

Sporadic dissemination of *mcr-8*-ST11 *Klebsiella pneumoniae* isolates in China



Diseminación esporádica de aislados de *Klebsiella pneumoniae* *mcr-8*-ST11 en China

Dear Editor:

Since the first plasmid-encoded colistin resistance gene (*mcr-1*) was described in *Escherichia coli* in China,¹ 9 others mobile colistin resistance genes have been identified (*mcr-2* to *mcr-10*).² A plasmid carrying *mcr-8* was first reported in 2018 but similar mechanisms of action for *mcr-8* and *mcr-2* indicated that *mcr-8* resistance genes may have appeared very early.² Notably, the *mcr-8* gene has been detected in patients from intensive care units as well as livestock² and *mcr-8*-bearing plasmids have been found primarily in *Klebsiella pneumoniae* and *Raoultella ornithinolytica*. In the current study, an IncFIA plasmid carrying *mcr-8* was identified in an ST11 *K. pneumoniae* strain isolated from wastewater in a duckery in Fujian, China. Further genetic analysis of this plasmid and others of the same type from global databases were then compared.

The *mcr-8*-harboring *K. pneumoniae* isolate ZZW20 was identified by PCR amplification and sequencing of the 16S rRNA. Susceptibility testing was performed following the CLSI³ and the EUCAST⁴ methodologies, and this strain was resistant to colistin (MIC 32 mg/L) and 6 other antimicrobials (Table S1). The whole genome sequence of ZZW20 was determined using a combination of Illumina (San Diego, CA, USA) and MinION (Nanopore, Oxford, UK) platforms and assembled using Unicycler.⁵ MLST analyses indicated that this *mcr-8*-positive *K. pneumoniae* belonged to the most prevalent clonal type (ST11) that carried virulence factors, that was present in Asia, especially China.⁶

A phylogenetic tree of ZZW20 was generated using 44 additional *K. pneumoniae* strains carrying *mcr-8* on plasmids. The 44 *mcr-8*-harboring *K. pneumoniae* strains were primarily from Asia but were distributed within different clades and isolated from humans ($n=17$) and animals ($n=27$), indicating a prevalence in animals. A total of 17 blow flies carrying *mcr-8* were trapped at three different locations in Northern Thailand: a local market in an urban community, a rural area and a suburb of the city Phitsanulok. Importantly, they were grouped in a cluster, suggesting that flies are likely to be one of the key vectors of the *mcr-8*. Compared with other strains carrying *mcr-8*, their ST type was different, indicating that the strains carrying *mcr-8* began to evolve toward diversification. Almost all strains of ST11 from China shared 6588 core-genome single nucleotide polymorphisms (cgSNPs). Interestingly, significantly distinct cgSNPs (range 1868–4216) were observed between ZZW20 and other database strains suggesting that they did not share a common transmission event (Fig. 1A).

The *mcr-8* gene was present in strain ZZW20 on an ~88 kb plasmid (pZZW20.80K; Acc. No. CP058962) that was identified by S1-PFGE and southern blotting⁷ using the appropriate probes (Figure S1), and was typed as IncFIA using the WGS data. BLAST analysis demonstrated that pZZW20.80K had a 129 kb backbone closely related to the *K. pneumoniae* plasmids p18-29mcr-8.2 (MK262711), p2018C01-046-1MCR8 (CP044369), p2018N16-148-2MCR8 (CP044395), p2018N17-066-2MCR8 (CP044391), pVnKp83 (LC549808) and PK91 (MG736312). Interestingly, there were only 44 nucleotides that differed between the plasmids in this group (>99.96% identity). Additionally, even though the 7 IncFIA ST11 *mcr-8*-positive *K. pneumoniae* isolates were unique (range 1868–4216 cgSNPs), the *mcr-8* plasmids they carried were highly similar,

especially for pZZW20.80K with 88 kb identity (81% query coverage and 99.9% identity) to *mcr-8* plasmid p18-29 (MK262711) from a human isolate in China. Then we used these plasmids to make a circular plasmid map with BRIG for further analysis.⁸ These plasmids shared the genetic context IS903B-ampC-hp-hp-hp-Giy-T-dgkA-baeS-copR-IS3-mcr-8-Gly-T-IS5. However, only the ZZW20 carried *bla*_{CTX-M-3} and *bla*_{TEM-1B} genes. There were also 2 copies of Δ Tn2 downstream of *bla*_{TEM-1B} as well as an intact Tn2 in pZZW20.88K indicating that the insertion of *bla*_{CTX-M-3} and *bla*_{TEM-1B} were most likely mediated by Tn2⁹ (Fig. 1B).

The pZZW20.80K plasmid also possessed the *tra/trb* gene cluster that mediates plasmid conjugative functions and we therefore conducted conjugation experiments to examine its potential transferability using *E. coli* EC600^{str} as recipient. The transconjugants that were isolated retained most of the antibiotic resistance genes from the plasmid including colistin resistance (Table S1). This was an additional confirmation that IncFIA plasmids carrying *mcr-8* may spread among *K. pneumoniae* strains.

The 6 virulence factors present on the chromosome of strain ZZW20 were identified using the VFDB database (<http://www.mgc.ac.cn/VFs/main.htm>). The genes necessary for enterobactin synthesis, transport and utilization (*entABCDEF*, *fepABCDG*, *fes*) as well as the gene responsible for salmochelin cleavage (*iroE*) were present on the plasmid. The enterobactin siderophore system is the primary iron-sequestering system for *Enterobacterales*. The presence of salmochelin is associated with invasive disease isolates and is common in highly virulent *K. pneumoniae* isolates.⁶ Strain ZZW20 also carried a Type VI Secretion System (T6SS) cluster that contributes to bacterial competition, cell invasion and *in vivo* colonization that can sensitize hosts to potentially fatal infections by other bacterial pathogens.¹⁰ These characteristics of ZZW20 indicated a likelihood that it was evolving into a hypervirulent strain.

In conclusion, ST11 *K. pneumoniae* strains possessing *mcr-8* IncFIA plasmids have been sporadically disseminated in China. These plasmids were highly similar and pZZW20.80K -*mcr-8* and were able to transfer to *E. coli* by conjugation. The presence of the highly efficient enterobactin system along with 5 other known virulence factors indicated a potential threat to public health. In particular, we found multiple copies of Δ IS66 transposases on the *mcr-8* plasmid revealing the potential transferability of *mcr-8*. Thus, there is an urgent need for further surveillance to understand the prevalence and dissemination of *mcr-8*-positive *K. pneumoniae* and prevent future disease outbreaks.

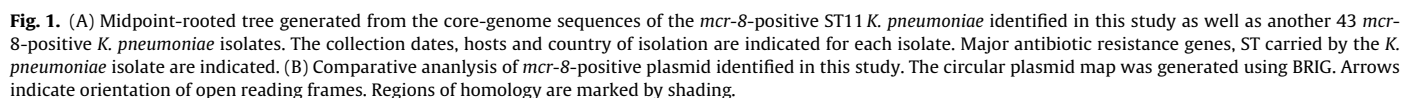
Accession number. Sequence of pZZW20-88K has been assigned GenBank accession number CP058962.

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Appendix A. Supplementary data

Supplementary data associated with this article can be found, in the online version, at [doi:10.1016/j.eimc.2021.01.007](https://doi.org/10.1016/j.eimc.2021.01.007).



References

1. Yi-Yun L, Yang W, Timothy R, Ling-Xian Y, Rong Z. Emergence of plasmid-mediated colistin resistance mechanism MCR-1 in animals and human beings in China: a microbiological and molecular biological study. *Lancet Infect Dis.* 2015;16:161–8, [http://dx.doi.org/10.1016/S1473-3099\(15\)00424-7](http://dx.doi.org/10.1016/S1473-3099(15)00424-7).
2. Ullah S, Ji K, Li J, Xu Y, Jiang C, Zhang H, et al. Characterization of NMCR-2, a new non-mobile colistin resistance enzyme: implications for an MCR-8 ancestor. *Environ Microbiol.* 2020, <http://dx.doi.org/10.1111/1462-2920.15171>.
3. Clinical and Laboratory Standards Institute (CLSI). Document M100. 28th edition; 2018. Wayne, PA, USA.
4. European Committee on Antimicrobial Susceptibility Testing (EUCAST). Breakpoint tables for interpretation of MICs and zone diameters; 2018. Version 8.0. (<http://www.eucast.org>).
5. Wick RR, Judd LM, Gorrie CL, Holt KE. Unicycler: resolving bacterial genome assemblies from short and long sequencing reads. *PLoS Comput Biol.* 2017;13:e1005595, <http://dx.doi.org/10.1371/journal.pcbi.1005595>.
6. Yang X, Wai-Chi Chan E, Zhang R, Chen S. A conjugative plasmid that augments virulence in *Klebsiella pneumoniae*. *Nat Microbiol.* 2019;4:2039–43, <http://dx.doi.org/10.1038/s41564-019-0566-7>.
7. Barton BM, Harding GP, Zuccarelli AJ. A general method for detecting and sizing large plasmids. *Anal Biochem.* 1995;226:235–40, <http://dx.doi.org/10.1006/abio.1995.1220>.
8. Alikhan NF, Petty NK, Ben Zakour NL, Beatson SA. BLAST Ring Image Generator (BRIG): simple prokaryote genome comparisons. *BMC Genomics.* 2011;12:402, <http://dx.doi.org/10.1186/1471-2164-12-402>.
9. Toleman MA, Walsh TR. Combinatorial events of insertion sequences and ICE in Gram-negative bacteria. *FEMS Microbiol Rev.* 2011;35:912–35, <http://dx.doi.org/10.1111/j.1574-6976.2011.00294.x>.
10. Perault AI, Chandler CE, Rasko DA, Ernst RK, Wolfgang MC, Cotter PA. Host adaptation predisposes *Pseudomonas aeruginosa* to type VI secretion system-mediated predation by the *Burkholderia cepacia* complex. *Cell Host Microbe.* 2020;28:534–47000, <http://dx.doi.org/10.1016/j.chom.2020.06.019>.

Shuan-Cheng Bai^{a,b,c,d,1}, Run-Bo Li^{a,b,c,d,1}, Yang-YU^{a,b,c,d}, Xiao-Ping Liao^{a,b,c,d,*}

^a National Risk Assessment Laboratory for Antimicrobial Resistance of Animal Original Bacteria, South China Agricultural University, Guangzhou, China

^b Laboratory of Veterinary Pharmacology, College of Veterinary Medicine, South China Agricultural University, Guangzhou, China

^c Guangdong Laboratory for Lingnan Modern Agriculture, Guangzhou, China

^d Guangdong Provincial Key Laboratory of Veterinary Pharmaceutics Development and Safety Evaluation, South China Agricultural University, Guangzhou, China

* Corresponding author.

E-mail address: xpliao@scau.edu.cn (X.-P. Liao).

¹ These authors contributed equally to this work.

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