

# Innate immunity and HPV: friends or foes

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Most human papillomavirus infections are readily cleared by the host immune response. However, in some individuals, human papillomavirus can establish a persistent infection. The persistence of high-risk human papillomavirus infection is the major risk factor for cervical cancer development. These viruses have developed mechanisms to evade the host immune system, which is an important step in persistence and, ultimately, in tumor development. Several cell types, receptors, transcription factors and inflammatory mediators involved in the antiviral immune response are viral targets and contribute to tumorigenesis. These targets include antigen-presenting cells, macrophages, natural killer cells, Toll-like receptors, nuclear factor kappa B and several cytokines and chemokines, such as interleukins, interferon and tumor necrosis factor. In the present review, we address both the main innate immune response mechanisms involved in HPV infection clearance and the viral strategies that promote viral persistence and may contribute to cancer development. Finally, we discuss the possibility of exploiting this knowledge to develop effective therapeutic strategies.

KEYWORDS: Innate Immune System; Human Papillomavirus; Cervical Cancer.

### ■ INTRODUCTION

Human papillomaviruses (HPVs) are an important group of viruses infecting the cutaneous and mucosal epithelia. HPVs cause diseases associated with high rates of morbidity and mortality, including benign lesions and cancer (1). According to the potential to cause cancer, HPV can be divided into two types: high-risk and low-risk. Low-risk types, which are mainly represented by HPV6 and HPV11, are associated with benign anogenital warts; although low-risk types are not usually associated with cancer, they may cause diseases associated with high morbidity. High-risk HPV (Hr-HPV) types, which are mainly represented by HPV16, HPV18, HPV31, HPV33, HPV35, HPV45 and other minor types, are related to cancer and precursor lesions, and Hr-HPV DNA sequences can be found in virtually all cervical tumors (2).

Cervical cancer is one of the most common cancers affecting women worldwide. According to the International Agency for Research on Cancer (IARC), the estimated number of new cases and deaths in 2012 were 527,600 and 265,700, respectively. The scenario is worse in less-developed countries, where cervical cancer represented the second most common cancer in women and was estimated to cause 230,200 of the

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total number of cervical cancer deaths in 2012, largely due to less-effective screening programs (3).

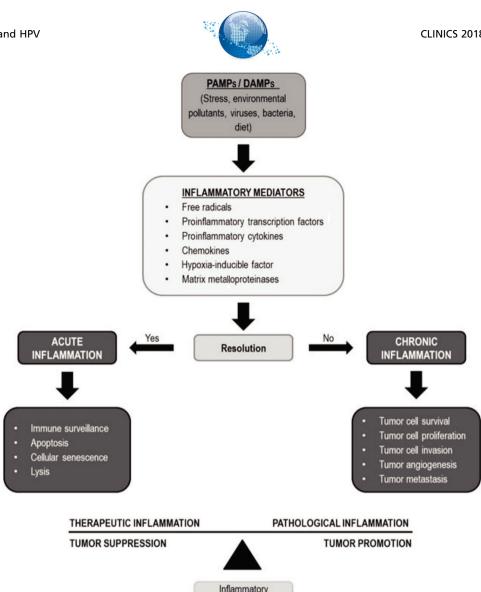
In addition, HPV may be associated with other anogenital tumors, although in a lower proportion. Indeed, HPV appears to be associated with 60-90% of all vaginal and anal cancers (4,5), and the virus can be found in 50% and 30-50% of vaginal and penile carcinoma cases, respectively (4,6). In addition, HPV is recognized as an etiologic cause of head and neck squamous cell carcinomas (HNSCCs); HNSCCs are mainly associated with HPV16, which is responsible for 40-80% of oropharyngeal cancers in the United States, a percentage that varies according to alcohol and tobacco use, poor oral hygiene and genetics (7,8). In general, the prognosis of HPV-positive vulvar and penile carcinomas as well as HPV-positive oral squamous cell carcinomas seems to be better than that of HPV-negative tumors (6,9).

Although most HPV infections are eliminated naturally, the persistence of Hr-HPV infection is the major risk factor for the development of high-grade cervical lesions and cervical cancer (10). To persist, HPV developed mechanisms to evade the host immune system; together with the oncogenic potential of HPV, persistence is the first step in the process leading to cancer (11).

The purpose of this review is to address the main innate immune response mechanisms involved in HPV infection clearance, HPV persistence and HPV-mediated cancer development, as well as to describe therapeutic possibilities.

### **HPV Elimination and Persistence**

Several immunological factors, especially those related to innate immunity pathways, which are the first line of defense against infections, are involved in HPV recognition and elimination. The efficient triggering of the immune response is a



**Figure 1** - Several stimuli, including HPV infection, can trigger the release of inflammatory mediators. The balance between these mediators may favor tumor suppression or tumor promotion. PAMPs: pathogen-associated molecular patterns; DAMPs: damage-associated molecular patterns.

signaling balance

turning point between viral clearance and persistence (12,13). As is generally observed in tumors, an inappropriate release of proinflammatory mediators and a chronic inflammatory response may contribute to cancer development (Figure 1).

In addition to innate immune cells, keratinocytes, which are both nonprofessional immune cells and targets of HPV infection, express pattern recognition receptors (PRRs). These receptors are able to identify microbial pathogens or damage signals, which are known as pathogen-associated molecular patterns (PAMPs) or damage-associated molecular patterns (DAMPs), respectively. PRRs include Toll-like receptors (TLRs), nucleotide binding oligomerization domain-like receptors (NLRs), retinoic acid-inducible gene I-like receptors (RLRs) and cytosolic DNA sensors (14,15).

Nucleic acids, which can accumulate during viral replication, are some of the microbial molecules recognized by PRRs (15). Previous studies showed that the high expression of TLR3, TLR7, TLR8 and TLR9, which recognize viral nucleic acids, is associated with HPV elimination and can be used as a predictor of clearance in HPV16-infected women. TLRs,

in combination with an HPV16 E6-specific effector response, are significantly associated with viral elimination (12,13). However, it has been shown that some types of Hr-HPV are capable of compromising the innate immune response, thus not only decreasing the expression of some TLRs but also impairing important pathways involving transcription factors such as nuclear factor kappa B (NF- $\kappa$ B) and interferon regulatory factor 3 (IRF3), which will be discussed below, and contributing to viral immune evasion and persistence (16-18).

Polymorphisms in innate immunity genes related to HPV infection, especially infection sensors and interleukins (ILs), have been reported in the literature. Although TLR9 has been recognized as a DNA sensor, TLR9 polymorphisms do not appear to be associated with viral clearance or persistence. However, polymorphisms in other innate immunity genes, such as interleukin 1 beta (IL-1β), interleukin 18 (IL-18), NLR1 and NLR3, were shown to be associated with HPV infection and persistence (19,20).

Another sensor of foreign DNA, interferon gamma inducible protein 16 (IFI16), has been shown to be capable of controlling



HPV18 replication and transcription through chromatin structure changes, thus reducing viral load and contributing to viral elimination (21).

Another antiviral mechanism involves the apolipoprotein B mRNA editing enzyme, catalytic polypeptide-like (APOBEC) protein family; members of this family are responsible for editing viral genomes, which can inhibit HPV infection. Interestingly, prolonged APOBEC activation during HPV infection can enhance genome mutagenesis, thus contributing to HPV-related cancer progression (22).

The efficiency of HPV in infection and persistence is associated with several mechanisms developed for immune response evasion. Mechanisms such as the modulation of cytokines and chemokines, downregulation of interferon (IFN) pathways, impairment of antigen presentation and reduction in the expression of adhesion molecules are directed mainly by the E6 and E7 oncoproteins (23). For example, Hr-HPV is able to inhibit cytokine production by blocking NF-κB activation upon innate immune system stimulation. Viral oncoproteins can increase the expression of interferon-related developmental regulator 1 (IFRD1), a protein responsible for recruiting histone deacetylase 1 (HDAC1) and HDAC3, thus leading to the deacetylation of NF-κB and inhibiting its ability to respond to immunological signals (17).

Additionally, to evade host innate immunity, Hr-HPV can inhibit PRR signaling through the induction of ubiquitin C-terminal hydrolase L1 (UCHL1) expression; UCHL1 blocks the activation of NF- $\kappa$ B and IRF3, both of which are transcription factors that induce the production of proinflammatory cytokines and chemokines (18). Additionally, Hr-HPV can downregulate the expression of inflammasome components and other downstream intermediaries of PRRs, such as genes in the IL-1 $\beta$  network, thus compromising not only innate immunity but also the activation of the adaptive response, which is also mediated by IL-1 $\beta$  (24). In addition, the direct effect of HPV16 E6 on IL-1 $\beta$  through its interaction with and recruitment of the E6AP ubiquitin-protein ligase, thus leading to cytokine degradation by the proteasome system, has been shown (25).

Other HPV proteins can play a role in immune escape; for example, HPV16 E2 can modulate the expression of 92 genes involved in the innate immune response, including the stimulator of interferon genes (STING), interferon kappa (IFN- $\kappa$ ) and interferon-stimulated genes (ISGs) (26).

The impairment of the IFN-mediated response is another resource used by HPV to evade the immune system. Viral oncoproteins reduce the secretion of IFN by keratinocytes, impair the phosphorylation of IFN pathway intermediates and compromise signaling through interferon regulatory factors (IRFs). For example, IRF1 expression is reduced in cervical tissues and cancer cell lines, and the reduced phosphorylation of IRF3 is associated with Hr-HPV infection. Thus, IFN is an important component of the innate immune response against Hr-HPV (18,27,28).

Finally, the potential for HPV to interfere with the migration and adhesion of innate immune cells and with the definition of cell phenotype must be emphasized. For example, the function of antigen-presenting cells (APCs), macrophages and natural killer (NK) cells can be compromised by HPV infection (29-32).

The abundance of dendritic cells (DCs), which are professional APCs that induce T cell-mediated immune responses, was found to be reduced in cervical intraepithelial neoplasias (CINs) (29). In addition, the expression of programmed

death-ligand 1 (PD-L1), which interacts with programmed cell death protein 1 (PD-1) in order to promote T cell anergy, is higher in DCs from Hr-HPV-positive patients than in DCs from Hr-HPV-negative patients (33,34).

Thus, an inefficient innate immune response can contribute to HPV persistence, which is a well-known cause of cell transformation and tumor progression initiated by HPV infection.

## **Tumor Progression: Cervical Cancer**

In cervical cancer, the expression of Hr-HPV oncoproteins is essential for cell transformation, and the integration of HPV DNA into the host genome contributes to this process (35,36). These oncoproteins, especially E6 and E7, induce continuous cell proliferation and prevent apoptosis, thus favoring the accumulation of mutations in the host genome due to the inhibition of pRb and p53 function (36). However, viral oncoproteins not only affect cell cycle regulatory mechanisms but also negatively impact the innate immune response (37). Together, viral persistence and the accumulation of cellular alterations allow the development of highgrade lesions and the progression of tumors.

The immune system plays a central role in determining the outcome of HPV infection, and immune system components are important in both viral clearance and tumorigenesis (38,39); this observation is true in HPV-related cancers as well as other cancers. For example, macrophages are present in the microenvironment of solid tumors, and although they can perform antitumor functions, they may also play an important role in cancer progression. Tumor-associated macrophages (TAMs) may promote cell proliferation and angiogenesis and may restrict immune defenses (40). These different roles can be explained by the identification of two macrophage phenotypes: the M1 proinflammatory macrophage phenotype and the M2 immunomodulatory macrophage phenotype. To help in tissue repair, M2 macrophages present a profile that elicits an increased production of vascular endothelial growth factor (VEGF) and matrix metalloproteinase 9 (MMP9). However, when activated by tumors, M2 macrophages can induce basement membrane disruption, tumor growth, and metastasis (41-43).

M2 macrophages are linked to protumor responses in many ways. The antitumor properties of M1 macrophages are primarily a result of the interleukin 12 (IL-12)-dominant cytokine milieu they produce (44). When macrophages assume an M2-like phenotype, they cannot produce IL-12, which is required for the activation of the antitumor response mediated by NK cells, T helper type 1 (Th1) cells and cytotoxic T lymphocytes (CTLs). Instead, M2 macrophages produce interleukin 10 (IL-10), which induces T helper type 2 (Th2) cell polarization, thus stimulating M2 macrophage polarization in a positive feedback loop mediated through interleukin 4 (IL-4) production (44,45). Furthermore, the immunoregulatory cytokine transforming growth factor beta (TGF-β) is linked to a Th2 response, and the precursor form of TGF-β can be processed by TAMs in order to release the active molecule (46,47).

Another important mechanism contributing to tumor expansion is the recruitment of regulatory T cells (Tregs) by M2-derived C-C motif chemokine ligand 22 (CCL22). In the tumor microenvironment, Treg activity is maintained by high local levels of IL-10 but is also induced by M2 macrophages, as mentioned previously. In addition, IL-10 promotes the



Table 1 - Potential prophylactic and therapeutic vaccines against HPV-related diseases.

Receptor	Agonist	HPV Antigen	Reference
Prophylactic Assays			
TLR2	Lipopeptide	E7 CTL epitope	(72)
TLR4	AS04, LPS derivative	VLP (L1)	(68)
TLR5	Flagellin	E6/E7 peptide	(77)
Therapeutic Assays	5	• •	
TLR3	Poly (I:C)	E7 peptide	(73)
TLR3/TLR7	Resiguimod/Poly (I:C)	E7 DNA	(75)
TLR7	Imiguimod	E7-calreticulin DNA	(76)
TLR7/TLR9	Imiquimod/CpG	E7 DNA	(74)
Prophylactic and Therapeutic Assays	·		
TLR2/TLR4	HMGB1 peptide	E7 DNA	(78)

differentiation of naive T cells to Tregs (45,48-50). Furthermore, TAMs expressing PD-L1 can directly induce T cell apoptosis by the binding of PD-L1 to its receptor (51).

Finally, the loss of antigen-presenting capabilities in macrophages is related to the downregulation of class II major histocompatibility complex (class II MHC) expression on M2 cells (52,53). In fact, when cervical lesions progress, the number of macrophages increases, and M2 macrophages are the main macrophage population in HPV-associated tumors (30,31).

NK cells are an important part of the innate immune response against viral attack and can lyse cancer cells even without the presentation of tumor antigens (54). NK cell activation occurs through an interaction between triggering receptors, such as NKp30, NKp44, NKp46 and natural killer group 2D (NKG2D), and tumor cell ligands, which is finely balanced between inhibitory receptors and coreceptors (55). In cervical cancer, the loss of class 1 MHC expression compromises the ability of tumor cells to present viral antigens to CTLs but may make tumor cells susceptible to NK cells. In addition, CD155, which is an activating NK cell receptor, was recently reported to be upregulated in squamous cervical carcinoma (56). However, another study showed that in high-grade squamous intraepithelial lesions (HSILs) and cervical cancer associated with HPV16 infection, the expression of the NK-activating receptors NKp30, NKp45 and NKG2D (only in cervical cancer) is considerably decreased, which affects the cytolytic functionality of cells and may contribute to tumor progression (32).

Whereas acute inflammation can promote antitumor immunity in the cervix, chronic inflammation can be associated with a protumor effect, partly through the provision of growth factors for use by the tumor. In addition, the production of inflammatory nitric oxide, cyclooxygenase (COX), IL-1 $\beta$  and tumor necrosis factor (TNF) enhances HPV-mediated tumorigenesis (40,57-60). Thus, the resistance of Hr-HPV-infected cells to the cytostatic or cytotoxic effect of some cytokines produced in a chronic inflammatory environment could be a key step in HPV-associated tumor development.

In cervical cancer, the expression of cyclooxygenase 2 (COX-2), which is an enzyme involved in the production of proinflammatory prostaglandins, is upregulated (61), and the induction of COX-2 by HPV16 E5 through NF-κB and activator protein-1 (AP-1) has been demonstrated (62). In addition, NF-κB levels may be elevated in cervical cancer epithelial cells, and this increase is associated with poor prognosis (63). Indeed, another study has shown that the HPV16 E6 and E7 oncoproteins induce an increase in NF-κB activity (64).

Previous studies showed that despite the antiproliferative effect of TNF on primary and HPV16-immortalized keratinocytes, HPV18-immortalized keratinocytes may be resistant to

this effect (65,66). Furthermore, genes associated with inflammatory responses, cell differentiation, cell death, proliferation, extracellular matrix remodeling and DNA repair were identified to be differentially expressed in HPV-immortalized keratinocytes with differential responses to the cytostatic effect of TNF (67). Taken together, these data support the idea that the acquisition of TNF resistance by HPV-infected cells may represent an important step towards malignancy.

The complete understanding of the immunological aspects of HPV infection and the cervical cancer microenvironment, including host immune components, HPV evasion and defense tactics and protumor factors, constitutes an important step in the development of new preventive and therapeutic options.

## **Prophylaxis and Treatment: Vaccines**

Considering the important role of the innate immune response in inflammatory processes, many studies have been carried out using receptor agonists to improve the immune response to prophylactic and therapeutic vaccines against HPV-related diseases (Table 1).

Indeed, there are three approved HPV vaccines, namely, Gardasil, Gardasil-9 and Cervarix; the first two use aluminum as the only adjuvant, and Cervarix also uses an LPS derivative that stimulates the innate immune system, thus activating TLR4 and helping to promote the death of HPV-infected cells through the activation of DCs and NK cells (68). Although Cervarix induces higher levels of neutralizing antibodies, no evidence supports Cervarix being more effective than Gardasil (69-71).

In previous studies, lipopeptides acting through TLR2 were used to stimulate the CTL response against HPV-associated tumors. Although the CTL response was useful in a prophylactic model, it could not inhibit tumor growth efficiently in a therapeutic context. The authors suggest that the depletion of immunosuppressive factors could improve the therapeutic effects of the vaccine (72). In another therapeutic vaccine model, a TLR3 agonist in a complex with an E7 peptide demonstrated a highly potent antitumor effect and induced a strong specific CTL response (73).

Interestingly, a DNA vector containing a virus-like particle sequence fused to a nononcogenic mutated E7 protein demonstrated an effect on established tumors in mice only when the vaccine was combined with TLR7 and TLR9 agonists (74). In addition, adding TLR3 and TLR7 agonists to a DNA vaccine containing the HPV16 E7 sequence promoted significant tumor regression in mice (75). Furthermore, the topical application of imiquimod (a TLR7 activator) associated with the intramuscular administration of a DNA vector containing HPV16 E7 fused to calreticulin increased the recruitment of CD8<sup>+</sup> T cells



to the genital tract in an orthotopic HPV16 E6/E7 syngeneic tumor model (76).

In addition, the use of flagellin as an adjuvant in order to induce a strong specific immune response through TLR5 activation has been studied as an alternative. When mixed with an intranasally administered antitumor vaccine, flagellin induced a cytotoxic response and conferred protection to mice challenged with HPV-transformed cells (77).

Additionally, the high mobility group box 1 (HMGB1) protein is related to innate immunity; HMGB1 activates TLR2/4 or the receptor for advanced glycation end products (RAGE) and can promote T cell activation. Moreover, an HMGB1 peptide displayed adjuvant properties in a mouse model when administered in combination with an E7 antigen. The data from the prophylactic and therapeutic assays indicated a substantial protective effect and showed the activation of the Th1 cellular immune response and the release of IFN- $\gamma$  (78).

Apart from the agonists used in prophylactic vaccines, no other vaccines are currently approved for clinical use in the treatment of HPV-positive tumors. Nevertheless, the potential of therapeutic vaccines must be investigated since an efficient therapeutic option will not be established soon.

Persistent HPV infection is associated with the modulation of immune cells, receptors, transcription factors, cytokines, chemokines and other immune mediators, all of which play a crucial role in inducing an effective immune response against HPV. Moreover, HPV has developed several mechanisms to evade or downregulate the innate immune response, including the modulation of the PRR response, the inhibition of antiviral molecules and the inhibition of the transcription of genes associated with the immune response. In addition, during HPV-associated tumor development, innate immune cells can contribute to the establishment and progression of such tumors. Because of the important role of the innate immune response in inflammatory processes and tumorigenesis, investing in studies that target this system to improve prophylaxis against HPV infection and to elicit efficient therapeutic responses in HPV-related tumors remains relevant.

## **■ AUTHOR CONTRIBUTIONS**

Nunes RA contributed to the abstract, the introduction, "HPV Elimination and Persistence", "Tumor Progression: Cervical Cancer" and the conclusion and revised and corrected all text and references. Silva GA contributed to "HPV Elimination and Persistence" and "Tumor Progression: Cervical Cancer". Morale MG contributed to "Prophylaxis and Treatment: Vaccines" and to organizing the references. Villa LL prepared, revised and corrected all text. Termini L prepared, revised and corrected all text.

## **■ REFERENCES**

- zur Hausen H. Papillomaviruses in the causation of human cancers a brief historical account. Virology. 2009;384(2):260-5, http://dx.doi.org/ 10.1016/j.virol.2008.11.046.
- Stanley MA, Pett MR, Coleman N. HPV: from infection to cancer. Biochem Soc Trans. 2007;35(Pt 6):1456-60, http://dx.doi.org/10.1042/BST0351456.
- Ferlay J, Soerjomataram I, Ervik M, Dikshit R, Eser S, Mathers C, et al. GLOBOCAN 2012 v1.0, Cancer Incidence and Mortality Worldwide: IARC CancerBase No. 11 [Internet]. Lyon, France: International Agency for Research on Cancer; 2013. Available from:http://globocan.iarc.f.
- Madsen BS, Jensen HL, van den Brule AJ, Wohlfahrt J, Frisch M. Risk factors for invasive squamous cell carcinoma of the vulva and vaginapopulation-based case-control study in Denmark. Int J Cancer. 2008; 122(12):2827-34, http://dx.doi.org/10.1002/ijc.23446.
- Frisch M, Biggar RJ. Aetiological parallel between tonsillar and anogenital squamous-cell carcinomas. Lancet. 1999;354(9188):1442-3, http://dx.doi. org/10.1016/S0140-6736(99)92824-6.

- 6. Lont AP, Kroon BK, Horenblas S, Gallee MP, Berkhof J, Meijer CJ, et al. Presence of high-risk human papillomavirus DNA in penile carcinoma predicts favorable outcome in survival. Int J Cancer. 2006;119(5):1078-81, http://dx.doi.org/10.1002/ijc.21961.
- D'Souza G, Kreimer AR, Viscidi R, Pawlita M, Fakhry C, Koch WM, et al. Case-control study of human papillomavirus and oropharyngeal cancer. N Engl J Med. 2007;356(19):1944-56, http://dx.doi.org/10.1056/NEJMoa 065497.
- Marur S, D'Souza G, Westra WH, Forastiere AA. HPV-associated head and neck cancer: a virus-related cancer epidemic. Lancet Oncol. 2010;11(8): 781-9, http://dx.doi.org/10.1016/S1470-2045(10)70017-6.
- Psyrri A, DiMaio D. Human papillomavirus in cervical and head-andneck cancer. Nat Clin Pract Oncol. 2008;5(1):24-31, http://dx.doi.org/ 10.1038/ncponc0984.
- Liaw KL, Hildesheim A, Burk RD, Gravitt P, Wacholder S, Manos MM, et al. A prospective study of human papillomavirus (HPV) type 16 DNA detection by polymerase chain reaction and its association with acquisition and persistence of other HPV types. J Infect Dis. 2001;183(1):8-15, http://dx.doi.org/10.1086/317638.
- Christensen ND. HPV disease transmission protection and control. Microb Cell. 2016;3(9):476-90, http://dx.doi.org/10.15698/mic2016.09.530.
- Daud II, Scott ME, Ma Y, Shiboski S, Farhat S, Moscicki AB. Association between toll-like receptor expression and human papillomavirus type 16 persistence. Int J Cancer. 2011;128(4):879-86, http://dx.doi.org/10.1002/ ijc.25400.
- Ścott ME, Ma Y, Farhat S, Moscicki AB. Expression of nucleic acid-sensing Toll-like receptors predicts HPV16 clearance associated with an E6-directed cell-mediated response. Int J Cancer. 2015;136(10):2402-8, http://dx.doi.org/ 10.1002/ijc.29283.
- Thompson MR, Kaminski JJ, Kurt-Jones EA, Fitzgerald KA. Pattern recognition receptors and the innate immune response to viral infection. Viruses. 2011;3(6):920-40, http://dx.doi.org/10.3390/v3060920.
- Jo EK, Kim JK, Shin DM, Sasakawa C. Molecular mechanisms regulating NLRP3 inflammasome activation. Cell Mol Immunol. 2016;13(2):148-59, http://dx.doi.org/10.1038/cmi.2015.95.
- Hasan UA, Bates E, Takeshita F, Biliato A, Accardi R, Bouvard V, et al. TLR9 expression and function is abolished by the cervical cancerassociated human papillomavirus type 16. J Immunol. 2007;178(5):3186-97, http://dx.doi.org/10.4049/jimmunol.178.5.3186.
- Tummers B, Goedemans R, Pelascini LP, Jordanova ES, van Esch EM, Meyers C, et al. The interferon-related developmental regulator 1 is used by human papillomavirus to suppress NFκB activation. Nat Commun. 2015;6:6537, http://dx.doi.org/10.1038/ncomms7537.
- Karim R, Tummers B, Meyers C, Biryukov JL, Alam S, Backendorf C, et al. Human papillomavirus (HPV) upregulates the cellular deubiquitinase UCHL1 to suppress the keratinocyte's innate immune response. PLoS Pathog. 2013;9(5):e1003384, http://dx.doi.org/10.1371/journal.ppat.1003384.
- Oliveira LB, Louvanto K, Ramanakumar AV, Franco EL, Villa LL; Ludwig-McGill Cohort Study. Polymorphism in the promoter region of the Toll-like receptor 9 gene and cervical human papillomavirus infection. J Gen Virol. 2013;94(Pt 8):1858-64, http://dx.doi.org/10.1099/vir.0.052811-0.
- Pontillo A, Bricher P, Leal VN, Lima S, Souza PR, Crovella S. Role of inflammasome genetics in susceptibility to HPV infection and cervical cancer development. J Med Virol. 2016;88(9):1646-51, http://dx.doi.org/ 10.1002/jmv.24514.
- Lo Cigno I, De Andrea M, Borgogna C, Albertini S, Landini MM, Peretti A, et al. The Nuclear DNA Sensor IFI16 Acts as a Restriction Factor for Human Papillomavirus Replication through Epigenetic Modifications of the Viral Promoters. J Virol. 2015;89(15):7506-20, http://dx.doi.org/10.1128/JVI.00013-15.
   Warren CJ, Westrich JA, Doorslaer KV, Pyeon D. Roles of APOBEC3A and
- Warren CJ, Westrich JA, Doorslaer KV, Pyeon D. Roles of APOBEC3A and APOBEC3B in Human Papillomavirus Infection and Disease Progression. Viruses. 2017;9(8):pii:E233, http://dx.doi.org/10.3390/v9080233.
   Kanodia S, Fahey LM, Kast WM. Mechanisms used by human papillo-
- Kanodia S, Fahey LM, Kast WM. Mechanisms used by human papillomaviruses to escape the host immune response. Curr Cancer Drug Targets. 2007;7(1):79-89, http://dx.doi.org/10.2174/156800907780006869.
   Karim R, Meyers C, Backendorf C, Ludigs K, Offringa R, van Ommen GJ,
- Karim R, Meyers Č, Backendorf C, Ludigs K, Offringa R, van Ommen GJ, et al. Human papillomavirus deregulates the response of a cellular network comprising of chemotactic and proinflammatory genes. PLoS One. 2011;6(3):e17848, http://dx.doi.org/10.1371/journal.pone.0017848.
- Niebler M, Qian X, Höfler D, Kogosov V, Kaewprag J, Kaufmann AM, et al. Post-translational control of IL-1β via the human papillomavirus type 16 E6 oncoprotein: a novel mechanism of innate immune escape mediated by the E3-ubiquitin ligase E6-AP and p53. PLoS Pathog. 2013; 9(8):e1003536, http://dx.doi.org/10.1371/journal.ppat.1003536.
- 9(8):e1003536, http://dx.doi.org/10.1371/journal.ppat.1003536.

  26. Sunthamala N, Thierry F, Teissier S, Pientong C, Kongyingyoes B, Tangsiriwatthana T, et al. E2 proteins of high risk human papillomaviruses down-modulate STING and IFN-к transcription in keratinocytes. PLoS One. 2014-9(3):e91473. http://dx.doi.org/10.1371/journal.pone.0091473.
- 2014;9(3):e91473, http://dx.doi.org/10.1371/journal.pone.0091473.

  27. Le Bon A, Tough DF. Links between innate and adaptive immunity via type I interferon. Curr Opin Immunol. 2002;14(4):432-6, http://dx.doi.org/10.1016/S0952-7915(02)00354-0.
- Um SJ, Rhyu JW, Kim EJ, Jeon KC, Hwang ES, Park JS. Abrogation of IRF-1 response by high-risk HPV E7 protein in vivo. Cancer Lett. 2002; 179(2):205-12, http://dx.doi.org/10.1016/S0304-3835(01)00871-0.



- 29. Mota FF, Rayment NB, Kanan JH, Singer A, Chain BM. Differential regulation of HLA-DQ expression by keratinocytes and Langerhans cells in normal and premalignant cervical epithelium. Tissue Antigens. 1998;52(3): 286-93, http://dx.doi.org/10.1111/j.1399-0039.1998.tb03046.x.
- Hammes LS, Tekmal RR, Naud P, Edelweiss MI, Kirma N, Valente PT, et al. Macrophages, inflammation and risk of cervical intraepithelial neoplasia (CIN) progression--clinicopathological correlation. Gynecol Oncol. 2007;105(1):157-65, http://dx.doi.org/10.1016/j.ygyno.2006.11.023.
   Lepique AP, Daghastanli KR, Cuccovia IM, Villa LL. HPV16 tumor
- Lepique AP, Daghastanli KR, Cuccovia IM, Villa LL. HPV16 tumor associated macrophages suppress antitumor T cell responses. Clin Cancer Res. 2009;15(13):4391-400, http://dx.doi.org/10.1158/1078-0432.CCR-09-0489.
- Garcia-Iglesias T, Del Toro-Arreola A, Albarran-Somoza B, Del Toro-Arreola S, Sanchez-Hernandez PE, Ramirez-Dueñas MG, et al. Low NKp30, NKp46 and NKG2D expression and reduced cytotoxic activity on NK cells in cervical cancer and precursor lesions. BMC Cancer. 2009;9:186, http://dx.doi. org/10.1186/1471-2407-9-186.
- Yang W, Song Y, Lu YL, Sun JZ, Wang HW. Increased expression of programmed death (PD)-1 and its ligand PD-L1 correlates with impaired cell-mediated immunity in high-risk human papillomavirus-related cervical intraepithelial neoplasia. Immunology. 2013;139(4):513-22, http://dx. doi.org/10.1111/imm.12101.
- Lyford-Pike S, Peng S, Young GD, Taube JM, Westra WH, Akpeng B, et al. Evidence for a role of the PD-1:PD-L1 pathway in immune resistance of HPV-associated head and neck squamous cell carcinoma. Cancer Res. 2013;73(6):1733-41, http://dx.doi.org/10.1158/0008-5472.CAN-12-2384.
- zur Hausen H. Papillomaviruses and cancer: from basic studies to clinical application. Nat Rev Cancer. 2002;2(5):342-50, http://dx.doi.org/10.1038/ nrc798.
- Doorbar J. Molecular biology of human papillomavirus infection and cervical cancer. Clin Sci (Lond). 2006;110(5):525-41.
- 37. Frazer IH. Interaction of human papillomaviruses with the host immune system: a well evolved relationship. Virology. 2009;384(2):410-4, http://dx.doi.org/10.1016/j.virol.2008.10.004.
- Ronco LV, Karpova AY, Vidal M, Howley PM. Human papillomavirus 16 E6 oncoprotein binds to interferon regulatory factor-3 and inhibits its transcriptional activity. Genes Dev. 1998;12(13):2061-72, http://dx.doi. org/10.1101/gad.12.13.2061.
- Senba M, Mori N. Mechanisms of virus immune evasion lead to development from chronic inflammation to cancer formation associated with human papillomavirus infection. Oncol Rev. 2012;6(2):e17, http://dx.doi. org/10.4081/oncol.2012.e17.
- Mazibrada J, Rittà M, Mondini M, De Andrea M, Azzimonti B, Borgogna C, et al. Interaction between inflammation and angiogenesis during different stages of cervical carcinogenesis. Gynecol Oncol. 2008;108(1): 112-20, http://dx.doi.org/10.1016/j.ygyno.2007.08.095.
- Martinez FÖ, Helming L, Gordon S. Alternative activation of macrophages: an immunologic functional perspective. Annu Rev Immunol. 2009;27: 451-83, http://dx.doi.org/10.1146/annurev.immunol.021908.132532.
- Quatromoni JG, Eruslanov E. Tumor-associated macrophages: function, phenotype, and link to prognosis in human lung cancer. Am J Transl Res. 2012;4(4):376-89.
- Tang X, Mo C, Wang Y, Wei D, Xiao H. Anti-tumour strategies aiming to target tumour-associated macrophages. Immunology. 2013;138(2):93-104, http://dx.doi.org/10.1111/imm.12023.
- Gabrilovich DI, Östrand-Rosenberg S, Bronte V. Coordinated regulation of myeloid cells by tumours. Nat Rev Immunol. 2012;12(4):253-68, http:// dx.doi.org/10.1038/nri3175.
- 45. DeNardo DG, Barreto JB, Andreu P, Vasquez L, Tawfik D, Kolhatkar N, et al. CD4(+) T cells regulate pulmonary metastasis of mammary carcinomas by enhancing protumor properties of macrophages. Cancer Cell. 2009;16(2):91-102, http://dx.doi.org/10.1016/j.ccr.2009.06.018.
- 46. Quatromoni JG, Wang Y, Vo DD, Morris LF, Jazirehi AR, McBride W, et al. T cell receptor (TCR)-transgenic CD8 lymphocytes rendered insensitive to transforming growth factor beta (TGFβ) signaling mediate superior tumor regression in an animal model of adoptive cell therapy. J Transl Med. 2012;10:127, http://dx.doi.org/10.1186/1479-5876-10-127.
- Chong H, Vodovotz Y, Cox GW, Barcellos-Hoff MH. Immunocytochemical localization of latent transforming growth factor-beta1 activation by stimulated macrophages. J Cell Physiol. 1999;178(3):275-83, http://dx.doi.org/10.1002/GICI)1097-4652(199903)178:3 < 275::AID-JCP1 > 3.0.CO;2-Q.
- Curiel TJ, Coukos G, Zou L, Alvarez X, Cheng P, Mottram P, et al. Specific recruitment of regulatory T cells in ovarian carcinoma fosters immune privilege and predicts reduced survival. Nat Med. 2004;10(9):942-9, http://dx.doi.org/10.1038/nm1093.
- Murai M, Turovskaya O, Kim G, Madan R, Karp CL, Cheroutre H, et al. Interleukin 10 acts on regulatory Tcells to maintain expression of the transcription factor Foxp3 and suppressive function in mice with colitis. Nat Immunol. 2009;10(11):178-84. http://dx.doi.org/10.1038/pi.1791
- Transcription factor rospo and suppressive function in the transcription factor rospo and suppressive function in the factor for factor factor for factor for factor f

- Kuang DM, Zhao Q, Peng C, Xu J, Zhang JP, Wu C, et al. Activated monocytes in peritumoral stroma of hepatocellular carcinoma foster immune privilege and disease progression through PD-L1. J Exp Med. 2009;206(6):1327-37, http://dx.doi.org/10.1084/jem.20082173.
- Eruslanov E, Daurkin I, Ortiz J, Vieweg J, Kusmartsev S. Pivotal Advance: Tumor-mediated induction of myeloid-derived suppressor cells and M2-polarized macrophages by altering intracellular PGE<sub>2</sub> catabolism in myeloid cells. J Leukoc Biol. 2010;88(5):839-48, http://dx.doi.org/10.1189/ jlb.1209821.
- Heusinkveld M, de Vos van Steenwijk PJ, Goedemans R, Ramwadhdoebe TH, Gorter A, Welters MJ, et al. M2 macrophages induced by prostaglandin E2 and IL-6 from cervical carcinoma are switched to activated M1 macrophages by CD4+ Th1 cells. J Immunol. 2011;187(3):1157-65, http:// dx.doi.org/10.4049/jimmunol.1100889.
- 54. Lanier LL. Evolutionary struggles between NK cells and viruses. Nat Rev Immunol. 2008;8(4):259-68, http://dx.doi.org/10.1038/nri2276.
- Moretta L, Bottino C, Pende D, Castriconi R, Mingari MC, Moretta A. Surface NK receptors and their ligands on tumor cells. Semin Immunol. 2006;18(3):151-8, http://dx.doi.org/10.1016/j.smim.2006.03.002.
- Textor S, Dürst M, Jansen L, Accardi R, Tommasino M, Trunk MJ, et al. Activating NK cell receptor ligands are differentially expressed during progression to cervical cancer. Int J Cancer. 2008;123(10):2343-53, http:// dx.doi.org/10.1002/jic.23733.
- 57. Woodworth CD, McMullin E, Iglesias M, Plowman GD. Interleukin 1 alpha and tumor necrosis factor alpha stimulate autocrine amphiregulin expression and proliferation of human papillomavirus-immortalized and carcinoma-derived cervical epithelial cells. Proc Natl Acad Sci U S A. 1995;92(7):2840-4, http://dx.doi.org/10.1073/pnas.92.7.2840.
- Young JL, Jazaeri AA, Darus CJ, Modesitt SC. Cyclooxygenase-2 in cervical neoplasia: a review. Gynecol Oncol. 2008;109(1):140-5, http://dx.doi.org/ 10.1016/j.ygyno.2008.01.008.
- Sappayatosok K, Maneerat Y, Swasdison S, Viriyavejakul P, Dhanuthai K, Zwang J, et al. Expression of pro-inflammatory protein, iNOS, VEGF and COX-2 in oral squamous cell carcinoma (OSCC), relationship with angiogenesis and their clinico-pathological correlation. Med Oral Patol Oral Cir Bucal. 2009;14(7):E319-24.
- Wei L, Gravitt PE, Song H, Maldonado AM, Ozbun MA. Nitric oxide induces early viral transcription coincident with increased DNA damage and mutation rates in human papillomavirus-infected cells. Cancer Res. 2009;69(11):4878-84, http://dx.doi.org/10.1158/0008-5472.CAN-08-4695.
- Khunamornpong S, Settakorn J, Sukpan K, Srisomboon J, Ruangvejvorachai P, Thorner PS, et al. Cyclooxygenase-2 expression in squamous cell carcinoma of the uterine cervix is associated with lymph node metastasis.
   Gynecol Oncol. 2009;112(1):241-7, http://dx.doi.org/10.1016/j.ygyno.2008.
- Kim SH, Oh JM, No JH, Bang YJ, Juhnn YS, Song YS. Involvement of NF-kappaB and AP-1 in COX-2 upregulation by human papillomavirus 16 E5 oncoprotein. Carcinogenesis. 2009;30(5):753-7, http://dx.doi.org/ 10.1093/carcin/bgp066.
- Beglin M, Melar-New M, Laimins L. Human papillomaviruses and the interferon response. J Interferon Cytokine Res. 2009;29(9):629-35, http:// dx.doi.org/10.1089/jir.2009.0075.
- Textor S, Accardi R, Havlova T, Hussain I, Sylla BS, Gissmann L, et al. NF-κ B-dependent upregulation of ICAM-1 by HPV16-E6/E7 facilitates NK cell/target cell interaction. Int J Cancer. 2011;128(5):1104-13, http://dx.doi.org/10.1002/ijc.25442.
- Villa LL, Vieira KB, Pei XF, Schlegel R. Differential effect of tumor necrosis factor on proliferation of primary human keratinocytes and cell lines containing human papillomavirus types 16 and 18. Mol Carcinog. 1992;6(1): 5-9, http://dx.doi.org/10.1002/mc.2940060103.
- Boccardo E, Noya F, Broker TR, Chow LT, Villa LL. HPV-18 confers resistance to TNF-alpha in organotypic cultures of human keratinocytes. Virology. 2004;328(2):233-43, http://dx.doi.org/10.1016/j.virol.2004.07.026.
- 67. Termini L, Boccardo E, Esteves GH, Hirata R Jr, Martins WK, Colo AE, et al. Characterization of global transcription profile of normal and HPV-immortalized keratinocytes and their response to TNF treatment. BMC Med Genomics. 2008;1:29, http://dx.doi.org/10.1186/1755-8794-1-29.
- Van den Bergh JM, Guerti K, Willemen Y, Lion E, Cools N, Goossens H, et al. HPV vaccine stimulates cytotoxic activity of killer dendritic cells and natural killer cells against HPV-positive tumour cells. J Cell Mol Med. 2014;18(7):1372-80, http://dx.doi.org/10.1111/jcmm.12284.
- Zurek Munk-Madsen M, Toft L, Kube T, Richter R, Ostergaard L, Søgaard OS, et al. Cellular immunogenicity of human papillomavirus vaccines Cervarix and Gardasil in adults with HIV infection. Hum Vaccin Immunother. 2018;14(4):909-16, http://dx.doi.org/10.1080/21645515.2017.1407896.
- ther. 2018;14(4):909-16, http://dx.doi.org/10.1080/21645515.2017.1407896.

  To. Einstein MH, Baron M, Levin MJ, Chatterjee A, Edwards RP, Zepp F, et al. Comparison of the immunogenicity and safety of Cervarix and Gardasil human papillomavirus (HPV) cervical cancer vaccines in healthy women aged 18-45 years. Hum Vaccin. 2009;5(10):705-19, http://dx.doi.org/10.4161/hv.5.10.9518.
- Haghshenas MR, Mousavi T, Kheradmand M, Afshari M, Moosazadeh M. Efficacy of Human Papillomavirus L1 Protein Vaccines (Cervarix and Gardasil) in Reducing the Risk of Cervical Intraepithelial Neoplasia: A Meta-analysis.



- Int J Prev Med. 2017;8:44, http://dx.doi.org/10.4103/ijpvm.IJPVM\_413\_16.
- Shen KY, Song YC, Chen IH, Chong P, Liu SJ. Depletion of tumorassociated macrophages enhances the anti-tumor immunity induced by a Toll-like receptor agonist-conjugated peptide. Hum Vaccin Immunother. 2014;10(11):3241-50. http://dx.doi.org/10.4161/hv.29275.
- 2014;10(11):3241-50, http://dx.doi.org/10.4161/hv.29275.
   Chen S, Ou R, Tang J, Deng X, Wu Y, van Velkinburgh JC, et al. Enhanced anti-tumor effects of HPV16E7(49-57)-based vaccine by combined immunization with poly(I:C) and oxygen-regulated protein 150. Cancer Epidemiol. 2013;37(2):172-8, http://dx.doi.org/10.1016/j.canep.2012.10.005.
- 2013;37(2):172-8, http://dx.doi.org/10.1016/j.canep.2012.10.005.
  74. Lescaille G, Pitoiset F, Macedo R, Baillou C, Huret C, Klatzmann D, et al. Efficacy of DNA vaccines forming e7 recombinant retroviral virus-like particles for the treatment of human papillomavirus-induced cancers. Hum Gene Ther. 2013;24(5):533-44, http://dx.doi.org/10.1089/hum.2012.037.
- Sajadian A, Tabarraei A, Soleimanjahi H, Fotouhi F, Gorji A, Ghaemi A. Comparing the effect of Toll-like receptor agonist adjuvants on the efficiency

- of a DNA vaccine. Arch Virol. 2014;159(8):1951-60, http://dx.doi.org/10.1007/s00705-014-2024-4.
- Soong RS, Song L, Trieu J, Knoff J, He L, Tsai YC, et al. Toll-like receptor agonist imiquimod facilitates antigen-specific CD8+ T-cell accumulation in the genital tract leading to tumor control through IFNγ. Clin Cancer Res. 2014;20(21):5456-67, http://dx.doi.org/10.1158/1078-0432. CCR-14-0344.
- 77. Nguyen CT, Hong SH, Ung TT, Verma V, Kim SY, Rhee JH, et al. Intranasal immunization with a flagellin-adjuvanted peptide anticancer vaccine prevents tumor development by enhancing specific cytotoxic T lymphocyte response in a mouse model. Clin Exp Vaccine Res. 2013;2(2):128-34, http://dx.doi.org/10.7774/cevr.2013.2.2.128.
- Talebi S, Bolhassani A, Sadat SM, Vahabpour R, Agi E, Shahbazi S. Hp91 immunoadjuvant: An HMGB1-derived peptide for development of therapeutic HPVn vaccines. Biomed Pharmacother. 2017;85:148-54, http://dx.doi.org/10.1016/j.biopha.2016.11.115.