

Revista Portuguesa de Estomatologia, Medicina Dentária e Cirurgia Maxilofacial



www.elsevier.pt/spemd

Research

Air quality assessment during dental practice: Aerosols bacterial counts in an universitary clinic

Patrícia Manarte-Monteiro^a, Alexandrine Carvalho^a, Cristina Pina^b, Hélder Oliveira^a, Maria Conceição Manso^{c,d,*}

- ^a Department of Medical Sciences, Faculty of Health Sciences, University Fernando Pessoa, Portugal
- ^b CEBIMED, Department of Microbiology, Faculty of Health Sciences, University Fernando Pessoa, Porto, Portugal
- ^c Faculty of Health Sciences, University Fernando Pessoa, Porto, Portugal
- ^d REQUIMTE, University of Porto, Portugal

ARTICLE INFO

Article history: Received 24 March 2012 Accepted 1 October 2012 Available online 21 February 2013

Keywords:
Aerosols
Microbiology
Dental cavity
preparation/instrumentation
Infection control,
dental/instrumentation

ABSTRACT

Objective: The main aim of this study was to assess the clinic atmosphere quality regarding the Index of Air Microbial contamination (IMA), according to dental aerosols bacterial counts, when different dentistry procedures are performed.

Material and methods: The aerosols generated by dentistry and endodontic procedures were analyzed in 26 dental units of the dental clinic. Blood agar plates (n = 244) were incubated at 37 °C/48 h and the colony forming units (CFU) were calculated.

All statistical analysis methods were conducted using the SPSS® vs.17.0 software (SPSS Inc., IL, Chicago, USA), using α = 0.05 for the comparison tests used (non parametric tests of Mann–Whitney and Wilcoxon).

Results: The IMA median value in the dental clinic was of 10.4 CFU/dm²/h. Aerosols' CFU counts were significantly higher at 0.5 m and during endodontic treatments. Longer treatment times were associated with higher CFU counts both in dentistry and endodontic procedures. The use and time of turbine use did not significantly affect the CFU counts. Micrococcus sp., Staphylococcus sp. and Streptococcus sp. were identified with presumptive tests after isolation of representative colonies.

Conclusion: Considering the IMA, the dental clinic atmosphere quality was found to be good.
© 2012 Sociedade Portuguesa de Estomatologia e Medicina Dentária. Published by
Elsevier España, S.L. All rights reserved.

Avaliação da qualidade do ar na prática clínica: contagens bacterianas em aerossóis numa clínica universitária

RESUMO

Palauras-chave: Aerossóis Objetivo: O objetivo principal deste trabalho foi determinar a qualidade do ar da clínica utilizando o Index of Air Microbial contamination (IMA), mediante utilização de contagens

^{*} Corresponding author.

Microbiologia
Preparação cavitária
dentária/instrumentação
Controlo de
infecção/instrumentação
dentária

bacterianas em aerossóis dentários produzidos durante a realização de diferentes procedimentos dentários.

Material e métodos: Foram analisados aerossóis gerados por procedimentos de dentística e endodontia em 26 unidades dentárias distribuídas pela clínica de medicina dentária. As placas de agar de sangue (n = 244) foram incubadas a 37 °C/48 h e foram calculadas a unidades formadoras de colónias (UFC).

A análise estatística foi realizada recorrendo ao software SPSS® vs. 17.0 software, utilizando α = 0.05 em todos os testes de comparação estatística realizados (testes não paramétricos de Mann–Whitney e de Wilcoxon).

Resultados: A mediana do IMA na clínica de medicina dentária foi 10,4 UFC/dm²/h. As contagens de UFC nos aerossóis foram significativamente superiores na distância 0,5 m e durante tratamentos endodônticos. Tratamentos mais longos estão associados a contagens mais elevadas de UFC, tanto para procedimentos de dentística como de endodontia. A utilização e tempo de utilização de turbina não afetou significativamente as contagens de UFC. Após isolamento de colónias representativas e utilizando de testes presumptivos, foi possível identificar Micrococcus sp., Staphylococcus sp. e Streptococcus sp.

Conclusão: A qualidade do ar na clínica de medicina dentária, utilizando o IMA, foi considerada como boa.

© 2012 Sociedade Portuguesa de Estomatologia e Medicina Dentária. Publicado por Elsevier España, S.L. Todos os direitos reservados.

Introduction

In dental clinic environment dentistry professionals and patients are daily exposed to a great variety of infectious agents and toxic substances transported by aerosols and droplets, produced during dental operative procedures,1 promoting an increased risk of cross infection.^{2,3} Aerosols are particles small enough to stay airborne for an extended period before they settle on environmental surfaces.^{4,5} As 75% of these particles drop on a desktop with a diameter of 2 m (meters) from the patient position, 6 the environment (water, air and surfaces) play an important role in this context.2 Mouth fluids are grossly contaminated with bacteria, mostly aerobic bacteria (streptococci and staphylococci) and viruses.^{2,7} Most dental procedures that use handpieces, turbines, ultrasonic scalers, air polishers and abrasion units removes material from the operative site, that becomes aerosolized^{3,7,8} by the instrument rotary action or the water sprays and compressed air combined actions¹; So, there is a strong possibility that aerosols, besides the presence of bacteria,9 will include viruses, blood, and supra- and sub-gingival plaque organisms. 10,11

Several studies^{12–16} have been conducted to determine which dental procedure produces the highest airborne bacterial contamination using nonselective bacterial growth media (blood agar). When an aerobic bacterium settles on the plate and grows as a colony, it is counted as a colony-forming unit, or CFU.¹ Although this method does not provide any differentiation between whether the bacteria are relatively benign or a pathogenic species, it is a sterile, easy to implement and low cost, reproducing the most common conditions and gives a good perspective regarding total airborne bacterial CFUs from a particular procedure.¹⁷

Pasquarella et al. 17 described the Index of Air Microbial contamination (IMA) based on the count of the microbial fallout on to Petri dishes left open to the air according to the 1/1/1 scheme (for 1h, 1m from the floor, at least 1m away from

walls or any obstacle). Classes of contamination and maximum acceptable levels have been established, and a threshold of 25 was considered adequate. ^{2,3} IMA has been tested in many different places and proved to be a reliable and useful tool for monitoring the microbial surface contamination settling from the air in any environment^{2,17,18} although its use is not consensual for critical environments such as operating theatres. ² A few studies on air quality microbial counts in dental clinics were presented in literature. ^{2,8,19}

The aim of this study was to assess the aerosols contamination, by quantifying the aerobic bacteria CFUs when different dental treatments were performed and to analyze the effect of type and time of treatment and time of turbine use on the quantitative bacterial variation of aerosols produced during restorative dentistry and endodontic treatments, and to identify the representative aerobic bacterial colonies. Another purpose of this study was to check the dental clinic atmosphere quality comparing with the IMA values.

Material and methods

This study was conducted in the teaching Dental Clinical of the Faculty of Health Sciences, University Fernando Pessoa (FHS-UFP). This clinic has a total area of $390\,\mathrm{m}^2$, divided into 37 dental units spaces (DUs), each one with $5\,\mathrm{m}^2$. The ventilation system was checked periodically and was not altered during the study period.

A baseline register of CFU counts was conducted before any appointment/dental procedure performed in the dental clinic. This control group was constituted by 12 opened plaques placed in three randomly selected DUs. IMA was assessed by passive sampling, using blood agar plates (bioMerieux Ref. 43041, Linda-a-Velha, Portugal) 9 cm (centimetre) in diameter that were exposed to air during dental treatments (two at 0.5 m and two at 2 m from the patient head position) (Fig. 1). The plates were opened at the beginning of each dental procedure and remained so for at least 1 h to a maximum of 4 h (duration

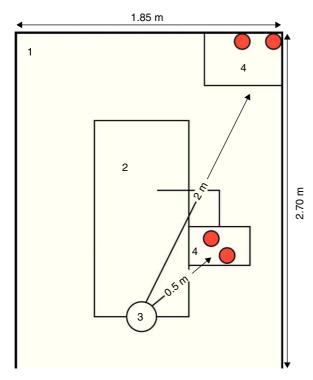


Figure 1 – Representation of the blood agar plates placement regarding the patient head position, on the dental unit space (5 m² area) during the operative procedures: (1) box; (2) dental unit; (3) patient head; (4) blood agar plates.

of the dental clinical appointments). Overall, 244 air samples in blood agar plates were collected from 26 DUs of the dental clinic.

The restorative dentistry procedures included cavities preparation of dental hard tissues disorders that were directly restored with adhesive or non-adhesive dental materials. All biomechanical preparation was done with manual instruments and high-speed handpieces (turbine) with watercooling. The restored surfaces were polished by rotary action, with water-cooling; Relative operatory field was done with cotton rolls and surgical aspirator.

All endodontic treatments, performed in mono- and multicanal teeth, were non-surgical and included biomechanical preparation with turbine (to prepare the access cavity) with water-cooling, manual instrumentation, disinfection with antimicrobial rinses and the root canal filling; Rubber dam was used for operatory field isolation.

To perform the microbial quantitative analysis of air all blood agar plates were incubated at $37\,^{\circ}\text{C}$ for $48\,\text{h}$. Colonies were counted by CFU/plate. The CFU/dm²/h was calculated to determine the IMA. Bacterial levels <39 CFU/dm²/h with passive sampling were considered as acceptable values for the air microbial contamination. The threshold value of $25\,\text{CFU/dm²/h}$ for IMA was considered as a good air performance.

The methodology employed for the qualitative analysis of microbial air followed the classical microbiologic identification. The isolation of representative colonies predominantly present in the blood agar plates was based on macroscopic colony morphology: colour, shape, size, texture and hemolytic characteristics. Further identification was determined using the Gram staining and cell morphology^{21,22}; and the enzymatic tests: catalase, oxidase, coagulase and DNAse. Based on these information it was possible to select appropriate procedures for identification of isolates by the usual commercial biochemical tests, using the API Staph (bioMérieux, Linda-a-Velha, Portugal) and API 20 Strep (bioMérieux, Linda-a-Velha, Portugal).

All statistical analysis methods were conducted using the SPSS® vs. 17.0 software (SPSS Inc., IL. Chicago, USA), using α = 0.05. The comparison of the median CFU value in dentistry procedures performed (restorative dentistry/endodontic), time of treatment (\leq 2 h/>2 h), use of turbine (yes/no) and time of turbine use (\leq 30 min/>30 min) was performed using the Mann–Whitney test (as the assumption of normality of the observations does neither hold nor the symmetry of the distributions). The Wilcoxon test was used for comparing median CFU values projected at two different distances (paired measurements, at 0.5 m and 2 m).

Results

The IMA value in the dental clinic was significantly (Mann–Whitney test, p < 0.001) lower before the opening and patient attendance (Mean (\pm SD); Median: 3.7 (\pm 1.0); 3.4 CFU/dm²/h) than during the clinical attendance (11.9 (\pm 6.7); 10.4 CFU/dm²/h). There were significant differences (Mann–Whitney test, p < 0.001) in the CFU/plate counts between the control group (7.0 (\pm 1.9); 6.4 CFU/dm²/h) and the experimental (restorative dentistry and endodontic treatments) group (17.3 (\pm 10.1); 15.0 CFU/dm²/h). The CFU/plate counts mean value was significantly (Mann–Whitney test, p < 0.001) higher for endodontic treatments (19.7 (\pm 10.8); 17.1 CFU/dm²/h) than for restorative dentistry (15.1 (\pm 8.9); 13.9 CFU/dm²/h) procedures.

During dentistry and endodontic treatments aerotransported CFU counts (Table 1) were significantly (Wilcoxon test, p < 0.001) higher at 0.5 m (19.0 (±11.5); 16.0) than at 2 m (15.6 (±8.2); 13.9) and significantly higher for endodontic treatment for both distances (Mann–Whitney test, p = 0.001, at 0.5 m and p = 0.001 at 2 m). During restorative dentistry procedures the CFU counts were significantly higher (Wilcoxon test, p < 0.001) at 0.5 m (16.6 (±10.4); 15.0) than at 2 m (13.6 (±6.9); 12.7).

Longer treatment times (≥ 2 h) were associated with higher CFU count (Mann–Whitney test, p < 0.005) in restorative dentistry (Mann–Whitney test, p < 0.004) and in endodontic treatments (Mann–Whitney test, p < 0.002) procedures, for both growth plate distances, at 0.5 m and at 2 m (Table 2). No significant differences were found in CFU/plate counts formed during the restorative dentistry or the endodontic treatments procedures, considering the use or no turbine use and the short time (≤ 30 min) or the longer time (> 30 min) of turbine use (Table 2).

Gram-positive cocci were predominant in the samples: Micrococcus sp. (99.9%), Staphylococcus capitis (99.8%), Streptococcus sp. (99.9%), Staphylococcus epidermidis (84.8%) and

endodontic treatments) projected at two distances (0.5 and 2 m).										
Plate distances ^a	Dentistry procedures performed	No. plates	Mean (±SD) ^b	Me (P25–P75) ^c	p ^d value	p ^e value				
0.5 m	All (restorative dentistry and endodontics)	122	19 (±11.5)	16A (12.3–22.6)		<0.001*				
	Restorative dentistry	64	16.6 (±10.4)	15bA (11.1–19.2)	0.011*	(All)				
2 m	Endodontic treatment	58	21.5 (±12.1)	18.6aA (13.1–28.4)		<0.001* (Dent)				
	All (restorative dentistry and endodontics)	122	15.6 (±8.2)	13.9B (9.8–19.4)		<0.001*				
	Restorative dentistry	64	13.6 (±6.9)	12.7bB (9.1–16.4)	0.006*	(Endo)				
	Endodontic treatment	58	17.8 (±9)	16.1aB (11.6-23.8)						

- ^a Blood agar plates placement regarding the patient head position, on the dental unit space (5 m² area); distance in meters (m).
- ^b Mean (±SD): mean and standard deviation of CFU/plate counts according to the dentistry procedures performed.
- ^c Me (P25–P75): median, 25 and 75 percentiles of CFU/plate counts according to the dentistry procedures performed.
- ^d Mann–Whitney test; different letters (a and b) after the median value stand for significant differences with treatment.
- ^e Wilcoxon test; different letters (A and B) after the median value stand for significant differences with distance, according to the Wilcoxon test, for Dentistry (Dent) and Endodontic (Endo) treatment.
- * Statistical significant difference.

Staphylococcus sp. (99.9%) colonies were identified (% of identification reliability given by the API system).

Discussion

In the present study the air quality of the dental clinic was found to be good, with acceptable values for the air microbial counts 17,20 since the bacteria levels (11.9 (\pm 6.7) CFU/dm²/h) registered were lower than 39 CFU/dm²/h. 2,17 Passive air sampling is one effective way of quantifying airborne bacteria once it measures the live microorganisms that can settle, growth and multiply, as used in the present study, reflects the extent of aerobic bacteria contamination on the surfaces, and highlights clinical areas of primary importance for cross infection prevention. 2,8,17

The results (Table 1) showed higher number of CFU counts during the endodontic treatments in which the rubber dam was used. One likely explanation is that during the restorative dentistry procedures, the operatory field was relatively isolated with cotton rolls and splatter aspiration. Although the use of rubber dam has been shown to be highly significant in reducing contamination of the atmosphere, 8,23 another data reported suggest that the use of rubber dam can concentrate and spread²⁴ the aerosols produced when high-speed rotating instruments are used or when endodontic treatment is performed, and momentarily contaminates the air; but as the bacteria settle on the surfaces, then the air quality increases. How far the aerosols spread and what level of contamination they cause in the dental surgery is of concern⁸ and should be analyzed.

During both dentistry and endodontic treatments CFU counts (Table 1) were significantly higher at $0.5\,\mathrm{m}$ than at $2\,\mathrm{m}$ and significantly higher in endodontic treatment for both

distances, results corroborated by Timmerman et al., ²⁵ but not by Rautemaa et al., ⁸ that registered higher CFU/m²/h counts at distances >1.5 m than at distances <1 m, explaining that may be related to the increased speed of instruments rotation, that gives a greater speed and higher angular trajectory of bacteria.

Longer treatment times ($\geq 2\,\mathrm{h}$) were associated with higher CFU count in dentistry and endodontic procedures, at both distances (0.5 m and 2 m) (Table 2), but the time of turbine use did not affect the aerosol CFU counts. These results are partially in agreement with Grenier (1995) study, regarding the level of air bacterial contamination that was higher at the end of dental treatments with about 6 h time duration,²⁴ but considering the use of turbine, are not in agreement with the Rautemaa et al.⁸ findings, where the CFU counts were less intense during dental treatments where high-speed and ultrasonic instruments were not used.

In the present study the most common microorganisms isolated were included in Micrococcus sp., Streptococcus sp., and Staphylococcus sp. genus, with a good identification by using the commercial biochemical test system, in spite of their own limits. Rautemaa et al.8 evaluated samples taken from personnel facial masks and surfaces in dental rooms where treatments were performed using high-speed rotating instruments, and registered Gram positive-cocci namely viridans streptococci (Streptococcus genus) and Staphylococcus, as the most common bacterial findings. Streptococcus is the main cause of bacterial endocarditis in compromised patients, and the normal habitat is the human upper respiratory tract and skin.⁷ Similarly, Staphylococcus is a normal commensal on the skin surface and anterior nares which has the characteristic of an opportunist pathogen and may cause catheter related sepsis and infection of artificial joints. Micrococcus are frequently present on normal skin.^{7,26} Despite the importance of air microbial level evaluation as a step toward cross infection prevention, the design studies variability limits the

Table 2 – Descriptive statistics of CFU/plate count formed during different dentistry procedures (restorative dentistry and endodontic treatments) projected at two distances (0.5 and 2 m) according to treatment duration, in hours (\leq 2 h and >2 h), and to length of turbine use, in minutes (0 minutes (no), \leq 30 min and >30 min).

Dentistry procedures performed ^a	Plates distance	Category	No. plates	CFU/plate count			
				Mean (±SD)	Me (P25–P75)	p ^b value	
Treatment duration (h)							
	0.5 m	≤2 h	60	15.9 (±10.2)	14.5b (10.6–18)	0.004*	
Restorative dentistry		>2 h	4	28.0 (±6.2)	27.7a (22.6–33.6)		
	2 m	$\leq 2h$	60	12.8 (±6.2)	11.9b (8.9–15.6)	0.002*	
		>2 h	4	26.3 (±2.2)	26.0a (24.4–28.5)		
	0.5 ***	≤2 h	42	18.8 (±11.2)	15.5b (10.4–25.0)	0.002*	
Endodontic treatment	0.5 m	>2 h	16	28.7 (±11.8)	24.3a (19.3–39.0)		
	2 m	≤2 h	42	15.3 (±7.5)	13.7b (10.0–19.8)	0.001*	
		>2 h	16	24.4 (±9.5)	23.5a (16.6–31.1)		
Time of turbine use (min)							
		≤30 min	16	13.1 (±5.4)	13.5 (8.3–18.0)	0.215 ^c	
Restorative dentistry	0.5 m	>30 min	46	17.1 (±11.1)	15.0 (11.7–19.1)		
		No (0 min)	2	34.1 (±1.3)	34.1 (33.2–35.0)	-	
		≤30 min	16	11.0 (±4.3)	11.1 (6.9–14.6)	0.179 ^c	
	2 m	>30 min	46	14.2 (±7.3)	12.7 (9.4–16.5)		
		No (0 min)	2	22.2 (±1.1)	22.2 (21.4–23.0)	-	
	0.5 m	≤30 min	44	23.1 (±12.3)	19.7 (15.1–27.8)	0.069	
Endodontic treatment		>30 min	14	16.8 (±10.4)	13.4 (8.4–30.1)		
	2 m	≤30 min	44	18.9 (±8.7)	17.0 (13.1–24.9)	0.058	
		>30 min	14	14.5 (±9.5)	11.8 (6.0–23.6)		

Different letters (a and b) after the median value stand for significant differences with duration of treatment according to the Mann–Whitney test

- ^a Blood agar plates placement regarding the patient head position on the dental unit space (5 m² area); distance in meters (m).
- ^b Mann–Whitney test.
- ^c Due to low "n" in the "no" category, *p*-values for these comparisons were not calculated or were calculated for the two other categories (out of three).
- * Statistical significant difference.

conclusions and difficult the comparison between published results regarding the quantity (acceptability levels of CFU) and quality (microbial identification) assessment of microbial contamination in dental aerosols. In the present study given the reduced dimension of the sample size, the presented statistical values are merely indicative, even though several variables were analyzed and the records obtained tend to suggest that dentistry procedures promote a risk of infection if the extent and nature of microbial aerosols created by high-speed handpieces are underestimated. A huge number of bacteria are frequently isolated from air in dental offices as well as from dental unit waterlines. Further studies with simultaneous qualitative and quantitative counts related

with usual dental treatments would be of the utmost importance.

The air quality of the UFP pedagogical dental clinic regarding bacterial counts was found to be good, but in dental clinic environments microbial transmission can occur by air, so preventive measures should be promoted to avoid pathogenic microorganisms spread.

Conclusions

CFU counts in aerosols are influenced by the dentistry procedures (restorative dentistry and endodontic), the operative site

distance and the treatment times performed. Qualitative and quantitative composition of dental aerosols probably varies with each patient and operative site. Dental aerosols produced during dentistry procedures should be controlled, to the greatest extent possible, to a health reassurance of patients and dental team in clinical work environments.

Ethical disclosures

Protection of human and animal subjects. The authors declare that no experiments were performed on humans or animals for this study.

Confidentiality of data. The authors declare that they have followed the protocols of their work center on the publication of patient data and that all the patients included in the study received sufficient information and gave their written informed consent to participate in the study.

Right to privacy and informed consent. The authors have obtained the written informed consent of the patients or subjects mentioned in the article. The corresponding author is in possession of this document.

Conflicts of interest

The authors have no conflicts of interest to declare.

Acknowledgment

Author M.C. Manso acknowledge Fundação para a Ciência e a Tecnologia through grant PEst-C/EQB/LA0006/2011.

REFERENCES

- Harrel SK, Molinari J. Aerosols and splatter in dentistry: a brief review of the literature and infection control implications. J Am Dent Assoc. 2004;135:429–37.
- Castiglia P, Liguori G, Montagna MT, Napoli C, Pasquarella C, Bergomi M, et al. Italian multicenter study on infection hazards during dental practice: control of environmental microbial contamination in public dental surgeries. BMC Public Health. 2008;8:187–94.
- Szymańska J. Dental bioaerosol as an occupational hazard in a dentist's workplace. Ann Agric Environ Med. 2007;14:203–7.
- Micik RE, Miller RL, Leong AC. Studies on dental aerobiology.
 Efficacy of surgical masks in protecting dental personnel from airborne bacterial particles. J Dent Res. 1971;50: 626–30.
- Micik RE, Miller RL, Mazzarella MA, Ryge G. Studies on dental aerobiology. I. Bacterial aerosols generated during dental procedures. J Dent Res. 1969;48:49–56.
- Checchi L, Montevecchi M, Moreschi A, Graziosi F, Taddei P, Violante FS. Efficacy of three face masks in preventing inhalation of airborne contaminants in dental practice. J Am Dent Assoc. 2005;136:877–82.
- Toroglu MS, Haytac MC, Koksal F. Evaluation of aerosol contamination during debonding procedures. Angle Orthod. 2001;71:299–306.

- 8. Rautemaa R, Nordberg A, Wuolijoki-Saaristo K, Meurman JH. Bacterial aerosols in dental practice a potential hospital infection problem? J Hosp Infect. 2006;64:76–81.
- Szymańska J, Sitkowska J. Bacterial hazards in a dental office: an update review. Afr J Microbiol Res. 2012;6: 1642–50.
- Rocas IN, Siqueira Jr JF. Identification of bacteria enduring endodontic treatment procedures by a combined reverse transcriptase-polymerase chain reaction and reverse-capture checkerboard approach. J Endod. 2010;36:45–52.
- Signoretto C, Bianchi F, Burlacchini G, Sivieri F, Spratt D, Canepari P. Drinking habits are associated with changes in the dental plaque microbial community. J Clin Microbiol. 2010;48:347–56.
- Barnes JB, Harrel SK, Rivera-Hidalgo F. Blood contamination of the aerosols produced by in vivo use of ultrasonic scalers. J Periodontol. 1998;69:434–8.
- 13. Bentley CD, Burkhart NW, Crawford JJ. Evaluating spatter and aerosol contamination during dental procedures. J Am Dent Assoc. 1994;125:579–84.
- Harrel SK. Clinical use of an aerosol-reduction device with an ultrasonic scaler. Compend Contin Educ Dent. 1996;17:1185–93, quiz 1194.
- Legnani P, Checchi L, Pelliccioni GA, D'Achille C. Atmospheric contamination during dental procedures. Quintessence Int. 1994;25:435–9.
- Muzzin KB, King TB, Berry CW. Assessing the clinical effectiveness of an aerosol reduction device for the air polisher. J Am Dent Assoc. 1999;130:1354–9.
- 17. Pasquarella C, Pitzurra O, Savino A. The index of microbial air contamination. J Hosp Infect. 2000;46:241–56.
- Pasquarella C, Sansebastiano GE, Ferretti S, Saccani E, Fanti M, Moscato U, et al. A mobile laminar airflow unit to reduce air bacterial contamination at surgical area in a conventionally ventilated operating theatre. J Hosp Infect. 2007;66: 313-9.
- Monarca S, Grottolo M, Renzi D, Paganelli C, Sapelli P, Zerbini I, et al. Evaluation of environmental bacterial contamination and procedures to control cross infection in a sample of Italian dental surgeries. Occup Environ Med. 2000;57: 721–6.
- Friberg B, Friberg S, Burman LG. Inconsistent correlation between aerobic bacterial surface and air counts in operating rooms with ultra clean laminar air flows: proposal of a new bacteriological standard for surface contamination. J Hosp Infect. 1999;42:287–93.
- 21. Al Maghlouth A, Al Yousef Y, Al Bagieh N. Qualitative and quantitative analysis of bacterial aerosols. J Contemp Dent Pract. 2004;5:91–100.
- Bennett AM, Fulford MR, Walker JT, Bradshaw DJ, Martin MV, Marsh PD. Microbial aerosols in general dental practice. Br Dent J. 2000;189:664–7.
- Logothetis DD, Martinez-Welles JM. Reducing bacterial aerosol contamination with a chlorhexidine gluconate pre-rinse. J Am Dent Assoc. 1995;126:1634–9.
- Grenier D. Quantitative analysis of bacterial aerosols in two different dental clinic environments. Appl Environ Microbiol. 1995;61:3165–8.
- 25. Timmerman MF, Menso L, Steinfort J, van Winkelhoff AJ, van der Weijden GA. Atmospheric contamination during ultrasonic scaling. J Clin Periodontol. 2004;31:458–62.
- Pankhurst C. Summary of: a pilot study of bioaerosol reduction using an air cleaning system during dental procedures. Br Dent J. 2010;209:408–9.
- Harrel SK. Airborne spread of disease the implications for dentistry. J Calif Dent Assoc. 2004;32:901–6.