



## Original Investigation

# Frequency of anti-neutrophil cytoplasmic antibodies in patients with systemic lupus erythematosus



Evelyn Santacruz-Sandoval<sup>a</sup>, Cristian C. Aragón<sup>b</sup>, Ivana Nieto-Aristizábal<sup>c</sup>,  
Ingrid Ruiz-Ordoñez<sup>c</sup>, Jhoan S. Quijano<sup>d</sup>, Daniela Ramírez-Castillo<sup>a</sup>,  
Karen Mondragón M<sup>a</sup>, Iván Posso-Osorio<sup>c</sup>, María Claudia Barrera<sup>b</sup>, Lady J. Rios-Serna<sup>b</sup>,  
Gabriel J. Tobón<sup>b,e,\*</sup>

<sup>a</sup> Department of Internal Medicine, Medical School, Universidad Icesi, Cali, Colombia

<sup>b</sup> Department of Internal Medicine and Rheumatology, Medical School, Centro CIRAT, Universidad Icesi, Cali, Colombia

<sup>c</sup> Clinical Research Center, Fundación Valle del Lili, Cali, Colombia

<sup>d</sup> Department of Internal Medicine, Medical School, Universidad del Valle, Cali, Colombia

<sup>e</sup> Unit of Rheumatology, Fundación Valle del Lili, Cali, Colombia

## ARTICLE INFO

### Article history:

Received 23 September 2020

Accepted 22 January 2021

### Keywords:

Antibodies, antineutrophil  
cytoplasmic  
Lupus erythematosus, systemic  
Immunofluorescence technique,  
indirect  
Enzyme-linked immunosorbent  
assay

## ABSTRACT

**Introduction:** Systemic lupus erythematosus (SLE) is a multisystem autoimmune disease. Patients with SLE exhibit multiple serum autoantibodies, including anti-neutrophil cytoplasmic antibodies (ANCA). There are two main techniques to detect ANCAs: indirect immunofluorescence (IIF) and enzyme-linked immunosorbent assay (ELISA). In this study, an attempt was made to determine the frequency and clinical associations of ANCAs in patients with SLE.

**Methods:** A cross-sectional study was conducted in a tertiary care hospital in Colombia that included 74 patients with SLE. The presence of ANCAs was assessed using IIF with ethanol-fixed slides, and ELISA was used to detect antibody specificities for myeloperoxidase (MPO)-ANCA and proteinase 3 (PR3)-ANCA.

**Results:** Of the 74 patients with SLE evaluated, 60 (81.1%) of them were ANCA-positive by IIF. By contrast, only one patient showed specificity for PR3-ANCA by ELISA. The relevance of ANCA positivity by IIF and clinical and serological features was significant for renal involvement ( $p = .0174$ ), and Systemic Lupus Erythematosus Disease Activity Index (SLEDAI) ( $p = .0308$ ).

**Conclusion:** ANCAs are common in the serum of patients with SLE, as detected by ethanol-fixed slides with IIF staining. However, detection of specificity to PR3 and/or MPO is rare, thus highlighting the importance of detecting these autoantibodies by different techniques.

© 2021 Asociación Colombiana de Reumatología. Published by Elsevier España, S.L.U. All rights reserved.

\* Corresponding author.

E-mail address: [gtobon1@yahoo.com](mailto:gtobon1@yahoo.com) (G.J. Tobón).

## Frecuencia de los anticuerpos anticitoplasma de neutrófilo en pacientes con lupus eritematoso sistémico

### R E S U M E N

#### Palabras clave:

Anticuerpos anticitoplasma de neutrófilos  
Lupus eritematoso sistémico  
Técnica del anticuerpo fluorescente indirecta  
Ensayo de inmunoadsorción enzimática

**Introducción:** El lupus eritematoso sistémico (LES) es una enfermedad autoinmune sistémica. Los pacientes con LES muestran múltiples autoanticuerpos séricos, incluyendo los anticuerpos anticitoplasma de neutrófilo (ANCA, por sus siglas en inglés). Existen 2 técnicas principales para la detección de ANCA: inmunofluorescencia indirecta (IFI) y ensayo por inmunoadsorción ligado a enzimas (ELISA). En este estudio nuestro objetivo fue determinar la frecuencia y las asociaciones clínicas de los ANCA en pacientes con LES.

**Métodos:** Realizamos un estudio transversal de 74 pacientes con LES en un hospital de alta complejidad de Colombia. La presencia de ANCA se evaluó por IFI, utilizando láminas con fijación de etanol, y con ELISA para determinar las especificidades para mieloperoxidasa (MPO)-ANCA y proteinasa 3 (PR3)-ANCA.

**Resultados:** Fueron evaluados 74 pacientes con LES, 60 (81,1%) de ellos fueron positivos para ANCA. Por el contrario, solo un paciente mostró especificidad para PR3-ANCA por ELISA. La relación entre la positividad para ANCA por IFI y las características clínicas y serológicas fue estadísticamente significativa para compromiso renal ( $p=0,0174$ ) y para el índice de actividad de la enfermedad (Systemic Lupus Erythematosus Disease Activity Index [SLEDAI]) ( $p=0,0308$ ).

**Conclusiones:** Los ANCA detectados mediante fijación con etanol por técnicas de IFI, son comunes en pacientes con LES. Sin embargo, la detección de especificidades para PR3 o MPO es rara; se destaca la importancia de la evaluación de estos autoanticuerpos mediante diferentes técnicas.

© 2021 Asociación Colombiana de Reumatología. Publicado por Elsevier España, S.L.U.

Todos los derechos reservados.

## Introduction

Systemic lupus erythematosus (SLE) is a multisystemic autoimmune disorder characterized by the presence of a wide range of serum autoantibodies such as antinuclear (ANA) and anti-double-stranded deoxyribonucleic acid (anti-dsDNA) antibodies. Furthermore, the serum of patients with SLE exhibits antineutrophil cytoplasmic antibodies (ANCAs).<sup>1</sup> ANCAs are a group of autoantibodies directed against proteins predominantly expressed in the cytoplasmic granules of neutrophils. Small vessels are the usual target of accumulating ANCA autoantibodies, which can affect any organ in the body in patients with ANCA-associated vasculitis.

Two main laboratory techniques are currently used to detect ANCAs. The standard method for ANCA detection in the serum is indirect immunofluorescence (IIF) of ethanol-fixed human neutrophils smeared onto glass slides. Other fixation methods (e.g., formalin) can also be used. Classical patterns evidenced in neutrophil IIF are cytoplasmic fluorescence with central interlobular accentuation (c-ANCA), which usually occurs with proteinase 3 (PR3)-ANCA specificity, and perinuclear fluorescence often with a nuclear extension (p-ANCA), which frequently occurs with myeloperoxidase (MPO)-ANCA specificity.<sup>2</sup> Additionally, atypical ANCA patterns can be detected in ethanol-fixed neutrophils in the presence of ANA and other autoantibodies against several antigens (such as lactoferrin, lysozyme, azurocidin, elastase, cathepsin G, bactericidal/permeability-increasing enzyme, among others) present in the cytoplasmic granules of the neutrophils.<sup>3,4</sup>

Enzyme-linked immunosorbent assay (ELISA) can detect more specific targets and is mainly used for MPO-ANCA and PR3-ANCA.<sup>5</sup> Likewise, in some studies, testing positive for ANCA by ELISA (MPO-ANCA and PR3-ANCA) has been associated with the worst prognosis in patients with SLE and lupus nephritis (LN).<sup>6</sup>

Given these techniques and the incomplete or conflicting information linking ANCA detection and patterns with clinical outcomes, we sought herein to determine the frequency and clinical associations of ANCAs in patients with SLE from a cohort of Colombian SLE patients.

## Materials and methods

We conducted a cross-sectional study of the serum of 74 patients with SLE evaluated between January 2018 and June 2019 in the Rheumatology service of the "Fundación Valle del Lili," a tertiary care hospital in Colombia. All patients signed an informed consent authorizing their participation in the study. Patient charts and immunological profiles were reviewed from clinical records. All patients met the SLE classification criteria of the Systemic Lupus International Collaborating Clinics (SLICC) 2012 and new SLE classification criteria of the American College of Rheumatology 2019. Renal involvement was defined as the presence of hematuria, proteinuria, or active urinary sediment. On the other hand, confirmed LN was grouped into proliferative and non-proliferative upon renal

biopsy reports. Combinations such as III + IV, III + V, IV + V, and IV + VI were classified as proliferative.

ANCAs were measured by IIF with the NOVA Lite® ANCA kit (Inova Diagnostics, San Diego, CA, USA) using ethanol-fixed slides. A 1:10 dilution of each sample was used for screening, and those showing fluorescence were classified as positive, and were also classified by intensity, ranging from 1+ to 4+, according to manufacturer's specifications. The c-ANCA, p-ANCA, or atypical patterns were recorded for each case. Antibody specificities for myeloperoxidase (MPO)-ANCA and proteinase 3 (PR3)-ANCA were determined using ELISA kits with an Alegria® automatic ELISA reader (Orgentec Diagnostika, Mainz, Germany) with a cutoff of  $\geq 5$  U/mL. The assays were performed according to the manufacturer's instructions.

### Statistical analysis

IIF and ELISA results were compared with the clinical characteristics, immunological profiles, and the Systemic Lupus Erythematosus Disease Activity Index (SLEDAI) assigned according to IIF results. Normality distribution data were evaluated using the Kolmogorov-Smirnov test. Continuous variables are shown as medians (interquartile range, IQR) or means ( $\pm$  standard deviation). Categorical variables are shown as frequencies and percentages and were compared by chi-squared test or Fisher's test as appropriate. The Mann-Whitney test was used for continuous variables. All *p* values were two-tailed, and *p* values  $< 0.05$  were considered statistically significant.

## Results

### ANCA by IIF

Seventy-four patients with SLE (94% women) were included, with a median age of 30 years (IQR 21–40).

Sixty (81.1%) patients showed ANCA positivity by IIF of ethanol-fixed slides. The majority of patients (*n*=31) presented a fluorescence intensity of 4+, followed by fifteen with 3+, six with 2+, and 8 with 1+. Fifty-eight (96.6%) of these 60 patients presented the p-ANCA pattern, while only two (3.33%) displayed the c-ANCA pattern. None of the patients exhibited an atypical pattern. The association between clinical and serological characteristics of patients with SLE according to their ANCA positivity is shown in Table 1. Renal involvement was found in 35 cases (58.3%) at inclusion. The median score of SLEDAI was 10 (IQR: 3–18). ANCA positivity by IIF was associated with renal involvement (*p*=0.0174) and lower values of SLEDAI (*p*=0.0308), compared to a median score of 11 in those with negative ANCAs by IIF; although there is a minimum difference between both scores, the statistical significance might be attributable to the number of patients compared (60 vs. 14) (Table 1).

In terms of LN, twenty-nine patients (39.2%) had the diagnosis confirmed by renal biopsy. Of those, 27 presented positive IIF test results. We then evaluated if there was any relation between ANCAs positivity and crescents, fibrinoid necrosis, vasculopathy, thrombosis; LN type; activity, or

chronicity index, but we did not find any significant result (Table 2).

Moreover, twenty-five patients with ANCAs by IIF of 3+ or 4+ did not have LN at the time of the blood sample. Of them, 17 had subsequent medical records of control appointments with a rheumatologist, of which three patients did develop LN; two of them had the confirmatory renal biopsy one month and 21 months after the blood sample, respectively. On the other hand, 14 did not develop LN by the last follow-up available, which were 21 (IQR 4.5–28.5) months apart from the blood sample.

### ANCAs by ELISA

ELISA testing produced only one case of PR3-ANCA with a result of 8.1 U/mL, and no patient displayed positivity for MPO-ANCA.

## Discussion

We carried out a cross-sectional study in 74 patients with SLE to evaluate the presence of ANCAs by IIF and ELISA. Because only one patient was positive by ELISA, we compared clinical and serological features of those with ANCA-positive IIF results. The high discrepancy between IIF and ELISA tests may result from different factors.

For IIF, ethanol is the most frequently used fixative agent. This technique causes solubilization of the granule membranes, allowing mobilization of their content. Cell changes can lead to the recognition of different antigens of several granulocyte components other than MPO or PR3, including lactoferrin, cathepsin G, elastase, lysozyme, bacterial permeability-increasing protein, catalase,  $\alpha$ -enolase, lamin B1, among others.<sup>4,5,7</sup> By contrast, the use of the cross-linking fixative formaldehyde prevents this redistribution.<sup>6</sup> The use of ethanol or formalin as fixative agents allows the distinction of true p-ANCA or c-ANCA patterns from those directed against other neutrophil targets. Looking for other antigen specificities by ELISA for routine purposes is not recommended.<sup>8</sup>

In the same way, ANAs positivity can interfere with ANCAs, yielding a p-ANCA pattern on the ethanol-fixed neutrophil substrate. The use of formalin-fixed neutrophils overcomes this problem, helping to distinguish similar fluorescent staining that occurs due to the presence of ANAs.<sup>9,10</sup> Herein, however, we used only ethanol as a fixative which could have caused the high proportion of positive ANCAs in our sample. The search for other antigen specificities or different fixation techniques may be pursued in other studies.

ANCAs detection may play a role in patients with SLE and LN.<sup>11</sup> Currently, different studies show an association with worse renal outcomes and poor prognosis in patients with LN and ANCAs positivity (Table 3).<sup>12</sup> The presence of ANCAs may, therefore, correlate with more severe clinical outcomes, histological necrosis, and crescentic glomerulonephritis. Indeed, ANCAs are directly implicated in the pathogenesis of this form of glomerular injury and are thought to activate cytokine-primed neutrophils and monocytes expressing ANCA-PR3 and MPO on their surfaces.<sup>13</sup>

**Table 1 – ANCA test results by IIF and presence of clinical characteristics and other autoantibodies in SLE patients.**

Clinical/Serological characteristics	Patients n = 74 (%)	Positive ANCA by IIF test n = 60 (%)	Negative ANCA by IIF test n = 14 (%)	p value
Age, year's	30 (21–40)**	30 (22–42)**	27 (20–35)**	–
Women, n (%)	70 (94.5)	57 (95)	13 (92.8)	–
Associated autoimmune disease n (%)				
APS	12 (16.2)	9 (15)	3 (21.4)	0.6873
Evans syndrome	4 (5.4)	3 (5)	1 (7.14)	1.0
RA	4 (5.4)	2 (14.2)	2 (3.3)	1.0
Clinical manifestations, n (%)				
Renal involvement	38 (51.3)	35 (58.3)	3 (21.4)	0.0174 <sup>+</sup>
Arthritis	21 (28.3)	17 (28.3)	4 (28.5)	1.0
Hematological Involvement	21 (28.3)	19 (31.6)	2 (14.2)	0.3239
Neurological Involvement	16 (21.6)	14 (23.3)	2 (14.2)	0.5121
Rash	11 (14.8)	10 (16.6)	1 (7.1)	0.4553
Fever	10 (13.5)	8 (13.3)	2 (14.2)	1.0
Oral Ulcers	7 (9.4)	6 (10)	1 (7.14)	1.0
Alopecia	6 (8.1)	6 (10)	0	0.3477
Serositis	6 (8.1)	6 (10)	0	0.3477
Vasculitis	6 (8.1)	6 (10)	0	0.3477
Raynaud's	2 (2.7)	2 (3.3)	0	1.0
Pulmonary involvement	2 (2.7)	2 (3.3)	0	1.0
Myositis	2 (2.7)	2 (3.3)	0	1.0
Renal biopsy	33 (44.5)	31 (57.6)	2 (14.2)	–
Lupus nephritis	29 (39.1)	27 (45)	2 (14.2)	0.065
Positive autoantibodies, n (%)				
Anti-dsDNA (IIF)	36 (48.6)	29 (48.3)	7 (50)	1.0
Anti-dsDNA (ELISA)	49 (66.2)	38 (63.3)	11 (78.5)	0.3575
Anti-Ro60/SSA	21 (28.3)	19 (31.6)	2 (14.2)	0.3239
Anti-La/SSB	5 (6.7)	4 (6.6)	1 (7.1)	1.0
Anti-RNP	23 (31)	22 (36)	1 (7.1)	0.051
Anti-Sm	24 (32.4)	21 (35)	3 (21.4)	0.3686
↓C3 mg/dL	44 (59.4)	38 (63.3)	6 (42.8)	0.2274
↓C4 mg/dL	32 (43.2)	26 (43.3)	6 (42.8)	1.0
SLEDAI	10 (3–18)**	10 (3–18)**	11 (4–18)**	0.0308 <sup>+</sup>

\*Mean (SD), \*\*median (IQR), <sup>+</sup> p values <0.05 statistically significant; low serum C3 < 80 mg/dL; low serum C4 < 10 mg/dL; APS: Antiphospholipid syndrome; RA: Rheumatoid Arthritis. IIF: indirect immunofluorescence; ELISA: enzyme-linked immunosorbent assay; ANA: antinuclear antibodies; Anti-dsDNA: antidouble-stranded deoxyribonucleic acid; Anti-RNP: Anti ribonucleoprotein antibodies; Anti-Sm: Anti-Smith. SLEDAI: Systemic Lupus Erythematosus Disease Activity Index; ANCA: anti-neutrophil cytoplasmic antibodies; SLE: Systemic Lupus Erythematosus.

**Table 2 – ANCA test by IIF in patients with LN.**

Renal biopsy	ANCA by IIF		p value
	Positive n = 27	Negative n = 2	
Crescents	5	1	0.222
Fibrinoid necrosis	5	1	0.222
Vasculopathy	1	0	1
Thrombosis	0	0	–
LN type			
Proliferative	23	2	1
Non proliferative	4	0	1
Activity index	4 (2–6.75)	10*	0.0859**
Chronicity index	3 (1–4.25)	3*	1**

IIF: indirect immunofluorescence; ANCA: anti-neutrophil cytoplasmic antibodies; LN: Lupus nephritis.

\* Median. As there are only two cases, no IQR is available.

\*\* Mann-Whitney U test.

**Table 3 – Studies of ANCA in patients with SLE and LN.**

Study	Number of patients with SLE	ANCA by IIF	p-ANCA	c-ANCA	ANCA by ELISA	MPO-ANCA	PR3-ANCA	MPO-ANCA and PR3-ANCA	Clinical correlation with LN
Santacruz-Sandoval, 2020 (Our results)	74	60	58	2	1	–	1	–	–
Li C, 2019	4077	–	–	–	95	69	23	–	+
Pyo JY, 2019	91	12	–	–	12	11	1	–	+
Sobral S, 2018	243	72	64	8	72	1	1	–	+
Amann, 2017	254	32	–	–	14%	82%	7%	11%	–
Turner-Stokes T, 2017	254	32	–	–	–	82%	7%	11%	+
Jarrot PA, 2016	40	12	9	3	12	3	2	–	–

IIF: indirect immunofluorescence; ELISA: enzyme-linked immunosorbent assay; SLE: Systemic Lupus Erythematosus; ANCA: anti-neutrophil cytoplasmic antibodies; LN: Lupus nephritis.

Moreover, some studies found an association between ANCAs and LN's classification of the International Society of Nephrology (ISN)/Renal Pathology Society.<sup>14</sup> These findings show that positive ANCAs serology in LN was associated with LN's class IV-S histological and clinical phenotype.<sup>15,16</sup> In other studies, MPO-ANCA mainly contributed to the chronicity index of LN<sup>17</sup> and was positively correlated with class IV LN.<sup>18</sup> Nevertheless, Olson et al. found that elevated dsDNA antibody was not associated with elevated MPO-ANCA levels.<sup>19</sup> Interestingly, our results did not show any of the statements found by the mentioned authors; instead, no relation was found between histological findings frequently seen in the presence of ANCA, nor associations with any LN classification, neither with activity or chronicity indexes. Pan et al. and Wang et al. had similar results regarding LN classification, indexes, presence of crescents and necrosis; as no difference was found between ANCA-positive vs. ANCA-negative patients in their cohorts.<sup>20,21</sup>

Some authors maintain that no significant clinical or pathological differences in SLE patients are associated with the presence or absence of ANCAs, and no significant correlation is present between clinical features and the status of either p-ANCA or c-ANCA.<sup>22</sup> In one study, the presence of ANCAs in patients with SLE rarely affected the clinical manifestations of SLE,<sup>23</sup> and in another, there was no association of ANCA with renal involvement.<sup>24</sup> In addition, Pan et al. studied SLE 120 patients, of which 45 had positivity in p-ANCA by IIF, 75 were negative for p-ANCA and none of them was positive in ELISA techniques; in this case, they did find more occurrence of LN and more Raynaud syndrome in patients with positive p-ANCA vs those that were negative.<sup>20</sup>

Despite this, it is essential to note that ANCAs in LN are associated with a higher SLE disease activity index score.<sup>25</sup> Li et al. found that ANCA-positive LN patients had higher SLEDAI scores, which might be associated with higher hematuria because of aggravated kidney injuries caused by ANCAs.<sup>26</sup> Herein, we obtained a significant correlation between SLEDAI index and renal outcomes (hematuria, proteinuria, and the presence of urinary sediment).

Other studies have shown that ANCAs can also be detected in healthy subjects and in several inflammatory or autoimmune diseases where its pathophysiological significance

remains debatable.<sup>27</sup> Nevertheless, ANCAs is an independent risk factor for poor renal outcomes in LN patients.<sup>28</sup>

## Conclusion

In our study, we observed that most patients with SLE tested positive for ANCAs by IIF, but this was combined with PR3 specificity in only one case. This finding may be due to the presence of other target antigens. We found a statistically significant correlation between SLEDAI index and renal outcomes in patients with SLE who are positive for ANCAs, but not with LN classes or indexes. Further studies are needed to determine the specificity of ANCAs detected by IIF in our patients and others with SLE.

## Funding

Universidad Icesi supported research.

## Conflict of interests

The authors declare that they have no competing interests.

## REFERENCES

- Li C, Wang JJ, Zhou ML, Liang DD, Yang J, Zhu HX, et al. Differences in clinico-pathological characteristics and outcomes between proteinase 3-ANCA positivity and myeloperoxidase-ANCA positivity in lupus nephritis. *Lupus*. 2019;28:1111–9, <http://dx.doi.org/10.1177/0961203319861680>.
- Goldschmeding R, van der Schoot CE, ten Bokkel Huinink D, Hack CE, van den Ende ME, M Kallenberg CG, et al. Wegener's granulomatosis autoantibodies identify a novel diisopropylfluorophosphate-binding protein in the lysosomes of normal human neutrophils. *J Clin Invest*. 1989;84:1577–87, <http://dx.doi.org/10.1172/JCI114335>.
- Halbwachs-Mecarelli L, Nusbaum P, Noel LH, Reumaux D, Erlinger S, Gronfeld JP, et al. Antineutrophil cytoplasmic antibodies (ANCA) directed against cathepsin G in ulcerative colitis, Crohn's disease and primary sclerosing cholangitis.



- Clin Exp Immunol. 1992;90:79-84, <http://dx.doi.org/10.1111/j.1365-2249.1992.tb05835.x>.
4. Stoffel MP, Csernok E, Herzberg C, Johnston T, Carroll SF, Gross WL. Anti-neutrophil cytoplasmic antibodies (ANCA) directed against bactericidal/permeability increasing protein (BPI): a new seromarker for inflammatory bowel disease and associated disorders. Clin Exp Immunol. 1996;104:54-9, <http://dx.doi.org/10.1046/j.1365-2249.1996.d01-654.x>.
  5. Schulte-Pelkum J, Radice A, Norman GL, Lopez Hoyos M, Lakos G, Buchner C, et al. Novel clinical and diagnostic aspects of antineutrophil cytoplasmic antibodies. J Immunol Res. 2014;2014:1-12, <http://dx.doi.org/10.1155/2014/185416>.
  6. Radice A, Bianchi L, Sinico RA. Anti-neutrophil cytoplasmic autoantibodies: methodological aspects and clinical significance in systemic vasculitis. Autoimmun Rev. 2013;12:487-95, <http://dx.doi.org/10.1016/j.autrev.2012.08.008>.
  7. Lin MW, Silvestrini RA, Culican S, Campbell D, Fulcher DA. A dual-fixed neutrophil substrate improves interpretation of antineutrophil cytoplasmic antibodies by indirect immunofluorescence. Am J Clin Pathol. 2014;142:325-30, <http://dx.doi.org/10.1309/AJCPG02FGQVAUSIU>.
  8. Sobral S, Ramassur K, Apsley E, Isenberg D. Do anti-neutrophil cytoplasmic antibodies play a role in systemic lupus erythematosus (SLE) patients? Analysis of the University College Hospital SLE cohort. Lupus. 2018;27:343-4, <http://dx.doi.org/10.1177/0961203317724218>.
  9. Martínez Téllez G, Torres Rives B, Rangel Velázquez S, Sánchez Rodríguez V, Ramos Ríos MA, Fuentes Smith LE. Anticuerpos contra el citoplasma del neutrófilo: positividad y correlación clínica. Reumatol Clin. 2015;11:17-21, <http://dx.doi.org/10.1016/j.reuma.2014.02.010>.
  10. Romero-Sánchez C, Benavides-Solarte M, Galindo-Ibáñez I, Ospina-Caicedo AI, Parra-Izquierdo V, Chila-Moreno L, et al. Frequency of positive ANCA test in a population with clinical symptoms suggestive of autoimmune disease and the interference of ANA in its interpretation. Reumatol Clin. 2019;1984:1-12, <http://dx.doi.org/10.1016/j.reuma.2018.09.007>.
  11. Yang J, Liang D, Zhang H, Liu Z, Le W, Zhou M, et al. Long-term renal outcomes in a cohort of 1814 Chinese patients with biopsy-proven lupus nephritis. Lupus. 2015;24:1468-78, <http://dx.doi.org/10.1177/0961203315593166>.
  12. Tobar-Marcillo M, Destrugue-Molina I, Torres-Orozco L, Santiago-Ramirez R. Lupus nephritis associated with cytoplasmic anti-neutrophil cytoplasmic antibodies. Reumatol Clin. 2018;14:246-8, <http://dx.doi.org/10.1016/j.reuma.2017.07.012>.
  13. Koenig KF, Kalbermatter SA, Menter T, Mayr M, Kiss D. Rapidly progressive lupus nephritis with extremely high levels of antineutrophil cytoplasmic antibodies. Case Reports Nephrol Dial. 2014;4:5-11, <http://dx.doi.org/10.1159/000358557>.
  14. Weening JJ, D'agati VD, Schwartz MM, Seshan SV, Alpers CE, Appel GB, et al. The classification of glomerulonephritis in systemic lupus erythematosus revisited. Kidney Int. 2004;65:521-30, <http://dx.doi.org/10.1111/j.1523-1755.2004.00443.x>.
  15. Turner-Stokes T, Wilson HR, Morreale M, Nunes A, Cairns T, Cook HT, et al. Positive antineutrophil cytoplasmic antibody serology in patients with lupus nephritis is associated with distinct histopathologic features on renal biopsy. Kidney Int. 2017;92:1223-31, <http://dx.doi.org/10.1016/j.kint.2017.04.029>.
  16. Amann K. Do ANCAs make the difference in lupus nephritis? Kidney Int. 2017;92:1048-50, <http://dx.doi.org/10.1016/j.kint.2017.07.011>.
  17. Pyo JY, Jung SM, Song JJ, Park YB, Lee SW. ANCA positivity at the time of renal biopsy is associated with chronicity index of lupus nephritis. Rheumatol Int. 2019;39:879-84, <http://dx.doi.org/10.1007/s00296-019-04263-2>.
  18. Said D, Rashad NM, Abdelrahman NS, Dawaa GA. Antineutrophil cytoplasmic antibody in lupus nephritis: correlation with clinicopathological characteristics and disease activity. Curr Rheumatol Rev. 2020, <http://dx.doi.org/10.2174/1573397116999201208213422>.
  19. Olson SW, Lee JJ, Poirier M, Little DJ, Prince LK, Baker TP, et al. Anti-myeloperoxidase antibodies associate with future proliferative lupus nephritis. Autoimmune Dis. 2017;1-11, <http://dx.doi.org/10.1155/2017/1872846>, 2017.
  20. Pan Y, He L. Perinuclear anti-neutrophil cytoplasmic antibody in systemic lupus erythematosus indicates more severe condition. Clin Biochem. 2020, <http://dx.doi.org/10.1016/j.clinbiochem.2020.12.004>.
  21. Wang S, Shang J, Xiao J, Zhao Z. Clinicopathologic characteristics and outcomes of lupus nephritis with positive antineutrophil cytoplasmic antibody. Ren Fail. 2020;42:244-54, <http://dx.doi.org/10.1080/0886022X.2020.1735416>.
  22. Jarrot PA, Chiche L, Hervier B, Daniel L, Vuiblet V, Bardin N, et al. Systemic lupus erythematosus and antineutrophil cytoplasmic antibody-associated vasculitis overlap syndrome in patients with biopsy-proven glomerulonephritis. Medicine (Baltimore). 2016;95:e3748, <http://dx.doi.org/10.1097/MD.0000000000003748>.
  23. Hirai Y, Iyoda M, Shibata T, Ashikaga E, Hosaka N, Suzuki H, et al. Lupus nephritis associated with positive MPO-ANCA in a patient with underlying autoimmune hemolytic anemia. Clin Exp Nephrol. 2008;12:393-7, <http://dx.doi.org/10.1007/s10157-008-0058-1>.
  24. Pan HF, Fang XH, Wu GC, Li WX, Zhao XF, Li XP, et al. Anti-neutrophil cytoplasmic antibodies in new-onset systemic lupus erythematosus and lupus nephritis. Inflammation. 2008;31:260-5, <http://dx.doi.org/10.1007/s10753-008-9073-3>.
  25. Sharmeen S, Cassol C, Kato H. ANCA-associated necrotizing glomerulonephritis overlapping with mesangial proliferative lupus nephritis refractory to plasmapheresis, steroid pulse therapy, and a combination of mycophenolate mofetil and rituximab. Case Rep. 2018;1-6, <http://dx.doi.org/10.1155/2018/3076806>, 2018.
  26. Li C, Zhou ML, Liang DD, Wang JJ, Yang J, Zeng CH, et al. Treatment and clinicopathological characteristics of lupus nephritis with anti-neutrophil cytoplasmic antibody positivity: a case-control study. BMJ Open. 2017;7:e015668, <http://dx.doi.org/10.1136/bmjopen-2016-015668>.
  27. Lemerle M, Garnier AS, Croue A, Chevailler A, Saint-André JP, Subra JF, et al. Clinico-pathological considerations in a 48-years-old female with acute kidney injury: is it lupus nephritis, ANCA-associated vasculitis or something else? BMC Nephrol. 2019;20:334, <http://dx.doi.org/10.1186/s12882-019-1531-7>.
  28. Wang Y, Huang X, Cai J, Xie L, Wang W, Tang S, et al. Clinicopathologic characteristics and outcomes of lupus nephritis with antineutrophil cytoplasmic antibody: a retrospective study. Medicine (Baltimore). 2016;95:e2580, <http://dx.doi.org/10.1097/MD.0000000000002580>.