

revista argentina de MICROBIOLOGÍA



www.elsevier.es/ram

SPECIAL ARTICLE

Statin and aspirin use in parasitic infections as a potential therapeutic strategy: A narrative review



Valentina Burgess^a, Juan D. Maya^{b,*}

- a Escuela de Medicina, Facultad de Medicina, Universidad de Chile, Independencia, Santiago, Chile
- ^b Programa de Farmacología Molecular y Clínica, Instituto de Ciencias Biomédicas, Facultad de Medicina, Universidad de Chile, Independencia, Santiago, Chile

Received 28 July 2022; accepted 26 January 2023 Available online 3 April 2023

KEYWORDS

Parasitic diseases; Resolution of inflammation; Lipoxins; Resolvins; Aspirin; Statins Abstract Infections, including zoonoses, constitute a threat to human health due to the spread of resistant pathogens. These diseases generate an inflammatory response controlled by a resolving mechanism involving specialized membrane lipid-derived molecules called lipoxins, resolvins, maresins, and protectins. The production of some of these molecules can be triggered by aspirin or statins. Thus, it is proposed that modulation of the host response could be a useful therapeutic strategy, contributing to the management of resistance to antiparasitic agents or preventing drift to chronic, host-damaging courses. Therefore, the present work presents the state of the art on the use of statins or aspirin for the experimental management of parasitic infections such as Chagas disease, leishmaniasis, toxoplasmosis or malaria. The methodology used was a narrative review covering original articles from the last seven years, 38 of which met the inclusion criteria. Based on the publications consulted, modulation of the resolution of inflammation using statins may be feasible as an adjuvant in the therapy of parasitic diseases. However, there was no strong experimental evidence on the use of aspirin; therefore, further studies are needed to evaluate its role inflammation resolution process in infectious diseases. © 2023 Asociación Argentina de Microbiología. Published by Elsevier España, S.L.U. This is an open access article under the CC BY-NC-ND license (http://creativecommons.org/licenses/bync-nd/4.0/).

PALABRAS CLAVE

Parasitosis; Resolución de la inflamación; Lipoxinas; Resolvinas; Uso de estatinas y aspirina en infecciones parasitarias como potencial estrategia terapéutica: Una revisión narrativa

Resumen Las infecciones, incluyendo las zoonosis, constituyen una amenaza a la salud humana debido a la diseminación de patógenos resistentes. Estas enfermedades generan una respuesta inflamatoria controlada por un mecanismo de resolución, en el que participan moléculas especializadas derivadas de lípidos de membrana llamadas lipoxinas, resolvinas, maresinas

E-mail address: jdmaya@uchile.cl (J.D. Maya).

^{*} Corresponding author.

Aspirina; Estatinas

y protectinas. La producción de algunas de estas moléculas puede ser gatillada por aspirina o estatinas. Así, se propone que la modulación de la respuesta del hospedero podría ser una estrategia terapéutica útil, que contribuye al manejo de la resistencia a agentes antiparasitarios o que puede prevenir la derivación hacia cursos crónicos, dañinos para el hospedero. En esta revisión se presenta una puesta al día sobre el uso de estatinas o aspirina para el manejo experimental de infecciones parasitarias, como enfermedad de Chagas, leishmaniasis, toxoplasmosis y malaria. Se hizo una revisión narrativa, buscando artículos originales de los últimos siete años, se encontraron 38 que cumplieron con los criterios de inclusión. De acuerdo con las publicaciones consultadas, la resolución de la inflamación modulada mediante estatinas podría ser un adyuvante en la terapia de enfermedades parasitarias. Por otro lado, no se observó una evidencia experimental fuerte con respecto al uso de aspirina, por lo que se recomiendan más estudios para evaluar su rol en el proceso de resolución de la inflamación en enfermedades infecciosas.

© 2023 Asociación Argentina de Microbiología. Publicado por Elsevier España, S.L.U. Este es un artículo Open Access bajo la licencia CC BY-NC-ND (http://creativecommons.org/licenses/by-nc-nd/4.0/).

Introduction

The treatment and prevention of infections focus on eradicating the microorganisms that cause them using a broad pharmacological arsenal of highly effective antibiotics and vaccines generating a sense of safety and confidence in reaching the end of infectious diseases⁴³. The COVID-19 pandemic and the spread of resistant pathogens capable of evading the immune system demonstrated that this goal is still far from being achieved. Moreover, the lack of safer and more effective drugs for the treatment of parasitic diseases, especially those causing chronic conditions, underscores this situation. Furthermore, the transformation of microbiological ecosystems and the global interconnection of societies have accelerated the evolution of microorganisms and infections^{43,106}, including the emergence of new threats. Moreover, the epidemiological behavior of traditional zoonotic diseases (as seen, for example, in the increase in congenital and transfusion transmission of Chagas disease in non-endemic countries) is changing. Thus, new therapeutic approaches are required for the treatment of infectious diseases.

The inflammatory response against pathogens is a selflimiting process. In general, tissue damage or invasion by pathogens triggers local activation of macrophages through recognizing pathogen-associated molecular (PAMPs) by specialized Toll-like receptors (TLRs)⁹⁹. Thus, the secretion of several interleukins (Ils), prostaglandins and leukotrienes, tumor necrosis factor-alpha (TNF- α), and gamma interferon (IFN- γ), among others, is promoted, depending on the type of pathogen involved. Free radicals are also generated, facilitating phagocytosis 10,97, triggering the inflammatory process. Although essential for eliminating pathogens, the prompt re-establishment of homeostasis is required through the resolution of inflammation. When this resolution is inadequate, inflammation gets out of control, causing further tissue damage or installing chronic processes, with or without active infection 97,102.

The resolution of inflammation is a highly regulated process orchestrated by multiple mediators, the so-called specialized pro-resolving mediators (SPMs)⁴⁶, a superfamily of lipid molecules derived from the ω -3 and ω -6 polyunsaturated fatty acids of the plasma membrane, grouped into four classes: lipoxins A4 and B4 (LXA4, LXB4), derived from arachidonic acid; resolvins (Rv) D1-6, protectins (PD) and maresins (MaR), derived from docosahexaenoic acid (DHA); and Rv E1-4, derived from eicosapentaenoic acid (EPA)^{18,48}. SPMs are described as potent anti-inflammatory molecules with an immunoregulatory role⁴⁸.

SPMs are synthesized by several lipooxygenases in different cell types, including neutrophils and macrophages, and act on various G protein-coupled receptors, such as Formyl Peptide Receptor-2 (FPR2) or G protein-coupled receptor 32 (GPR32)⁸⁷. The process is initiated during the exudate formation in the active phase of inflammation. The classic initiators, prostaglandins and leukotrienes, are key in the subsequent switch of lipid mediators⁹⁰, activating the expression of the enzymes necessary for producing SPMs⁵⁹. During this process, LXs⁸⁷ appear first, followed by Rvs and, finally, PDs and MaRs⁸⁸; however, the resolution is more a result of the concerted action of these mediators than of the time course of their secretion; although it is clear that their early appearance guarantees the self-limiting nature of the inflammation⁹².

The resolution of inflammation is aimed, in part, at clearing the pathogenic load through the SPMs. These are also responsible for promoting local neutrophil apoptosis, reducing the systemic inflammatory response by decreasing the production of cytokines and other Nuclear Factor κB (NF κB)-associated products, and increasing the production of IL-10 and nitric oxide in macrophages⁴⁷. In addition, they increase phagocytosis of apoptotic leukocytes (efferocytosis) and pathogen clearance by tissue macrophages^{47,102}. For these reasons, SPMs have high therapeutic potential in managing inflammation⁸⁹, including the possibility of circumventing the adverse reactions and immunosuppression

associated with the treatment of hyperinflammatory states triggered by severe infections, such as sepsis or COVID-19⁷⁴.

Interestingly, the synthesis of SPMs can be triggered by the prostaglandin production inhibitor aspirin since cyclooxygenase 2 (COX2) acetylation blocks prostaglandin synthesis and, at the same time, confers the ability to produce the epimer intermediates 15R-HETE from arachidonic acid, 18R-HEPE from EPA and 17R-HDHA from DHA. Neutrophils transform these intermediates into lipoxins, resolvins, and protectins triggered by aspirin⁹¹. Likewise, statins, inhibitors of 3-hydroxy-methyl-glutaryl coenzyme A reductase, lower blood cholesterol levels and have other effects, called pleiotropic, by decreasing the synthesis of isoprenoids, intermediates in the mevalonate pathway⁸⁶. These effects include regulation of endothelial function, coagulation, and anti-inflammatory actions such as reduced leukocyte migration and proinflammatory cytokine generation⁵⁵. Statins induce COX2 nitrosylation, generating the same epimer intermediates of SPMs as aspirin but causing S-nitrosylation⁴⁹, explaining, in part, the antiinflammatory effects of statins²¹.

SPMs are relevant in acute infections because they shorten the inflammatory interval and increase the elimination of bacteria and other microorganisms; eventually, allowing the decrease in antibiotic doses^{17,90}. Therefore, the relationship between infection and resolution is attractive due to the immunosuppressive potential of some anti-inflammatory agents³⁰. For example, in self-limited Escherichia coli infections, resolution programs are activated, and PD1 and RvD1, and D5 levels are elevated^{21,22}. In addition, LXA4, Rv D1, and MaR1 increase the survival of mice with experimental sepsis^{16,50,107}; moreover, LXA4 decreases biofilm formation by Pseudomonas aeruginosa while increasing the ability of ciprofloxacin and imipenem to kill this bacterium¹⁰¹. Interestingly, a recent report links the protective effect of the antimalarial artesunate to the activation of GPR32, another SPM receptor, mimicking the effects of PD1 on macrophages in the context of murine Plasmodium infections and Listeria sepsis⁶. Thus, given the importance of antimicrobial resistance, a pharmacological intervention of resolution could provide a different approach to decreasing exposure to antibiotics¹⁷.

Modulating the host response with statins or aspirin may be helpful in the therapeutics of infections with significant inflammatory components. It could contribute to managing antimicrobial resistance or prevent referral to potentially harmful chronic courses. Thus, it is interesting to ask whether the modulation of relevant host factors with aspirin or a statin could be beneficial for the adjuvant treatment of parasitic infections caused by systemic protozoa such as the causative agents of malaria, leishmaniasis, toxoplasmosis or Chagas disease. Therefore, we performed a systematic search and a narrative review of the state-of-ofthe-art in the use of statins or aspirin as potential therapies to modify host factors in treating parasitic diseases, the most neglected of all infectious diseases, to identify the potential benefits of aspirin or statins in the treatment of these parasitic diseases, based on their properties for modulating the inflammatory response.

Methodology

The present work was structured as a narrative review based on a systematic search for articles on the use of aspirin or statins in experimental models or clinical studies of parasitic infections caused by *Plasmodium*, *Leishmania*, *Toxoplasma* or *Trypanosoma*.

The database used for the search was PUBMED (https://pubmed.ncbi.nlm.nih.gov/). The parasitic diseases included in this study were Chagas disease, leishmaniasis, malaria, and toxoplasmosis. These infectious diseases were included because of their epidemiological importance or the difficulties in finding an effective therapy.

The search strategy was designed using terms included in "all fields" or as a medical subject heading (MeSH). The subheading "pharmacological action" was used as an additional descriptor.

For the aspirin search, the terms "aspirin" or "acetylsalicylic acid" were used. For the statin search, "statin" OR "lovastatin", "simvastatin", "pravastatin", and "pitavastatin".

For the parasitic diseases, the keywords used were "Trypanosoma cruzi", "Chagas disease", "Trypanosoma brucei", "African trypanosomiasis", "Toxoplasma gondii", "toxoplasmosis", "malaria", "Plasmodium falciparum", "Plasmodium vivax", "Plasmodium malariae", "Plasmodium ovale", "leishmaniasis", "Leishmania tropica", "Leishmania donovani", "Leishmania major", "Leishmania Mexicana", "Leishmania amazonensis" and "Leishmania infantum".

Inclusion criteria were (1) English language of publication, (2) articles published between 2015 and 2022, (3) original article, (4) *in vitro* or *in vivo* study design. The *in vivo* studies included animal or human models (prospective or retrospective clinical trials).

Two hundred thirty-two (232) hits were retrieved, and after applying the inclusion criteria, 38 studies were obtained and included in this narrative review (Table 1).

When necessary, works before 2011 were mentioned to contextualize the concepts discussed.

Role of aspirin and statins in the therapy of some parasitic diseases of medical interest

Malaria infections

Malaria, caused by protozoa of the *Plasmodium* genus, is the most important parasitic disease in the world, with a high burden of disease and mortality⁷⁹. *P. falciparum* is responsible for 95% of morbidity and mortality, including severe manifestations derived from brain involvement, reaching a mortality rate of 10–20%⁷⁹. Although its incidence has been decreasing²⁰, one of the main problems of this disease is the emergence of resistance to pharmacological treatment and insecticides used for vector control; therefore, its control has not been easy^{104,109}. Moreover, it is necessary to create new treatment strategies with consistent efficiency and new

			Total HITS: 232	5. 737			
	Aspirin 108					Statins 124	
Chagas disease 36	Leishmaniasis 9	Leishmaniasis Toxoplasmosis Malaria 9 53	Malaria 53	Chagas disease 34	Leishmaniasis 24	Leishmaniasis Toxoplasmosis Malaria 24 53	Malaria 13
After applying the inclusion criteria 1214,15,19,22,34,56,63,72,73,77,78,94,96	eria 6 15	0	1100	74,11,12,22,33,36,110 323,46,75	323,46,75	6 26,53,60,71,82-84	626,33,60,71,82-84 *813,24,25,68,80,95,98,105

drug combinations to protect and reduce severe and resistant forms of malaria¹⁰⁹. Thus, statins or aspirin could play a role in modulating the inflammatory response to infection by the *Plasmodium* parasite.

From 2015 to date, only two articles were found reporting the effectiveness of lovastatin or atorvastatin in reducing the cerebral inflammatory response to *P. berghei* infection in a cerebral malaria model^{13,68}. For this reason, the search was extended to cover a longer period of 10 years (2011–2022). This strategy yielded 20 articles, only eight of which studied the modulation of host factors with statins. The remaining articles fell beyond the focus of the present review because they report the activity of statins or their analogs directly on *Plasmodium in vitro* models.

The concept of the use of statins in malaria is not without controversy. On the one hand, in 2009, Helmers et al. reported that statins do not protect against cerebral malaria in an experimental murine model⁴⁰; moreover, in a preclinical trial, the administration of atorvastatin did not alter the course of infection with P. bergei, although it acted synergistically with methylene blue²⁴. On the other hand, preliminary in vitro studies suggest a direct effect of statins on the parasite⁷⁶. However, importantly, there is preclinical evidence pointing to the control of neuroinflammation by decreasing endothelial damage 13,68,98 and even improving cognitive alterations caused by cerebral malaria⁸⁰. Moreover, the combined therapy of atorvastatin with artemeter¹⁰⁵, dihydroartemisinin²⁵ or mefloquine⁹⁵ can alter the course of cerebral infection, reducing mortality from this cause in murine models infected with P. bergei^{25,95}.

Only one article met inclusion criteria for aspirin in malaria. Although it sounds logical to think that prostaglandin synthesis inhibition could influence the host response in malaria, considering the endothelial and platelet alterations observed during plasmodial infection, studies in this regard are scarce and controversial. Indeed, it has been suggested that prostaglandins may protect against endothelial damage¹⁰⁸; moreover, following a prospective, randomized study in 97 patients with *P. falciparum* infection, aspirin did not alter the course of the disease⁴¹. However, in a recent study in a murine model of *P. yoelii*-induced renal failure, aspirin prevented renal cell damage, especially in mice subjected to monocytic depletion¹⁰⁰. Therefore, further preclinical studies are needed to support aspirin as an adjuvant in malaria therapy.

Toxoplasma gondii infections

Toxoplasma gondii is an apicomplexan intracellular parasite that can infect humans, producing a generally asymptomatic infection, but with the persistence of the parasite in tissues in the form of cysts⁵³, which can be activated by immunosuppression⁴². The most severe consequences of congenital infection in the fetus are observed during pregnancy⁶⁴, in addition to severe ocular sequelae in immunosuppressed patients²⁷. It is a public health concern whose pharmacological treatment is ineffective due to the persistence of cysts in infected tissues. However, in symptomatic and congenital cases, a combination of pyrimethamine and sulfadiazine is used, which blocks folate synthesis, with the consequent risk of causing

hematological toxicity in the host due to the high doses required and the prolonged duration of treatments³⁷. However, trimethoprim-sulfamethoxazole, which is less toxic, is an affordable alternative for some patients¹⁰³. Therefore, the search for alternative treatments is necessary¹. Modifying host factors has been proposed owing to the manipulation performed by the parasite for its continuous replication and survival in the host cell⁵⁴. However, few studies analyze the effect of aspirin and statins in toxoplasmosis.

The use of statins as an experimental pharmacological strategy for the modification of host factors in models of toxoplasmosis is reported in six articles published in the last seven years. The availability of isoprenoids in both parasite and host had been previously suggested⁵⁴, being essential for establishing the pathogen-host relationship, which can be altered using atorvastatin. However, it has been recently reported that only high doses of atorvastatin inhibited enteric cell invasion, suggesting that de novo synthesis of isoprenoids is not essential for replication⁶⁰. Moreover, when atorvastatin is combined with bisphosphonates, there is a synergistic effect on isoprenoid production and, therefore, on decreasing the parasite load by reducing its replicative capacity^{53,54}. Furthermore, rosuvastatin inhibited intracellular replication of T. gondii in HeLa cells⁸⁴. Still, in addition, pravastatin and simvastatin, combined with low doses of pyrimethamine-sulfadiazine, synergistically decreased tachyzoite infection in HeLa cells⁸³, reducing the host levels of cytokines IL-6 and IL-7, responsible, in part, for the spread of the disease⁸². These findings are supported by a recent study in which rosuvastatin decreased brain parasite load and local inflammation in a murine model of toxoplasmosis⁷¹ and improved the neurological alterations produced by the infection, including memory disorders²⁶. Those findings suggest that rosuvastatin could be helpful in the treatment of chronic toxoplasmosis.

The search strategy for the use of aspirin did not yield results in the last seven years. However, preliminary reports link PGE2 production and IL-10 secretion with a neuroprotective effect against *T. gondii* infection⁸¹. In addition, this same effect could contribute to regulating the innate immune response since when mice were treated with *T. gondii* extracts, a significant increase in the production of lipoxin A4 was reported. Lipoxin A4-epimer can be induced by aspirin, related to the suppression of cytokine signaling mediated by SOCS2⁶¹.

Leishmania spp. infections

Leishmaniasis is a group of endemic diseases caused by different species of parasites of the genus *Leishmania*. They are obligate intracellular parasites infecting host macrophages, causing significant morbidity and mortality worldwide⁴⁶. They are transmitted to humans by the bite of female insects of the *Phlebotomus* and *Lutzomyia* genus, generating a cutaneous infection in most cases and, less frequently, affecting the liver and spleen, which can be fatal if left untreated^{2,43}. To date, no effective vaccines exist, and the pharmacological treatment includes pentavalent antimonial agents⁴³. However, the antifungal amphotericin B, the aminoglycoside paromomycin, and the phospholipid metabolism inhibitor miltefosine have also shown efficacy. They are directed

against the parasite but require monitoring and evaluation of serious adverse effects or the emergence of resistance⁴³. Therefore, it is of utmost importance for searching drugs with antiparasitic activity, as well as adjuvants or those that control the inflammatory process of the host and could intervene in the parasite invasion in macrophages³⁸.

In this regard, only three studies were identified using the search strategy described above in the context of Leishma*nia* infections. An *in vitro* study with *L. donovani* highlights the importance of an adequate cholesterol level in host cells for optimal parasite entry because chronic cholesterol depletion caused by lovastatin reduced promastigote binding to the macrophage surface, with a consequently lower intracellular amastigote load⁴⁶. This finding suggests that lowering cholesterol content with statins in the host cell membrane could be effective for infection control⁴⁶. Moreover, Haughan et al. previously reported that inhibition of sterol synthesis with lovastatin and miconazole is synergistic³⁹. However, Ghosh et al. found that an atherogenic diet in mice could be protective against L. donovani infection since the parasite extracts cholesterol from the membrane preventing T-cell activation³²; thus, statininduced cholesterol depletion could increase susceptibility to infection. However, the intracellular parasite load did not improve, suggesting a role of statin in controlling parasite growth³², as sterol synthesis by the parasite can be inhibited by mevastatin, the first statin of its kind, affecting cell replication²³. The use of simvastatin in a model of cutaneous leishmaniasis caused by L. major decreased the local parasite load and inflammation by accelerating phagosome maturation and enhancing the oxidative burst in macrophages⁷⁵; which had been previously described with pravastatin in a murine model with L. amazoniensis^{44,45,70}.

Only one recent report suggests that aspirin decreases the parasite load in macrophages, improves the phagocytic activity of these cells and modulates cytokine secretion⁵. However, more research on this drug is needed for leishmaniasis.

Trypanosoma cruzi infection

T. cruzi is the protozoan responsible for Chagas disease, an endemic illness in Latin America, which is treated with benznidazole and nifurtimox²⁸. Both agents have high toxicity and limited efficacy, especially in the chronic phase of the disease⁷⁸. In 30% of cases, without adequate treatment, the disease may progress to a chronic inflammatory stage, causing heart failure, arrhythmias, and death³. Different therapeutic strategies have been sought, where statins and aspirin could be candidates for repositioning⁸.

Seven experimental studies had been identified since 2015 to date, in which a statin was tested in murine models. Considering that *T. cruzi* has an affinity for cholesterol and that the parasite uses the LDL receptor as part of the mechanism to infect cells, the role of atorvastatin in lipid metabolism is studied, reporting a deleterious effect on the course of the disease, especially in subjects fed a high-fat diet¹¹⁰. However, Soares de Souza et al. observed that simvastatin can mitigate disease progression in a similar model²². Notwithstanding the role that statins may have on the mechanics of *T. cruzi* infection, it is more

likely that their actions in the chronic phase are related to the modulation of inflammation induced by the persistence of the parasite. Thus, it has been suggested that simvastatin could have anti-inflammatory potential in models of acute Chagas disease⁹³. Moreover, Campos-Estrada et al. proposed that simvastatin triggers the production of 15epi-LXA4 in endothelial cell models¹², although without causing a synergistic effect with benznidazole. However this synergism could exist, according to Araujo-Lima et al., who also highlights the low cardiotoxigenic potential of atorvastatin⁴, probably facilitating parasite clearance in cardiac tissue induced by the resolution of inflammation³³. Moreover, treating T. cruzi-infected human umbilical vein endothelial cells (HUVEC) with simvastatin promotes the differential expression of inflammation-related genes. Furthermore, simvastatin treatment of human umbilical vein endothelial cells infected with T. cruzi promotes the differential expression of inflammation-related genes. It also influences the Notch111 pathway, which is related to cardiac development in the embryo and may also participate in the cardiac protective effect of simvastatin during the chronic phase of the disease35.

Cyclooxygenase (COX) is a relevant player in the pathophysiology of T. cruzi infection³⁴, and much has been studied since the publication of the relationship of prostaglandins and the phagocytosis of apoptotic bodies²⁹. Since 2015, 12 studies have been published linking aspirin as a potential modulator of the course of T. cruzi infection. During the acute phase and as an evasive measure, the parasite modifies the macrophage response, generating an anti-inflammatory environment resulting from the production of prostaglandin E2 and the activation of TGF- β , especially in the presence of apoptotic T lymphocyte apoptotic bodies²⁹. Additionally, COX inhibition has been reported to facilitate parasite survival in the host^{67,69}. Furthermore, COX involvement in the invasion process has been recently demonstrated in a process inhibited by aspirin^{15,56,69}.

In an in vivo model of chronic Chagas disease, it was observed that aspirin treatment during the acute phase was able to prevent damage to esophageal nitrergic myenteric neurons⁶³. Those findings were corroborated in another study comparing aspirin administration during the acute or chronic phase, which showed a significant decrease in colonic inflammatory foci associated with a neuroprotective effect on myenteric neurons^{63,72,96}. Moreover, it has been reported that aspirin could have some beneficial impact on the neurological manifestations of Chagas disease, as it prevents behavioral alterations in mice acutely infected with T. cruzi94. Moreover, the early use of aspirin in combination with benznidazole proved effective in preventing chronic heart disease⁷⁸. Its use during the chronic phase of the disease can prevent the progression of cardiomyopathy, decreasing endothelial activation, cardiac inflammatory infiltrate, and fibrosis, probably by generating pro-resolving lipid mediators of inflammation such as 15-epi-LXA4 and AT-RvD1, among others^{14,58,65,66,73}. In a pilot study in humans, aspirin showed efficacy in reducing the symptoms associated with microvascular abnormalities caused by Chagas heart disease⁷⁷.

Despite the controversy about the role of COX inhibitors in *T. cruzi* infection¹⁹, there is abundant evidence supporting the immunomodulatory role of aspirin in the context

of Chagas disease 57,62 . However, there are still no clinical studies to corroborate this. On the other hand, there are no studies evaluating the use of statins or aspirin in the context of *T. brucei* infections during the period analyzed.

Conclusions

The relationship between a pathogen and its host determines the course of infection and defines whether the infection progresses or is effectively contained. Progression of infection can have deleterious consequences for the host because it can lead to disability or death. Immunity is one of these major determinants in the host-pathogen relationship. Parasitosis is not excluded from this interaction. However, therapeutic development to expand the antiparasitic pharmacological arsenal is insufficient, slow, or non-existent. Programs such as the Drugs for Neglected Diseases initiative seek to correct this scenario. The active search for compounds capable of modifying key aspects of the inflammatory process that are already approved for use in humans, have proven to be safe and reasonably priced, and a sound strategy⁹. In the present review, we present state-of-the-art agents such as aspirin and statins, which induce the production of inflammation-resolving agents and. therefore, could eventually change the course of infections, not only parasitic, as we have seen, but also bacterial or fungal infections. The uncontrolled nature of the inflammatory response in severe SARS-CoV2 infections, leading to patient death in many cases⁷⁴, boosted the field for exploring the use of agents to modulate the host factors⁸⁵. The prevention of organ damage induced by the persistence of the pathogen and the functional recovery of the innate immune response that prevents chronic inflammation are elements that can help to improve the efficacy of specific anti-infective treatments.

Undoubtedly other strategies to modify the host response would make more sense because they are more direct and probably broader in the spectrum, such as the direct blockade of TNF- α action or interferon- γ administration¹¹¹. However, these agents are not without adverse events, producing increased susceptibility to infections or severe arthropathy, respectively. Moreover, in many cases, they are also expensive. On the other hand, the use of vaccines for managing and preventing parasitic infections is far from optimal, and little progress has been made in this field, except for the RTS S/ASO1 malaria vaccine²⁰.

Unfortunately, the evidence supporting the utility of aspirin in inducing inflammation resolution is not strong. Moreover, the potential for severe reactions such as, for example, bleeding or gastric intolerance considerably diminishes the attractiveness of its use in humans, especially in parasitic diseases, mainly if they are chronic such as in Chagas disease or leishmaniasis. However, this drug may be helpful for further study of the phenomena of inflammation resolution in parasitic infections, as well as other immunopathogenic mechanisms that can be further modified with immunomodulatory drugs.

Furthermore, although our literature review mainly focused on *in vitro* and *in vivo* experimental models, a beneficial effect is glimpsed with statins as adjuvant therapy through their cholesterol lowering-independent effects, also

called pleiotropic. Although the isoprenoid metabolism in parasites can be directly inhibited by statins, halting their growth, there is no doubt that modulating the host response to the pro-inflammatory effects of infection is an attractive alternative. This effect must necessarily be proven in human studies. The study of modulating the resolution of inflammation with statins is a very active field in which they have also been tested for viral⁵¹ and bacterial infections⁷, including tuberculosis⁵², COVID-19⁸⁵, and septicemia³¹.

Thus, the modulation of host factors is an attractive tool to help combat these diseases in a context of emerging treatment resistance and spread to non-endemic areas, with the consequent risk of severe epidemic outbreaks. Because it is still controversial³⁰, it is necessary to improve the strength of evidence, determine the most effective statin, and, most importantly, analyze the clinical outcomes to further understand statin use in parasitic diseases.

Funding

This work was funded by Agencia Nacional de Investigacion y Desarrollo (ANID) programa FONDECYT, grant number 1210359.

Conflict of interest

The authors declare that they have no conflicts of interest.

References

- Alday PH, Doggett JS. Drugs in development for toxoplasmosis: advances, challenges, and current status. Drug Des Devel Ther. 2017;11:273-93, http://dx.doi.org/10.2147/DDDT.S60973.
- 2. Alidosti M, Heidari Ζ, ShahnaziF Н., Alavijeh F. Behaviors and perceptions related to cutaneous leishmaniasis in endemic areas of the a review. Acta Trop. 2021;223:106090, http://dx.doi.org/10.1016/j.actatropica.2021.106090.
- Álvarez-Hernández DA, García-Rodríguez-Arana R, Ortiz-Hernández A, Álvarez-Sánchez M, Wu M, Mejia R, Martínez-Juárez LA, Montoya A, Gallardo-Rincon H, Vázquez-López R, Fernández-Presas AMA. Systematic review of historical and current trends in Chagas disease. Ther Adv Infect Dis. 2021;8, http://dx.doi.org/10.1177/20499361211033715, 20499361211033715.
- Araujo-Lima CF, Peres RB, Silva PB, Batista MM, Aiub CAF, Felzenszwalb I, Soeiro MNC. Repurposing strategy of atorvastatin against *Trypanosoma cruzi*: in vitro monotherapy and combined therapy with benznidazole exhibit synergistic trypanocidal activity. Antimicrob Agents Chemother. 2018;62, http://dx.doi.org/10.1128/AAC.00979-18.
- Banerjee S, Bose D, Das S, Chatterjee N, Mishra S, Das Saha K. *Leishmania donovani* infection induce extracellular signal-regulated kinase (1/2) (ERK(1/2)) mediated lipid droplet generation in macrophages. Mol Immunol. 2022;141:328–37, http://dx.doi.org/10.1016/j.molimm.2021.12.008.
- Bang S, Donnelly CR, Luo X, Toro-Moreno M, Tao X, Wang Z, Chandra S, Bortsov AV, Ji ER, Derbyshire RR. Activation of GPR37 in macrophages confers protection against infection-induced sepsis and painlike behaviour in mice. Nat Commun. 2021;12:1704, http://dx.doi.org/10.1038/s41467-021-21940-8.
- 7. Bedi P, Chalmers JD, Graham C, Clarke A, Donaldson S, Doherty C, Govan JRW, Davidson DJ, Rossi AG, Hill AT.

- A randomized controlled trial of atorvastatin in patients with bronchiectasis infected with *Pseudomonas aeruginosa*: a proof of concept study. Chest. 2017;152:368–78, http://dx.doi.org/10.1016/j.chest.2017.05.017.
- Brindah J, Balamurali MM, Chanda K. An overview on the therapeutics of neglected infectious diseases – leishmaniasis and chagas diseases. Front Chem. 2021;9:622286, http://dx.doi.org/10.3389/fchem.2021.622286.
- Brindha J, Balamurali MM, Chanda K. An overview on the therapeutics of neglected infectious diseases – leishmaniasis and Chagas diseases. Front Chem. 2021;9:622286, http://dx.doi.org/10.3389/fchem.2021.622286.
- Buckley CD, Gilroy DW, Serhan CN. Proresolving lipid mediators and mechanisms in the resolution of acute inflammation. Immunity. 2014;40:315–27, http://dx.doi.org/10.1016/j.immuni.2014.02.009.
- Campos-Estrada C, Gonzalez-Herrera F, Greif G, Carillo I, Guzman-Rivera D, Liempi A, Robello C, Kemmerling U, Castillo C, Maya JD. Notch receptor expression in *Try-panosoma cruzi*-infected human umbilical vein endothelial cells treated with benznidazole or simvastatin revealed by microarray analysis. Cell Biol Int. 2020;44:1112–23, http://dx.doi.org/10.1002/cbin.11308.
- Campos-Estrada C, Liempi A, Gonzalez-Herrera F, Lapier M, Kemmerling U, Pesce B, Ferreira J, Lopez-Munoz R, Maya JD. Simvastatin and benznidazole-mediated prevention of *Trypanosoma cruzi-*induced endothelial activation: role of 15-epi-lipoxin A4 in the action of simvastatin. PLoS Negl Trop Dis. 2015;9:e0003770, http://dx.doi.org/10.1371/journal.pntd.0003770.
- 13. Canavese M, Crisanti A. Vascular endothelial growth factor (VEGF) and lovastatin suppress the inflammatory response to *Plasmodium berghei* infection and protect against experimental cerebral malaria. Pathog Glob Health. 2015;109:266–74, http://dx.doi.org/10.1179/2047773215Y.0000000021.
- 14. Carrillo I, Rabelo RAN, Barbosa C, Rates M, Fuentes-Retamal S, Gonzalez-Herrera F, Guzman-Rivera D, Quintero H, Kemmerling U, Castillo C, Machado FS, Diaz-Araya G, Maya JD. Aspirin-triggered resolvin D1 reduces parasitic cardiac load by decreasing inflammation in a murine model of early chronic Chagas disease. PLoS Negl Trop Dis. 2021;15:e0009978, http://dx.doi.org/10.1371/journal.pntd.0009978.
- 15. Carvalho de Freitas R, Lonien SCH, Malvezi AD, Silveira GF, Wowk PF, da Silva RV, Yamauchi LM, Yamada-Ogatta SF, Rizzo LV, Bordignon J, Pinge-Filho P. *Trypanosoma cruzi*: inhibition of infection of human monocytes by aspirin. Exp Parasitol. 2017;182:26–33, http://dx.doi.org/10.1016/j.exppara.2017.09.019.
- 16. Chen F, Fan XH, Wu YP, Zhu JL, Wang F, Bo LL, Li JB, Bao R, Deng XM. Resolvin D1 improves survival in experimental sepsis through reducing bacterial load and preventing excessive activation of inflammatory response. Eur J Clin Microbiol Infect Dis. 2014;33:457-64, http://dx.doi.org/10.1007/s10096-013-1978-6.
- 17. Chiang N, Fredman G, Backhed F, Oh SF, Vickery T, Schmidt BA, Serhan CN. Infection regulates pro-resolving mediators that lower antibiotic requirements. Nature. 2012;484:524–8, http://dx.doi.org/10.1038/nature11042.
- Chiang N, Serhan CN. Specialized pro-resolving mediator network: an update on production and actions. Essays Biochem. 2020;64:443-62, http://dx.doi.org/10.1042/EBC20200018.
- Cossentini LA, Da Silva RV, Yamada-Ogatta SF, Yamauchi LM, De Almeida Araujo EJ, Pinge-Filho P. Aspirin treatment exacerbates oral infections by Trypanosoma cruzi. Exp Parasitol. 2016;164:64–70, http://dx.doi.org/10.1016/j.exppara.2016.01.008.
- 20. Cotter C, Sturrock HJ, Hsiang MS, Liu J, Phillips AA, Hwang J, Gueye CS, Fullman N, Gosling RD, Feachem RG.

- The changing epidemiology of malaria elimination: new strategies for new challenges. Lancet. 2013;382:900–11, http://dx.doi.org/10.1016/S0140-6736(13)60310-4.
- Dalli J, Chiang N, Serhan CN. Elucidation of novel 13-series resolvins that increase with atorvastatin and clear infections. Nat Med. 2015;21:1071-5, http://dx.doi.org/10.1038/nm.3911.
- 22. de Souza DMS, de Paula Costa G, Leite ALJ, de Oliveira DS, de Castro Pinto KM, Farias SEB, Simoes NF, de Paiva NCN, de Abreu Vieira PM, da Silva CAM, Figueiredo VP, de Jesus Menezes AP, Talvani A. A high-fat diet exacerbates the course of experimental *Trypanosoma cruzi* infection that can be mitigated by treatment with simvastatin. Biomed Res Int. 2020;2020:1230461, http://dx.doi.org/10.1155/2020/1230461.
- Dinesh N, Soumya N, Singh S. Antileishmanial effect of mevastatin is due to interference with sterol metabolism. Parasitol Res. 2015;114:3873–83, http://dx.doi.org/10.1007/s00436-015-4618-5.
- Dormoi J, Briolant S, Desgrouas C, Pradines B. Impact of methylene blue and atorvastatin combination therapy on the apparition of cerebral malaria in a murine model. Malar J. 2013;12:127, http://dx.doi.org/10.1186/1475-2875-12-127.
- Dormoi J, Briolant S, Pascual A, Desgrouas C, Travaille C, Pradines B. Improvement of the efficacy of dihydroartemisinin with atorvastatin in an experimental cerebral malaria murine model. Malar J. 2013;12:302, http://dx.doi.org/10.1186/1475-2875-12-302.
- 26. Evangelista FF, Costa-Ferreira W, Mantelo FM, Beletini LF, de Souza AH, de Laet Sant'Ana P, de Lima KK, Crestani CC, Falavigna-Guilherme AL. Rosuvastatin revert memory impairment and anxiogenic-like effect in mice infected with the chronic ME-49 strain of *Toxoplasma gondii*. PLoS One. 2021;16:e0250079, http://dx.doi.org/10.1371/journal.pone.0250079.
- Fabiani S, Caroselli C, Menchini M, Gabbriellini G, Falcone M, Bruschi F. Ocular toxoplasmosis, an overview focusing on clinical aspects. Acta Trop. 2022;225:106180, http://dx.doi.org/10.1016/j.actatropica.2021.106180.
- Ferraz LRM, Silva L, Souza ML, Alves LP, Sales VAW, Barbosa I, Andrade MC, Santos WMD, Rolim LA, Rolim-Neto PJ. Drug associations as alternative and complementary therapy for neglected tropical diseases. Acta Trop. 2022;225:106210, http://dx.doi.org/10.1016/j.actatropica.2021.106210.
- Freire-de-Lima CG, Nascimento DO, Soares MB, Bozza PT, Castro-Faria-Neto HC, de Mello FG, DosReis GA, Lopes MF. Uptake of apoptotic cells drives the growth of a pathogenic trypanosome in macrophages. Nature. 2000;403:199–203, http://dx.doi.org/10.1038/35003208.
- Fullerton JN, O'Brien AJ, Gilroy DW. Lipid mediators in immune dysfunction after severe inflammation. Trends Immunol. 2014;35:12–21, http://dx.doi.org/10.1016/j.it.2013.10.008.
- 31. Ghayda RA, Lee JY, Yang JW, Han CH, Jeong GH, Yoon S, Hong SH, Lee KH, Gauckler P, Kronbichler A, Kang W, Shin JI. The effect of statins on all-cause and cardiovascular mortality in patients with non-dialysis chronic kidney disease, patients on dialysis, and kidney transplanted recipients: an umbrella review of meta-analyses. Eur Rev Med Pharmacol Sci. 2021;25:2696–710, http://dx.doi.org/10.26355/eurrev_202103_25433.
- 32. Ghosh J, Das S, Guha R, Ghosh D, Naskar K, Das A, Roy S. Hyperlipidemia offers protection against *Leishmania donovani* infection: role of membrane cholesterol. J Lipid Res. 2012;53:2560–72, http://dx.doi.org/10.1194/jlr.M026914.
- 33. Gonzalez-Herrera F, Cramer A, Pimentel P, Castillo C, Liempi A, Kemmerling U, Machado FS, Maya JD. Simvastatin attenuates endothelial activation through 15-epi-lipoxin A4 production in murine chronic chagas

- cardiomyopathy. Antimicrob Agents Chemother. 2017;61, http://dx.doi.org/10.1128/AAC.02137-16.
- 34. Guerrero NA, Camacho M, Vila L, Iniguez MA, Chillon-Marinas C, Cuervo H, Poveda C, Fresno M, Girones N. Cyclooxygenase-2 and prostaglandin E2 signaling through prostaglandin receptor EP-2 favor the development of myocarditis during acute *Trypanosoma cruzi* infection. PLoS Negl Trop Dis. 2015;9:e0004025, http://dx.doi.org/10.1371/journal.pntd.0004025.
- 35. Guzman-Rivera D, Liempi A, Gonzalez-Herrera F, Fuentes-Retamal S, Carrillo I, Abarca P, Castillo C, Kemmerling U, Pesce B, Maya JD. Simvastatin improves cardiac function through Notch 1 activation in BALB/c mice with chronic chagas cardiomyopathy. Antimicrob Agents Chemother. 2020;64, http://dx.doi.org/10.1128/AAC.02141-19.
- 36. Guzmán-Rivera D, Liempi A, González-Herrera F, Fuentes S, Carrillo I, Abarca P, Castillo C, Kemmerling U, Pesce B, Maya JD. Simvastatin improves cardiac function through Notch1 activation in BALB/c mice with chronic Chagas cardiomyopathy. Antimicrob Agents Chemother. 2020;64, http://dx.doi.org/10.1128/AAC.02141-19, e02141-19.
- 37. Harrell M, Carvounis PE. Current treatment of toxoplasma retinochoroiditis: an evidence-based review. J Ophthalmol. 2014;2014:273506, http://dx.doi.org/10.1155/2014/273506.
- 38. Hartley MA, Kohl K, Ronet C, Fasel N. The therapeutic potential of immune cross-talk in leishmaniasis. Clin Microbiol Infect. 2013;19:119–30, http://dx.doi.org/10.1111/1469-0691.12095.
- 39. Haughan PA, Chance ML, Goad LJ. Synergism in vitro of lovastatin and miconazole as anti-leishmanial agents. Biochem Pharmacol. 1992;44:2199–206, http://dx.doi.org/10.1016/0006-2952(92)90347-l.
- Helmers AJ, Gowda DC, Kain KC, Liles WC. Statins fail to improve outcome in experimental cerebral malaria and potentiate Toll-like receptor-mediated cytokine production by murine macrophages. Am J Trop Med Hyg. 2009;81:631–7, http://dx.doi.org/10.4269/ajtmh.2009.09-0204.
- 41. Hemmer CJ, Kern P, Holst FG, Nawroth PP, Dietrich M. Neither heparin nor acetylsalicylic acid influence the clinical course in human *Plasmodium falciparum* malaria: a prospective randomized study. Am J Trop Med Hyg. 1991;45:608–12, http://dx.doi.org/10.4269/ajtmh.1991.45.608.
- 42. Israelski DM, Remington JS. Toxoplasmosis in patients with cancer. Clin Infect Dis. 1993;17 Suppl. 2:S423–35, http://dx.doi.org/10.1093/clinids/17.supplement_2.s423.
- Kourbeli V, Chontzopoulou E, Moschovou K, Pavlos D, Mavromoustakos T, Papanastasiou IP. An overview on target-based drug design against kinetoplastid protozoan infections: human African trypanosomiasis, Chagas disease and leishmaniases. Molecules. 2021;26, http://dx.doi.org/10.3390/molecules26154629.
- 44. Kuckelhaus CS, Kuckelhaus SA, Muniz-Junqueira MI. Influence of long-term treatment with pravastatin on the survival, evolution of cutaneous lesion and weight of animals infected by Leishmania amazonensis. Exp Parasitol. 2011;127:658–64, http://dx.doi.org/10.1016/j.exppara.2010.12.003.
- Kuckelhaus 45. Kuckelhaus CS, SA, Tosta CE, modulates Junqueira MI. Pravastatin macrophage functions of Leishmania (L.) amazonensis-infected Parasitol. BALB/c mice. Exp 2013;134:18-25, http://dx.doi.org/10.1016/j.exppara.2013.01.020.
- 46. Kumar GA, Roy S, Jafurulla M, Mandal C, Chattopadhyay A. Statin-induced chronic cholesterol depletion inhibits *Leishmania donovani* infection: relevance of optimum host membrane cholesterol. Biochim Biophys Acta. 2016;1858:2088–96, http://dx.doi.org/10.1016/j.bbamem.2016.06.010.

- 47. Lee HN, Kundu JK, Cha YN, Surh YJ. Resolvin D1 stimulates efferocytosis through p50/p50-mediated suppression of tumor necrosis factor-alpha expression. J Cell Sci. 2013;126 Pt 17:4037–47, http://dx.doi.org/10.1242/jcs.131003.
- 48. Leuti A, Maccarrone M, Chiurchiu V. Proresolving lipid mediators: endogenous modulators of oxidative stress. Oxid Med Cell Longev. 2019;2019:8107265, http://dx.doi.org/10.1155/2019/8107265.
- Levy BD. Myocardial 15-epi-lipoxin A4 generation provides a new mechanism for the immunomodulatory effects of statins and thiazolidinediones. Circulation. 2006;114:873-5, http://dx.doi.org/10.1161/CIRCULATIONAHA.106.647925.
- Li R, Wang Y, Ma Z, Ma M, Wang D, Xie G, Yin Y, Zhang P, Tao K. Maresin 1 mitigates inflammatory response and protects mice from sepsis. Mediators Inflamm. 2016;2016:3798465, http://dx.doi.org/10.1155/2016/3798465.
- 51. Li X, Sheng L, Liu L, Hu Y, Chen Y, Lou L. Statin and the risk of hepatocellular carcinoma in patients with hepatitis B virus or hepatitis C virus infection: a meta-analysis. BMC Gastroenterol. 2020;20:98, http://dx.doi.org/10.1186/s12876-020-01222-1.
- Li X, Sheng L, Lou L. Statin use may be associated with reduced active tuberculosis infection: a meta-analysis of observational studies. Front Med (Lausanne). 2020;7:121, http://dx.doi.org/10.3389/fmed.2020.00121.
- 53. Li ZH, Li C, Szajnman SH, Rodriguez JB, Moreno SNJ. Synergistic activity between statins and bisphosphonates against acute experimental toxoplasmosis. Antimicrob Agents Chemother. 2017;61, http://dx.doi.org/10.1128/AAC.02628-16.
- 54. Li ZH, Ramakrishnan S, Striepen B, Moreno SN. Toxoplasma gondii relies on both host and parasite isoprenoids and can be rendered sensitive to atorvastatin. PLoS Pathog. 2013;9:e1003665, http://dx.doi.org/10.1371/journal.ppat.1003665.
- 55. Liberale L, Carbone F, Montecucco F, Sahebkar A. Statins reduce vascular inflammation in atherogenesis: a review of underlying molecular mechanisms. Int J Biochem Cell Biol. 2020;122:105735, http://dx.doi.org/10.1016/j.biocel.2020.105735.
- 56. Lonien SCH, Malvezi AD, Suzukawa HT, Yamauchi LM, Yamada-Ogatta SF, Rizzo LV, Bordignon J, Pinge-Filho P. Response to *Trypanosoma cruzi* by human blood cells enriched with dentritic cells is controlled by cyclooxygenase-2 pathway. Front Microbiol. 2017;8:2020, http://dx.doi.org/10.3389/fmicb.2017.02020.
- 57. Lopez-Munoz R, Faundez M, Klein S, Escanilla S, Torres G, Lee-Liu D, Ferreira J, Kemmerling U, Orellana M, Morello A, Ferreira A, Maya JD. *Trypanosoma cruzi*: in vitro effect of aspirin with nifurtimox and benznidazole. Exp Parasitol. 2010;124:167–71, http://dx.doi.org/10.1016/j.exppara.2009.09.005.
- Lopez-Munoz RA, Molina-Berrios A, Campos-Estrada C, Abarca-Sanhueza P, Urrutia-Llancaqueo L, Pena-Espinoza M, Maya JD. Inflammatory and pro-resolving lipids in Trypanosomatid infections: a key to understanding parasite control. Front Microbiol. 2018;9:1961, http://dx.doi.org/10.3389/fmicb.2018.01961.
- 59. Loynes CA, Lee JA, Robertson AL, Steel MJ, Ellett F, Feng Y, Levy BD, Whyte MKB, Renshaw SA. PGE2 production at sites of tissue injury promotes an anti-inflammatory neutrophil phenotype and determines the outcome of inflammation resolution in vivo. Sci Adv. 2018;4:eaar8320, http://dx.doi.org/10.1126/sciadv.aar8320.
- 60. Luu L, Johnston LJ, Derricott H, Armstrong SD, Randle N, Hartley CS, Duckworth CA, Campbell BJ, Coombes JM, Wastling JL. An open-format enteroid culture system for interrogation of interactions between *Toxoplasma gondii* and the

- intestinal epithelium. Front Cell Infect Microbiol. 2019;9:300, http://dx.doi.org/10.3389/fcimb.2019.00300.
- 61. Machado FS, Johndrow JE, Esper L, Dias A, Bafica A, Serhan CN, Aliberti J. Anti-inflammatory actions of lipoxin A4 and aspirin-triggered lipoxin are SOCS-2 dependent. Nat Med. 2006;12:330-4, http://dx.doi.org/10.1038/nm1355.
- 62. Malvezi AD, da Silva RV, Panis C, Yamauchi LM, Lovo-Martins MI, Zanluqui NG, Tatakihara VL, Rizzo LV, Verri WA Jr, Martins-Pinge MC, Yamada-Ogatta SF, Pinge-Filho P. Aspirin modulates innate inflammatory response and inhibits the entry of *Trypanosoma cruzi* in mouse peritoneal macrophages. Mediators Inflamm. 2014;2014:580919, http://dx.doi.org/10.1155/2014/580919.
- 63. Massocatto CL, Moreira NM, Muniz E, Pinge-Filho P, Rossi RM, Araujo EJ, Sant'Ana DM. Aspirin prevents atrophy of esophageal nitrergic myenteric neurons in a mouse model of chronic Chagas disease. Dis Esophagus. 2017;30:1–8, http://dx.doi.org/10.1111/dote.12449.
- 64. Megli CJ, Coyne CB. Infections at the maternal-fetal interface: an overview of pathogenesis and defence. Nat Rev Microbiol. 2022;20:67-82, http://dx.doi.org/10.1038/s41579-021-00610-y.
- 65. Molina-Berrios A, Campos-Estrada C, Henriquez N, Faundez M, Torres G, Castillo C, Escanilla S, Kemmerling U, Morello A, Lopez-Munoz RA, Maya JD. Protective role of acetylsalicylic acid in experimental *Trypanosoma cruzi* infection: evidence of a 15-epi-lipoxin A(4)-mediated effect. PLoS Negl Trop Dis. 2013;7:e2173, http://dx.doi.org/10.1371/journal.pntd.0002173.
- 66. Molina-Berrios A, Campos-Estrada C, Lapier M, Duaso J, Kemmerling U, Galanti N, Ferreira J, Morello A, Lopez-Munoz R, Maya JD. Protection of vascular endothelium by aspirin in a murine model of chronic Chagas' disease. Parasitol Res. 2013;112:2731-9, http://dx.doi.org/10.1007/s00436-013-3444-x.
- KC, Diniz LF, 67. Moraes Bahia MT. Role of cyclooxygenase-2 in Trypanosoma survival cruzi in the early stages of parasite host-cell interac-Mem Inst Oswaldo Cruz. http://dx.doi.org/10.1590/0074-02760140311.
- Mota S, Bensalel J, Park DH, Gonzalez S, Rodriguez A, Gallego-Delgado J. Treatment reducing endothelial activation protects against experimental cerebral malaria. Pathogens. 2022;11, http://dx.doi.org/10.3390/pathogens11060643.
- 69. Mukherjee S, Machado FS, Huang H, Oz HS, Jelicks LA, Prado CM, Koba W, Fine EJ, Zhao D, Factor SM, Collado JE, Weiss LM, Tanowitz HB, Ashton AW. Aspirin treatment of mice infected with *Trypanosoma cruzi* and implications for the pathogenesis of Chagas disease. PLoS One. 2011;6:e16959, http://dx.doi.org/10.1371/journal.pone.0016959.
- Muniz-Junqueira MI, Karnib SR, de Paula-Coelho VN, Junqueira LF Jr. Effects of pravastatin on the in vitro phagocytic function and hydrogen peroxide production by monocytes of healthy individuals. Int Immunopharmacol. 2006;6:53–60, http://dx.doi.org/10.1016/j.intimp.2005.07.010.
- Nishi L, Santana PL, Evangelista FF, Beletini LF, Souza AH, Mantelo FM, Souza-Kaneshima AM, Costa IN, Falavigna-Guilherme AL. Rosuvastatin reduced brain parasite burden in a chronic toxoplasmosis in vivo model and influenced the neuropathological pattern of ME-49 strain. Parasitology. 2020;147:303-9, http://dx.doi.org/10.1017/S0031182019001604.
- Oda JY, Belem MO, Carlos TM, Gouveia R, Luchetti BFC, Moreira NM, Massocatto CL, Araujo SM, Sant Ana DMG, Buttow NC, Pinge-Filho P, Araujo EJA. Myenteric neuroprotective role of aspirin in acute and chronic experimental infections with *Trypanosoma cruzi*. Neurogastroenterol Motil. 2017;29:1–13, http://dx.doi.org/10.1111/nmo.13102.

- 73. Ogata H, Teixeira MM, Sousa RC, Silva MV, Correia D, Rodrigues Junior V, Levy BD, Rogerio Ade P. Effects of aspirin-triggered resolvin D1 on peripheral blood mononuclear cells from patients with Chagas' heart disease. Eur J Pharmacol. 2016;777:26–32, http://dx.doi.org/10.1016/j.ejphar.2016.02.058.
- Panigrahy D, Gilligan MM, Huang S, Gartung A, Cortes-Puch I, Sime PJ, Phipps RP, Serhan CN, Hammock BD. Inflammation resolution: a dual-pronged approach to averting cytokine storms in COVID-19? Cancer Metastasis Rev. 2020;39:337–40, http://dx.doi.org/10.1007/s10555-020-09889-4.
- 75. Parihar SP, Hartley MA, Hurdayal R, Guler R, Brombacher F. Topical simvastatin as host-directed therapy against severity of cutaneous leishmaniasis in mice. Sci Rep. 2016;6:33458, http://dx.doi.org/10.1038/srep33458.
- Parquet V, Briolant S, Torrentino-Madamet M, Henry M, Almeras L, Amalvict R, Baret E, Fusai T, Rogier C, Pradines B. Atorvastatin is a promising partner for antimalarial drugs in treatment of *Plasmodium falciparum* malaria. Antimicrob Agents Chemother. 2009;53:2248–52, http://dx.doi.org/10.1128/AAC.01462-08.
- 77. Pavao RB, Moreira HT, Pintya AO, Haddad JL, Badran AV, Lima-Filho MO, Lago IM, Chierice JRA, Schmidt A, Marin-Neto JA. Aspirin plus verapamil relieves angina and perfusion abnormalities in patients with coronary microvascular dysfunction and Chagas disease: a pilot nonrandomized study. Rev Soc Bras Med Trop. 2021;54:e0181, http://dx.doi.org/10.1590/0037-8682-0181-2021.
- 78. Pereira RS, Malvezi AD, Lovo-Martins MI, Lucchetti BFC, Santos JP, Tavares ER, Verri WA Jr, de Almeida Araujo EJ, Yamauchi LM, Yamada-Ogatta SF, Martins-Pinge MC, Pinge-Filho P. Combination therapy using benznidazole and aspirin during the acute phase of experimental chagas disease prevents cardiovascular dysfunction and decreases typical cardiac lesions in the chronic phase. Antimicrob Agents Chemother. 2020;64, http://dx.doi.org/10.1128/AAC.00069-20.
- 79. Plewes K, Leopold SJ, Kingston HWF, Dondorp AM. Malaria: what's new in the management of malaria? Infect Dis Clin North Am. 2019;33:39-60, http://dx.doi.org/10.1016/j.idc.2018.10.002.
- 80. Reis PA, Estato V, da Silva TI, d'Avila JC, Siqueira LD, Assis EF, Bozza PT, Bozza FA, Tibirica EV, Zimmerman GA, Castro-Faria-Neto HC. Statins decrease neuroinflammation and prevent cognitive impairment after cerebral malaria. PLoS Pathog. 2012;8:e1003099, http://dx.doi.org/10.1371/journal.ppat.1003099.
- 81. Rozenfeld C, Martinez R, Figueiredo RT, Bozza MT, Lima FR, Pires AL, Silva PM, Bonomo A, Lannes-Vieira J, De Souza W, Moura-Neto V. Soluble factors released by *Toxoplasma gondii*-infected astrocytes down-modulate nitric oxide production by gamma interferon-activated microglia and prevent neuronal degeneration. Infect Immun. 2003;71:2047–57, http://dx.doi.org/10.1128/IAI.71.4.2047-2057.2003.
- 82. Sanfelice R, da Silva SS, Bosqui LR, Machado LF, Miranda-Sapla MM, Panagio LA, Navarro IT, Conchon-Costa I, Pavanelli WR, Almeida RS, Costa IN. Pravastatin and simvastatin pretreatment in combination with pyrimethamine and sulfadiazine reduces infection process of *Toxoplasma gondii* tachyzoites (RH strain) in HeLa cells. Acta Parasitol. 2019;64:612–6, http://dx.doi.org/10.2478/s11686-019-00076-2.
- 83. Sanfelice RA, da Silva SS, Bosqui LR, Miranda-Sapla MM, Barbosa BF, Silva RJ, Ferro EAV, Panagio LA, Navarro IT, Bordignon J, Conchon-Costa I, Pavanelli WR, Almeida RS, Costa IN. Pravastatin and simvastatin inhibit the adhesion, replication and proliferation of *Toxoplasma gondii* (RH strain) in HeLa cells. Acta Trop. 2017;167:208–15, http://dx.doi.org/10.1016/j.actatropica.2016.12.006.

- 84. Sanfelice RA, Machado LF, Bosqui LR, Miranda-Sapla MM, Tomiotto-Pellissier F, de Alcantara Dalevedo G, Ioris D, Reis GF, Panagio LA, Navarro IT, Bordignon J, Conchon-Costa I, Pavanelli WR, Almeida RS, Costa IN. Activity of rosuvastatin in tachyzoites of *Toxoplasma gondii* (RH strain) in HeLa cells. Exp Parasitol. 2017;181:75–81, http://dx.doi.org/10.1016/j.exppara.2017.07.009.
- 85. Santosa A, Franzen S, Natman J, Wettermark B, Parmryd I, Nyberg F. Protective effects of statins on COVID-19 risk, severity and fatal outcome: a nationwide Swedish cohort study. Sci Rep. 2022;12:12047, http://dx.doi.org/10.1038/s41598-022-16357-2.
- Satny M, Hubacek JA, Vrablik M. Statins and inflammation. Curr Atheroscler Rep. 2021;23:80, http://dx.doi.org/10.1007/s11883-021-00977-6.
- 87. Serhan CN. Lipoxins and aspirin-triggered 15-epilipoxins are the first lipid mediators of endogenous anti-inflammation and resolution. Prostaglandins Leukot Essent Fatty Acids. 2005;73:141-62, http://dx.doi.org/10.1016/j.plefa.2005.05.002.
- Serhan CN. Pro-resolving lipid mediators are leads for resolution physiology. Nature. 2014;510:92–101, http://dx.doi.org/10.1038/nature13479.
- 89. Serhan CN. Resolution phase of inflammation: novel endogenous anti-inflammatory and proresolving lipid mediators and pathways. Annu Rev Immunol. 2007;25:101–37, http://dx.doi.org/10.1146/annurev.immunol.25.022106. 141647.
- 90. Serhan CN, Chiang N, Dalli J, Levy BD. Lipid mediators in the resolution of inflammation. Cold Spring Harb Perspect Biol. 2014;7:a016311, http://dx.doi.org/10.1101/cshperspect.a016311.
- 91. Serhan CN, Fredman G, Yang R, Karamnov S, Belayev LS, Bazan NG, Zhu M, Winkler JW, Petasis NA. Novel proresolving aspirin-triggered DHA pathway. Chem Biol. 2011;18:976–87, http://dx.doi.org/10.1016/j.chembiol.2011.06.008.
- 92. Serhan CN, Levy BD. Resolvins in inflammation: emergence of the pro-resolving superfamily of mediators. J Clin Invest. 2018;128:2657-69, http://dx.doi.org/10.1172/JCI97943.
- 93. Silva RR, Shrestha-Bajracharya D, Almeida-Leite CM, Leite R, Bahia MT, Talvani A. Short-term therapy with simvastatin reduces inflammatory mediators and heart inflammation during the acute phase of experimental Chagas disease. Mem Inst Oswaldo Cruz. 2012;107:513–21, http://dx.doi.org/10.1590/s0074-02762012000400012.
- 94. Silvero-Isidre A, Morinigo-Guayuan S, Meza-Ojeda A, Mongelos-Cardozo M, Centurion-Wenninger C, Figueredo-Thiel SD, Sanchez F, Acosta N. Protective effect of aspirin treatment on mouse behavior in the acute phase of experimental infection with *Trypanosoma cruzi*. Parasitol Res. 2018;117:189–200, http://dx.doi.org/10.1007/s00436-017-5693-6.
- 95. Souraud JB, Briolant S, Dormoi J, Mosnier J, Savini H, Baret E, Amalvict R, Soulard R, Rogier C, Pradines B. Atorvastatin treatment is effective when used in combination with mefloquine in an experimental cerebral malaria murine model. Malar J. 2012;11:13, http://dx.doi.org/10.1186/1475-2875-11-13.
- 96. Souza ND, Belin BS, Massocatto CL, Araujo SM, Sant'ana DMG, Araujo EJA, Filho PP, Nihei OK, Moreira NM. Effect of acetylsalicylic acid on total myenteric neurons in mice experimentally infected with *Trypanosoma cruzi*. An Acad Bras Cienc. 2019;91:e20180389, http://dx.doi.org/10.1590/0001-3765201920180389.
- 97. Tang S, Wan M, Huang W, Stanton RC, Xu Y. Maresins: specialized proresolving lipid mediators and their potential role in inflammatory-related diseases. Mediators Inflamm. 2018;2018:2380319, http://dx.doi.org/10.1155/2018/2380319.

- 98. Taoufiq Z, Pino P, N'Dilimabaka N, Arrouss I, Assi S, Soubrier F, Rebollo A, Mazier D. Atorvastatin prevents *Plasmodium falciparum* cytoadherence and endothelial damage. Malar J. 2011;10:52, http://dx.doi.org/10.1186/1475-2875-10-52.
- 99. Tartey S, Takeuchi O. Pathogen recognition and Toll-like receptor targeted therapeutics in innate immune cells. Int Rev Immunol. 2017;36:57–73, http://dx.doi.org/10.1080/08830185.2016.1261318.
- 100. Terkawi MA, Nishimura M, Furuoka H, Nishikawa Y. Depletion of phagocytic cells during nonlethal *Plasmodium yoelii* infection causes severe malaria characterized by acute renal failure in mice. Infect Immun. 2016;84:845–55, http://dx.doi.org/10.1128/IAI.01005-15.
- 101. Thornton JM, Walker JM, Sundarasivarao PYK, Spur BW, Rodriguez A, Yin K. Lipoxin A4 promotes reduction and antibiotic efficacy against *Pseudomonas aeruginosa* biofilm. Prostaglandins Other Lipid Mediat. 2021;152:106505, http://dx.doi.org/10.1016/j.prostaglandins.2020.106505.
- 102. Thornton JMK, Yin K. Role of specialized proresolving mediators in modifying host defense and decreasing bacterial virulence. Molecules. 2021;26, http://dx.doi.org/10.3390/molecules26226970.
- 103. Torre D, Speranza F, Martegani R, Zeroli C, Banfi M, Airoldi M. A retrospective study of treatment of cerebral toxoplasmosis in AIDS patients with trimethoprim-sulphamethoxazole. J Infect. 1998;37:15–8, http://dx.doi.org/10.1016/s0163-4453(98)90217-1.
- **104.** WHO. World malaria report 2020: 20 years of global progress and challenges. Geneva: World Health Organization; 2020.
- 105. Wilson NO, Solomon W, Anderson L, Patrickson J, Pitts S, Bond V, Liu M, Stiles JK. Pharmacologic inhibition of CXCL10 in combination with anti-malarial therapy eliminates mortality associated with murine

- model of cerebral malaria. PLoS One. 2013;8:e60898, http://dx.doi.org/10.1371/journal.pone.0060898.
- 106. Wolfe ND, Dunavan CP, Diamond J. Origins of major human infectious diseases. Nature. 2007;447:279–83, http://dx.doi.org/10.1038/nature05775.
- 107. Wu B, Capilato J, Pham MP, Walker J, Spur B, Rodriguez A, Perez LJ, Yin K. Lipoxin A4 augments host defense in sepsis and reduces *Pseudomonas aeruginosa* virulence through quorum sensing inhibition. FASEB J. 2016;30:2400–10, http://dx.doi.org/10.1096/fj.201500029R.
- 108. Xiao L, Patterson PS, Yang C, Lal AA. Role of eicosanoids in the pathogenesis of murine cerebral malaria. Am J Trop Med Hyg. 1999;60:668-73, http://dx.doi.org/10.4269/ajtmh.1999.60.668.
- 109. Yang T, Ottilie S, Istvan ES, Godinez-Macias KP, Lukens AK, Baragana B, Campo B, Walpole C, Niles JC, Chibale K, Dechering KJ, Llinas M, Lee MCS, Kato N, Wyllie S, McNamara CW, Gamo FJ, Burrows J, Fidock DA, Goldberg DE, Gilbert IH, Wirth DF, Winzeler EAC, Malaria Drug Accelerator. MalDA, accelerating malaria drug discovery. Trends Parasitol. 2021;37:493–507, http://dx.doi.org/10.1016/j.pt.2021.01.009.
- 110. Zhao D, Lizardo K, Cui MH, Ambadipudi K, Lora J, Jelicks LA, Nagajyothi JF. Antagonistic effect of atorvastatin on high fat diet induced survival during acute Chagas disease. Microbes Infect. 2016;18:675–86, http://dx.doi.org/10.1016/j.micinf.2016.06.006.
- 111. Zumla A, Rao M, Wallis RS, Kaufmann SHE, Rustomjee R, Mwaba P, Vilaplana C, Yeboah-Manu D, Chakaya J, Ippolito G, Azhar E, Hoelscher M, Maeurerc M, Host-Directed Therapies Network. Host-directed therapies for infectious diseases: current status, recent progress, and future prospects. Lancet Infect Dis. 2016;16:e47-63, http://dx.doi.org/10.1016/S1473-3099(16)00078-5.