

ENDOCRINOLOGÍA Y NUTRICIÓN



www.elsevier.es/endo

ORIGINAL ARTICLE

High-fat diets and body composition over two generations. An experimental study $^{\!\!\!\!\!\!\!\!/}$

Cristina Suarez, Andrea Ferreira Monteiro, Elisa Macri, Ana Chiarenza, Susana Zeni, Patricia Rodríguez, Patricia Boyer, Silvia Friedman*

Cátedra de Bioquímica General y Bucal, Facultad de Odontología, Universidad de Buenos Aires, Buenos Aires, Argentina

Received 29 June 2011; accepted 9 January 2012 Available online 27 May 2012

KEYWORDS

Body composition; Diet; Rat

Abstract

Introduction and objective: Despite recent findings reported on the nutritional factors that induce epigenetic changes, little information is available at early ages. This study analyzed in an experimental model, over two generations, potential changes in body composition and potential expression of epigenetic changes as the result of the intake of isoenergetic diets with different fat levels.

Materials and methods: At weaning, Wistar female rats were divided into two groups that were fed either a control diet (fat = 7%, w/w) or a high-fat diet (15%, w/w). Rats were mated at 70 days (M_1) and their pups (P_1) were the first generation; P_1 rats were mated at 70 days (M_2) and their pups (P_2) represented the second generation. At weaning, mothers and pups (M_1 , M_2 and P_1 , P_2) were measured body weight (W) and composition (% body fat, %BF), and total skeleton bone mineral content (BMC), expressed as %BMC, using chemical and DXA methods, respectively.

Results: At weaning, high-fat diet groups M_2 and P_2 showed significant increases in W and %BF (p < 0.05); increased %BF values were already found in the M_1 and P_1 groups (p < 0.001). By contrast, %BMC significantly decreased in M_2 and P_2 rats (p < 0.001).

Conclusion: This study demonstrates the need to review certain eating habits to avoid perpetuation of unhealthy patterns generation after generation.

© 2011 SEEN. Published by Elsevier España, S.L. All rights reserved.

PALABRAS CLAVE

Composición corporal; Dieta; Rata Dietas ricas en grasa y composición corporal a lo largo de dos generaciones. Estudio experimental

Resumen

Antecedentes y objetivo: A pesar de los últimos avances acerca de los factores nutricionales que inducen modificaciones epigenéticas, la información en edades tempranas es escasa.

E-mail address: friedman@odon.uba.ar (S. Friedman).

[†] Please cite this article as: Suarez C, et al. Dietas ricas en grasa y composición corporal a lo largo de dos generaciones. Estudio experimental. Endocrinol Nutr. 2012;59:232–8.

^{*} Corresponding author.

El presente trabajo estudió en un modelo experimental a lo largo de dos generaciones las posibles modificaciones en la composición corporal, la posible expresión de cambios epigenéticos, y el resultado del consumo de dietas isocalóricas con niveles de grasa diferentes.

Materiales y métodos: Ratas Wistar hembras al destete se dividieron en dos grupos que recibieron una dieta con 7 y 15% de grasa (rica en grasa). A los 70 días se aparearon (M_1) y sus crías (C_1) constituyeron la primera generación; C_1 a los 70 días fueron apareadas (M_2) y sus crías (C_2) constituyeron la segunda generación. Al destete, se evaluaron tanto las madres como las crías (M_1 , M_2 y C_1 , C_2), el peso (P) y composición corporales % de grasa (% Gra), por método químico y contenido mineral óseo de esqueleto total (CMO) por densitometría, expresado como %CMO. *Resultados*: Al destete, en los grupos con dieta rica en grasa M_2 y C_2 (15% Gra) se observó un incremento significativo del P y % Gra (p < 0,05), mientras que el aumento en el % Gra ya se observó en C_1 y M_1 (p < 0,001). Por el contrario, el % CMO de M_2 y C_2 disminuyó significativamente (p < 0,001).

Conclusión: Este estudio pone de manifiesto la potencial necesidad de modificar ciertos hábitos alimentarios que eviten repetir patrones distorsionados de generación en generación.

© 2011 SEEN. Publicado por Elsevier España, S.L. Todos los derechos reservados.

Introduction

There is evidence to suggest that food choices by the mother during pregnancy may lay the foundations for food acceptance in postnatal life. The amniotic fluid surrounding the fetus, which maintains fetal temperature, is a rich source of sensory exposure to the fetus. Many flavors present in maternal diet are also present in amniotic fluid, 1,2 and since taste and smell are already functional during fetal life, they represent the first experiences of contact before birth. Exposure to these "transmissible" flavors influences their postnatal acceptance. 3,4 This early experience, by providing a "flavor bridge" and making the fetus familiar with flavors in maternal diet, plays a key role in food acquisition and preferences. 4,5

After birth, parents participate in the early experiences of children with food. Breast milk, with its flavors, facilitates the transition to adult diet. As omnivorous young humans, they are willing to learn and eat an adult diet and, despite the wide range of food products available to them, they develop the capacity to repeat the dietary pattern of the family at an early age. These food preferences acquired by children reflect the available and accessible food products, and pave the way for the dietary cultural pattern of their own future offspring.

There is evidence that intrauterine metabolic disturbances may influence gene expression and modulate the phenotype during adult life. Exposure in early life to inadequate nutritional factors may be critical for body tissue growth and development, and is a potential risk factor for diseases due to inadequate dietary patterns by default^{6,7} or by excess.⁸ Despite recent advances in the understanding of nutritional factors inducing epigenetic changes, little information is available for early age.⁹

This study examined in an experimental model, over two generations, potential changes in body composition resulting from the use of isocaloric diets with different fat levels.

Materials and methods

Animal population

Ninety Wistar female rats with a mean initial weight of $44.7\pm0.7\,\mathrm{g}$ (mean \pm standard error, SE), from the department of general and oral biochemistry of the stomatology school of Universidad de Buenos Aires, were studied from weaning (22 \pm 1 days of age). The protocol submitted met the internationally agreed specifications for the use and care of laboratory animals. ¹⁰

The animals were placed in galvanized cages with mesh floors and suspended in order to maintain hygienic conditions and avoid coprophagia. A room temperature of $21\pm1\,^{\circ}\text{C}$ was maintained using a thermostatized system, and relative humidity was 50–60%. A system for strictly maintaining light-dark periods of $12\,\text{h}/12\,\text{h}$ was also used, because changes in these periods may cause changes in animal behavior and feeding rhythm. 11

At weaning, the animals were divided into three groups with no differences in baseline mean weights (p > 0.05). A group of 10 animals (C_0) were killed to measure baseline values. Two groups of 40 animals each were fed ad libitum one of two diets throughout the experimental period: a control diet containing 7 g of fat/100 g (7%) and an experimental diet containing 15 g/100 g (15%).

At 28, 35, and 49 days of age, 10 animals from each group were killed to measure body composition. At 70 days of age, 10 rats from both groups were mated, and the pups were weaned at 110 days. The mothers of this first generation ($M_{1/7\%}$ and $M_{1/15\%}$) and a subgroup of pups ($C_{1/7\%}$ and $C_{1/15\%}$) were killed. The remaining subgroup continued with the diet of the previous generation. They were mated at 70 days, and the pups were weaned at 110 days of age. The mothers ($M_{2/7\%}$ and $M_{2/15\%}$) and pups ($C_{2/7\%}$ and $C_{2/15\%}$) were then killed (Fig. 1).

During the experimental period, the animals were fed ad libitum one of the following isocaloric diets (4kcal/g), the composition of which is detailed in Table 1. Drinking water

C. Suarez et al.

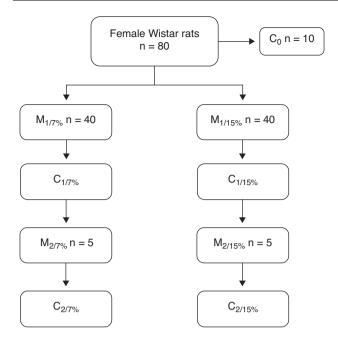


Figure 1 Design of the study procedure. M_1 : first generation mothers; M_2 : second generation mothers. C_0 : baseline group; C_1 : first generation pups; C_2 : second generation pups. Proportions of 7% and 15% indicate contents of 7 g of fat/100 g of diet and 15 g of fat/100 g of diet, respectively.

Table 1 Percent composition of diets.

Control	Experimental
28.7	28.7
7.0	15.0
55.0	37.0
3.5	3.5
1.0	1.0
0.71	0.71
4.1	14.1
	28.7 7.0 55.0 3.5 1.0 0.71

^a Potassium caseinate: 87% protein (Inmobal Nutrer S.A., Argentina).

was provided ad libitum, a continuous supply being ensured because of the close correlation between fluid intake and feeding behavior.¹²

Parameters tested

Body weight: this was measured using a Mettler PC 4000 analytical scale with a precision of $\pm 1\,\text{mg}$. Measurements were made after fasting for at least 2 h and up to 4 h. Weight was recorded in grams.

Body composition: analysis of body composition included body fat content (% fat) and total skeleton bone mineral content (expressed as %BMC (g/100 g body weight)). Body fat content was measured on the dry carcass using a chemical method consisting of intermittent extraction with petroleum ether (Soxhlet, AOAC).¹³ Results were expressed in g of fat/100 g body weight. Total skeleton bone mineral content was assessed by ex vivo densitometry using Lunar DPX equipment (DPX Alpha 8034, Small Animal Software, Lunar Radiation Corp. Madison, USA). Equipment precision was assessed by five measurements in the same rat, with animal replacement during the same and on a different day. The coefficient of variation (CV) for bone mineral content (BMC) was 3%.

Results were expressed as mean ± SE, and were analyzed statistically using ANOVA to compare the groups. When a statistically significant difference was found, a Student-Newman-Keuls was performed. Tests for normality (Wilk-Shapiro) and homoscedasticity (Bartlett) of variances were performed in all cases. Results were considered significant at the 5% level. The software used included GraphPad Prism version 3.0 (GraphPad Software Inc., San Diego, CA, USA) and SPSS version 9.0 (SPSS Inc., Chicago, USA).

Results

An analysis of body weight changes in the groups $M_{1/15\%}$ (Fig. 2a) and $M_{2/15\%}$ (Fig. 2b) over the experimental period showed no significant differences between them and their respective controls at 28, 35, 49, 70, and 90 days of age. At 110 days of age, a significant body weight increase was only seen in the $M_{2/15\%}$ vs the $M_{2/7\%}$ group (p < 0.05).

Fig. 3 shows the body weight of pups from the first $(C_{1/7\%}, C_{1/15\%})$ and second $(C_{2/7\%}, C_{2/15\%})$ generations at weaning. A significant body weight increase was seen in the second pup generation $(C_{2/15\%})$ born to mothers $(M_{2/15\%})$ who received a diet containing 15% of fat. No significant body weight changes were seen in pups fed the control diet over the generations.

Body fat content was significantly greater in $M_{1/15\%}$ and $M_{2/15\%}$ as compared to their respective controls at 110 days of age (p < 0.001) (Fig. 4).

An analysis of body fat in pups from the first and second generations at weaning showed significantly higher values in $C_{1/15\%}$ and $C_{2/15\%}$ born to mothers ($M_{1/15\%}$ and $M_{2/15\%}$) that received a diet containing 15% of fat. No significant body weight changes were seen in pups fed the control diet over the generations (p > 0.05) as compared to C_0 (7.66 \pm 1.42%) (Fig. 5).

Figs. 6 and 7 show changes over time in total skeleton bone mineral content. While no significant changes were seen in $M_{1/15\%}$ vs $M_{1/7\%}$ and between their respective pups ($C_{1/15\%}$ vs $C_{1/7\%}$), BMC significantly decreased in $M_{2/15\%}$ vs $M_{2/7\%}$ and $C_{2/15\%}$ vs $C_{2/7\%}$ (p < 0.001).

Discussion

In prior studies, our work group analyzed the effect of isocaloric diets with different fat and carbohydrate levels on the body composition of growing rats over one generation. However, the effect of such diets, rich in n-6 polyunsaturated fatty acids (n-6 PUFAs), and potential changes in body composition over two generations have not been assessed yet. There is evidence to show that maternal exposure to high cholesterol and fat diets causes epigenetic changes that

b Salt mix: AIN-93G Mineral Mix. AIN-93 (ICN Biomedicals Inc., Ohio, USA) (AIN-93 Purified Diets for Laboratory Rodents for growth, pregnancy and lactational phases).

^c Vitamin mix: AIN-93-VX (ICN Biomedicals Inc., Ohio, USA) (AIN-93 Purified Diets for Laboratory Rodents).

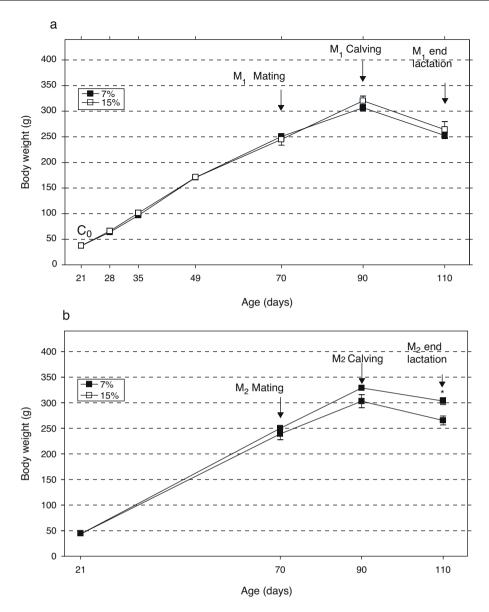


Figure 2 Change in body weight (mean \pm body weight) with age over two generations (1a: first generation; 1b: second generation) in animals fed diets containing 7% (control group) or 15% (experimental group) of fat. C_0 : baseline group; M_1 : first generation mothers; M_2 : second generation mothers. *p < 0.05 between control and experimental groups.

modulate gene expression leading to the development of obesity in the offspring. 15

When the body weights of each of the generations were compared, higher values were always found in groups fed diets with higher lipid contents, both in mothers and pups. ¹⁶ Weight increase was associated with an increased body fat percentage. This may specifically be due to an imbalance in the n-6/n-3 ratio favoring n-6 PUFAs and their metabolites. These include arachidonic acid, which acts as a potent adipogenic agent precursor of prostacyclins, increasing the risk of excessive adipose tissue development. ¹⁷ Other authors found increases in inguinal white adipose tissue and adipocyte size already in the first generation, ¹⁸ in agreement with our results, which show an impact in the first generation, which is increased in the second generation.

An imbalance in the n-6/n-3 ratio and a decreased presence of PUFAs in corn oil may account for the increased body weight and fat in animals on an experimental diet. These results agree with those reported by other authors who found that an increased maternal intake of n-3 PUFA leads to an adipose tissue mass reduction in rats¹⁸ and mice.¹⁹ It is known that the presence of n-6 PUFAs in adequate amounts has benefits on bone activity²⁰ and growth cartilage formation.²¹

In this study, the use of a diet rich in n-6 PUFAs was shown to decrease %BMC, which was inversely related to body fat in both mothers and second generation pups. This reduction in %BMC may have been due to an increased production of arachidonic acid, leading to increased prostaglandin E_2 levels in bone and decreased serum levels of insulin-like growth factor-1, 22,23 with a resultant reduction in the bone

C. Suarez et al.

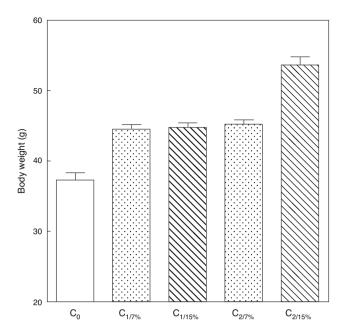


Figure 3 Body weight of pups from two generations of rats fed diets containing 7% (control group) or 15% (experimental group) of fat. C_0 : baseline group; C_1 : first generation pups; C_2 : second generation pups. Mean \pm standard error. ANOVA, overall significance level <0.001; a posteriori, a Student–Newman–Keuls test found the following differences between the groups (different letters indicate significant differences between the groups): C_0 vs $C_1/7\%$ vs $C_1/15\%$; p < 0.05; C_0 vs $C_2/7\%$; p < 0.01; C_0 vs $C_2/15\%$; p < 0.001; $C_1/7\%$ vs $C_2/7\%$; p > 0.05; C_0 vs $C_2/15\%$; p < 0.001; $C_1/7\%$ vs $C_2/7\%$; p < 0.05; $C_1/15\%$ vs $C_2/15\%$; p < 0.001; $C_2/7\%$ vs $C_2/15\%$; p < 0.001.

formation rate. 24 By contrast, a diet rich in n-3 PUFAs promotes bone formation by maintaining adequate arachidonic acid levels in bone. 23

Increased lipid dietary content, associated with increased body fat, was not reflected in the %BMC content

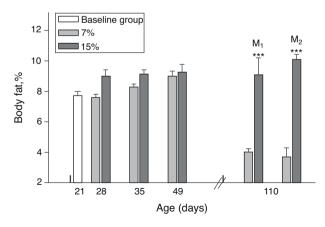


Figure 4 Body fat (mean \pm standard error) as a function of age in animals fed diets containing 7% (control group) and 15% (experimental group) of fat. C_0 : baseline group; M_1 : first generation mothers; M_2 : second generation mothers. t test between groups for each experimental time. Differences between the indicated groups: *p < 0.05; ***p < 0.001.

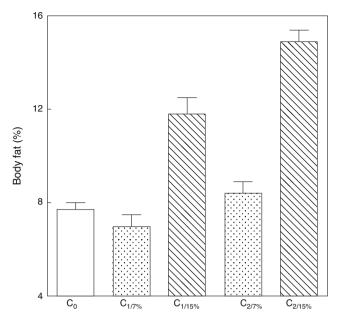


Figure 5 Body weight of pups from two generations of rats fed diets containing 7% (control group) or 15% (experimental group) of fat. C_0 : baseline group; C_1 : first generation pups; C_2 : second generation pups. Mean \pm standard error. ANOVA, overall significance level <0.001; a posteriori, a Student-Newman-Keuls test found the following differences between the groups (different letters indicate significant differences between the groups): C_0 vs $C_1/7\%$ vs $C_2/7\%$; p > 0.05; C_0 , $C_1/7\%$, $C_2/7\%$ vs $C_1/15\%$; p < 0.001; C_0 , $C_1/7\%$, $C_2/7\%$ vs $C_2/15\%$; p < 0.001.

in first generation mothers and pups; in the mothers, this may have been due to bone mass preservation at the expense of a decreased bone turnover rate.²⁵

Studies in experimental animals have shown that exposure to fat-rich diets during pregnancy and lactation predisposes pups to develop a phenotype similar to

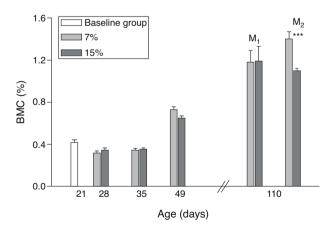


Figure 6 Bone mineral content (%) in total skeleton (mean \pm standard error) as a function of age in animals fed diets containing 7% (control group) and 15% (experimental group) of fat. C_0 : baseline group; M_1 : first generation mothers; M_2 : second generation mothers. t test between groups for each experimental time: *p < 0.05; ***p < 0.001.

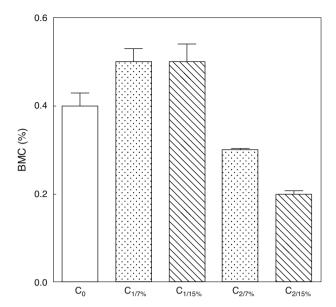


Figure 7 Bone mineral content in total skeleton of pups from two generations of rats fed diets containing 7% (control group) or 15% (experimental group) of fat. C_0 : baseline group; C_1 : first generation pups; C_2 : second generation pups. Mean \pm standard error. ANOVA, overall significance level <0.001; a posteriori, a Student–Newman–Keuls test found the following differences between the groups (different letters indicate significant differences between the groups): C_0 vs $C_1/7\%$ vs $C_1/15\%$; p > 0.05; C_0 , $C_1/7\%$, $C_1/15\%$ vs $C_2/15\%$; p < 0.001; C_0 , $C_1/7\%$, $C_1/15\%$ vs $C_2/15\%$; p < 0.001; C_0 , $C_1/7\%$, $C_1/15\%$ vs $C_2/15\%$; p < 0.001.

metabolic syndrome and to endothelial and cardiovascular dysfunction, among other conditions. Body composition changes may induce the development of conditions changes may induce the development of conditions changes may induce the development of conditions and autoimmune diseases, and cancer, and inflammatory and autoimmune diseases, and bone diseases. Small changes in dietary habits such as the simple addition of α -linolenic acid would allow for fat mass reduction by the suppression of lipogenesis. 37,38

Bearing in mind the significance of cultural patterns in the quality of life of individuals and of the fact that corn oil is a significant component of the Argentinian diet,³⁹ this study shows the potential need for changing some dietary habits to avoid repeating distorted patterns generation after generation.

Conflicts of interest

The authors state that they have no conflicts of interest.

Acknowledgements

Authors thank Mr. Ricardo Orzuza, a technician from Bioterio, for animal care and for his technical support. This study was funded by UBACyT 20020100100613 and C 002.

References

- Hauser GJ, Chitayat D, Berns L, Braver D, Muhlbauer B. Peculiar odours in newborns and maternal pernatal ingestion of spicy foods. Eur J Pediatr. 1985;144:403.
- Mennella JA, Johnson A, Beauchamp GK. Garlic ingestion by pregnant women alters the odor of amniotic fluid. Chem Senses. 1995;20:207-9.
- Schaal B, Marlier L, Soussignan R. Human foetuses learn odours from their pregnant mother's diet. Chem Senses. 2000;25:729-37.
- Mennella JA, Coren P, Jagnow MS, Beauchamp GK. Prenatal and postnatal flavor learning by human infants. Pediatrics. 2001;107:88-94.
- Savage JS, Fisher JO, Birch LL. Parental influence on eating behavior: conception to adolescence. J Law Med Ethics. 2007;35:22-34.
- Lifshitz F. Children on adult diet. Bibl Nutr Dieta. 1996;53:55–9 [review].
- Friedman SM, Rodriguez PN, Boyer PM, Lifshitz F. Decreased energy expenditure—an adaptive mechanism of nutritional growth retardation. Nutr Res. 2006;26:345–9.
- 8. Rising R, Lifshitz F. Relationship between maternal obesity and infant feeding-interactions. Nutr J. 2005;4:17.
- Canani RB, Di Costanzo M, Leone L, Bedogni G, Brambilla P, Cianfarani S, et al. Epigenetic mechanisms elicited by nutrition in early life. Nutr Res Rev. 2011;1:8, doi:10.1017/S09544224110001020.
- 10. National Institutes of Health Laboratory Animal Welfare. Public Health Service policy on humane care and use of laboratory animals by awardees institutions; 1985, revised 1990. NIH "Guidelines for the care and use of laboratory animals".
- NRC. National Research Council. Committee on the guide for laboratory animals facilities and care. Guide for laboratory facilities and care. Washington, DC: Department of Health, Education and Welfare. National Academy of Sciences. US Government Printing Office; 1965.
- Mercer P, Bukhardt SS. The relationship between food intake, weight gain and dietary nutrient concentration in the rat. Nutr Rep Int. 1982;26:121–9.
- 13. AOAC. Official methods for analysis. 15th ed. Washington, DC: US Government Printing Office; 1990.
- 14. Gamba C, Friedman S, Rodriguez P, Macri E, Vacas M, Lifshitz F. Metabolic status in growing rats fed isocaloric diets with increased carbohydrate-to-fat ratio. Nutrition. 2005;21:249-54.
- 15. Buckley AJ, Keseru B, Briody J. Altered body composition and metabolism in the male offspring of high fat-fed rats. Metabolism. 2005;54:500-7.
- Korotkova M, Gabrielsson B, Holmäng A, Larsson B, Hanson L, Strandvik B. Gender-related long-term effects in adult rats by perinatal dietary ratio of n-6/n-3 fatty acids. Am J Physiol Regul Integr Comp Physiol. 2005;288:R575-9.
- Ailhaud G, Guesnet P, Cunnane S. An emerging risk factor for obesity: does disequilibrium of polyunsaturated fatty acid metabolism contribute to excessive adipose tissue development. Br J Nutr. 2008;100:461–70.
- Kortkova M, Gabrielsson B, Lönn M, Hanson L, Strandvik B. Leptin levels in rat offspring are modified by the ratio of linoleic to alpha-linolenic acid in the maternal diet. J Lipid Res. 2002;43:1743-9.
- Massiéra F, Saint-Marc P, Seydoux J, Murata T, Kobayashi T, Narumiya S, et al. Arachidonic acid and prostacyclin signaling promote adipose tissue development: a human health concern. J Lipid Res. 2003;44:271–9.
- 20. Corwin R. Effects of dietary fats on bone health in advanced age. Prostaglandins Leukot Essent Fatty Acids. 2003;68:379–86.

238 C. Suarez et al.

 Xu H, Watkins BA, Adkisson HD. Dietary lipids modify the fatty acid composition of cartilage, isolated chondrocytes and matrix vesicles. Lipids. 1994;29:619–25.

- 22. Watkins BA, Shen CL, Allen KG, Seifert MF. Dietary (n-3) and (n-6) polyunsaturated and acetylsalicylic acid alter ex vivo PGE2 biosynthesis, tissue IGF-I levels, and bone morphometry in chicks. J Bone Miner Res. 1996;11:1321–32.
- Watkins B, Li Y, Lippman H, Seifert M. Omega-3 polyunsaturated fatty acids and skeletal health. Exp Biol Med Vol. 2001;226:485–97.
- 24. Watkins BA, Li Y, Allen KG, Hoffmann WE, Seifert MF. Ratio of (n-6)/(n-3) polyunsaturated fatty acids alters the fatty acid composition of bone compartments and biomarkers of bone formation in rats. J Nutr. 2000;130:2274–84.
- 25. Zeni SN, Weisstaub S, Di Gregorio S, Ronaine de Ferrer P, De Portela ML. Bone mass changes in vivo during the entire reproductive cycle in rats feeding different dietary calcium and calcium/phosphorous ratio content. Calcif Tissue Int. 2003;73:594-600.
- 26. Simopoulos A. The importance of the ratio of omega-6/omega-3 essential fatty acids. Biomed Pharmacother. 2002;56:365–79.
- Hibbeln J, Nieven L, Blasbag T, Riggs J, Lanas W. Healthy intakes of n-3 and n-6 fatty acids: estimations considering worldwide diversity. Am J Clin Nutr. 2006;83:14835–935.
- 28. Wang C, Chung M, Lichtenstein A. Effects of omega-3 fatty acids on cardiovascular disease. Evid Rep Technol Assess. 2004;94:1–8.
- 29. Ramsden CE, Hibbeln JR, Majchrzak SF, Davis JM. n-6 Fatty acidspecific and mixed polyunsaturated dietary interventions have different effects on CHD risk: a meta-analysis of randomized controlled trials. Br J Nutr. 2010;104:1586-600.
- Zeghichi-Hamri S, De Lorgeril M, Salen P, Chibane M, De Leiris J, Boucher F, et al. Protective effect of dietary n-3 polyunsaturated fatty acids on myocardial resistance to ischemia-reperfusion injury in rats. Nutr Res. 2010;30: 849-57.

- 31. Lou YR, Peng QY, Li T, Medvecky CM, Lin Y, Shih WJ, et al. Effects of high fat diets rich in either omega-3 or omega-6 fatty acids on UVB-induced skin carcinogenesis in SKH-1 mice. Carcinogenesis. 2011;32:1078–84.
- 32. Williams CD, Whitley BM, Hoyo C, Grant DJ, Iraggi JD, Newman KA, et al. A high ratio of dietary n-6/n-3 polyunsaturated fatty acids is associated with increased risk of prostate cancer. Nutr Res. 2011;31:1–8.
- 33. Murff HJ, Shu XO, Li H, Yang G, Wu X, Cai H, et al. Dietary polyunsaturated fatty acids and breast cancer risk in Chinese women: a prospective cohort study. Int J Cancer. 2011;128:1434–41.
- 34. MacLean CH, Mojica WA, Morton SC, Pencharz J, Hasenfeld Garland R, Tu W, et al. Effects of omega-3 fatty acids on lipids and glycemic control in type II diabetes and the metabolic syndrome and on inflammatory bowel disease, rheumatoid arthritis, renal disease, systemic lupus erythematosus, and osteoporosis. Evid Rep Technol Assess. 2004;89:1–4.
- 35. Hou JK, Abraham B, El-Serag H. Dietary intake and risk of developing inflammatory bowel disease: a systematic review of the literature. Am J Gastroenterol. 2011;106:563–73.
- 36. Maggio M, Artoni A, Lauretani F, Borghi L, Nouvenne A, Valenti G, et al. The impact of omega-3 fatty acids on osteoporosis. Curr Pharm Des. 2009;15:4157-64.
- 37. Willumsen B, Hexeberg S, Skorve J, Lundquist M, Berge R. Docosahexaenoic acid shows no triglyceride-lowering effects but increases the peroxisomal fatty acid oxidation in liver of rats. J Lipid Res. 1993:34:13–22.
- 38. Ukropec J, Reseland J, Gasperikova D, Demcakova E, Madsen L, Berge R, et al. The hypotriglyceridemic effect of dietary n-3 FA is associated with increased β-oxidation and reduced leptin expression. Lipids. 2003;38:1023-9.
- 39. Friedman S, Rodriguez P, Portela ML, Riba Sicart M, Almajano Pablos MP. Estudio comparativo entre los hábitos alimentarios de estudiantes universitarios argentinos y catalanes: aspectos nutricionales. Rev Esp Nutr Comunitaria. 2008;14:210–8.