

## CLINICAL SCIENCE

HUMAN SAPHENOUS VEIN ORGAN CULTURE  
UNDER CONTROLLED HEMODYNAMIC CONDITIONS

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**INTRODUCTION:** Saphenous vein grafting is still widely used to revascularize ischemic myocardium. The effectiveness of this procedure is limited by neointima formation and accelerated atherosclerosis, which frequently leads to graft occlusion. A better understanding of this process is important to clarify the mechanisms of vein graft disease and to aid in the formulation of strategies for prevention and/or therapeutics.

**OBJECTIVE:** To develop an *ex vivo* flow system that allows for controlled hemodynamics in order to mimic arterial and venous conditions.

**METHODS:** Human saphenous veins were cultured either under venous (flow: 5 ml/min) or arterial hemodynamic conditions (flow: 50 ml/min, pressure: 80 mmHg) for 1-, 2- and 4-day periods. Cell viability, cell density and apoptosis were compared before and after these intervals using MTT, Hoechst 33258 stain, and TUNEL assays, respectively.

**RESULTS:** Fresh excised tissue segments were well preserved prior to the study. Hoechst 33258 and MTT stains showed progressive losses in cell density and cell viability in veins cultured under arterial hemodynamic conditions from 1 to 4 days, while no alterations were observed in veins cultured under venous conditions. Although the cell density from 1-day cultured veins under arterial conditions was similar to that of freshly excised veins, the TUNEL assay indicated that most of these cells were undergoing apoptosis.

**CONCLUSION:** The results observed resemble the events taking place during early *in vivo* arterial-vein grafting and provide evidence that an *ex vivo* perfusion system may be useful for the identification of new therapeutic targets that ameliorate vein graft remodeling and increase graft patency over time.

**KEYWORDS:** Saphenous vein graft; *Ex vivo* organ culture; Vascular biology.

## INTRODUCTION

Coronary artery bypass graft (CABG) is the standard procedure for the revascularization of ischemic heart tissue. Saphenous vein grafts were the first vascular conduits used in CABG, but their effectiveness has been limited by accelerated atherosclerosis that develops within the vein conduit.<sup>1,2</sup> In contrast, arterial conduits display better long-term patency and have been increasingly used instead

of venous grafts. Despite the increased usage of arterial conduits, full arterial revascularizations are carried out in only a small minority of eligible patients; currently, CABG surgery utilizes a combination of arterial and venous grafts.<sup>3,4</sup> However, a better understanding of the molecular events associated with the early arterialization of vein grafts may give rise to alternative approaches to overcoming the poor long-term patency of saphenous vein grafts.

*In vivo*, vein grafts are immediately subjected to increased hemodynamic stress, and the vein's adaptive response may predispose later bypass occlusion. The morphological changes associated with the arterialization of veins have been well characterized, and several *in vivo* and *in vitro* studies have been performed to identify the molecular events taking place when stretch and shear

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stresses are increased.<sup>5-10</sup> *Ex vivo* organ culture has also been used in vascular biology studies; most of the protocols make use of vessel rings that may be maintained up to 8 days in culture media without tissue degeneration.<sup>11,12</sup> More recently, perfusion organ culture systems have been developed for the study of vessels under well-controlled hemodynamic conditions. Arteries kept in organ culture systems have been shown to be viable and with active vasomotion tone for up to 7 days.<sup>13,14</sup> In contrast, vein culture studies have been limited to no more than 24 hours of culture.<sup>15,16</sup>

In the present investigation, we developed an *ex vivo* perfusion system, which mimics *in vivo* hemodynamic conditions, allowing for the study of viable vein grafts under venous or arterial hemodynamic conditions for up to 4 days. We demonstrated that this *ex vivo* system reproduces early events observed in *in vivo* animal models during the arterialization of vein grafts.

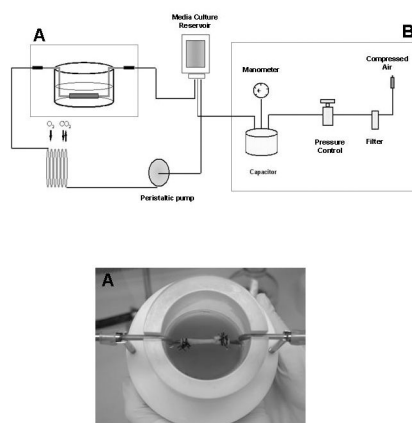
## MATERIALS AND METHODS

### Saphenous vein preparation

Saphenous veins were obtained from patients undergoing aortocoronary bypass surgery at the Heart Institute (InCor), University of São Paulo Medical School. The vein segments were kept in physiological solution until mounting in the culture system. This study protocol was approved by the local ethical committee (SDC – 1834/01/22, CAPPesq – 299/01).

### Organ culture system

The organ culture system is schematically represented in Figure 1. The vessel chamber was connected to a perfusion



**Figure 1** - *Ex vivo* perfusion system. Schematic representation of the system used for organ culture. It consists of an organ bath (A), a peristaltic pump station and a pressure apparatus controller (B)

system where the media from the reservoir was pumped at a set flow rate. This circuit was adapted from the CELLMAX Artificial Capillary System (Spectrum Laboratories, Inc. Rancho Dominguez, CA) according to previously described methods.<sup>13</sup> Vessel chambers and a pressure device were developed in-house to properly place the vessel segments and to enable pressure and flow to be controlled independently. The flow circuit was maintained in a humidified CO<sub>2</sub> incubator at 37°C.

Saphenous vein segments (approximately 2 cm each) were placed in the chamber filled with Dulbecco's modified Eagle's medium containing 10% fetal bovine serum, 100 U/ml penicillin, and 100 µg/ml streptomycin. Vein segments from the same patient were connected to the perfusion system and cultured either under venous conditions (flow: 5 ml/min) or arterial conditions (flow: 50 ml/min, pressure: 80 mmHg) for 1, 2, or 4 days. Fresh, non-mounted segments were also collected for analysis.

All culture solutions were purchased from GIBCO™ (Invitrogen Corporation, Carlsbad, CA).

### Histological analysis

Tissue samples were fixed in 4% phosphate-buffered formalin and embedded in paraplast (Oxford, St. Louis, MO). Sections of 3-µm thickness were stained with haematoxylin-eosin (HE) and Verhoeff's-Van Gieson (VVG) for elastin. The sections were also used for Hoechst staining and TUNEL assays as described below.

### Hoechst staining

Cell nuclei were labeled with Hoechst 33258 (Sigma Chemical Co., St. Louis, MO) as described previously.<sup>17</sup> Briefly, tissue sections were deparaffinized in citrisolv (Fisher Scientific Company, Pittsburgh, PA), rehydrated in serial alcohol dilutions, and treated with 0.5% Triton X-100 for 15 min at 20 °C. The staining reaction was performed using Hoechst 33258 20 µg/ml diluted in assay buffer (mmol/L): NaCl 137, KCl 5, Na<sub>2</sub>HPO<sub>4</sub> 0.4, NaHCO<sub>3</sub> 4, glucose 5.5, MgCl<sub>2</sub> 2, EGTA 2, PIPES, pH 6.1. The sections were visualized under fluorescence microscopy (Carl Zeiss Inc., Thornwood, NY) and quantification was performed by an observer who was blinded to the hemodynamic conditions and duration of the cultures. Each section was measured at 10 different randomly selected points and positive nuclei stains were quantified. Tissue area was quantified using Leica Qwin software (version 2.2) calibrated as per the manufacturer's instructions (40X magnification, 1 pixel = 0.228 µm). The cellular density was expressed as the total cell number per vessel area.

### Terminal transferase-mediated dUTP nick end labeling (TUNEL) assay

DNA fragmentation was detected using the In Situ Cell Death Detection kit, AP (Roche Molecular Biochemicals, Mannheim, Germany) according to the manufacturer's instructions. Briefly, tissue sections were deparaffinized in citrisolv, rehydrated in serial alcohol dilutions, and permeabilized with 0.5% Triton X-100 in 0.1% sodium citrate. The reaction with terminal deoxynucleotidyl transferase and alkaline phosphatase conversion was performed and the cross-sections were examined by light microscopy. Quantification was performed using Leica QWin software (version 2.2). TUNEL-positive cells and total cells were quantified by an observer who was blinded to the hemodynamic conditions and the duration of culture. The results were expressed as a percentage of TUNEL-positive cells among all cells evaluated.

### MTT stain

To verify tissue viability, the vein segments were stained with methylthiazol tetrazolium (MTT - Sigma Chemical Co., St. Louis, MO) as described elsewhere.<sup>14</sup> When reduced by active mitochondria and other cellular enzymes, the MTT dye gives rise to a dark purple precipitate only in viable areas within the tissue fragments, making them readily visible. Briefly, the vein was incubated at 37°C for 1 hour with MTT 0.5 mg/ml in PBS solution; then, pictures were taken from the samples submitted to both hemodynamic conditions.

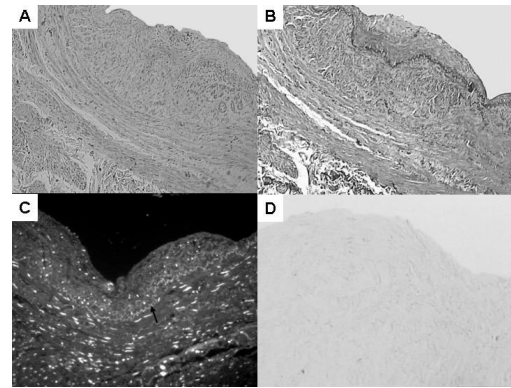
### Statistical Analyses

Data analyses were performed by two-way ANOVA, followed by Tukey's post hoc test, as appropriate. Values for  $p < 0.05$  were considered statistically significant.

### RESULTS

The human saphenous vein segments were cultured under venous and arterial hemodynamic conditions for 1, 2 and 4 days. Before and after these periods, analysis of cell viability, cell density, and apoptosis were performed. In the fresh excised segment of human saphenous vein, tissue morphology and integrity were verified by HE and VVG staining, and also by Hoechst 33258 nuclei stain and TUNEL assay (Figure 2). Furthermore, there was no evidence for apoptotic process indicating that the vessels used were well preserved (Figure 2).

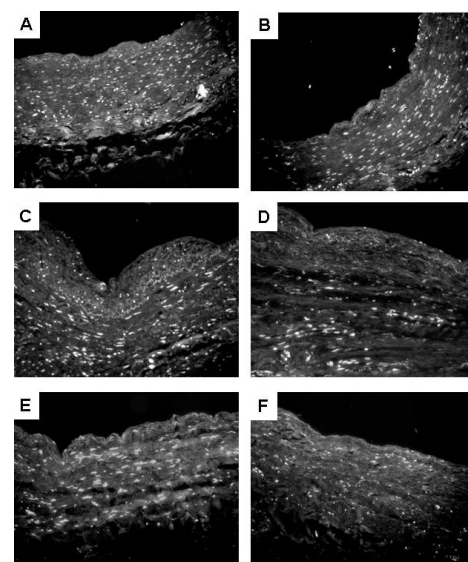
The HE staining was also performed in vein segments cultured under venous and arterial conditions, and the



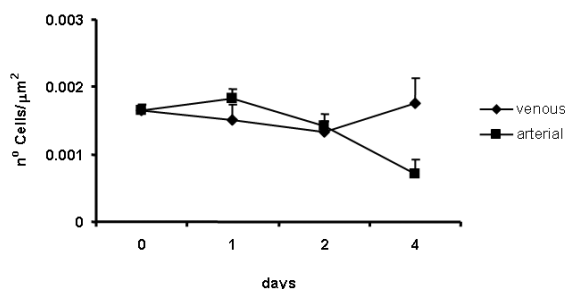
**Figure 2** - Morphological and integrity analyses of fresh excised human saphenous vein. HE stain (A) and VVG stain (B) demonstrating the tissue structure. Hoechst 33258-labeled cell nuclei (C) and TUNEL assay (D) showing tissue cellular density with no presence of apoptotic processes. 100X magnification

number of nuclei stained by hematoxylin gradually diminished from 1 to 4 days when cultured in the arterial condition. No alteration was observed in veins cultured under venous conditions (data not shown). To further confirm the HE staining results, Hoechst 33258 nuclei staining was used to verify cell density in the vessel segments cultured under venous and arterial stimuli. As noted before, cellular density tended to decrease in specimens cultured under arterial conditions for 4 days, whereas no modification was observed in the veins cultured under venous conditions (Figures 3 and 4).

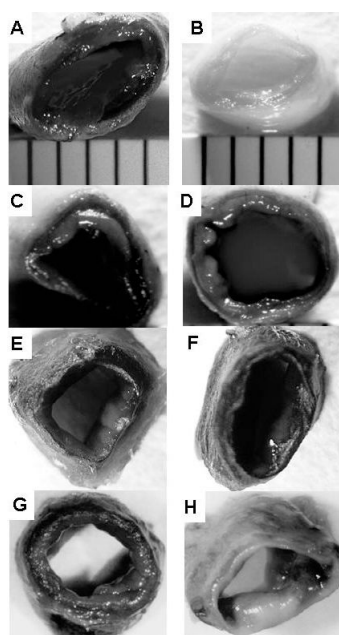
Tissue viability of the segments was examined by MTT staining (Figure 5). A control experiment was performed



**Figure 3** - Hoechst 33258 nuclei stain of human saphenous vein cultured in the *ex vivo* perfusion system. The segments were cultured under venous (A, C, E) and arterial (B, D, F) hemodynamic conditions for 1 (A, B), 2 (C, D) or 4 (E, F) days. 100X magnification



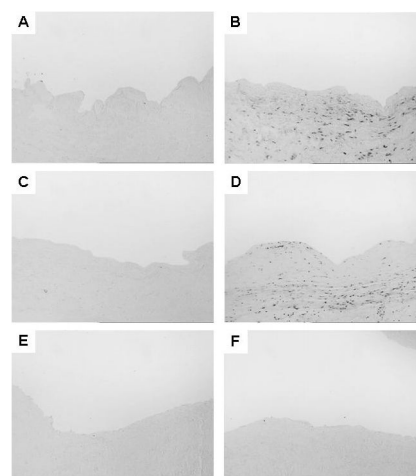
**Figure 4** - Analysis of cellular density of human saphenous vein cultured in the *ex vivo* perfusion system. The results are expressed as mean  $\pm$  standard error of total cell number per vessel area of 3-5 samples stained with Hoechst 33258. (♦) represents human saphenous vein cultured under venous conditions and (■) under arterial conditions



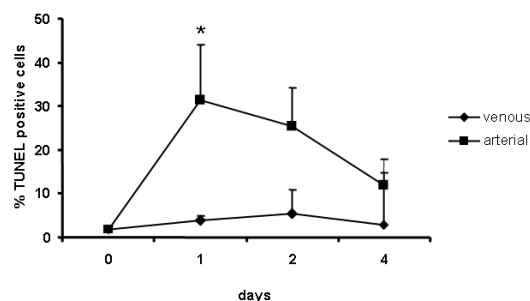
**Figure 5** - MTT stain of human saphenous vein cultured in the *ex vivo* perfusion system. The segments were cultured under venous (C, E, G) and arterial (D, F, H) hemodynamic conditions for 1 (C, D), 2 (E, F) or 4 (G, H) days. The freshly excised tissue (A) and dead samples (B) were used as positive and negative controls for the visualization of tissue viability. 1.5X magnification

using a freshly isolated saphenous segment and a dead, refrigerated vessel. Since MTT stains only living cells, the fresh cross-section was completely stained, while no staining was observed in the dead vein segment (Figures 5A and 5B). The segments cultured under venous conditions from 1 to 4 days contained viable cells throughout the vessel wall, but few viable cells were verified in segments cultured under arterial conditions for 4 days (Figure 5H). To examine if the decreased cell density and viability were due to apoptotic events, a TUNEL assay was performed to detect *in situ* DNA fragmentation (Figures 6 and 7). Almost no apoptotic cells were observed in the saphenous vein cultured under venous

conditions for up to 4 days. In contrast, cultured segments maintained for 1 and 2 days under arterial hemodynamic stimuli showed portions with several TUNEL-positive cells (Figures 6B and 6D), suggesting that the cellular losses observed in the segments cultured for 4 days may have been a result of apoptosis.



**Figure 6** - TUNEL assay of human saphenous vein cultured in the *ex vivo* perfusion system. The segments were cultured under venous (A, C, E) and arterial (B, D, F) hemodynamic conditions for 1 (A, B), 2 (C, D) or 4 (E, F) days. 100X magnification



**Figure 7** - Analysis of apoptosis in human saphenous vein cultured in the *ex vivo* perfusion system. The results are expressed as mean  $\pm$  standard error of percentage of TUNEL-positive cells of 3-5 samples. (♦) represents human saphenous vein cultured under venous conditions and (■) under arterial conditions. \* indicates  $p < 0.05$  vs. venous condition

## DISCUSSION

In this work, we present a perfusion organ culture system developed to mimic the arterialization of human saphenous vein grafts. This *ex vivo* culture system reproduces the initial events taking place when the grafted vein is submitted to arterial hemodynamic conditions and may be a valuable approach to the identification of molecular mechanisms underlying the early stages of bypass grafting, which have been validated in our lab using an *in situ* rat model (data not shown).

Several *in vivo* studies have demonstrated that cell apoptosis is induced during the initial period of arterial-vein graft due to increased tensile stress and strain.<sup>5,6,18,19</sup> In our system, the cellular density and tissue viability decreased gradually in the human saphenous vein cultured under arterial conditions, whereas no changes in morphology or viability were observed when the veins were cultured up to 4 days under venous hemodynamic conditions (Figures 3 and 5). It is interesting to note that the cell density in veins cultured for 1 day under arterial conditions was similar to that in freshly excised samples, but a substantial number of these cells were TUNEL-positive, indicating the presence of apoptotic processes (Figures 3 and 6), which may explain the cellular losses observed in saphenous veins cultured under arterial hemodynamic conditions. It has been described that *in vivo* models of increased tensile stress induce cell death in vein grafts during the first 24 hours, and this process seems to be associated with increased caspase 3 activity.<sup>5,20</sup> More recently, Goldman et al.<sup>16</sup> demonstrated that rat vena cava cultured under arterial conditions showed an increase in caspase 3 activity at 6 hours of culture and progressive -actin filament degradation and decreases in smooth muscle cell density over 48 hours, all of which are thought to be mediated by p38 MAPK.

It is generally accepted that vascular remodeling results from an imbalance between cell death and proliferation.<sup>6,20</sup> In arterial-vein grafts, apoptosis occurs in response to increased mechanical stress, which is counterbalanced by subsequent proliferation processes. In rat vein graft models 10 to 30 days after surgery, it has been shown that early decreases in cellular density may be compensated for or may even increase in comparison to control veins.<sup>19</sup> Liu et al.<sup>20</sup> suggested that the initial cell death observed in experimental vein grafts possibly

mediates the subsequent cellular proliferation. The absence of proliferation in saphenous veins cultured under arterial hemodynamic conditions in our system is not understood, but it may involve a lack of specific growth factors in the media culture used. Another possibility is that, under the present conditions, the proliferation process may require additional time beyond the 4-day observation period. However, it is important to emphasize that the lack of proliferation did not preclude the reproduction of other early events that have been observed in *in vivo* vein grafts, such as apoptotic processes and progressive decreases in cellular density and viability that may be associated with increased tensile stress.

The system presented here reproduced the early events observed in *in vivo* vein graft models with the advantage of working with human samples in very well controlled hemodynamic conditions. Although other components of blood circulation have an important role in vein graft failure, this *ex vivo* organ culture system provides the opportunity to study the effects of hemodynamic stimuli alone. We believe that the *ex vivo* organ culture is a useful model for characterizing the early molecular events taking place in the arterialization of human vein grafts and to provide new approaches to prolonging its lifespan.

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