong I. Sepsis-induced cardiac dysfunction: a review of pathophysiology. *Acute Crit Care*. 2020;35:57-66.
- Nedeva C. Inflammation and cell death of the innate and adaptive immune system during sepsis. *Biomolecules*. 2021;11:1011.
- Iqbal U, Bashir K, Khan M, et al. Cross-sectional study of covid-19 patients and their inflammatory markers in tertiary care hospitals of Peshawar, Pakistan. *Bull Biol All Sci Res*. 2021;2021:31.
- Ullah I, Ullah A, Rehman S, et al. Prevalence and risk factors of *Helicobacter pylori* infection among individuals with tobacco consumption habits in district Peshawar: a cross-sectional study. *Bull Biol All Sci Res*. 2023;2023:42.
- Sheema, Bashir K, Fiaz S, et al. Molecular identification of HCV genotypes among injecting drug users having HCV and HIV co-infection. *Bull Biol All Sci Res*. 2024;2024:71.
- Ullah W, Ullah A, Khan M, et al. Microbial profile and nutritional evaluation of broiler and domestic chicken meat from selected districts of Khyber Pakhtunkhwa, Pakistan. *Bull Biol All Sci Res*. 2023;2023:34.
- Catarina AV, Branchini G, Bettoni L, De Oliveira JR, Nunes FB. Sepsis-associated encephalopathy: from pathophysiology to progress in experimental studies. *Mol Neurobiol*. 2021;58:2770-2779.
- Sygitowicz G, Sitkiewicz D. Molecular mechanisms of organ damage in sepsis: an overview. *Braz J Infect Dis*. 2021;24:552-560.
- van der Slikke EC, An AY, Hancock RE, Bouma HR. Exploring the pathophysiology of post-sepsis syndrome to identify therapeutic opportunities. *EBioMedicine*. 2020;61:103044.
- Schlapbach LJ, Kisson N, Alhawsawi A, et al. *World Sepsis Day: a global agenda to target a leading cause of morbidity and mortality*. Bethesda, MD: American Physiological Society; 2020:L518-L522.
- McBride MA, Owen AM, Stothers CL, et al. The metabolic basis of immune dysfunction following sepsis and trauma. *Front Immunol*. 2020;11:1043.
- Li J, Li M, Li L, Ma J, Yao C, Yao S. Hydrogen sulfide attenuates ferroptosis and stimulates autophagy by blocking mTOR signaling in sepsis-induced acute lung injury. *Mol Immunol*. 2022;141:318-327.
- Yang H, Zhang Z. Sepsis-induced myocardial dysfunction: the role of mitochondrial dysfunction. *Inflamm Res*. 2021;70:379-387.
- Kashiouris MG, L'Heureux M, Cable CA, Fisher BJ, Leichtle SW, Fowler AA. The emerging role of vitamin C as a treatment for sepsis. *Nutrients*. 2020;12:292.
- Pérez-Hernández EG, Delgado-Coello B, Luna-Reyes I, Mas-Oliva J. New insights into lipopolysaccharide inactivation mechanisms in sepsis. *Biomed Pharmacother*. 2021;141:111890.
- Hortová-Kohoutková M, Tidu F, De Zuani M, Šrámek V, Helán M, Fric J. Phagocytosis-inflammation crosstalk in sepsis: new avenues for therapeutic intervention. *Shock*. 2020;54:606-614.
- Gu M, Mei XL, Zhao YN. Sepsis and cerebral dysfunction: BBB damage, neuroinflammation, oxidative stress, apoptosis and autophagy as key mediators and the potential therapeutic approaches. *Neurotox Res*. 2021;39:489-503.
- Gabarin RS, Li M, Zimmer PA, Marshall JC, Li Y, Zhang H. Intracellular and extracellular lipopolysaccharide signaling in sepsis: avenues for novel therapeutic strategies. *J Innate Immun*. 2021;13:323-332.
- Vanderhaeghen T, Vandewalle J, Libert C. Hypoxia-inducible factors in metabolic reprogramming during sepsis. *FEBS J*. 2020;287:1478-1495.
- Lin H, Wang W, Lee M, Meng Q, Ren H. Current status of septic cardiomyopathy: basic science and clinical progress. *Front Pharmacol*. 2020;11:210.
- Fernández-Sarmiento J, Salazar-Peláez LM, Carcillo JA. The endothelial glycocalyx: a fundamental determinant of vascular permeability in sepsis. *Pediatr Crit Care Med*. 2020;21:e291.
- Gao Q, Hernandes MS. Sepsis-associated encephalopathy and blood-brain barrier dysfunction. *Inflammation*. 2021;44:2143-2150.
- Sun B, Luan C, Guo L, Zhang B, Liu Y. Low expression of microRNA-328 can predict sepsis and alleviate sepsis-induced cardiac dysfunction and inflammatory response. *Braz J Med Biol Res*. 2020;53:e9501.
- Gaudino SJ, Kumar P. Cross-talk between antigen presenting cells and T cells impacts intestinal homeostasis, bacterial infections, and tumorigenesis. *Front Immunol*. 2019;10:360.
- Kaufmann SH, Schaible UE. Antigen presentation and recognition in bacterial infections. *Curr Opin Immunol*. 2005;17:79-87.
- Wardell CM, MacDonald KN, Levings MK, Cook L. Cross talk between human regulatory T cells and antigen-presenting cells: lessons for clinical applications. *Eur J Immunol*. 2021;51:27-38.
- Pai S, Muruganandah V, Kupz A. What lies beneath the airway mucosal barrier? Throwing the spotlight on antigen-presenting cell function in the lower respiratory tract. *Clin Transl Immunol*. 2020;9:e1158.
- Muntjewerff EM, Meesters LD, Van den Bogaart G. Antigen cross-presentation by macrophages. *Front Immunol*. 2020;11:1276.
- Wang F, Ullah A, Fan X, et al. Delivery of nanoparticle antigens to antigen-presenting cells: from extracellular specific targeting to intracellular responsive presentation. *J Control Release*. 2021;333:107-128.
- Atitey K, Anchang B. Mathematical modeling of proliferative immune response initiated by interactions between classical antigen-presenting cells under joint antagonistic IL-2 and IL-4 signaling. *Front Mol Biosci*. 2022;9:777390.
- Pejic A, Andjelkovic Z, Marjanovic D, et al. Comparative analysis of antigen-presenting cells in gingival tissues in healthy and periodontitis patients. *J Clin Pathol*. 2024;77:702-708.

47. Bellini R, Bonacina F, Norata GD. Crosstalk between dendritic cells and T lymphocytes during atherogenesis: focus on antigen presentation and break of tolerance. *Front Cardiovasc Med*. 2022;9, 934314.
48. Farzi R, Aghbash PS, Eslami N, et al. The role of antigen-presenting cells in the pathogenesis of COVID-19. *Pathol Res Pract*. 2022;233, 153848.
49. Ashaolu TJ. Immune boosting functional foods and their mechanisms: a critical evaluation of probiotics and prebiotics. *Biomed Pharmacother*. 2020;130, 110625.
50. Xiao Z, Deng Q, Zhou W, Zhang Y. Immune activities of polysaccharides isolated from *Lycium barbarum* L. What do we know so far? *Pharmacol Ther*. 2022;229, 107921.
51. Shaikat A, Hanif S, Shaikat I, et al. Upregulated-gene expression of pro-inflammatory cytokines, oxidative stress and apoptotic markers through inflammatory, oxidative and apoptosis mediated signaling pathways in Bovine Pneumonia. *Microb Pathog*. 2021;155, 104935.
52. Martinez GJ, Appleton M, Kipp ZA, Loria AS, Min B, Hinds TD Jr. Glucocorticoids, their uses, sexual dimorphisms, and diseases: new concepts, mechanisms, and discoveries. *Physiol Rev*. 2024;104:473–532.
53. Li M, Zhang R, Li J, Li J. The role of C-type lectin receptor signaling in the intestinal microbiota–inflammation–cancer axis. *Front Immunol*. 2022;13, 894445.
54. Grobler C, van Tongeren M, Gettemans J, Kell DB, Pretorius E. Alzheimer's disease: a systems view provides a unifying explanation of its development. *J Alzheimers Dis*. 2023;91:43–70.
55. Jose AM, Rasool M. Choline kinase: an underappreciated rheumatoid arthritis therapeutic target. *Life Sci*. 2022;121031.
56. Bikomeye JC, Beyer AM, Kwarteng JL, Beyer KM. Greenspace, inflammation, cardiovascular health, and cancer: a review and conceptual framework for greenspace in cardio-oncology research. *Int J Environ Res Public Health*. 2022;19:2426.
57. Chang SK, Alasalvar C. Improvement of the immune system by dietary supplements and natural products. Dietary supplements with antioxidant activity: understanding mechanisms and potential health benefits; 2023:226.
58. Sausen D, Poirier M, Spiers L, Smith E. Mechanisms of T cell evasion by Epstein-Barr virus and implications for tumor survival. *Front Immunol*. 2023;14:1289313.
59. Straub RH. Chronic immune system activation. Early trauma as the origin of chronic inflammation: a psychoneuroimmunological perspective. Springer; 2023:135–231.
60. Goh M, Joy C, Gillespie AN, Soh QR, He F, Sung V. Asymptomatic viruses detectable in saliva in the first year of life: a narrative review. *Pediatr Res*. 2023;95:508–531.
61. Tenchov R, Sasso JM, Wang X, Zhou QA. Antiaging strategies and remedies: a landscape of research progress and promise. *ACS Chem Neurosci*. 2024;15:408–446.
62. Londt RS. Development of an autologous human dendritic cell vaccine against *Mycobacterium tuberculosis* in patients with extensively drug-resistant tuberculosis; 2022.
63. Hung PHS. Study on the intestinal conditions of chicken orally administered with *Lactobacillus acidophilus* strain L-55 under the parasite infection or virus vaccination; 2021.
64. Simpson C. The relationship between circadian rhythms and neurodegenerative disease; 2022.
65. Balato A, Zink A, Babino G, et al. The impact of psoriasis and atopic dermatitis on quality of life: a literature research on biomarkers. *Life*. 2022;12:2026.
66. Houghton CA. The rationale for sulforaphane favourably influencing gut homeostasis and gut–organ dysfunction: a clinician's hypothesis. *Int J Mol Sci*. 2023;24:13448.
67. Ilesanmi-Oyelere BL. The role of dietary patterns, inflammatory status and gut microbiome in bone health maintenance of postmenopausal women: a cross-sectional study: a thesis presented in fulfilment of the requirements for the degree of Doctor of Philosophy in Nutritional Science, School of Health Sciences. Palmerston North: Massey University; 2020.
68. Monzón-Atienza L, Bravo J, Serradell A, Montero D, Gómez-Mercader A, Acosta F. Current status of probiotics in European sea bass aquaculture as one important Mediterranean and Atlantic commercial species: a review. *Animals*. 2023;13:2369.
69. Kumari P, Sharma S. ACE2: a double-edged sword against SARS CoV-2 associated cardiovascular complications and endothelial dysfunction. Research and Scientific Innovation Society (RSIS International); 2021.
70. Henriquez FL, Mooney R, Bandel T, et al. Paradigms of protist/bacteria symbioses affecting human health: *Acanthamoeba* species and *Trichomonas vaginalis*. *Front Microbiol*. 2021;11:616213.
71. Kim JB, Prunicki M, Haddad F, et al. Cumulative lifetime burden of cardiovascular disease from early exposure to air pollution. *J Am Heart Assoc*. 2020;9:e014944.
72. Spence T. Associations between maternal diet, markers of inflammation and oxidative stress in pregnancy: implications for maternal health and child outcomes. Ulster University; 2021.
73. Sibanda M. Evaluation of the therapeutic potential of Green rooibos (*Aspalathus linearis*) extract in neurological disease. Stellenbosch: Stellenbosch University; 2021.
74. Khurana N. Vascular pathophysiology characterization in chronic rhinosinusitis for controlled drug delivery. The University of Utah; 2021.
75. Roy Hierro DI. Development of a closed-loop for measuring and stimulating peripheral nervous system; 2022.
76. Jeddi M. Identification of a novel method for differentiating human monocytic cell line into macrophages. London Metropolitan University; 2020.
77. Basile DP, Sreedharan R, Basu RK, Van Why SK. Pathogenesis of acute kidney injury. *Pediatric nephrology*. Springer; 2022:1555–1592.
78. Belov Kirdajova D, Kriska J, Tureckova J, Anderova M. Ischemia-triggered glutamate excitotoxicity from the perspective of glial cells. *Front Cell Neurosci*. 2020;14:51.
79. Bedoui S, Herold MJ, Strasser A. Emerging connectivity of programmed cell death pathways and its physiological implications. *Nat Rev Mol Cell Biol*. 2020;21:678–695.
80. Kist M, Vucic D. Cell death pathways: intricate connections and disease implications. *EMBO J*. 2021;40:e106700.
81. Chen Y, Luo X, Xu B, Bao X, Jia H, Yu B. Oxidative stress-mediated programmed cell death: a potential therapy target for atherosclerosis. *Cardiovasc Drugs Ther*. 2022;38:819–832.
82. Yuan C, Ma Z, Xie J, et al. The role of cell death in SARS-CoV-2 infection. *Signal Transduct Target Ther*. 2023;8:357.
83. Tuo Q, Zhang S, Lei P. Mechanisms of neuronal cell death in ischemic stroke and their therapeutic implications. *Med Res Rev*. 2022;42:259–305.
84. Liu X, Miao M, Sun J, Wu J, Qin X. PANoptosis: a potential new target for programmed cell death in breast cancer treatment and prognosis. *Apoptosis*. 2023;29:277–288.
85. Cui J, Zhao S, Li Y, et al. Regulated cell death: discovery, features and implications for neurodegenerative diseases. *Cell Commun Signal*. 2021;19:1–29.
86. Li Z, Li D, Chen R, Gao S, Xu Z, Li N. Cell death regulation: a new way for natural products to treat osteoporosis. *Pharmacol Res*. 2023;187, 106635.
87. Verdile N, Pasquariello R, Scolari M, Scirè G, Brevini TA, Gandolfi F. A detailed study of rainbow trout (*Onchorhynchus mykiss*) intestine revealed that digestive and absorptive functions are not linearly distributed along its length. *Animals*. 2020;10:745.
88. Berger RM, Weck JM, Kempe SM, et al. Nanoscale FasL organization on DNA origami to decipher apoptosis signal activation in cells. *Small*. 2021;17, 2101678.
89. Mirzayans R, Murray D. Do TUNEL and other apoptosis assays detect cell death in preclinical studies? *Int J Mol Sci*. 2020;21:9090.
90. Alhoshani A, Alatawi FO, Al-Anazi FE, et al. BCL-2 inhibitor venetoclax induces autophagy-associated cell death, cell cycle arrest, and apoptosis in human breast cancer cells. *Onco Targets Ther*. 2022;13:13357–13707.
91. Perugini CA, Kaneko N, Maehara T, et al. CD4+ and CD8+ cytotoxic T lymphocytes may induce mesenchymal cell apoptosis in IgG4-related disease. *J Allergy Clin Immunol*. 2021;147: 368–382.
92. Silva EE, Skon-Hegg C, Badovinac VP, Griffith TS. The calm after the storm: implications of sepsis immunoparalysis on host immunity. *J Immunol*. 2023;211:711–719.
93. He X, Hong W, Yang J, et al. Spontaneous apoptosis of cells in therapeutic stem cell preparation exert immunomodulatory effects through release of phosphatidylserine. *Signal Transduct Target Ther*. 2021;6:270.
94. Ibrahim SA, Kulshrestha A, Katara GK, Riehl V, Sahoo M, Beaman KD. Cancer-associated V-ATPase induces delayed apoptosis of protumorigenic neutrophils. *Mol Oncol*. 2020;14:590–610.
95. Bessou M, Lopez J, Gadet R, et al. The apoptosis inhibitor Bcl-xL controls breast cancer cell migration through mitochondria-dependent reactive oxygen species production. *Oncogene*. 2020;39:3056–3074.
96. Bray MA, Sartain SE, Gollamudi J, Rumbaut RE. Microvascular thrombosis: experimental and clinical implications. *Transl Res*. 2020;225:105–130.
97. Silva Andrade B, Siqueira S, de Assis Soares WR, et al. Long-COVID and post-COVID health complications: an up-to-date review on clinical conditions and their possible molecular mechanisms. *Viruses*. 2021;13:700.
98. Bellomo R, Ronco C, Mehta RL, et al. Acute kidney injury in the ICU: from injury to recovery: reports from the 5th Paris International Conference. *Ann Intens Care*. 2017;7:1–40.
99. Xu H, Sheng S, Luo W, Xu X, Zhang Z. Acute respiratory distress syndrome heterogeneity and the septic ARDS subgroup. *Front Immunol*. 2023;14:1277161.
100. Bhargavi Sindhuja N. A study on hyperuricemia as an early marker for severity of illness in sepsis in IMCU of a Tertiary Care Centre. Vellore: Government Vellore Medical College; 2020.
101. Priyanka B. A study on correlation between microalbuminuria and Sofa Score and Apache II Score as a marker of sepsis and treatment efficacy in patients admitted to a medical intensive care unit of a tertiary health care centre. Chennai: Madras Medical College; 2022.
102. Dabiru VA, Müller L, Schönborn L, Greinacher A. Vaccine-induced immune thrombocytopenia and thrombosis (VITT) – insights from clinical cases, in vitro studies and murine models. *J Clin Med*. 2023;12:6126.
103. Lopes-Pires ME, Frade-Guanaes JO, Quinlan GJ. Clotting dysfunction in sepsis: a role for ROS and potential for therapeutic intervention. *Antioxidants*. 2021;11:88.
104. Unar A, Bertolino L, Patauner F, Gallo R, Durante-Mangoni E. Decoding sepsis-induced disseminated intravascular coagulation: a comprehensive review of existing and emerging therapies. *J Clin Med*. 2023;12:6128.
105. Neuenfeldt FS, Weigand MA, Fischer D. Coagulopathies in intensive care medicine: balancing act between thrombosis and bleeding. *J Clin Med*. 2021;10:5369.
106. Papageorgiou C, Jourdi G, Adjambri E, et al. Disseminated intravascular coagulation: an update on pathogenesis, diagnosis, and therapeutic strategies. *Clin Appl Thromb Hemost*. 2018;24(Suppl.):8S–28S.
107. Unar A, Bertolino L, Patauner F, Gallo R, Durante-Mangoni E. Pathophysiology of disseminated intravascular coagulation in sepsis: a clinically focused overview. *Cells*. 2023;12:2120.
108. Jacobi J. The pathophysiology of sepsis – 2021 update: part 1, immunology and coagulopathy leading to endothelial injury. *Am J Health Syst Pharm*. 2022;79:329–337.
109. Heuberger DM, Schuepbach RA. Protease-activated receptors (PARs): mechanisms of action and potential therapeutic modulators in PAR-driven inflammatory diseases. *Thromb J*. 2019;17:1–24.
110. Vieceli Dalla Segà F, Fortini F, Licastro D, et al. Serum from COVID-19 patients promotes endothelial cell dysfunction through protease-activated receptor 2. *Inflamm Res*. 2023;73:117–130.
111. Hamilton JR, Frauman AG, Cocks TM. Increased expression of protease-activated receptor-2 (PAR2) and PAR4 in human coronary artery by inflammatory stimuli unveils endothelium-dependent relaxations to PAR2 and PAR4 agonists. *Circ Res*. 2001;89:92–98.

112. Rommel MG, Milde C, Eberle R, Schulze H, Modlich U. Endothelial-platelet interactions in influenza-induced pneumonia: a potential therapeutic target. *Anat Histol Embryol.* 2020;49:606–619.
113. Emekli N. Can tissue factor, a multifactorial molecule of the hemostasis, be used as a biomarker for thrombosis, inflammation and cancer? *ACTA Pharm Sci.* 2017;55, <http://dx.doi.org/10.23893/1307-2080.APS.05521>.
114. Krudysz-Amblo J, Mann KG, Butenas S. Tissue factor structure and coagulation. In: Ercan E, Ece G, eds. *Thrombosis and inflammation in acute coronary syndromes*. Sharjah: Bentham Science Publishers; 2015:23–57.
115. Eilertsen KE, Østerud B. Tissue factor: (patho) physiology and cellular biology. *Blood Coagul Fibrinol.* 2004;15:521–538.
116. Araldi RP, Prezoto BC, Gonzaga V, et al. Advanced cell therapy with low tissue factor loaded product NestaCell® does not confer thrombogenic risk for critically ill COVID-19 heparin-treated patients. *Biomed Pharmacother.* 2022;149, 112920.
117. Cimmino G, Cirillo P. Tissue factor: newer concepts in thrombosis and its role beyond thrombosis and hemostasis. *Cardiovasc Diagn Ther.* 2018;8:581.
118. Tracy PB, Peterson JM, Nesheim ME, McDuffie FC, Mann KG. Interaction of coagulation factor V and factor Va with platelets. *J Biol Chem.* 1979;254:10354–10361.
119. Bos MH, Camire RM. Blood coagulation factors V and VIII: molecular mechanisms of procofactor activation. *J Coagul Disord.* 2010;2:19.
120. Salem HH, Broze GJ, Miletich JP, Majerus PW. Human coagulation factor Va is a cofactor for the activation of protein C. *Proc Natl Acad Sci.* 1983;80:1584–1588.
121. Kalafatis M, Egan JO, van't Veer C, Cawthern KM, Mann KG. The regulation of clotting factors. *Crit Rev Eukaryot Gene Expr.* 1997;7:241–280.
122. Camire R, Bos M. The molecular basis of factor V and VIII procofactor activation. *J Thromb Haemost.* 2009;7:1951–1961.
123. Silva E, de Figueiredo LF, Colombari F. Prowess-shock trial: a protocol overview and perspectives. *Shock.* 2010;34(Suppl. 1):48–53.
124. Lai PS, Thompson BT. Why activated protein C was not successful in severe sepsis and septic shock: are we still tilting at windmills? *Curr Infect Dis Rep.* 2013;15:407–412.
125. Tang SC, Lai KN. *History and development of anticoagulation. dialysis: history, development and promise*. World Scientific; 2012:223–231.
126. Singh SK, Rajoria K. Medical leech therapy in Ayurveda and biomedicine – a review. *J Ayurveda Integr Med.* 2020;11:554–564.
127. Beshay JE, Morgan H, Madden C, Yu W, Sarode R. Emergency reversal of anticoagulation and antiplatelet therapies in neurosurgical patients: a review. *J Neurosurg.* 2010;112:307–318.
128. Núñez-Navarro NE, Santana FM, Parra LP, Zacconi FC. Surfing the blood coagulation cascade: insight into the vital factor Xa. *Curr Med Chem.* 2019;26:3175–3200.
129. Cheng S, Tu M, Liu H, Zhao G, Du M. Food-derived antithrombotic peptides: preparation, identification, and interactions with thrombin. *Crit Rev Food Sci Nutr.* 2019;59(Suppl. 1):S81–S95.
130. Reynolds HN, Haupt MT, Thill-Baharozian MC, Carlson RW. Impact of critical care physician staffing on patients with septic shock in a university hospital medical intensive care unit. *JAMA.* 1988;260:3446–3450.
131. Brun-Buisson C, Doyon F, Carlet J, et al. Incidence, risk factors, and outcome of severe sepsis and septic shock in adults: a multicenter prospective study in intensive care units. *JAMA.* 1995;274:968–974.
132. Sakr Y, Jaschinski U, Wittebole X, et al. *Sepsis in intensive care unit patients: worldwide data from the intensive care over nations audit*. US: Oxford University Press; 2018.
133. Marshall JC, Bosco L, Adhikari NK, et al. What is an intensive care unit? A report of the task force of the World Federation of Societies of Intensive and Critical Care Medicine. *J Crit Care.* 2017;37:270–276.
134. Azabou E, Magalhaes E, Braconnier A, et al. Early standard electroencephalogram abnormalities predict mortality in septic intensive care unit patients. *PLoS One.* 2015;10, e0139969.
135. Schoenberg M, Weiss M, Radermacher P. Outcome of patients with sepsis and septic shock after ICU treatment. *Langenbecks Arch Surg.* 1998;383:44–48.
136. Giuliano KK. Physiological monitoring for critically ill patients: testing a predictive model for the early detection of sepsis. *Am J Crit Care.* 2007;16:122–130.
137. Inoue S, Hatakeyama J, Kondo Y, et al. Post-intensive care syndrome: its pathophysiology, prevention, and future directions. *Acute Med Surg.* 2019;6:233–246.
138. Degoricija V, Sharma M, Legac A, Gradiser M, Sefer S, Vucicevic Z. Survival analysis of 314 episodes of sepsis in medical intensive care unit in university hospital: impact of intensive care unit performance and antimicrobial therapy. *Croat Med J.* 2006;47:385–397.
139. Hu-Lieskovan S, Bhaumik S, Dhodapkar K, et al. SITC cancer immunotherapy resource document: a compass in the land of biomarker discovery. *J Immunother Cancer.* 2020;8:e000705.
140. Maecker HT, Lindstrom TM, Robinson WH, et al. New tools for classification and monitoring of autoimmune diseases. *Nat Rev Rheumatol.* 2012;8:317–328.
141. Anayyat U, Ahad F, Muluh TA, et al. Immunotherapy: constructive approach for breast cancer treatment. *Breast Cancer Targets Ther.* 2023;15:925–951.
142. DePriest BP, Vieira N, Bidgoli A, Paczesny S. An overview of multiplexed analyses of CAR T-cell therapies: insights and potential. *Expert Rev Proteom.* 2021;18:767–780.
143. Kaur S, Alley SC, Szapacs M, et al. 2021 white paper on recent issues in bioanalysis: mass spec of proteins, extracellular vesicles, CRISPR, chiral assays, oligos; nanomedicines bioanalysis; ICH M10 section 7.1; non-liquid & rare matrices; regulatory inputs (part 1A – recommendations on endogenous compounds, small molecules, complex methods, regulated mass spec of large molecules, small molecule, PoC & part 1B – regulatory agencies' inputs on bioanalysis, biomarkers, immunogenicity, gene & cell therapy and vaccine. *Bioanalysis.* 2022;14:505–580.
144. Desaulniers D, Vasseur P, Jacobs A, Aguila MC, Ertych N, Jacobs MN. Integration of epigenetic mechanisms into non-genotoxic carcinogenicity hazard assessment: focus on DNA methylation and histone modifications. *Int J Mol Sci.* 2021;22:10969.
145. Tran T, Blanc C, Granier C, Saldmann A, Tanchot C, Tartour E. *Therapeutic cancer vaccine: building the future from lessons of the past*. Springer; 2019.
146. Advani D, Sharma S, Kumari S, Ambasta RK, Kumar P. Precision oncology, signaling, and anticancer agents in cancer therapeutics. *Anticancer Agents Med Chem.* 2022;22:433–468.
147. Song FX, Xu X, Ding H, et al. Recent progress in nanomaterial-based biosensors and theranostic nanomedicine for bladder cancer. *Biosensors.* 2023;13:106.
148. Grimaldi D, Llitjos J, Pene F. Post-infectious immune suppression: a new paradigm of severe infections. *Med Mal Infect.* 2014;44:455–463.
149. Bahreyni A, Mohamud Y, Luo H. Recent advancements in immunotherapy of melanoma using nanotechnology-based strategies. *Biomed Pharmacother.* 2023;159:114243.
150. Gupta B, Sadaria D, Warriar VU, et al. *Plant lectins and their usage in preparing targeted nanovaccines for cancer immunotherapy*. Elsevier; 2022.
151. Galon J, Bruni D. Tumor immunology and tumor evolution: intertwined histories. *Immunity.* 2020;52:55–81.
152. Rifai N, Gillette MA, Carr SA. Protein biomarker discovery and validation: the long and uncertain path to clinical utility. *Nat Biotechnol.* 2006;24:971–983.
153. Karsdal M, Henriksen K, Leeming D, et al. Biochemical markers and the FDA Critical Path: how biomarkers may contribute to the understanding of pathophysiology and provide unique and necessary tools for drug development. *Biomarkers.* 2009;14:181–202.
154. Lotan Y, Shariat SF, Schmitz-Dräger BJ, et al. *Considerations on implementing diagnostic markers into clinical decision making in bladder cancer*. Elsevier; 2010.
155. Puntmann V. How-to guide on biomarkers: biomarker definitions, validation and applications with examples from cardiovascular disease. *Postgrad Med J.* 2009;85:538–545.
156. Lin J, Bunn V. Comparison of multi-arm multi-stage design and adaptive randomization in platform clinical trials. *Contemp Clin Trials.* 2017;54:48–59.
157. García-Gutiérrez MS, Navarrete F, Sala F, Gasparyan A, Austrich-Olivares A, Manzanares J. Biomarkers in psychiatry: concept, definition, types and relevance to the clinical reality. *Front Psychiatry.* 2020;11:432.
158. Weickert CS, Weickert TW, Pillai A, Buckley PF. Biomarkers in schizophrenia: a brief conceptual consideration. *Dis Markers.* 2013;35:3.
159. Xia J, Broadhurst DI, Wilson M, et al. Translational biomarker discovery in clinical metabolomics: an introductory tutorial. *Metabolomics.* 2013;9:280–299.
160. Lipkovich I, Dmitrienko A. Strategies for identifying predictive biomarkers and subgroups with enhanced treatment effect in clinical trials using SIDES. *J Biopharm Stat.* 2014;24:130–153.
161. Depledge MH. *The rational basis for the use of biomarkers as ecotoxicological tools. Nondestructive biomarkers in vertebrates*. CRC Press; 2020:271–295.
162. Reinhart K, Bauer M, Riedemann NC, et al. New approaches to sepsis: molecular diagnostics and biomarkers. *Clin Microbiol Rev.* 2012;25:609–634.
163. Gyawali B, Ramakrishna K, Dhamoon AS. Sepsis: the evolution in definition, pathophysiology, and management. *SAGE Open Med.* 2019;7, 2050312119835043.
164. Delano MJ, Ward PA. Sepsis-induced immune dysfunction: can immune therapies reduce mortality? *J Clin Invest.* 2016;126:23–31.
165. Delano MJ, Ward PA. The immune system's role in sepsis progression, resolution, and long-term outcome. *Immunol Rev.* 2016;274:330–353.
166. Mühlen S, Dersch P. *Anti-virulence strategies to target bacterial infections. How to overcome the antibiotic crisis: facts, challenges, technologies and future perspectives*; 2016:147–183.
167. Hanif F, Muzaffar K, Perveen K, et al. Glioblastoma multiforme: a review of its epidemiology and pathogenesis through clinical presentation and treatment. *Asian Pac J Cancer Prevent.* 2017;18:3.