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ORIGINAL ARTICLE

The wide distribution of an extremely thermoacidophilic microorganism in the copper mine at ambient temperature and under acidic condition and its significance in bioleaching of a chalcopyrite concentrate



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KEYWORDS

Chalcopyrite; Thermoacidophilic; Bioleaching; Copper recovery; Antagonistic reactions

Abstract Thermoacidophiles can exist in a state of dormancy both in moderate temperatures and even in cold conditions in heap leaching. Sulphide mineral ores such as chalcopyrite produce sulfuric acid when exposed to the air and water. The produced sulfuric acid leads to the decrease of pH and exothermic reactions in heap leaching causing the temperature to increase up to 55 °C and the activation of thermoacidophilic microorganisms. The aim of the present study was to isolate indigenous extreme thermoacidophilic microorganisms at ambient temperature from Sarcheshmeh Copper Complex, to adapt them to the high pulp density of a chalcopyrite concentrate, and to determine their efficiency in chalcopyrite bioleaching in order to recover copper. In this study samples were collected at ambient temperature from Sarcheshmeh Copper Complex in Iran. Mixed samples were inoculated into the culture medium for enrichment of the microorganisms. Pure cultures from these enrichments were obtained by subculture of liquid culture to solid media. Morphological observation was performed under the scanning electron microscope. Isolates were adapted to 30% (w/v) pulp density. For the bioleaching test, the experiments were designed with DX7 software. Bioleaching experiments were carried out in Erlenmeyer flasks and a stirred tank reactor. The highest copper recovery in Erlenmeyer flasks was 39.46% with pulp 15%, inoculums 20%, size particle 90 µm and 160 rpm. The lowest recovery was 3.81% with pulp 20%, inoculums 20%, size particle $40\,\mu m$ and $140\,rpm$ after 28 days. In the reactor, copper recovery was 32.38%. Bioleaching residues were analyzed by the X-ray

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diffraction (XRD) method. The results showed no jarosite ($KFe_3(SO_4)_2(OH)_6$) had formed in the bioleaching experiments. It seems that the antagonistic reactions among various species and a great number of planktonic cells in Erlenmeyer flasks and the stirred tank reactor are the reasons for the low recovery of copper in our study.

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PALABRAS CLAVE

Calcopirita; Termoacidófilo; Biolixiviación; Recuperación de cobre; Reacciones antagónicas La amplia distribución de un microorganismo termoacidófilo en el mineral de una mina cuprífera a temperatura ambiente y en condición ácida, y su importancia en la biolixiviación del concentrado de calcopirita

Resumen Los microorganismos termoacidófilos pueden estar en estado latente tanto a temperatura moderada como baja, en lixiviación en pilas. Los minerales sulfurosos, como la calcopirita, producen ácido sulfúrico cuando se exponen al aire y al agua. El ácido sulfúrico producido conduce a la disminución del pH y a reacciones exotérmicas durante la lixiviación en pilas, lo que hace que la temperatura aumente hasta 55 °C y se activen los microorganismos termoacidófilos. El objetivo del presente estudio fue aislar del complejo de cobre Sarcheshmeh (Irán) microorganismos termoacidófilos extremos que proliferan a temperatura ambiente e investigar su adaptación a la alta densidad de pulpa del concentrado de calcopirita, así como su eficiencia para biolixiviarese mineral, con el objeto de recuperar el cobre. Se recogieron muestras a temperatura ambiente del citado complejo, y luego muestras mixtas se inocularon en un medio de cultivo de enriquecimiento. A partir de estos enriquecimientos, mediante el subcultivo del cultivo líquido a medio sólido, se obtuvieron cultivos puros. La observación morfológica se realizó bajo microscopio electrónico de barrido. Los aislados estaban adaptados al 30% p/v de densidad de pulpa. Para la prueba de biolixiviación, los experimentos fueron diseñados con el software DX7. Los experimentos de biolixiviación se llevaron a cabo en Erlenmeyers y en un reactor tanque con agitación. La mayor recuperación de cobre en los Erlenmeyers fue del 39,46% y se obtuvo con la pulpa al 15%, un inóculo del 20%, un tamaño de partícula de 90 µm y una agitación de 160 rpm. La menor recuperación fue del 3,81% y se obtuvo con la pulpa al 20%, un inóculo del 20%, un tamaño de partícula de 40 µm y una agitación de 140 rpm, después 28 días. En el reactor, la recuperación del cobre fue del 32,38%. El análisis de difracción de rayos X (XRD) no mostró que se formara jarosita (KFe₃[SO₄]₂[OH]₆) en los experimentos de biolixiviación. Dicha técnica sirve para determinar la estructura cristalina de una sustancia desconocida. Al parecer, las reacciones antagónicas entre las diversas especies y el mayor número de células planctónicas en los Erlenmeyers y en el reactor fueron las causas de la baja recuperación de cobre observada en este estudio.

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Introduction

Chalcopyrite (CuFeS₂), is the main copper-containing sulfide mineral and the most persistent of the copper sulfides in the lithosphere^{8,13,17,18,35,37,38,40}. Moreover, it is economically the most important resource of copper^{19,38}. In the pyrometallurgical processes used for chalcopyrite, more than 90% copper recovery can be achieved; however, serious pollution to the environment is always produced such as sulfur dioxide and arsenic trioxide^{2,37}. The use of microorganisms to extract copper from chalcopyrite has several advantages such as easy operation, obvious economic benefits, being more environmentally friendly than pyrometallurgical processes^{8,10,41}. At first, it was believed that the most important microorganisms involved in the leaching

of chalcopyrite were the mesophilic acidophiles^{9,25}. However, they have been less successful in oxidizing chalcopyrite mostly due to slow kinetics, poor extraction, long leaching time and passivation of the mineral by ferric precipitates and/or sulfur. For this reason, their industrial use is still a major challenge in the field of biohydrometallurgy^{12,14,38}. In order to overcome these problems, various methods proposed by researchers include the control of redox levels via the addition of catalysts such as silver and bismuth, ultra-fine grinding of the concentrate prior to bioleaching and the addition of chloride^{1,4,12,13}. It seems that thermophilic microbes are much more efficient in increasing the rate of chemical ferric oxidation of chalcopyrite and other minerals². Therefore, another way to overcome this problem is to use thermophilic microorganism strains instead of

mesophilic microorganisms^{2,14,24,40}. This approach has garnered much attention in recent years and has been assessed greatly in batch tests at laboratory scale¹³.

In recent years, new species of thermophilic microorganisms with greater capacity for solution of metal sulfides and more resistance to different toxic metal cations have been isolated^{3,14,26}. However, all thermoacidophilic microorganisms do not confer the same degree of effectiveness, and vary from one species to another². Therefore, it is imperative to find new species capable of giving high dissolution percentages and tolerating higher pulp density. Although several researchers have reported that pulp densities higher than 15-20% (w/v) are injurious to cell viability and oxidizing activity, both work with mesophiles and thermophiles². Moreover, chalcopyrite concentrates produced from distinctive ores may contain different concentrations of contaminants such as metals and organic chemical materials. Mostly, the presence of various concentrations of metals and chemical materials is highly undesirable for bioleaching and may prevent the growth of standard microorganisms¹⁷. Autochthonous microbes are not only more resistant than metals and chemical materials, but can also multiply for several generations within the copper sulfide ore and may effectively increase the extraction of copper during bioleaching. In addition, the isolation of such microbes would eliminate the necessity of using standard laboratory cultures, specifically for largescale studies in industrial processing of mineral sulfide ores or concentrates 18,24.

The aim of this study was to isolate indigenous extremely thermoacidophilic microorganisms at ambient temperature from Sarcheshmeh Copper Complex in Iran, determine their efficiency in the bioleaching of chalcopyrite for the recovery of copper and answer the key question of whether the above mentioned microorganisms can tolerate higher pulp density than other extreme thermophilic microorganisms or not.

Material and methods

Site description and samples

Samples were collected at the Sarcheshmeh Copper Complex. The mine is located 160 km southwest of Kerman city in Iran and has been continuously exploited for more than 40 years. Sample collection was carried out in January 2015. Ambient temperature was about 6 °C. Eight samples were collected from mine soil at the surface layer (0–10 cm). Pregnant leach solution (PLS) samples were taken from the pond located under heap number one. Five samples were also taken from running waters around the mine. All samples were collected in several plastic bags and carried to the laboratory with no temperature control within a day. All samples were mixed in one bucket. pH value was 2.4 and was measured by analyzer (Metrohm model 827).

Culture medium

Mixed samples were inoculated into the culture medium for enrichment and cultured in autoclaved *Acidianus brierleyi* medium [DSMZ (http://dsmz.de/media/media150.htm)]. Experiments were performed in 500 ml Erlenmeyer flasks.

Each flask contained 150 ml culture medium and 50 ml mixed samples.

Initial pH was adjusted to 1.98 with Merck sulfuric acid. Cultures were run in a thermostatic rotary agitator (Innova, New Brunswick Scientific Innova 4200 incubator shaker) at 70 °C and 125 rpm. Water loss by evaporation was compensated by the daily addition of distilled water. ORP (oxidation–reduction potential = Eh) using Eh meter (WTW model 325, Electrode SenTix ORP 0–100/3 mol/l KCl) and pH value was monitored in successive intervals.

Isolation of microorganism

The mixed culture was subcultured based on growth rate at 7–18 day intervals in the new medium with the above condition in an Erlenmeyer flask. Cell density and morphology of the microorganism in the solution were analyzed according to the methods described by Keeling et al.¹⁸ and Zeng et al.³⁷ The pure culture was obtained after four successive subcultures in one month under aerobic condition. The redox potential and pH of each culture were monitored and adjusted as described above.

Growth on solid media

To ensure that a pure culture was achieved, pure cultures from these enrichments were obtained by subculture of 0.1 ml liquid culture to solid media solidified with 0.6% Gelrite in aerobic condition. The solid medium was prepared with the above composition; only sulfur was replaced by sodium thiosulfate.

Morphological observation

The morphology of the isolated culture grown on Gelrite was observed under a Scanning Electron Microscope PHE-NOM Pro X (Max Magnification: $45\,000\times$, Resolution: $25\,\text{nm}$, Detectors: BSD, SED and SDD, working voltage: $5\,\text{kV}$, $10\,\text{kV}$ and $15\,\text{kV}$).

Concentrate preparation and chemical analysis

A representative sample of chalcopyrite concentrate was obtained from the Sarcheshmeh Copper Complex of Iran. Sampling took two months. The samples were placed in the laboratory on a sheet to dry. The dried samples were rolled and sifted successively until a sample for testing could be obtained. The d90 of the sample achieved 90 μm using a cyclosizer. This concentrate was then pulverized to a particle size of d90 = 40 μm . The chemical analysis of the concentrate was performed using X-ray fluorescence (XRF) and X-ray diffraction (XRD) methods.

Adaptation of microbes to chalcopyrite prior to leaching tests

The adaptation of isolates to the chalcopyrite concentrate was carried out according to the procedures described by Astudillo and Acevedo². Fifteen transfers were done aerobically at 7 day-intervals, at 70 °C for about four months

onto the abovementioned medium in an orbital incubator shaker agitated at 125–175 rpm. Pulp densities were 2, 4, 6, 8, 10, 12, 14, 16, 18, 20, 22, 24, 26, 28 and 30% (w/v). This adapted population was used as inoculum to new cultures of increasing pulp density.

Bioleaching experiments

Experiment design

The experiments were designed with DX7 software²³. Four parameters, inoculum percentage, pulp density, agitation and size particle of concentrate, were considered in this investigation and consequently, 31 experiments were carried out using the following levels: inoculum percentage of bacteria (5, 10, 20%); pulp density (10, 15, 20%); agitation (120, 140, 160 rpm); particle size (90, $40 \mu m$).

Shake flask

Bioleaching experiments were carried out in autoclaved 500 ml Erlenmeyer flasks. The total volume of each flask was 200 ml containing medium, adapted culture containing about 1.0×10^7 cells/ml and pulp density. Erlenmeyer flasks were placed in a rotary shaking incubator at $70\,^{\circ}$ C. Control experiments were conducted without inoculum.

The evaporated water in the medium was supplemented with sterilized deionized water and culture medium. Evaporation losses were determined by weighing all flasks before and after sampling. The leaching experiments lasted for 28 days. The experiment was accomplished at 160, 120 and 140 rpm respectively.

Stirred tank reactor

The best result obtained in the shake flask was tested in the stirred tank reactor for better recovery of copper. Bioleaching experiments were carried out in a 1l glass reactor at 70 °C. Oxygen was introduced, also into the base of the reactor, at an approximate rate of 1l/min (Fig. 1).

Analyses

During the experiment, pH and ORP values were measured. Aliquots of the solution were sampled, and the concentration of Cu²⁺, Fe²⁺, total iron and acid in the solution was measured in successive intervals. Copper and total iron concentrations in the solution were determined by atomic absorption spectrophotometer; variant AA220. At the end of the bioleaching tests, solids were removed by filtration, dried and their chemical composition was analyzed by XRF and XRD. XRD was used for jarosite examination. The acid consumption was compensated by sulfuric acid to keep pH value around 2 in the early stages. All analyses were performed in the central laboratory of Sarcheshmeh Copper Complex.

Statistics

The statistical analysis of the bioleaching and control experiments in the shake flask were performed using One-Sample t-test. $p \le 0.05$ was accepted as statistically significant.



Figure 1 Glass reactor with a mechanic stirrer operating.

Result and discussion

Measure of pH and Eh

pH and Eh-related curve before bioleaching over 12 days from one of the sub-cultures was shown in Figure 2. pH increased from an initial value of 1.96–2.4 after 2 days of incubation, followed by a decrease up to the 5th day. The curve was followed by an increase and then a decrease. Eh increased from 390 mV initially on the first day to 427 mV after 4 days of incubation. This curve was followed by an increase and a decrease.

Growth on solid media

After incubation at $70\,^{\circ}$ C for about 30 days, very small colonies were observed. In order to prevent the plate from drying out, the plastic lacuna was used. Inoculated plates

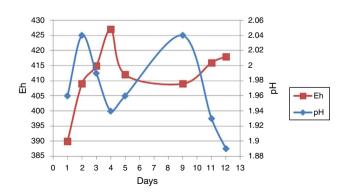


Figure 2 pH and Eh-related curve before bioleaching in culture medium.

were inspected daily for microbial growth and water evaporation.

Growth in anaerobic and aerobic conditions

When the colonies grown on Gelrite were cultured in liquid media, growth was poor and the number of microbes did not increase over time. Our results showed that in addition to the compounds used to culture the microorganisms, mine soil should also be analyzed to provide trace elements for growth. If possible, trace elements should be identified to enhance microorganism growth, because the microorganisms grow very well as long as they are in the presence of their natural ingredients.

Morphological observation

The cell morphology in the scanning electron microscope was similar to *Sulfolobus*-like microorganisms, with lobed irregular cocci, variable shape and size (Fig. 3). The isolate was motile.

Chemical composition of chalcopyrite concentrate

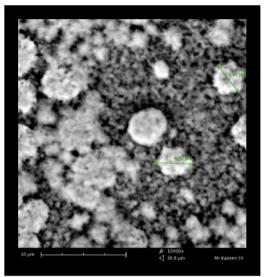
The chemical composition of the concentrate with X-ray fluorescence (XRF) analysis is presented in Table 1. X-ray diffraction (XRD) analysis reported chalcopyrite 95.5% and pyrite 4.4%.

Bioleaching experiments and copper recovery in shake flask

According to the residue analysis, the result of the experiment design based on four selected control parameters and their application in the study are shown in Table 2. The results showed that different runs had different percentage recovery of copper after 28 days. As it is seen, the highest recovery is run 20 with pulp 15%, inoculums 20%, particle size 90 μm and 160 rpm with 39.46% recovery and the lowest recovery is run 23 with pulp 20%, inoculums 20%, particle size 40 μm and 140 rpm with 3.81% recovery. Run 32 is chemical leaching (control).

The results of the statistical analysis in relation to the extracted copper in different processes of bioleaching test (31 stage) relative to the control stage (chemical leaching) showed that the mean extracted copper in bioleaching test were 12.83. The results of One-Sample t-test showed that there is a significant difference between bioleaching and chemical leaching (p = 0.001).

The aim of bioleaching processes is the biodegradation of ore or concentrate using organisms which do so effectively. Those microorganisms which can degrade ores more effectively are the ones with the fastest growth and the fastest



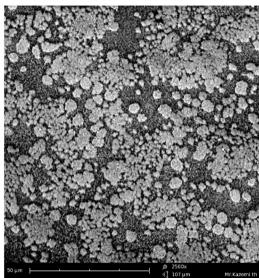


Figure 3 The morphological characteristic under Scanning Electron Microscope ($10\,000 \times = a$) and ($2500 \times = b$).

doubling time. Therefore, there is a strong positive natural selection for this type of microorganisms, which would become the microbial dominant community in bioleaching environment while the others would be eliminated or washed away. In this study, the chalcopyrite concentrate did not sterilize so that the microbial community could contribute to the bioleaching like the mineral processes. Some studies like the present study have shown at laboratory scale that extreme thermoacidophilic culturing contains *Sulfolobus*-like microorganisms and can be successfully effective in chalcopyrite concentrate bioleaching^{6,7,28}. In this study, the mixed microorganisms were confirmed by

Table 1	Composition of the chalcopyrite concentrate used in bioleaching experiments by X-ray fluorescence (XRF)										
Element	Cu	Fe	Мо	SiO ₂	S	Al_2O_3	ZnO	TiO ₂	PbO	K ₂ O	CaO
%	24.10	28.40	0.08	5.87	31.03	2.43	0.47	0.12	0.10	0.43	0.26

Run no.	Initial pH	Final pH		Para	Cu-extract (%)	Fe-extract (%)		
			Pulp density (9	6) Inoculation (%)	Particle size (μm)	Agitation (rpm)		
1	1.92	1.80	20	5	40	140	5.45	11.44
2	1.87	1.48	20	10	90	140	5.91	11.34
3	1.92	1.63	15	5	40	120	11.05	8.58
4	1.93	1.75	10	5	40	140	12.39	5.72
5	1.95	1.57	10	5	40	160	17.93	5.72
6	1.87	1.72	10	5	40	120	11.60	5.72
7	1.91	1.72	15	20	40	140	11.58	8.58
8	1.95	1.87	20	5	40	160	16.77	11.44
9	1.94	1.99	20	20	90	160	17.01	11.34
10	1.98	1.71	10	10	40	160	27.73	5.72
11	1.96	1.82	15	5	90	120	11.88	8.5
12	1.97	2.0	15	10	90	140	13.52	8.5
13	1.94	2.03	10	5	90	120	16.69	5.67
14	1.94	1.62	15	10	40	160	12.65	8.58
15	1.93	1.57	20	20	40	120	7.50	11.44
16	1.94	1.91	10	20	90	140	11.19	5.67
17	1.87	2.12	20	10	40	120	9.09	11.44
18	1.92	1.60	15	20	90	120	8.12	8.5
19	1.97	2.03	10	20	40	160	14.25	5.72
20	1.98	1.80	15	20	90	160	39.46	8.5
21	1.90	1.83	15	5	90	160	12.84	8.5
22	1.95	1.60	15	10	40	140	12.66	8.5
23	1.91	1.75	20	20	40	140	3.81	11.44
24	1.92	1.89	10	5	90	140	13.58	5.67
25	1.98	1.70	10	10	90	160	10.92	5.67
26	1.97	1.85	20	5	90	120	9.9	11.34
27	1.97	1.82	10	20	40	140	12.38	5.72
28	1.89	1.78	20	5	40	120	4.50	11.44
29	1.95	1.69	10	10	90	120	19.6	5.67
30	1.87	1.83	15	10	40	120	12.85	8.5
31	1.80	1.85	10	10	40	120	7.30	5.72
32	1.90	1.75	20	0	40	140	8.53	11.44

microscopic (Fig. 3) and metagenomic (results not shown) techniques.

Most of the work on thermophilic microorganisms has been confined to rather low pulp densities of less than 5%18,21,30. Similar to Guo and Rubio, in this work, thermophilic microorganisms can be adapted to 30% pulp density^{15,29}. In some papers it was mentioned that the cell wall of extreme thermoacidophiles is not as rigid as that for the mesophilic species, therefore they are demolished when exposed to disturbance at high pulp densities^{30,36}. In another article, the cell wall structure of thermophilic microorganisms was reported to lack peptidoglycan and cell membranes, which are much more flexible than those of mesophilic microorganisms. This means that this type of microorganism can only endure low pulp densities, which is one of the principal problems for its industrial use^{14,34}. These statements are not correct. Firstly, because all cells have cell membrane and secondly, because these very specialized membrane lipids present in thermoacidophiles are critical for survival in extremes of both pH and temperature 16,26.

Some researchers propose that very tiny concentrates may not be necessary to obtain acceptable copper extraction rates and levels 13,32,33 . Similar to these statements, we also take the best result in $90\,\mu m$ particle size in run 20 and 10. Conversely, some researchers believe that the thin particle size of the used ore assists growth, by preparing better accessibility for the microorganisms to the mineral sulfides 27 .

It seems that the attachment of the microorganisms to the surface of the chalcopyrite particles is more important than particle size, because, the highest copper dissolution is achieved when the inoculated microorganisms are able to access the chalcopyrite surface and adhere to it via extracellular polymeric substances (EPS)^{11,36,38}. Similar to Plumb et al., in our result, the greatest number of cells that were observed in each of the flask cultures were planktonic cells²⁷. This issue is one of the reasons we did not achieve greater recovery. Similar to Zeng et al., we recommend, the attached microorganisms on the chacopyrite surface be detected by atomic force microscope (AFM)³⁹.

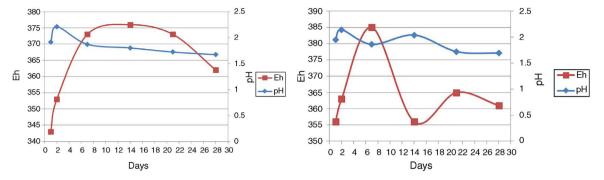


Figure 4 pH and Eh-related curve after bioleaching. (Left): Run 20. (Right): Run23.

Our result shows that the highest recovery was at 160 rpm and the lowest was at 140 rpm. By increasing agitation, recovery also increased and the reason why the 140 rpm had the lowest recovery was due to the successive subculturing of isolates from 160 and 120 rpm. The successive subculturing decreased its ability to bioleach over time. Hence, it is recommended that at each stage of bioleaching previous adapted microbes that are well grown should be used¹⁸. In this survey, the number of microorganisms was high during 28 days with initial inoculums between 2.4×10^7 and 4.8×10^7 cells/ml. Keeling et al. kept their microorganisms alive, without additional nutrients or growth supplements for over 90 days¹⁸. The microorganisms used in this study were also able to remain alive for more than 28 days; however, if bioleaching was too long, there would be no benefit for industrial recovery.

pH and Eh

As seen in Figure 4, run 20 and run 23 both represent the increase and decrease in pH. However, there are more fluctuations in run 23, although run 23 at 14–28 days and run 20 at 2–28 days are associated with reduced pH. As a result of redox potential, run 23 is also associated with more fluctuations, as can be seen in Figure 4.

In the bioleaching experiments, all microorganisms utilized were acidophilic. Hence, maintaining the pH in the correct range was the most important factor for the extractive process to yield copper, iron and other elements^{22,31}. Similar to the study of Zhou et al., an initial pH increase was seen in the early days, then the pH started to decrease, but it was not continuous⁴¹. Our result indicated the lowest copper recovery when pH was below 1.57 and above 2.12 (results not shown). Therefore, at a pH value lower than 1.57, a slight growth was observed and it was the end of the bioleaching process.

In run 20, the redox potential value showed an increase of 353–376 mV from the beginning to the end of the bioleaching process. No correlation was seen between the decrease in the pH value and the increase in redox potential. Our result shows that the increase in redox potential does not reflect an increase in the concentration of Cu and Fe in solution, because the passivation of chalcopyrite occurs at elevated potentials⁴. Furthermore, similar to Sandstrom et al., the redox potential was relatively indifferent to changes in retention time^{20,30}. Therefore, ORP value does not increase over time.

Ferrous and ferric concentrations

As has been shown in Table 2, in run 20 and run 23 the total extracted iron is 8.5% and 11.44% respectively. Initial ferric iron concentrations were low in both runs. Ferric iron concentration in run 20 started to increase from the second day to 21 days later, but decreased from the 21st to 28th days. Moreover, in run 23, the ferric iron concentration increased in 14 days and decreased from the 21st to 28th day. Ferrous iron concentration in run 20 started to increase from the first day to the second day, but decreased from the 2nd to the 21st day. From the 21st to 28th days, the iron concentration increased slightly. In run 23, the ferrous iron concentration also increased from the beginning to day 21st and from the 21st to 28th days the amount significantly decreased (Fig. 5).

Chemical leaching (control)

The pH of control increased gradually until the 21st day and then decreased to 28th days (Fig. 6, left). Changes in soluble ferric and ferrous iron concentration in leachates are shown in Figure 6, right. A mixture of thymol with methanol was added to the control flasks to prevent microbial growth; however, due to evaporation at high temperatures, chalcopyrite was sterilized in the oven at 150 °C for 2 h instead. In chemical leaching with the conversion of ferrous iron to ferric iron, from the 1st to the 14th day, the amount of ferrous iron decreased and the amount of ferric iron increased. From the 14th to 21st day due to the lowering of oxygen and accumulation of ferric iron, the converting process from ferrous iron to ferric iron decreases and results in the increasing of ferrous iron from the 14th to the 21st day. The decrease of ferrous iron and ferric iron from the 21st and 28th day shows the termination of leaching reactions. The high oxidation-reduction potential in chemical leaching relative to bioleaching is due to the higher production of ferric iron (8 g/l) in chemical leaching. In chemical leaching the presence of ferric iron is necessary to trigger the leaching reactions as in bioleaching⁵.

Reactor leaching

Bioleaching of chalcopyrite concentrate by extreme thermoacidophiles was performed at 15% pulp density for 28 days in a stirred tank reactor (Run 20). pH value in the reactor was between 1.74 and 2.17 and Eh value was between 298 and 419. Copper recovery was 32.38% (result not shown). The result of the tested solutions cannot be trusted due to a device error, a sampling error due to water evaporation,

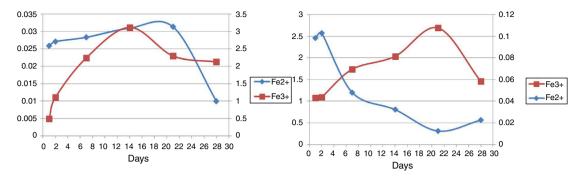


Figure 5 Ferrous and ferric concentrations. (Left): Run 23. (Right): Run20.

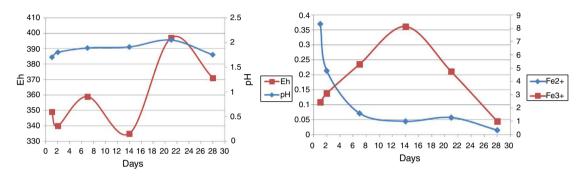


Figure 6 pH and Eh-related curve in chemical leaching (Left). Ferrous and ferric concentrations (Right).

Table 3 X-ray diffraction (XRD) peak for chalcopyrite, pyrite and jarosite.											
Mineral				Pe							
Chalcopyrite	3.04	1.86	1.59	1.87	1.58	2.64	1.21	1.08			
Pyrite	1.63	2.71	2.43	2.21	1.92	3.13	1.45	1.04			

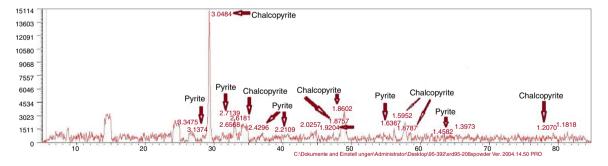


Figure 7 X-ray diffractogram of chalcopyrite concentrate with pyrite mineral.

hence no report was filed. If the tests are not done as soon as the specimens arrive at the laboratory, Fe²⁺ becomes Fe³⁺. Therefore, the residual results are more reliable. As has been shown in Table 3 and Figure 7, the XRD result showed no jarosite formed in the bioleaching experiments.

Conclusion

Based on the results obtained in this research, the following conclusions could be summarized:

It is not surprising that extreme thermoacidophilic microorganisms can be separated from the environments at ambient temperature. Hyperthermophilic enzymes are inactive in an acidic environment and low temperature and these microorganisms are inactive in these environments. Although mineral oxidation is an exothermic process, most of the bioleaching microorganisms grow and activate in a temperature higher than ambient temperature.

As the pH, temperature and aeration system are under control under laboratory condition, the shake flask environment and the reactor are highly homogeneous. However,

the solution metals and metalloids increase and can have a significant effect on the variation and number of indigenous microbial species. It has been argued in some studies that the microbial population is completely archaea at a temperature above 70 °C and consists of species of the order Sulfolobales. Sulfolobus and Acidianus are example of Sulfolobales. It has been observed in some studies that antagonistic reactions between Acidianus sp. (DSMZ 29038) and Sulfolobus metallicus inhibit bioleaching to some extent. The antagonistic effects have been observed during pre-colonization tests. Releasing some antagonistic compounds by S. metallicus has also been observed. It seems that quorum sensing signals and chemotactic responses also have a significant role in bioleaching. We use an unsterilized concentrate for bioleaching in these studies, because the sterilization of chalcopyrite concentrate is not cost effective in industry. It seems that the antagonistic relationship among various species of the order Sulfolobales is another reason for the low recovery of copper in our study.

The survival of the microorganism inside the ore depends on the expression of metal resistance genes. The horizontal transition of such genes in the environment has been well documented. Therefore, the adaptability of indigenous microorganisms with chalcopyrite concentrate in various percentages before bioleaching seems unnecessary. As bioleaching starts, metal concentration in leachate increases and species resistant to the high concentration of metals survive at the end of the bioleaching process. These microorganisms contain resistant and adaptable mechanisms toward metals such as active efflux or trapping of the metal ions by metal chaperones, gene duplications, the presence of genomic islands (GI), passive mechanisms for pH and cations homeostasis in acidic condition, as well as an inorganic polyphosphate (polyP)-driven metal resistance mechanism.

Authors' contribution

All authors had equal role in design, work, statistical analysis, and manuscript writing.

Conflict of interest

The authors declare no conflict of interest.

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References

- Abdollahi H, Noaparast M, Shafaei SZ, Manafi Z, Muñoz JA, Tuovinen OH. Silver-catalyzed bioleaching of copper, molybdenum and rhenium from a chalcopyrite molybdenite concentrate. Int Biodeterior Biodegradation. 2015;104:194–200.
- Astudillo C, Acevedo F. Adaptation of Sulfolobus metallicus to high pulp densities in the biooxidation of a flotation gold concentrate. Hydrometallurgy. 2008;92:11–5.

 Bharadwaj A, Ting Y-P. Bioleaching of spent hydrotreating catalyst by acidophilic thermophile *Acidianus brierleyi*: leaching mechanism and effect of decoking. Bioresour Technol. 2013;130:673–80.

- Cancho L, Blázquez M, Ballester A, González F, Muñoz J. Bioleaching of a chalcopyrite concentrate with moderate thermophilic microorganisms in a continuous reactor system. Hydrometallurgy. 2007;87:100–11.
- Córdoba E, Munoz J, Blázquez M, González F, Ballester A. Leaching of chalcopyrite with ferric ion. Part IV. The role of redox potential in the presence of mesophilic and thermophilic bacteria. Hydrometallurgy. 2008;93:106–15.
- d'Hugues P, Foucher S, Galle-Cavalloni P, Morin D. Continuous bioleaching of chalcopyrite using a novel extremely thermophilic mixed culture. Int J Miner Process. 2002;66:107–19.
- 7. Demergasso CS, Castillo D, Casamayor EO. Molecular characterization of microbial populations in a low-grade copper ore bioleaching test heap. Hydrometallurgy. 2005;80:241–53.
- 8. Ding J, Zhang R, Yu Y, Jin D, Liang C, Yi Y, Zhu W, Xia J. A novel acidophilic, thermophilic iron and sulfur-oxidizing archaeon isolated from a hot spring of Tengchong, Yunnan, China. Braz J Microbiol. 2011;42:514–25.
- Dopson M, Baker-Austin C, Hind A, Bowman JP, Bond PL. Characterization of ferroplasma isolates and *Ferroplasma acidarmanus* sp. nov., extreme acidophiles from acid mine drainage and industrial bioleaching environments. Appl Environ Microbiol. 2004;70:2079–88.
- Escobar B, Godoy I. Enumeration of Acidithiobacillus ferrooxidans adhered to agglomerated ores in bioleaching processes. Appl Environ Microbiol. 2002;18:875–9.
- 11. Gautier V, Escobar B, Vargas T. Cooperative action of attached and planktonic cells during bioleaching of chalcopyrite with *Sulfolobus metallicus* at 70 °C. Hydrometallurgy. 2008;94:121-6.
- **12.** Gericke M, Govender Y, Pinches A. Tank bioleaching of low-grade chalcopyrite concentrates using redox control. Hydrometallurgy. 2010;104:414-9.
- Gericke M, Pinches A, Van Rooyen J. Bioleaching of a chalcopyrite concentrate using an extremely thermophilic culture. Int J Miner Process. 2001;62:243–55.
- **14.** Gomez E, Ballester A, Gonzalez F, Blazquez M. Leaching capacity of a new extremely thermophilic microorganism, *Sulfolobus rivotincti*. Hydrometallurgy. 1999;52:349–66.
- **15.** Guo Z, Zhang L, Cheng Y, Xiao X, Pan F, Jiang K. Effects of pH, pulp density and particle size on solubilization of metals from a Pb/Zn smelting slag using indigenous moderate thermophilic bacteria. Hydrometallurgy. 2010;104:25–31.
- **16.** Jacquemet A, Barbeau J, Lemiegre L, Benvegnu T. Archaeal tetraether bipolar lipids: structures, functions and applications. Biochimie. 2009;91:711–7.
- Johnson DB, Okibe N, Wakeman K, Yajie L. Effect of temperature on the bioleaching of chalcopyrite concentrates containing different concentrations of silver. Hydrometallurgy. 2008;94: 42–7.
- Keeling S, Davies K, Palmer M-L, Townsend D, Watkin E, Johnson J, Watling H. Utilisation of native microbes from a spent chalcocite test heap. Hydrometallurgy. 2006;83: 124-31
- **19.** Konishi Y, Tokushige M, Asai S, Suzuki T. Copper recovery from chalcopyrite concentrate by acidophilic thermophile *Acidianus brierleyi* in batch and continuous-flow stirred tank reactors. Hydrometallurgy. 2001;59:271–82.
- **20.** Liang C-l, Xia J-l, Nie Z-y, Yu S-j, Xu B-q. Effect of initial pH on chalcopyrite oxidation dissolution in the presence of extreme thermophile *Acidianus manzaensis*. Trans Nonferrous Met Soc China. 2014;24:1890–7.
- 21. Liu YG, Zhou M, Zeng GM, Wang X, Li X, Fan T, Xu W-H. Bioleaching of heavy metals from mine tailings by indigenous

- sulfur-oxidizing bacteria: effects of substrate concentration. Bioresour Technol. 2008:99:4124–9.
- 22. Navarro CA, Bernath Dv, Jerez CA. Heavy metal resistance strategies of acidophilic bacteria and their acquisition: importance for biomining and bioremediation. Biol Res. 2013;46:363–71.
- 23. Noori Felegari Z, Nematdoust Haghi B, Amoabediny G, Mousavi S, Amouei Torkmahalleh M. An Optimized Integrated Process for the Bioleaching of a Spent Refinery Processing Catalysts. Int J Environ Res. 2014;8:621-34.
- 24. Norris PR, Burton NP, Clark DA. Mineral sulfide concentrate leaching in high temperature bioreactors. Miner Eng. 2013;48:10-9.
- **25.** Okibe N, Gericke M, Hallberg KB, Johnson DB. Enumeration and characterization of acidophilic microorganisms isolated from a pilot plant stirred-tank bioleaching operation. Appl Environ Microbiol. 2003;69:1936–43.
- **26.** Plumb J, Gibbs B, Stott M, Robertson W, Gibson J, Nichols P, Watling H, Franzmann P. Enrichment and characterisation of thermophilic acidophiles for the bioleaching of mineral sulphides. Miner Eng. 2002;15:787–94.
- Plumb J, McSweeney N, Franzmann P. Growth and activity of pure and mixed bioleaching strains on low grade chalcopyrite ore. Miner Eng. 2008;21:93–9.
- 28. Rawlings DE. Characteristics and adaptability of iron- and sulfur-oxidizing microorganisms used for the recovery of metals from minerals and their concentrates. Microb Cell Fact. 2005;4:13.
- 29. Rubio A, Frutos FG. Bioleaching capacity of an extremely thermophilic culture for chalcopyritic materials. Miner Eng. 2002;15:689–94.
- Sandström Å, Petersson S. Bioleaching of a complex sulphide ore with moderate thermophilic and extreme thermophilic microorganisms. Hydrometallurgy. 1997;46:181–90.
- Sobral LS, De Barros Lima R, Ferreira Leite SG, et al. Bioleaching of copper sulfide flotation concentrate in batch reaction system using mesophile and thermophile microorganisms. DYNA. 2011;79:133–40.
- Valencia P, Acevedo F. Are bioleaching rates determined by the available particle surface area concentration? World J Microbiol Biotechnol. 2009;25:101-6.

- Vuković M, Štrbac N, Sokić M, Grekulović V, Cvetkovski V. Bioleaching of polymetallic sulphide concentrate using thermophilic bacteria. Hem Ind. 2014;68:575–83.
- 34. Wang J, Tao L, Zhao H, Hu M, Zheng X, Peng H, Gan X, Xiao W, Cao P, Qin W. Cooperative effect of chalcopyrite and bornite interactions during bioleaching by mixed moderately thermophilic culture. Miner Eng. 2016;95:116–23.
- Wang Y, Zeng W, Qiu G, Chen X, Zhou H. A moderately thermophilic mixed microbial culture for bioleaching of chalcopyrite concentrate at high pulp density. Appl Environ Microbiol. 2014;80:741–50.
- 36. Wheaton G, Counts J, Mukherjee A, Kruh J, Kelly R. The confluence of heavy metal biooxidation and heavy metal resistance: implications for bioleaching by extreme thermoacidophiles. Minerals. 2015;5:397–451.
- Zeng WM, Wu CB, Zhang RB, Hu PL, Qiu GZ, Gu GH, Zhou HB. Isolation and identification of moderately thermophilic acidophilic iron-oxidizing bacterium and its bioleaching characterization. Trans Nonferrous Met Soc China. 2009;19: 222-7.
- **38.** Zeng W, Qiu G, Zhou H, Peng J, Chen M, Tan SN, Chao W, Liu X, Zhang Y. Community structure and dynamics of the free and attached microorganisms during moderately thermophilic bioleaching of chalcopyrite concentrate. Bioresour Technol. 2010;101:7068–75.
- Zeng W, Tan S, Chen M, Qiu G. Detection and analysis of attached microorganisms on the mineral surface during bioleaching of pure chalcopyrite with moderate thermophiles. Hydrometallurgy. 2011;106:46-50.
- Zhou HB, Zeng WM, Yang ZF, Xie YJ, Qiu GZ. Bioleaching of chalcopyrite concentrate by a moderately thermophilic culture in a stirred tank reactor. Bioresour Technol. 2009;100: 515–20.
- 41. Zhou HB, Zeng WM, Yang ZF, Xie YJ, Qiu GZ. Isolation and characterization of *Ferroplasma thermophilum* sp. nov., a novel extremely acidophilic, moderately thermophilic archaeon and its role in bioleaching of chalcopyrite. J Appl Microbiol. 2008;105:591-601.