

Revista Colombiana de REUMATOLOGÍA



www.elsevier.es/rcreuma

Original Investigation

Association of FokI polymorphism of the VDR gene with systemic lupus erythematosus in an adolescent population of the Colombian Caribbean Asociación del poRkI del gen VDR y lupus eritematoso sistémico en población adolescente del Caribe colombiano



Gloria Garavito^{a,b}, Luis Fang^a, Alex Domínguez-Vargas^a, Ana Moreno-Woo^a, Guillermo López-Luch^c, Antonio Iglesias^a, Gustavo Aroca^{a,b}, Eduardo Egea^{a,*}

- ^a Departamento de Medicina, División de Ciencias de la Salud, Universidad del Norte, Barranquilla, Colombia
- ^b Facultad de Ciencias de la Salud, Universidad Simón Bolívar, Barranquilla, Colombia
- ^c Centro Andaluz de Biología del Desarrollo (CABD), Universidad Pablo de Olavide-CSIC, Sevilla, Spain

ARTICLE INFO

Article history: Received 22 January 2021 Accepted 8 April 2021 Available online 14 March 2023

Keywords: Systemic lupus erythematosus VDR Vitamin D Polymorphism

ABSTRACT

Introduction: Vitamin D and vitamin D receptor (VDR) polymorphisms are associated with autoimmune diseases including systemic lupus erythematosus (SLE). The aim of this study is to assess the genetic association between VDR polymorphisms: TaqI, ApaI, Bsml and FokI and SLE with serum levels of Vitamin D in the Colombian Caribbean population.

Method: Case and control study. One hundred and thirty-three patients with SLE and 100 healthy individuals were included. VDR polymorphism were genotyped by RT-PCR and Taqman[®] probes. Allelic, genotypic and haplotype associations were estimated. Serum vitamin D concentrations were quantified by Elisa. Values of 30 to 100 ng/ml were established as a normal reference range. P values <.05 were considered statistically significant.

Results: A high prevalence of SLE was observed in women (94%) and was associated with a higher risk of SLE [OR: 10.8; 95% CI: 4.7–24.6] (p<.05). Moreover, higher risk of SLE was observed in individuals with FokI VDR [rs2228570] [OR: 1.58; 95% CI: 1.05–2.36] in allelic models. The ACCA Haplotype of TaqI/ApaI/Bsml/FokI polymorphisms was associated with higher risk of SLE [OR = 2.28, 95% CI = 1.12–4.66, psim <.01]. Vitamin D deficiency was evidenced in 11.3% of the patients.

^{*} Corresponding author.

Conclusion: In this study, the VDR rs2228570 polymorphism and ACCA haplotype were associated with higher SLE risk in an adolescent population.

© 2021 Asociación Colombiana de Reumatología. Published by Elsevier España, S.L.U. All rights reserved.

RESUMEN

Palabras clave: Lupus eritematoso sistémico VDR Vitamina D Polimorfismo Introducción: La vitamina D y los polimorfismos en el receptor de vitamina D (VDR) se asocian con enfermedades autoinmunes, incluido el lupus eritematoso sistémico (LES). El objetivo de este estudio es analizar la asociación genética entre los polimorfismos de VDR: TaqI, ApaI, Bsml y FokI, y la susceptibilidad al LES, así como su relación con los niveles séricos de vitamina D en población del Caribe colombiano.

Metodología: Estudio de casos y controles. Se incluyeron 133 pacientes adultos con diagnóstico de LES y 100 individuos sanos. Los polimorfismos VDR fueron genotipados por RT-PCR y sondas Taqman[®]. Se estimaron asociaciones alélicas, genotípicas y haplotípicas. Las concentraciones séricas de vitamina D fueron cuantificadas por Elisa. Se establecieron valores de 30 a 100 ng/mL como rango normal de referencia. Valores p < 0.05 fueron considerados estadísticamente significativos.

Resultados: Se observó una alta prevalencia de LES en pacientes femeninas (94%) y se asoció a mayor riesgo de LES [OR: 10,8; IC 95%: 4,7–24,6] (p<0,05). Se evidenció mayor riesgo de LES en individuos con polimorfismo FokI del gen VDR [rs2228570] [OR: 1,58; IC 95%: 1,05–2,36] en modelos alélicos. El haplotipo ACCA de los polimorfismos TaqI, ApaI, Bsml yFokI se asoció a mayor riesgo de LES [OR: 2,28, IC 95%: 1,12-4,66, psim< 0,01]. Se evidenció deficiencia de vitamina D en 11,3% de los pacientes.

Conclusión: En este estudio, el polimorfismoVDR rs22228570 y el haplotipo ACCA se asociaron a mayor riesgo de LES en población adolescente.

© 2021 Asociación Colombiana de Reumatología. Publicado por Elsevier España, S.L.U.

Todos los derechos reservados.

Introduction

Systemic lupus erythematosus (SLE) is a chronic autoimmune disease characterized by its wide epidemiological, clinical, and immunological heterogeneity. The pathophysiology is associated with immunological, hormonal, genetic, and environmental factors^{1–3}. Various studies have reported the effects of single nucleotide polymorphisms (SNPs) on the susceptibility to SLE^{4–6}, including the vitamin D receptor (VDR) gene; however, its effects on VDR activity are still poorly understood^{7–9}.

The VDR is the mediator of the biological activity of vitamin D. It is located in the nucleus of target cells, including immune cells (antigen-presenting cells, natural killer cells, B and T lymphocytes) $^{9-11}$. The VDR gene is located on chromosome 12q13.11 and regulates gene expression in various tissues that respond to vitamin $D^{10,12}$.

In addition to the regulation of bone mineral homeostasis, 1,25-dihydroxyvitamin D3 [1,25(OH)2D3] is also involved in interleukin 2 (IL-2) inhibition, modulation of T-cell phenotypes ^{13,14}, the increase in the development of regulatory T- cells ^{15,16}, and the production of immunoglobulins ¹⁷. 1,25(OH)2D3 inhibits interferon secretion and regulates the expression of IL-12 and nuclear factor kappa-B ^{11,18}. An in vitro study demonstrated that 1,25(OH)2D3 has a protective effect in autoimmune diseases ¹⁹. Therefore, vitamin D3 is considered a regulator of the immune system ^{11,20}.

Several polymorphic markers of unknown functional consequence within the VDR gene have been studied for their potential role in SLE risk. There are four important polymorphisms of the VDR gene: TaqI (rs731236), located in exon 9; ApaI (rs7975232) in intron 8, BsmI (rs1544410) in intron 9, and FokI (rs10735810) at the initiation codon^{8,9,21–23}.

However, the results of the association between these VDR gene polymorphisms and the risk of SLE are inconsistent in different studies carried out in various population groups, mainly due to different linkage disequilibrium coefficients and haplotype blocks in the populations, as well as to differences in sample size, population stratification, and variation in environmental factors between geographically separated areas⁸.

Population-based studies have shown that patients with SLE have significantly low serum levels of vitamin $D^{18,24}$; likewise, VDR gene polymorphisms are associated with higher activity/chronicity indices and an augmented risk of organic damage^{9,18,25}.

This study aims to analyze the genetic association between VDR polymorphisms (TaqI, ApaI, BsmI, and FokI) and susceptibility to SLE, as well as its relationship with serum vitamin D levels in the Colombian Caribbean population.

Methods

A case-control study was conducted. A total of 133 patients diagnosed with SLE and 100 healthy individuals were included,

in the adolescence (10–19 years), age group according to the World Health Organization (WHO), from the city of Barranquilla, on the Colombian Caribbean Coast. The diagnosis of SLE was performed by the presence of at least 4 of the 11 American College of Rheumatology (ACR) criteria²⁶. Subjects were assessed both by specialized consultation and in a high-complexity level institution in the city of Barranquilla, Colombia.

Patients had no history of systemic diseases, other rheumatic disorders, infections, or malignancies. The control group with a negative ANA test had no history of autoimmune diseases or family relationships with patients with SLE. All patients and controls approved in writing by assent or informed consent depending on whether they were minors or adults in their voluntary participation in the study. This study was approved by the institutional ethics committee of the Universidad del Norte (Resolution No. 05).

Genomic DNA extraction

A team composed of phlebotomists, researchers, and physicians collected 10 ml of peripheral venous blood in Vacutainer[®] tubes, either with serum separator tube (SST) or with dipotassium EDTA (Beckton Dickinson & Company, Franklin Lakes, USA; Ref: 367856) for each participant, to obtain serum and the subsequent extraction of genomic DNA, respectively.

DNA was extracted from buffy coat (leukoplatelet fraction) of anticoagulated blood using the modified salting out protocol and precipitation with sodium perchlorate²⁷. DNA concentration and quality were measured using the Nanodrop 2000/2000c spectrophotometer (Thermo Scientific, Waltham, MA, USA). Genomic DNA and serum samples were aliquoted and stored at $-70\,^{\circ}$ C until further analysis.

Serum vitamin D concentrations

Serum 25-hydroxyvitamin D concentrations were measured using enzyme immunoassay techniques: IDS-iSYS 25-Hydroxy Vitamin D (Immunodiagnosis Systems Ltd. Boldon, Tyne & Wear, United Kingdom). Reference values were established according to the Institute of Medicine (IOM) criteria, where <10 ng/ml corresponds to deficient levels; 10-29 ng/ml is defined as insufficient vitamin D; \geq 30- and <100 ng/ml are sufficient levels, and \geq 100 ng/ml corresponds to possible vitamin D intoxication²⁸.

SNP genotyping

The VDR gene polymorphisms (TaqI [rs731236 A/G], ApaI [rs7975232 A/C], FokI [rs2228570 A/G], and BsmI [rs1544410 C/T]) were genotyped by real-time PCR (RT-PCR), using commercially available kits: TaqMan[®] SNP Genotyping (Assay ID: C__16021387_20, C__2404008_10; C__28977635_10; C__12060045_20, and C__8716062_10; respectively [Applied Biosystems, Foster City, CA, USA]).

Genotyping assays were performed on an ABI Prism 7300 RT-PCR equipment from Applied Biosystems, using a total volume per reaction of 5 μ l (2.4 μ l of DNA at an approximate

concentration of $10 \text{ ng/}\mu\text{l}$, 2.5 μl of Master Mix-2x, and 0.125 μl of Taqman Genotyping- $40\times$ probes specific to each SNP).

All PCR reactions were performed in duplicate. The cycling program consisted of an initial step of 10 min at 95 °C, followed by 40 cycles from 15 s to 92 °C, and 1 min at 60 °C per cycle. The genotypic assignment was performed automatically by applying Allelic Discrimination from the Applied Biosystems system, which considered an amplification quality \geq 90% per sample.

Statistical analysis

Allelic, genotypic, and haplotypic frequencies were estimated in the study groups. In this study, non-probabilistic sampling was performed for convenience. Allele frequencies were used to estimate the Hardy-Weinberg genetic balance; in addition, the genetic association analysis with SLE was carried out at the allele level using the Arlequin v3.5 software.

The association analysis at the genotype level was performed using the Pearson \mathbf{x}^2 test, with Bonferroni correction of the p-value, in the statistical software SPSS v20 (IBM® SPSS® Statistics 20; IBM Corp., USA.). For these two analyses, the risk of SLE associated with each genotype or allele was also estimated, depending on the case, by calculating odds ratios (OR) and their corresponding 95% confidence intervals, using logistic regression models adjusted for the variable gender.

Association analyses between polymorphisms and serum vitamin D concentrations (quantitative data) were performed using the Mann-Whitney U and Kruskal Wallis tests, as appropriate, expressing mean values \pm standard deviation (SD). On the other hand, the analyses with the categorical data of vitamin D were carried out using Pearson's x^2 test or Fisher's exact test in cases with expected values $\,$ <5, with Bonferroni correction of the p-value.

For the haplotype association analysis, the statistical package "Haplo.stats v1.6.8" (R version 3.0.2; http://www.r-project.org)²⁹ was used; With this package, the Haplo.cc functions were used (logistic regression models for risk estimation: OR [95% CI], excluding haplotypes with frequencies <1%). In addition, 10,000 simulations were run to refine the p-value, represented as [psim]) and Haplo.glm (generalized linear regression model for estimating associations with numerical traits). Statistical significance was interpreted as p < 0.05.

Results

In the sample studied, the average age was 16.19 ± 3.48 years; 94% (n = 125) were female; this was associated as a risk factor for SLE (OR: 10.8; 95% CI: 4.7–24.6).

The Hardy-Weinberg equilibrium distribution showed that the VDR polymorphisms: ApaI [rs7975232] and BsmI [rs1544410] were not found in genetic equilibrium for the control and case groups, respectively (p < 0.05) (Table 1).

When comparing the genotypic and allele frequencies of the VDR polymorphisms in patients with SLE and the control group, a significantly higher frequency of the A allele of the FokI polymorphism [rs2228570] was found in cases than in controls (41 vs. 30%), associated with a 1.58 greater risk of SLE (OR: 1.58; 95% CI: 1.05-2.36; p=0.025) (Table 1).

Table 1 – Comparison of the genotypic and allelic frequencies of the vitamin D receptor polymorphisms in patients with systemic lupus erythematosus and the control group.

Cases	H-W	Controls	H-W	OR	95% CI	pª
(n = 133)		(n = 100)				
20 (18)		12 (12)		2.12	(0.9-4.8)	0.081
51 (45.9)		37 (37)		1.75	(0.9-3.17)	
40 (36)		51 (51)		0.47	(0.20-1.07)	
91 (41)	1.00	61 (30.5)	1.00	1.58	(1.05-2.36)	0.025*
131 (59)		139 (69.5)		0.63	(0.42-0.94)	
25 (22.5)		24 (24)		0.61	(0.2-1.4)	0.317
59 (53.2)		60 (60)		0.58	(0.2–1.1)	
27 (24.3)		16 (16)		1.62	(0.7–3.7)	
109 (49.1)	1.00	108 (54)	0.03*	0.82	(0.56-1.20)	0.314
113 (50.9)		92 (46)		1.21	(0.83-1.78)	
67 (60.4)		56 (56)		2.39	(0.6-8.3)	0.371
` '		` '		2.22	,	
` '		` '		0.41	(0.1–1.46)	
174 (78.4)	1.00	148 (74)	1.00	1.27	(0.81–1.99)	0.291
48 (21.6)		52 (26)		0.78	(0.5–1.23)	
55 (49 5)		55 (55)		2 33	(0 5–9 4)	0.169
, ,		, ,			` '	0.103
				0.42	,	
` '	0.04*		1.00	0.97	` '	0.893
• •		, ,		1.03	(0.6–1.59)	
	(n = 133 20 (18) 51 (45.9) 40 (36) 91 (41) 131 (59) 25 (22.5) 59 (53.2) 27 (24.3) 109 (49.1) 113 (50.9) 67 (60.4) 40 (36) 4 (3.6) 174 (78.4)	(n = 133) 20 (18) 51 (45.9) 40 (36) 91 (41) 1.00 131 (59) 25 (22.5) 59 (53.2) 27 (24.3) 109 (49.1) 113 (50.9) 67 (60.4) 40 (36) 4 (3.6) 174 (78.4) 48 (21.6) 55 (49.5) 53 (47.7) 3 (2.7) 163 (73.4) 0.04*	(n = 133) (n = 10) 20 (18) 12 (12) 51 (45.9) 37 (37) 40 (36) 51 (51) 91 (41) 1.00 61 (30.5) 131 (59) 139 (69.5) 25 (22.5) 24 (24) 59 (53.2) 60 (60) 27 (24.3) 16 (16) 109 (49.1) 1.00 108 (54) 113 (50.9) 92 (46) 67 (60.4) 56 (56) 40 (36) 36 (36) 4 (3.6) 36 (36) 4 (3.6) 8 (8) 174 (78.4) 1.00 148 (74) 48 (21.6) 55 (55) 53 (47.7) 38 (38) 3 (2.7) 7 (7) 163 (73.4) 0.04 148 (74)	(n = 133) (n = 100) 20 (18) 12 (12) 37 (37) 40 (36) 51 (51) 91 (41) 1.00 61 (30.5) 1.00 131 (59) 139 (69.5) 25 (22.5) 24 (24) 59 (53.2) 60 (60) 27 (24.3) 16 (16) 109 (49.1) 1.00 108 (54) 0.03* 113 (50.9) 92 (46) 67 (60.4) 56 (56) 40 (36) 36 (36) 4 (3.6) 8 (8) 174 (78.4) 1.00 148 (74) 1.00 55 (49.5) 53 (47.7) 38 (38) 7 (7) 163 (73.4) 0.04* 148 (74) 1.00	(n = 133) (n = 100) 20 (18) 12 (12) 2.12 51 (45.9) 37 (37) 1.75 40 (36) 51 (51) 0.47 91 (41) 1.00 61 (30.5) 1.00 1.58 131 (59) 139 (69.5) 0.63 25 (22.5) 24 (24) 0.61 0.58 27 (24.3) 16 (16) 1.62 109 (49.1) 1.00 108 (54) 0.03* 0.82 113 (50.9) 92 (46) 1.21 67 (60.4) 56 (56) 2.39 40 (36) 36 (36) 2.22 4 (3.6) 8 (8) 0.41 174 (78.4) 1.00 148 (74) 1.00 1.27 48 (21.6) 52 (26) 0.78 55 (49.5) 55 (55) 2.33 53 (47.7) 38 (38) 3.25 3 (2.7) 7 (7) 0.42 163 (73.4) 0.04* 148 (74) 1.00 0.97	(n=133) (n=100) 20 (18) 12 (12) 2.12 (0.9-4.8) 51 (45.9) 37 (37) 1.75 (0.9-3.17) 40 (36) 51 (51) 0.47 (0.20-1.07) 91 (41) 1.00 61 (30.5) 1.00 1.58 (1.05-2.36) 131 (59) 139 (69.5) 0.63 (0.42-0.94) 25 (22.5) 24 (24) 0.61 (0.2-1.4) 59 (53.2) 60 (60) 0.58 (0.2-1.1) 27 (24.3) 16 (16) 1.62 (0.7-3.7) 109 (49.1) 1.00 108 (54) 0.03* 0.82 (0.56-1.20) 113 (50.9) 92 (46) 1.21 (0.83-1.78) 67 (60.4) 56 (56) 2.39 (0.6-8.3) 40 (36) 36 (36) 2.22 (0.6-8.0) 4 (3.6) 8 (8) 0.41 (0.1-1.46) 174 (78.4) 1.00 148 (74) 1.00 1.27 (0.81-1.99) 48 (21.6) 52 (26) 0.78 (0.5-1.23) 55 (49.5) 53 (47.7) 38 (38) 3.25 (0.7-13.4) <td< td=""></td<>

H-W: Hardy-Weinberg equilibrium test, enhanced with 100,000 steps of the Markov chain. OR [95% CI]: logistic regression models.

^{*} p < 0.05 with Bonferroni correction.

Table 2 – Haplotype analysis of vitamin D receptor polymorphisms in patients with systemic lupus erythematosus and control group.												
TaqI [rs731236]	ApaI [rs7975232]	BsmI [rs1544410]	FokI [rs2228570]	Controls (n = 100) (%)	Cases (n = 133) (%)	pª	dummy p ^b	OR ^c	95% CI			
Α	A	С	G.	0.23	0.158	0.052	0.058	0.90	(0.4-1.7)			
G.	Α	T	G.	0.13	0.107	0.114	0.107	0.95	(0.4-2.2)			
A	C.	С	G.	0.298	0.215	0.135	0.137	1	_			
G.	Α	T	Α	0.103	0.0726	0.666	0.669	0.94	(0.4-2.2)			
A	Α	С	Α	0.0601	0.0642	0.593	0.600	1.28	(0.4-3.6)			
A	C	C	Α	0.14	0.259	0.00*	0.00*	2.28	(1.12-4.66)			
Α	Α	Т	G	0.0094	0.0575	0.00*	0.00*	14.7	(0.2–767)			

^a Empirical p- value.

Haplotype analysis showed that, at the VDR locus, the ACCA haplotype of the TaqI/ApaI/BsmI/FokI polymorphisms was associated with a 2.28-fold increased risk of SLE (OR: 2.28; 95% CI: 1.12-4.66; p=0.0) (Table 2).

When analyzing the serum expression of vitamin D, it was found that the cases showed significantly higher serum concentrations, compared to the controls $(49.5\pm24.06\,\text{ng/ml})$ vs. $38.6\pm13.23\,\text{ng/ml})$ (p=0.00) (Fig. 1). Vitamin D deficiency was evidenced in 11.3% of the patients (Fig. 2).

Discussion

In the current study, a higher frequency of the A allele of the FokI polymorphism [rs2228570] was observed in the cases, which was associated with a greater risk of SLE. There was no statistical significance between the ApaI [rs7975232], TaqI [rs731236], and BsmI [rs1544410] polymorphisms and SLE in the codominant or allelic models. The ACCA haplotype of

^a χ^2 likelihood ratio.

^b Simulated p-value.

 $^{^{\}rm c}\,$ Reference value for the logistic regression model.

 $^{^{*}}$ p < 0.05 with Bonferroni correction.

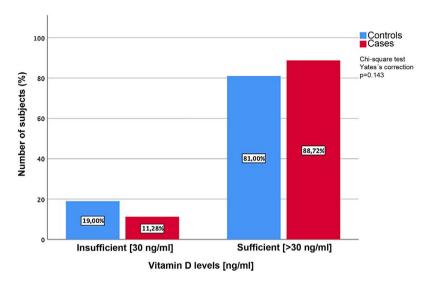


Figure 1 - Comparison of serum vitamin D levels (ng/ml) between cases with systemic lupus erythematosus and controls.

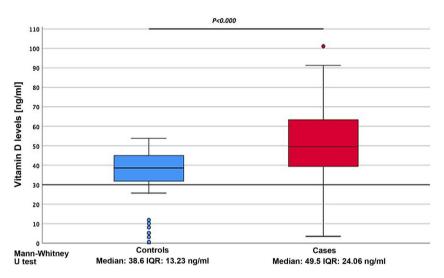


Figure 2 – Comparison of serum vitamin D levels (ng/ml) according to reference values between cases with systemic lupus erythematosus and controls.

the TaqI, ApaI, BsmI, and FokI polymorphisms was associated with a higher risk of SLE. Additionally, individuals with SLE had significantly higher serum concentrations of vitamin D.

SLE is associated with the overproduction of cytokines. Shifts in the balance of Th1 and Th2 toward the overproduction of Th2 cytokines mediate hyperactivity of B cells, which promote plasma cell differentiation and antibody production^{30–32}. The increase in the generation of autoantibodies and the deposition of immune complexes are the cornerstones of the clinical manifestations of SLE, such as lupus nephritis, cytopenias, and arthralgias^{33–35}.

Vitamin D allows for greater tolerance by limiting the production of pro-inflammatory cytokines such as IL-12, which is a polarizing cytokine of T cells in Th1 cells and increases the production of anti-inflammatory cytokines such as IL-10. Consequently, it decreases autoreactivity and is one of the many mechanisms by which vitamin D can decrease the expression of autoimmune diseases^{9,10,36}.

Abou-Raya et al. 37 evaluated the effects of vitamin D supplementation on inflammatory cytokines (IL-1, IL-6 and TNF α), as well as the production of anti-dsDNA and anti-Sm in patients with SLE with vitamin D insufficiency. The levels of IL-1, IL-6, and TNF α decreased significantly, and the Systemic Lupus Erythematosus Activity Index (SLEDAI) scores improved in patients with SLE who received vitamin D supplementation, compared to the placebo group The immunomodulatory role of vitamin D is highlighted, demonstrating that deficiency is associated with higher rates of activity in autoimmune diseases, including SLE 9,10,36 .

The effects of vitamin D are mediated by the high affinity of the VDR which binds to the active form of vitamin D 1,25(OH)2D3 and is distributed in various tissues^{17,36}. The VDR gene is highly polymorphic; however, the polymorphisms rs7975232 and rs7975232 in intron 8, rs731236 in exon 9, and rs2228570 in the start codon are the most studied variants³⁹. In recent years, different studies have yielded conflicting results

on the association between VDR gene polymorphisms and the risk of SLE^{9,10,23,40,41}. Possible reasons for this disparity may be small sample sizes, low statistical power, or clinical heterogeneity⁸.

One meta-analysis included 11 studies with 1621 cases and 1883 controls that associated the BsmI, FokI, ApaI, or TaqI polymorphisms of the VDR gene with the risk of SLE⁴¹. An association between the BsmI B allele and the occurrence of SLE was observed in the general (OR: 1.726; 95% CI: 1.214–2.455) and Asian (OR: 1.952; 95% CI: 1.135–3.355) populations. The FokI T/C and TaqI polymorphisms were not associated with the risk of SLE in Caucasians. There was no significant association between the ApaI polymorphism and the risk of SLE for the general, Asian, and Caucasian populations⁴¹. In a study carried out by Monticielo et al.⁴² the VDR rs1544410 and rs2228570 polymorphisms were not associated with SLE in the European population, but higher concentrations of 1,25(OH)2D3 were found in patients carrying the rs2228570 ff42 genotype.

In our population, no significant associations were found between the BsmI, ApaI, and TaqI polymorphisms and the risk of SLE. However, greater susceptibility was shown through the FokI polymorphism in allelic models, only in agreement with series in the Arab population, such as those described by Imam et al.⁴³ and Emerah and El-Shal⁴⁰, with risks of susceptibility to SLE with OR: 1.617; 95% CI: 1.282–2.041; p < 0.000, and 1.948; 95% CI: 1.330–2.853; p < 0.001, respectively.

Emerah and El-Shal⁴⁰ conducted a study to assess the VDR gene polymorphisms and haplotypes (ApaI, BsmI, and FokI) as risk factors or markers of SLE activity, finding that the ApaI AA genotype, the BsmI allele B, Bb, BB genotypes, FokI F allele, and VDR genotype FF were increased in the SLE group. In addition, serum levels of vitamin D 1,25(OH)2D3 were increased in SLE patients carrying the FokI ff genotype compared to subjects carrying the FF genotype.

In another meta-analysis (13 studies), Zhou et al. 44 observed the association between the rs1544410B and rs2228570f alleles and susceptibility to SLE in the general population, Asians, and Africans, but not in Caucasians. In Africans, the rs1544410 BB/bb, rs2228570 ff, and rs7975232 AA/aa genotypes were associated with SLE. However, there was no significant association between VDR rs1544410, rs2228570, rs7975232, and rs731236 and susceptibility to SLE in Caucasians.

Exposure to sunlight in SLE has a double effect, since ultraviolet B rays are essential for the cutaneous synthesis of vitamin D and, at the same time, is a possible trigger for a SLE flare^{45,46}. Children with SLE are prone to vitamin D deficiency, as they avoid sunlight due to photosensitivity, use of sunscreen, physical inactivity secondary to pain or fatigue, and chronic use of medications such as corticosteroids and antimalarials that can improve vitamin D clearance^{47–51}.

Hypovitaminosis D is associated with higher complement consumption and higher rates of disease activity. Ospina-Caicedo et al. 24 , in a cross-sectional study of 69 patients with SLE, found that 36.2% of them had low levels of vitamin D, which was higher than our study (11.3%); the lowest levels were found in patients with moderate to severe activity by SLEDAI score. In addition, higher consumption of complement proteins and high anti-dsDNA titers (p = 0.006) were

evidenced. Patients who received steroids $(0.5-1\,\text{mg/kg})$ had lower mean levels of vitamin D compared with those who did not (p=0.048). Therefore, it is important to consider vitamin D supplementation in patients with SLE, especially when receiving corticosteroids.

Azab et al.51 showed that hypovitaminosis D is highly prevalent (57%) among Egyptian children and adolescents with SLE. Moreover, Yap et al. 50 detected a vitamin D deficiency in more than a quarter of Australian adult individuals with SLE, with a high proportion of Caucasians in their cohort. A multicenter French study by Schoindre et al. 49 reported similar findings, and the authors concluded that suboptimal vitamin D status was observed in more than 80% of SLE subjects. However, contrary to most reports, in our study higher levels of vitamin D were found in cases with SLE than in controls, which suggests that our population resides in areas that benefit from sufficient sunlight; in addition, it is also a signal of an adequate vitamin D supplementation scheme in corticotherapy; moreover, it could suggest a lower presence of anti-vitamin D antibodies between cases, which requires evaluation in future studies⁵².

In conclusion, the present study demonstrated the association between the VDR FokI rs2228570 polymorphism in allelic models and the ACCA haplotype of the TaqI, ApaI, BsmI, and FokI polymorphisms and an increased risk of SLE in the adolescent population. More studies are needed to assess the effects of these polymorphisms in SLE, especially their functional manifestations in other ethnic groups with larger sample sizes to confirm or refute our findings.

Limitations

To our knowledge, this is the first study that analyses VDR gene polymorphisms in adolescents with SLE from the Colombian Caribbean. However, some limitations in our study may affect the results. One of them was the failure to obtain clinical and immunological variables that would limit the establishment of associations between vitamin D levels, VDR gene polymorphisms, and SLE activity; furthermore, the sample size was relatively small. Because the results of the association between VDR polymorphisms and SLE have been different in different races and ethnic groups, more research is needed to distinguish such variables in our region.

Financing

The study was co-financed by Minciencias, Universidad Simon Bolivar, and Universidad del Norte, through project No. 840 of 2017, call 777 in the city of Barranquilla, Colombia.

Conflict of interests

None.

Acknowledgments

The authors wish to express their gratitude to the Universidad Simon Bolivar and the Universidad del Norte for funding this study. We would also like to express our gratitude to all the participating patients.

REFERENCES

- Lightstone L. Lupus nephritis: Where are we now? Curr Opin Rheumatol. 2010;22:252–6, http://dx.doi.org/10.1097/BOR.0b013e3283386512.
- Davidson A. What is damaging the kidney in lupus nephritis? Nat Rev Rheumatol. 2016;12:143–53, http://dx.doi.org/10.1038/nrrheum.2015.159.
- 3. Kashgarian M. Lupus nephritis: Pathology, pathogenesis, clinical correlations, and prognosis. In: Wallace D, Hahn B, Dubois E, editors. Dubois' Lupus erythematosus. 4th ed. Lippincott Williams & Wilkins; 2002. p. 1061–76.
- Salimi S, Mohammadoo-Khorasani M, Tabatabai E, Sandoughi M, Zakeri Z, Naghavi A. XRCC1 Arg399Gln and Arg194Trp polymorphisms and risk of systemic lupus erythematosus in an Iranian population: A pilot study. Biomed Res Int. 2014;2014:1–5, http://dx.doi.org/10.1155/2014/492956.
- Moudi B, Salimi S, Farajian Mashhadi F, Sandoughi M, Zakeri Z. Association of FAS and FAS ligand genes polymorphism and risk of systemic lupus erythematosus. Sci World J. 2013;2013:1–6, http://dx.doi.org/10.1155/2013/176741.
- Cui Y, Sheng Y, Zhang X. Genetic susceptibility to SLE: Recent progress from GWAS. J Autoimmun. 2013;41:25–33, http://dx.doi.org/10.1016/j.jaut.2013.01.008.
- Narooie-Nejad M, Moossavi M, Torkamanzehi A, Moghtaderi A, Salimi S. Vitamin D receptor gene polymorphism and the risk of multiple sclerosis in South Eastern of Iran. J Mol Neurosci. 2015;56:572–6, http://dx.doi.org/10.1007/s12031-015-0513-x.
- Bae SC, Lee YH. Vitamin D receptor Fokl, Taql, and Apal polymorphisms and susceptibility to systemic lupus erythematosus: an updated meta-analysis. Clin Rheumatol. 2018;37:1529–37, http://dx.doi.org/10.1007/s10067-018-4036-z.
- Monticielo OA, Teixeira TDM, Chies JAB, Brenol JCT, Xavier RM. Vitamin D and polymorphisms of VDR gene in patients with systemic lupus erythematosus. Clin Rheumatol. 2012;31:1411–21, http://dx.doi.org/10.1007/s10067-012-2021-5.
- Watad A, Neumann SG, Soriano A, Amital H, Shoenfeld Y. Vitamin D and systemic lupus erythematosus: Myth or reality? Isr Med Assoc J. 2016;18:177–82.
- Boonstra A, Barrat FJ, Crain C, Heath VL, Savelkoul HF, O'Garra A. 1alpha,25-Dihydroxyvitamin d3 has a direct effect on naive CD4(+) T cells to enhance the development of Th2 cells. J Immunol. 2001;167:4974–80, http://dx.doi.org/10.4049/jimmunol.167.9.4974.
- Miyamoto K, Kesterson RA, Yamamoto H, Taketani Y, Nishiwaki E, Tatsumi S, et al. Structural organization of the human vitamin D receptor chromosomal gene and its promoter. Mol Endocrinol. 1997;11:1165–79, http://dx.doi.org/10.1210/mend.11.8.9951.
- von Essen MR, Kongsbak M, Schjerling P, Olgaard K, Odum N, Geisler C. Vitamin D controls T cell antigen receptor signaling and activation of human T cells. Nat Immunol. 2010;11:344–9, http://dx.doi.org/10.1038/ni.1851.
- Smith-Garvin JE, Koretzky GA, Jordan MS. T cell activation. Annu Rev Immunol. 2009;27:591–619, http://dx.doi.org/10.1146/annurev.immunol.021908.132706.
- Ghoreishi M, Bach P, Obst J, Komba M, Fleet JC, Dutz JP. Expansion of antigen-specific regulatory T cells with the topical vitamin d analog calcipotriol. J Immunol. 2009;182:6071–8, http://dx.doi.org/10.4049/jimmunol.0804064.
- 16. Gorman S, Kuritzky LA, Judge MA, et al. Topically applied 1,25-dihydroxyvitamin D3 enhances the suppressive activity

- of CD4+CD25+ cells in the draining lymph nodes. J Immunol. 2007;179:6273–83, $\,$
- http://dx.doi.org/10.4049/jimmunol.179.9.6273.
- 17. Chen S, Sims GP, Chen XX, Gu YY, Chen S, Lipsky PE. Modulatory effects b of 1,25-dihydroxyvitamin D3 on human B cell differentiation. J Immunol. 2007;179:1634–47, http://dx.doi.org/10.4049/jimmunol.179.3.1634.
- Sabio JM, Vargas-Hitos JA, Martinez-Bordonado J, Navarrete-Navarrete N, Díaz-Chamorro A, Olvera-Porcel C, et al. Association between low 25-hydroxyvitamin D, insulin resistance and arterial stiffness in nondiabetic women with systemic lupus erythematosus. Lupus. 2015;24:155–63, http://dx.doi.org/10.1177/0961203314551811.
- Koizumi T, Nakao Y, Matsui T, Nakagawa T, Matsuda S, Komoriya K, et al. Effects of corticosteroid and 1,24R-dihydroxy-vitamin D3 administration on lymphoproliferation and autoimmune disease in MRL/MP-lpr/lpr mice. Int Arch Allergy Appl Immunol. 1985;77:396–404, http://dx.doi.org/10.1159/000233815.
- 20. Di Rosa M, Malaguarnera M, Nicoletti F, Malaguarnera L. Vitamin D3: A helpful immuno-modulator. Immunology. 2011;134:123–39, http://dx.doi.org/10.1111/j.1365-2567.2011.03482.x.
- Cauci S, Maione V, Buligan C, Linussio M, Serraino D, Stinco G. BsmI (rs1544410) and FokI (rs2228570) vitamin D receptor polymorphisms, smoking, and body mass index as risk factors of cutaneous malignant melanoma in northeast Italy. Cancer Biol Med. 2017;14:302–18, http://dx.doi.org/10.20892/j.issn.2095-3941.2017.0064.
- Jiang LL, Zhang C, Zhang Y, Ma F, Guan Y. Associations between polymorphisms in VDR gene and the risk of osteoporosis: A meta-analysis. Arch Physiol Biochem. 2020:1–8, http://dx.doi.org/10.1080/13813455.2020.1787457.
- Zhang WT, Jin TF, Chen L. Associations of four common VDR polymorphisms with rheumatoid arthritis and systemic lupus erythematosus: Evidence from a meta-analysis. Lupus. 2020;29:364–70, http://dx.doi.org/10.1177/0961203320909 432.
- 24. Ospina-Caicedo AI, Cardona-Rincón AD, Bello-Gualtero JM, Valle-Oñate R, Romero-Sánchez C, Chalem-Choueka P, et al. Lower levels of vitamin D associated with disease activity in Colombian patients with systemic lupus erythematosus. Curr Rheumatol Rev. 2019;15:146–53, http://dx.doi.org/10.2174/1573397114666181015161547.
- De Azevedo Silva J, Addobbati C, Sandrin-Garcia P, Crovella S. Systemic lupus erythematosus: old and new susceptibility genes versus clinical manifestations. Curr Genomics. 2014;15:52–65, http://dx.doi.org/10.2174/138920291501140306113715.
- Hahn BH, McMahon MA, Wilkinson A, Wallace WD, Daikh DI, Fitzgerald JD, et al. American College of Rheumatology guidelines for screening, treatment, and management of lupus nephritis. Arthritis Care Res (Hoboken). 2012;64:797–808, http://dx.doi.org/10.1002/acr.21664.
- Miller SA, Dykes DD, Polesky HF. A simple salting out procedure for extracting DNA from human nucleated cells. Nucleic Acids Res. 1988;16:1215, http://dx.doi.org/10.1093/nar/16.3.1215.
- Ross AC, Manson JE, Abrams SA, Aloia JF, Brannon PM, Clinton SK, et al. The 2011 report on dietary reference intakes for calcium and vitamin D from the Institute of Medicine: What clinicians need to know. J Clin Endocrinol Metab. 2011;96:53–8, http://dx.doi.org/10.1210/jc.2010-2704.
- 29. R Core Team. https://www.R-project.org/, 2018.
- Muhammad Yusoff F, Wong KK, Mohd Redzwan N. Th1, Th2, and Th17 cytokines in systemic lupus erythematosus. Autoimmunity. 2020;53:8–20, http://dx.doi.org/10.1080/08916934.2019.1693545.

- 31. Akahoshi M, Nakashima H, Tanaka Y, Kohsaka T, Nagano S, Ohgami E, et al. Th1/Th2 balance of peripheral T helper cells in systemic lupus erythematosus. Arthritis Rheum. 1999;42:1644–8,
 - http://dx.doi.org/10.1002/1529-0131(199908)42:8<1644::AID-ANR12>3.0.2012;21:43-52, http://dx.doi.org/10.1177/0961203311421798.
- 32. Chan RW, Lai FM, Li EK, Tam LS, Chow KM, Li PK, et al. Imbalance of Th1/Th2 transcription factors in patients with lupus nephritis. Rheumatology. 2006;45:951–7, http://dx.doi.org/10.1093/rheumatology/kel029.
- 33. Moulton VR, Suarez-Fueyo A, Meidan E, Li H, Mizui M, Tsokos GC. Pathogenesis of human systemic lupus erythematosus: a cellular perspective. Trends Mol Med. 2017;23:615–35, http://dx.doi.org/10.1016/j.molmed.2017.05.006.
- Wigren M, Nilsson J, Kaplan MJ. Pathogenic immunity in systemic lupus erythematosus and atherosclerosis: common mechanisms and possible targets for intervention. J Intern Med. 2015;278:494–506, http://dx.doi.org/10.1111/joim.12357.
- 35. Han S, Zhuang H, Shumyak S, Yang L, Reeves WH.

 Mechanisms of autoantibody production in systemic lupus
 erythematosus. Front Immunol. 2015;6:228,
 http://dx.doi.org/10.3389/fimmu.2015.00228.
- Aranow C. Vitamin D and the immune system. J Investig Med. 2011;59:881–6, http://dx.doi.org/10.2310/JIM.0b013e31821b8755.
- 37. Abou-Raya A, Abou-Raya S, Helmii M. The effect of vitamin D supplementation on inflammatory and hemostatic markers and disease activity in patients with systemic lupus erythematosus: A randomized placebo-controlled trial. J Rheumatol. 2013;40:265–72, http://dx.doi.org/10.3899/jrheum.111594.
- 38. Uribe AG, Vilá LM, McGwin G, Sanchez ML, Reveille JD, Alarcón GS. The systemic lupus activity measure-revised, the Mexican systemic lupus erythematosus disease activity Index (Sledai), and a modified Sledai-2K are adequate instruments to measure disease activity in systemic lupus erythematosus. J Rheumatol. 2004;31:1934–40 http://www.ncbi.nlm.nih.gov/pubmed/15468356
- 39. Xiong DH, Xu FH, Liu PY, Shen H, Long JR, Elze L, et al. Vitamin D receptor gene polymorphisms are linked to and associated with adult height. J Med Genet. 2005;42:228–34, http://dx.doi.org/10.1136/jmg.2004.024083.
- Emerah AA, El-Shal AS. Role of vitamin D receptor gene polymorphisms and serum 25-hydroxyvitamin D level in Egyptian female patients with systemic lupus erythematosus. Mol Biol Rep. 2013;40:6151–62, http://dx.doi.org/10.1007/s11033-013-2726-9.
- 41. Mao S, Huang S. Association between vitamin D receptor gene BsmI, FokI, ApaI and TaqI polymorphisms and the risk of systemic lupus erythematosus: A meta-analysis. Rheumatol Int. 2014;34:381–8, http://dx.doi.org/10.1007/s00296-013-2898-6.

- 42. Monticielo OA, Brenol JC, Chies JA, Longo MG, Rucatti GG, Scalco R, et al. The role of BsmI and FokI vitamin D receptor gene polymorphisms and serum 25-hydroxyvitamin D in Brazilian patients with systemic lupus erythematosus. Lupus.
- 43. Imam AA, Ibrahim HE, Farghaly MAA, Alkholy UM, Gawish HH, Abdalmonem N, et al. Vitamin D receptor gene FokI polymorphism in Egyptian children and adolescents with SLE: A case-control study. Lupus. 2017;26:1426–34, http://dx.doi.org/10.1177/0961203317725588.
- 44. Zhou TB, Jiang ZP, Lin ZJ, Su N. Association of vitamin D receptor gene polymorphism with the risk of systemic lupus erythematosus. J Recept Signal Transduct Res. 2015;35:8–14, http://dx.doi.org/10.3109/10799893.2014.922577.
- Barbhaiya M, Costenbader KH. Ultraviolet radiation and systemic lupus erythematosus. Lupus. 2014;23:588–95, http://dx.doi.org/10.1177/0961203314530488.
- Barbhaiya M, Costenbader KH. Environmental exposures and the development of systemic lupus erythematosus. Curr Opin Rheumatol. 2016;28:497–505, http://dx.doi.org/10.1097/BOR.000000000000318.
- Ahluwalia J, Marsch A. Photosensitivity and photoprotection in patients with lupus erythematosus. Lupus. 2019;28:697–702, http://dx.doi.org/10.1177/0961203319839486.
- Zandman-Goddard G, Solomon M, Rosman Z, Peeva E, Shoenfeld Y. Environment and lupus-related diseases. Lupus. 2012;21:241–50, http://dx.doi.org/10.1177/0961203311426568.
- 49. Schoindre Y, Jallouli M, Tanguy ML, Ghillani P, Galicier L, Aumaître O, et al. Lower vitamin D levels are associated with higher systemic lupus erythematosus activity, but not predictive of disease flare-up. Lupus Sci Med. 2014;1:e000027, http://dx.doi.org/10.1136/lupus-2014-000027.
- 50. Yap KS, Northcott M, Hoi AB-Y, Morand EF, Nikpour M. Association of low vitamin D with high disease activity in an Australian systemic lupus erythematosus cohort. Lupus Sci Med. 2015;2:e000064, http://dx.doi.org/10.1136/lupus-2014-000064.
- 51. Azab SF, Ali YF, Farghaly MAA, Hamed ME, Allah MAN, Emam AA, et al. Vitamin D receptor gene BsmI polymorphisms in Egyptian children and adolescents with systemic lupus erythematosus: A case-control study. Medicine (Baltimore). 2016;95:e5233,
 - http://dx.doi.org/10.1097/MD.000000000005233.
- 52. Carvalho JF, Blank M, Kiss E, Tarr T, Amital H, Shoenfeld Y. Anti-vitamin D, vitamin D in SLE: Preliminary results. Ann N Y Acad Sci. 2007;1109:550–7, http://dx.doi.org/10.1196/annals.1398.061.