+Model RAM-691; No. of Pages 6

ARTICLE IN PRESS

Revista Argentina de Microbiología xxx (xxxx) xxx-xxx



revista argentina de MICROBIOLOGÍA



www.elsevier.es/ram

BRIEF REPORT

Azospirillum argentinense REC3 and its glycosylated flagellin AzFlap reduce lesions caused by *Botrytis* cinerea on strawberry leaves

María A. Correa Deza^a, Stefanía G. Venegas Tarancón^b, Gustavo Martínez Zamora^c, Josefina A. Amigo ^(b) a,*, Sergio M. Salazar^{a,d}, María P. Filippone^{a,b}, Raúl O. Pedraza^a

Received 15 April 2025; accepted 15 October 2025

KEYWORDS

Azospirillum argentinense REC3; Botrytis cinerea; Flagellin; Fragaria ananassa Abstract Among the pathogenic fungi of strawberry (*Fragaria ananassa* Duch.), *Botrytis cinerea*, a filamentous necrotrophic fungus that causes gray mold, stands out worldwide for its ubiquity and prevalence. Chemical control has been the most effective method used for years to manage *B. cinerea* in strawberry crops. However, the frequent use of numerous fungicides has increased issues related to pathogen resistance, resurgence, and toxic residues. In this study, we propose the use of the plant growth-promoting bacterium *Azospirillum argentinense* REC3 and its flagellin AzFlap, whether non-glycosylated or glycosylated, for the control of *B. cinerea*. We observed that only REC3 inhibited the mycelial growth of *B. cinerea in vitro*, whereas AzFlap had no inhibitory effect. Moreover, REC3 and only its glycosylated flagellin AzFlap contributed to reducing lesions caused by *B. cinerea* on detached strawberry leaves.

© 2025 The Author(s). Published by Elsevier España, S.L.U. on behalf of Asociación Argentina de Microbiología. This is an open access article under the CC BY-NC-ND license (http://creativecommons.org/licenses/by-nc-nd/4.0/).

E-mail address: josefinaamigo1112@gmail.com (J.A. Amigo).

https://doi.org/10.1016/j.ram.2025.10.002

0325-7541/© 2025 The Author(s). Published by Elsevier España, S.L.U. on behalf of Asociación Argentina de Microbiología. This is an open access article under the CC BY-NC-ND license (http://creativecommons.org/licenses/by-nc-nd/4.0/).

Please cite this article as: M.A. Correa Deza, S.G. Venegas Tarancón, G. Martínez Zamora et al., *Azospirillum argentinense* REC3 and its glycosylated flagellin AzFlap reduce lesions caused by *Botrytis cinerea* on strawberry leaves, Revista Argentina de Microbiología, https://doi.org/10.1016/j.ram.2025.10.002

^a Facultad de Agronomía, Zootecnia y Veterinaria, Universidad Nacional de Tucumán, Av. Kirchner 1900, (4000) San Miguel de Tucumán, Tucumán, Argentina

^b Consejo Nacional de Investigaciones Científicas y Tecnológicas (CONICET), Av. Kirchner 1900, (4000) San Miguel de Tucumán, Tucumán, Argentina

^c Instituto Superior de Investigaciones Biológicas (INSIBIO), CONICET-UNT, and Instituto de Química Biológica ''Dr. Bernabé Bloj'', Facultad de Bioquímica, Química y Farmacia, Universidad Nacional de Tucumán, San Miguel de Tucumán, Argentina

d Estación Experimental Agropecuaria Famaillá, Instituto Nacional de Tecnología Agropecuaria, Tucumán, Argentina

^{*} Corresponding author.

+Model RAM-691; No. of Pages 6

ARTICLE IN PRESS

M.A. Correa Deza, S.G. Venegas Tarancón, G. Martínez Zamora et al.

PALABRAS CLAVE

Azospirillum argentinense REC3; Botrytis cinerea; Flagelina; Fragaria ananassa

Azospirillum argentinense REC3 y su flagelina AzFlap glicosilada reducen las lesiones causadas por Botrytis cinerea en hojas de frutilla

Resumen Entre los patógenos de frutilla (*Fragaria ananassa* Duch.), *Botrytis cinerea*, un hongo filamentoso necrotrófico que causa la enfermedad del moho gris, se destaca mundialmente por su ubicuidad y prevalencia. El control químico se ha utilizado durante años y ha sido el método más efectivo para manejar *B. cinerea* en el cultivo de frutilla. Sin embargo, el uso frecuente de numerosos fungicidas ha incrementado los problemas relacionados con la resistencia a los patógenos, el resurgimiento y los residuos tóxicos. En este trabajo proponemos el uso de la bacteria promotora del crecimiento vegetal *Azospirillum argentinense* REC3 y su flagelina AzFlap, ya sea no glicosilada o glicosilada, para el control de *B. cinerea*. Observamos que solo REC3 inhibió el crecimiento micelial de *B. cinerea in vitro*, mientras que AzFlap no produjo inhibición. Además, REC3 y solo su flagelina glicosilada AzFlap contribuyeron a reducir las lesiones causadas por *B. cinerea* en hojas desprendidas de frutilla.

© 2025 Los Autores. Publicado por Elsevier España, S.L.U. en nombre de Asociación Argentina de Microbiología. Este es un artículo Open Access bajo la licencia CC BY-NC-ND (http://creativecommons.org/licenses/by-nc-nd/4.0/).

Among the pathogenic fungi of strawberry (*Fragaria ananassa* Duch.), *Botrytis cinerea*, a filamentous necrotrophic fungus that causes gray mold, stands out worldwide for its ubiquity and prevalence². Chemical control has been the most effective method used for years to manage *B. cinerea* in strawberry crops; however, the frequent use of numerous fungicides has increased issues related to pathogen resistance, resurgence, and toxic residues³. Among the possible alternatives, based on bioinputs, successful use has been reported with *Bacillus cereus*¹⁵, *Paenibacillus polymyxa*¹², the application of extracts from the fungus *Colletotrichum acutatum*⁹, or of *Streptomyces* sp. sdu1201 and its derived compound actinomycin D¹⁴.

For the control of B. cinerea, we propose the use of the plant growth-promoting bacterium Azospirillum argentinense REC3⁵, and as a novel finding, its flagellin AzFlap, obtained from its polar flagellum⁶. Regardless of the presence of A. argentinense REC3, it was observed that its flagellin AzFlap was capable of controlling Macrophomina phaseolina, eliciting defense responses in the plant similar to those triggered when it was inoculated with REC36. REC3 flagellin is a glycosylated protein (100 kDa) with elicitor capacity for defense responses in strawberry plants⁶. However, to date, it remains unknown whether this flagellin confers a differential response in strawberry, specifically in the control of B. cinerea, when applied in its glycosylated or non-glycosylated form. It is known that glycosylated proteins provide mechanisms for controlling signal transduction, protein folding, stability, cell-to-cell interaction, and immune response in the host⁷. Therefore, based on the background of REC3 and its flagellin, our working hypothesis is that A. argentinense REC3 and its flagellin AzFlap, in its glycosylated form, contribute to reducing lesions caused by B. cinerea on strawberry leaves. To investigate this further, we propose as study objectives to evaluate in vitro the inhibition of B. cinerea growth caused by REC3 and AzFlap, and to assess the incidence of gray mold on strawberry leaves treated with REC3 or its flagellin AzFlap, whether non-glycosylated or glycosylated.

A pure culture of A. argentinense REC3 was grown in NFb liquid medium 1 , supplemented with 1% NH4Cl for 24h at 30 °C and 120 rpm. Cells were then centrifuged at $2000\times g$ for 10 min and washed twice with sterile double distilled water at pH 7.0 to remove culture medium residues that might interfere with the assays; cells were suspended in sterile double distilled water and the concentration for strawberry leaves inoculation was $\sim\!10^6\,\text{CFU/ml}$ (OD560 nm 0.2).

Flagellin AzFlap was obtained as described previously⁶. After denaturing polyacrylamide gel electrophoresis, following standard procedures, two bands, one of 100 kDa and another of 70 kDa, were excised from the gel, electroeluted in Tris-glycine buffer pH 8.3 at 40 V for 12 h, dialyzed, lyophilized, and suspended in distilled water to a 200 nM concentration. Both bands correspond to flagellin AzFlap, with the 70 kDa band being the deglycosylated version, derived from the extraction process. This was previously confirmed by Western blot using the antibody antiflagellin of *A. brasilense* Sp7 and by sugar staining using the Shiff method⁶.

The BMM strain of *B. cinerea* was used. It was cultivated on potato dextrose agar (PDA) medium (39 g/l) at 24 °C with 16 h of light (200 μ mol/m/s). After 8–10 days of growth, a conidia suspension (1 × 10⁴ conidia/ml) was prepared by filtration through gauze and quantified using a Neubauer chamber⁸.

The antifungal activity against *B. cinerea* growth was assessed on PDA plates (9 cm in diameter). One hundred microliters of either AzFlap 70 kDa solution (200 nM), AzFlap 100 kDa solution (200 nM), or REC3 suspension (10⁶ UFC/ml) was spread evenly over the surface of the PDA plates using a Drigalski spatula until fully absorbed. Afterward, $5\,\mu l$ of the conidia suspension (1 \times 10⁴ conidia/ml) was placed in the center of each plate and incubated in darkness at 24 °C. Fungal colony diameter (cm) was measured with a caliber at 24, 48, and 72 h postinoculation. The experiment was conducted three times, with three replicates per treatment.

+Model RAM-691; No. of Pages 6

ARTICLE IN PRESS

Revista Argentina de Microbiología xxx (xxxx) xxx-xxx

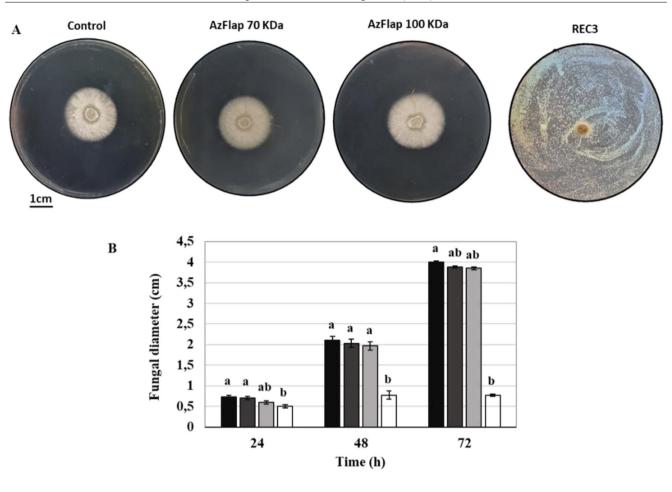


Figure 1 In vitro antifungal activity of Azospirillum argentinense REC3, Azflap 70 kDa, and Azflap 100 kDa. (A) Qualitative examples of mycelial growth inhibition 72 h after the assay. (B) Diameter of mycelial growth of *B. cinerea* observed at 24, 48, and 72 h. Vertical bars represent the standard errors of the means. Different letters above the bars indicate statistically significant differences at p < 0.05.

■Control ■AzFlap 70Kda ■AzFlap 100Kda □REC3

Biocontrol assays were conducted to evaluate the incidence of gray mold on strawberry leaves treated with REC3 and its flagellins. B. cinerea grown on APG medium and detached strawberry leaves, var. Pájaro, 3 months old, were used¹³. The leaves were surface disinfected with 70% ethanol, and the leaflets were separated and mixed; then, three leaflets were randomly selected for each treatment and placed in Petri dishes on filter paper moistened with sterile water. The treatments were applied by spraying both sides of the leaflet until runoff: sterile water (control): REC3 (\sim 10⁶ CFU/ml); flagellin 70 kDa (200 nM); flagellin 100 kDa (200 nM). Three days after the treatments were applied, the leaflets were inoculated on the adaxial side with 0.5 cm diameter plugs containing B. cinerea (10⁴ conidia/ml). The Petri dishes were placed in a phytotron at photon flux density of 150 µmol/m/s (12/12 h light/dark cycle) at a relative humidity of 60–70% and 25 \pm 2 $^{\circ}$ C for 18 days. At 9, 14, and 18 days post-infection (dpi), the diameter of the lesions caused by B. cinerea was measured. At 18 dpi, disease severity was determined, which consists of the ratio between the dam-

aged area and the total surface of the leaflet, multiplied by 100.

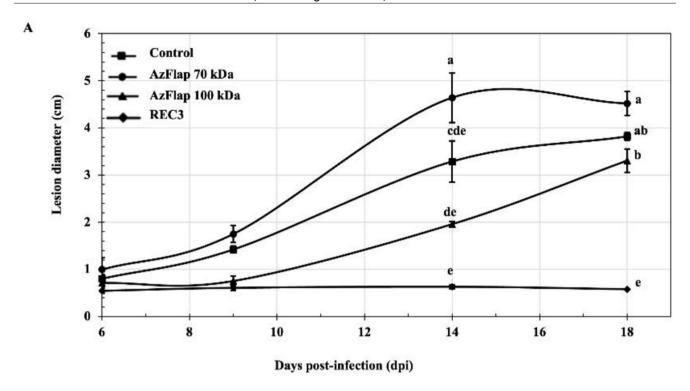
After verifying the normal distribution of the data, the general linear ANOVA model was applied using Infostat statistical software version 2018^4 . Significant differences between mean values were determined by Fisher's pairwise comparisons (p < 0.05)⁴.

In this study, after conducting the *in vitro* challenge to evaluate the inhibition of *B. cinerea* growth caused by *A. argentinense* REC3 and its glycosylated or non-glycosylated flagellin, it was observed that the bacterium inhibits fungal growth (Figs. 1A and B). This was also previously observed in the inhibition of mycelial growth of *C. acutatum* and *M. phaseolina*^{6,10}. In the case of flagellin, neither the glycosylated nor the non-glycosylated form inhibited the growth of *B. cinerea* (Figs. 1A and B).

Previously, we had reported that flagellin AzFlap, in its glycosylated form, induces a defense response in strawberry plants against the fungus *M. phaseolina*⁶. However, to date, AzFlap in its deglycosylated form has not been evaluated. In

ARTICLE IN PRESS

M.A. Correa Deza, S.G. Venegas Tarancón, G. Martínez Zamora et al.



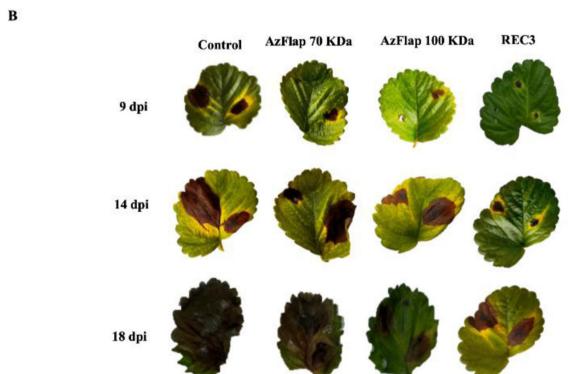


Figure 2 Development of lesions caused by *B. cinerea* on strawberry leaves, expressed as lesion diameter (cm), at 9, 14 and 18 dpi (A). Vertical bars represent the standard errors of the means. Different letters above the bars indicate statistically significant differences at p < 0.05. Bioassay of *A. argentinense* REC3 and its flagellin AzFlap against *B. cinerea* on detached strawberry leaves. Representative pictures of symptoms on strawberry leaves under different treatments at 9, 14 and 18 days post-treatments (B).

the biocontrol assay, it was observed that when strawberry leaves were treated with water (control), they developed lesions caused by *B. cinerea*, expressed as lesion diameter

(cm), which increased in size over the 9, 14, and 18 days of the test evaluation (Fig. 2A). However, when REC3 was applied to the leaves, the bacterium was able to control

Revista Argentina de Microbiología xxx (xxxx) xxx-xxx

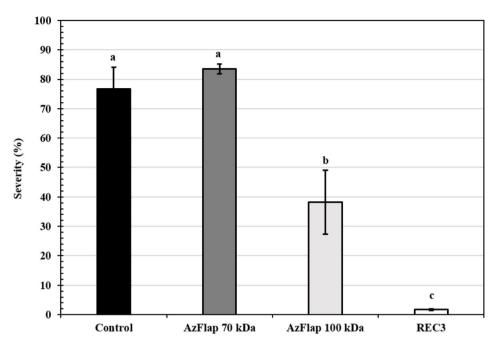


Figure 3 Severity of the disease on detached strawberry leaves. The treatments applied were: sterile water (control); flagellin 70 kDa (200 nM); flagellin 100 kDa (200 nM); A. argentinense REC3 (\sim 106 CFU/ml). Three days after the treatments were applied, the leaflets were inoculated on the adaxial side with 0.5 cm diameter plugs containing B. cinerea (104 conidia/ml). Vertical bars represent the standard errors of the means. Different letters above the bars indicate statistically significant differences at p < 0.05.

the development of the lesions caused by the fungus during the evaluation period. Figure 2B shows representative pictures of symptoms on strawberry leaves at 9, 14 and 18 days' post-treatments. This behavior was also observed in strawberry plants inoculated with REC3 and challenged with *C. acutatum*¹¹ and *M. phaseolina*⁶. The mechanisms involved in those cases included plant defense responses expressed at the structural, biochemical, and molecular levels. These mechanisms are also likely involved in the observations with *B. cinerea*. Regarding the treatments with flagellin AzFlap, it was observed that its deglycosylated form (70 kDa) did not reduce the lesions caused by *B. cinerea* and behaved like the control treatment. However, its glycosylated form (100 kDa) significantly (*p* < 0.05) reduced the lesion diameter, although not as much as with the application of REC3 (Figs. 2A and B).

With regard to the severity of the disease, in the treatments with AzFlap, its deglycosylated form (70 kDa) did not significantly reduce (83.5%, p < 0.05) the lesions caused by $B.\ cinerea$ and behaved like the control treatment (76.65%, p < 0.05). However, its glycosylated form (100 kDa) significantly reduced the severity of the disease (38.39%, p < 0.05), although not as much as with the application of REC3 (1.67%, p < 0.05) (Fig. 3).

This is the first study to evaluate deglycosylated AzFlap flagellin in a pathological test on strawberry. While the mechanisms involved in these responses have not yet been evaluated, these results suggest that the glycosylation of AzFlap is important in controlling gray mold in strawberry. This, in turn, opens up new possibilities in the search for bioproducts for the control of phytopathogens, independent of bacterial presence, that is, not dependent on the viability of the bacteria to exert a biocontrol effect. Additionally, as observed in the *in vitro* growth inhibition response of *B*.

cinerea, where AzFlap did not inhibit its mycelial growth, the pathological tests revealed an elicitor action only with glycosylated AzFlap (100 kDa), similar to that observed with M. phaseolina⁶. Therefore, for the biocontrol of B. cinerea, it is important that flagellin AzFlap remains in its glycosylated form. However, further studies on plants will be necessary, including structural analysis of strawberry leaf tissues, as well as the biochemical and molecular aspects involved in the defense response.

Funding

This study was partially funded by Secretaria de Ciencia, Arte y Tecnología, Universidad Nacional de Tucumán (PIUNT A726), and by Agencia Nacional de Promoción Científica y Tecnológica (PICT-2019-02199).

Conflict of interest

The authors declare no conflicts of interest.

References

- Baldani JI, Reis VM, Videira SS, Boddey LH, Baldani VLD. The art of isolating nitrogen-fixing bacteria from non-leguminous plants using N-free semi-solid media: a practical guide for microbiologists. Plant Soil. 2014;384:413–31, http://dx.doi.org/10.1007/s11104-014-2186-6.
- Breen J, Mur LAJ, Sivakumaran A, Akinyemi A, Wilkinson MJ, Rodriguez Lopez CM. Botrytis cinerea loss and restoration of virulence during in vitro culture follows flux in global DNA methylation. Int J Mol Sci. 2022;23:3034, http://dx.doi.org/10.3390/ijms23063034.

ARTICLE IN PRESS

M.A. Correa Deza, S.G. Venegas Tarancón, G. Martínez Zamora et al.

- de Moura GGD, de Barros AV, Machado F, Martins AD, da Silva CM, Durango LGC, Forim M, Alves E, Pasqual M, Doria J. Endophytic bacteria from strawberry plants control gray mold in fruits via production of antifungal compounds against *Botrytis cinerea* L. Microbiol Res. 2021;251:126793, http://dx.doi.org/10.1016/j.micres.2021.126793.
- Di Rienzo JA, Casanoves F, Balzarini MG, González L, Tablada M, Robledo CW. InfoStat versión 2018. Argentina: Centro de Transferencia InfoStat, FCA, Universidad Nacional de Córdoba; 2020 [cited 16 Oct 2024]. Available from: http://www.infostat.com.ar
- dos Santos Ferreira N, Coniglio A, Puente M, Hayashi Sant' Anna F, Maroniche G, Garcia J, Molina R, Nievas S, Gazolla Volpiano C, Ambrosini A, Pereira Passaglia LM, Pedraza RO, Massena Reis V, Zilli JE, Cassán F. Genome-based reclassification of Azospirillum brasilense Az39 as the type strain of Azospirillum argentinense sp. nov. Int J Syst Evol Microbiol. 2022;72:1099, http://dx.doi.org/10.1099/ijsem.0.005475.
- Elías JM, Ramírez-Mata A, Albornoz PL, Baca BE, Díaz-Ricci JC, Pedraza RO. The polar flagellin of Azospirillum brasilense REC3 induces a defense response in strawberry plants against the fungus Macrophomina phaseolina. J Plant Growth Regul. 2021;41:2992–3008, http://dx.doi.org/10.1007/s00344-021-10490-4.
- Merino S, Tomás JM. Gram-negative flagella glycosylation. Int J Mol Sci. 2014;15, http://dx.doi.org/10.3390/ijms15022840.
- Qin GZ, Zong YY, Chen QL, Hua DL, Tian SP. Inhibitory effect of boron against *Botrytis cinerea* on table grapes and its possible mechanisms of action. Int J Food Microbiol. 2010;138:145–50, http://dx.doi.org/10.1016/j.ijfoodmicro.2009.12.018.
- Tomas-Grau RH, Hael-Conrad V, Requena-Serra FJ, Perato SM, Caro MdP, Salazar SM, Díaz-Ricci JC. Biological control of strawberry grey mold disease caused

- by *Botrytis cinerea* mediated by *Colletotrichum acutatum* extracts. BioControl. 2020;65:461–73, http://dx.doi.org/10.1007/s10526-020-10003-4.
- Tortora ML, Díaz-Ricci JC, Pedraza RO. Azospirillum brasilense siderophores with antifungal activity against Colletotrichum acutatum. Arch Microbiol. 2011;193:275–86, http://dx.doi.org/10.1007/s00203-010-0672-7.
- 11. Tortora ML, Díaz-Ricci JC, Pedraza tion of strawberry plants (Fragaria ananassa Duch.) by against anthracnose disease induced Azospir-Plant 2012:356:279-90. illum brasilense. Soil. http://dx.doi.org/10.1007/s11104-011-0916-6.
- 12. Tsai SH, Chen YT, Yang YL, Lee BY, Huang CJ, Chen CY. The potential biocontrol agent *Paenibacillus polymyxa* TP3 produces *Fusaricidin* type compounds involved in the antagonism against gray mold pathogen *Botrytis cinerea*. Phytopathology. 2022;112:775–83, http://dx.doi.org/10.1094/phyto-04-21-0178-r.
- Vellicce GR, Diaz Ricci JC, Hernández L, Castagnaro AP. Enhanced resistance to *Botrytis cinerea* mediated by the transgenic expression of the chitinase gene ch5B in strawberry. Transgenic Res. 2006;15:57–68.
- 14. Yong D, Li Y, Gong K, Yu Y, Zhao S, Duan Q, Ren C, Li A, Fu J, Ni J, Zhang Y, Li R. Biocontrol of strawberry gray mold caused by *Botrytis cinerea* with the termite associated *Streptomyces* sp. sdu1201 and actinomycin D. Front Microbiol. 2022;13, http://dx.doi.org/10.3389/fmicb.2022.1051730, 1051730.
- Yu YY, Dou GX, Sun XX, Chen L, Zheng Y, Xiao HM, Wang YP, Li HY, Guo JH, Jiang CH. Transcriptome and biochemical analysis jointly reveal the effects of *Bacillus cereus* AR156 on postharvest strawberry gray mold and fruit quality. Front Plant Sci. 2021;12, http://dx.doi.org/10.3389/fpls.2021.700446, 700446.