

CLÍNICA E INVESTIGACIÓN EN ARTERIOSCLEROSIS



www.elsevier.es/arterio

REVIEW ARTICLE

Update of HDL in atherosclerotic cardiovascular disease



Leonie Schoch^{a,b}, Sebastián Alcover^a, Teresa Padró^a, Soumaya Ben-Aicha^c, Guiomar Mendieta^d, Lina Badimon^{a,e,f}, Gemma Vilahur^{a,f,*}

- ^a Cardiovascular Program, Institut de Recerca, Hospital de la Santa Creu I Sant Pau, IIB Sant Pau, 08025 Barcelona, Spain
- ^b Faculty of Medicine, University of Barcelona (UB), 08036 Barcelona, Spain
- ^c Imperial College, London, United Kingdom
- d Cardiology Unit, Cardiovascular Clinical Institute, Hospital Clínic de Barcelona, Barcelona, Spain
- e Cardiovascular Research Chair, UAB, 08025 Barcelona, Spain
- ^f CiberCV, Institute of Health Carlos III, Madrid, Spain

Received 13 September 2023; accepted 13 October 2023

KEYWORDS

HDL; Atherosclerosis; Cardiovascular disease; Therapy **Abstract** Epidemiologic evidence supported an inverse association between HDL (high-density lipoprotein) cholesterol (HDL-C) levels and atherosclerotic cardiovascular disease (ASCVD), identifying HDL-C as a major cardiovascular risk factor and postulating diverse HDL vascular- and cardioprotective functions beyond their ability to drive reverse cholesterol transport. However, the failure of several clinical trials aimed at increasing HDL-C in patients with overt cardiovascular disease brought into question whether increasing the cholesterol cargo of HDL was an effective strategy to enhance their protective properties. In parallel, substantial evidence supports that HDLs are complex and heterogeneous particles whose composition is essential for maintaining their protective functions, subsequently strengthening the "HDL quality over quantity" hypothesis.

The following state-of-the-art review covers the latest understanding as per the roles of HDL in ASCVD, delves into recent advances in understanding the complexity of HDL particle composition, including proteins, lipids and other HDL-transported components and discusses on the clinical outcomes after the administration of HDL-C raising drugs with particular attention to CETP (cholesteryl ester transfer protein) inhibitors.

© 2023 Sociedad Española de Arteriosclerosis. Published by Elsevier España, S.L.U. All rights reserved.

E-mail address: gvilahur@santpau.cat (G. Vilahur).

^{*} Corresponding author.

PALABRAS CLAVE

HDL; Aterosclerosis; Enfermedad cardiovascular; Terapia

Nuevos conceptos de las HDL en la enfermedad cardiovascular aterosclerótica

Resumen Estudios epidemiológicos respaldan una asociación inversa entre los niveles de colesterol de lipoproteínas de alta densidad (c-HDL) y la enfermedad cardiovascular aterosclerótica (ECVA), identificando el c-HDL como un importante factor de riesgo cardiovascular y postulando diversas funciones vasculares y cardioprotectoras de las HDL más allá de su capacidad para promover el transporte reverso del colesterol. Sin embargo, el fracaso de varios ensayos clínicos dirigidos a aumentar el c-HDL en pacientes con enfermedad cardiovascular manifiesta, puso en duda el concepto que incrementar la carga de c-HDL fuera una estrategia eficaz para potenciar sus propiedades protectoras. Paralelamente, numerosos estudios han evidenciado que las HDL son partículas complejas y heterogéneas cuya composición es esencial para mantener sus funciones protectoras, lo que refuerza la hipótesis de que «la calidad de las HDL prima sobre la cantidad».

En el siguiente manuscrito revisamos el estado del arte sobre los últimos avances en torno a las funciones de las HDL en la ECVA, nos adentramos en los avances recientes en la comprensión de la complejidad de la composición de las partículas de HDL, incluidas las proteínas, los lípidos y otros componentes transportados por las HDL, y revisamos los resultados clínicos tras la administración de inductores del c-HDL, especialmente los inhibidores de la proteína transportadora del colesterol esterificado (CETP).

© 2023 Sociedad Española de Arteriosclerosis. Publicado por Elsevier España, S.L.U. Todos los derechos reservados.

Introduction

Epidemiology of cardiovascular disease

Despite recent advances in preventing, detecting, and treating cardiovascular disease (CVDs), the global burden is increasing due to population growth and ageing. Within the last 30 years, cardiovascular (CV) mortality has risen by 53%, 1 remaining the leading cause of death worldwide, accounting for 32% of all deaths, of which the two most prevalent causes of death are ischemic heart disease (16%) and stroke (12%). 2 The probability of suffering from any CVD increases with risk factors (Fig. 1).

As a largely asymptomatic condition, elevated blood pressure is estimated to be responsible for around 50% of all deaths from ischemic heart disease and stroke.3 It is thus considered the single largest contributor to the global burden of disease and mortality. Being overweight $(BMI \ge 25 \text{ kg/m}^2)$ and subsequently obese $(BMI \ge 30 \text{ kg/m}^2)$ are continuously on the rise and have become a global health crisis due to their increased risk of suffering from numerous diseases, including CVD. Directly linked to obesity are physical inactivity and an unhealthy and unbalanced diet with high amounts of sodium (>5 g/day), saturated fat, cholesterol, and red or processed meat but low intake of fruits, vegetables, whole grains, legumes, fibre, nuts, seeds, omega-3 and polyunsaturated fats which increase CV risk and thus contribute to overall CV deaths. 1,4 Consequently, these dietary habits can lead to dyslipidaemia, an imbalance of serum lipids. As such, high total cholesterol, low-density lipoprotein cholesterol (LDL-C), triglyceride levels, and lower high-density lipoprotein cholesterol (HDL-C) concentrations are strongly associated with the development of atherosclerotic CVDs (ACVDs). Furthermore, besides developing a proatherogenic profile,⁵ serum lipid levels may induce a prothrombotic state.⁶ Likewise, lifestyle-related hyperglycaemia and diabetes mellitus are increasing problems since age-standardized mean fasting plasma glucose levels have continuously risen during the past 30 years, regardless of sex, age, and country.^{7,8} Compared to 1990, there has been an increase in diabetes of over 186%.⁷ Finally, smoking is one of the more difficult-to-grasp risk factors due to the complexity of the chemical constituents of the smoke. Metal components in cigarette smoke have been shown to damage vascular endothelial cells by inducing oxidative stress and inflammation, which are involved in the development of ACVDs progression.⁹

Brief overview of the pathogenesis of atherosclerosis

Atherosclerosis is a lipid-driven inflammatory disease characterised by the deposition of fatty, fibrous, and calcified material in the innermost layer of large and medium-sized arteries. Its development starts with the activation of the arterial endothelium, followed by an inflammatory cascade, continuous lipid build-up in the vessel wall, and plaque formation, which ultimately will disrupt and induce thrombosis (i.e., atherothrombosis) with impaired or even blocked blood flow (Fig. 2).

The first step in the development of atherosclerosis is endothelial activation and dysfunction due to hemodynamic forces within the arteries. ¹⁰ Lesion-prone regions such as arterial bifurcations are mainly characterised by disturbed laminar flow and reduced shear stress, ^{11,12} which disrupt the endothelial barrier and facilitate the retention of LDL particles, ^{13,14} but also upregulate the expression of genes involved in the attraction and recruitment of

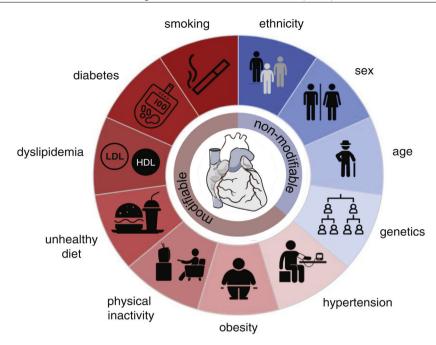


Figure 1 Risk factors for atherosclerotic cardiovascular diseases. Modifiable (red) and non-modifiable (blue) risk factors. This figure was partly generated using Servier Medical Art, provided by Servier, licensed under a Creative Commons Attribution 3.0 unported license (heart), and The Noun Project (symbols). Donut chart by google docs (excel).

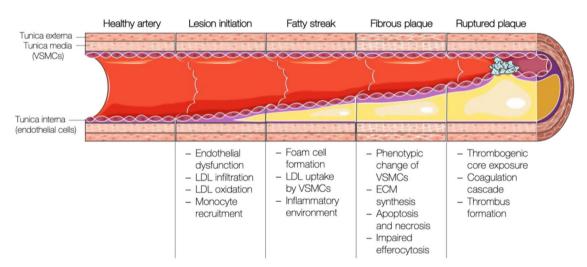


Figure 2 Schematic representation of the different stages of atherosclerotic plaque formation. VSMCs: vascular smooth muscle cells; LDL: low-density lipoprotein; ECM: extracellular matrix. This figure was partly generated using Servier Medical Art, provided by Servier, licensed under a Creative Commons Attribution 3.0 unported license.

immune cells (MCP-1, ¹⁵ VCAM-1, ¹⁶ ICAM-1, ¹⁷ PECAM-1¹⁸). On the other hand, fluid mechanical forces also affect the expression and activity of the endothelial nitric oxide synthase (eNOS), the enzyme catalysing the conversion from L-arginine to nitric oxide (NO). Endothelium-derived NO production and bioavailability are key for the homeostasis of the vascular endothelium, adjacent smooth muscle cell vasore-laxation, and inhibition of platelet activation, adhesion, and aggregation. ¹⁹

High levels of LDL-C favour LDL particle infiltration into the arterial intima by diffusion, paracellular crossing,

or transcytosis, 20 where they suffer modifications. Circulating monocytes are then attracted and extravasate to the intima to engulf these modified LDL particles and become foam cells. Foam cells secrete pro-inflammatory cytokines (mainly interleukin (IL)-1, IL-6, and tumour necrosis factor-alpha (TNF α)) and chemokines (MCP-1), which further promote monocyte recruitment and inflammatory response propagation. 21

Due to a cocktail of mainly macrophage-derived growth factors and cytokines, vascular smooth muscle cells (VSMCs) are recruited to the luminal side of the

lesion and secrete extracellular matrix components, matrix metalloproteases, and pro-inflammatory cytokines.²² As atherosclerotic plagues mature and more cholesterol is deposited, foam cells and intimal VSMCs cannot cope with the excessive cholesterol burden and undergo apoptosis.²³ Cellular debris, cholesterol crystals, and thrombogenic, inflammatory, and oxidative material accumulate in an extending necrotic core. Osteochondrogenic VSMC-derived calcifications gradually extend from the necrotic centre to the surrounding matrix, which, together with increasing ECM production, causes arterial stiffness. 24,25 Upon rising apoptosis rates in the intimal VSMC population and matrix metalloprotease activity, the plague becomes vulnerable, 10 which is generally considered to include a large necrotic core, a thin fibrous cap, and an increased inflammatory infiltrate due to continuous exposure to the pro-atherogenic milieu.¹⁰ When the fibrous cap that protects circulation from the content of the necrotic core fissures or ruptures, the thrombogenic and pro-inflammatory plaque content is exposed to the blood. Upon contact with blood components (platelets and coagulant factors), sub-occlusive or occlusive thrombosis may occur, leading to acute coronary syndromes.²⁶

Lipoprotein particles

Lipoproteins are complex particles that enable the transportation of non-polar lipids such as triglycerides and cholesteryl esters in their hydrophobic core, protected by an outer hydrophilic layer containing free cholesterol, phospholipids, and apolipoproteins. They primarily differ in size, density, and composition, as summarised in Table 1.

Chylomicrons are the largest and least dense lipoprotein particles, taking dietary triglycerides and cholesterol (exogenous origin from diet) from the intestine to transport to muscle and adipose tissue. The enzymatic action of the lipoprotein lipase at the luminal surface of the adipose and muscle tissues capillary endothelium hydrolyses the triglycerides into glycerol and free fatty acids (lipolysis). Free fatty acids are either used as local energy substrates (betaoxidation and Krebs cycle) or for energy storage in their re-esterified form (triglycerides). The remaining chylomicron remnants are taken up by hepatic remnant receptors or the LDL receptor (LDLr), and the endogenous lipoprotein pathway is initiated by forming very low-density lipoproteins (VLDLs). VLDLs mainly carry endogenous triglycerides, which either have been synthesised by the liver de novo or produced by the re-esterification of free fatty acids. VLDL-transported triglycerides are hydrolysed at peripheral tissues by lipoprotein lipase, forming VLDL remnants or intermediate-density lipoproteins (IDLs). IDLs are transformed to low-density lipoproteins (LDLs) by further active removal of triglycerides mediated by the cholesteryl ester transport protein (CETP) (to high-density lipoproteins (HDL)) and hepatic triglyceride lipase (to the liver). LDLs are the main cholesterol carrier in circulation, provided for cellular needs by LDLr-mediated uptake and lysosomal digestion of the LDL particle. Excess of apolipoprotein (Apo)-B-containing lipoproteins are considered pro-atherogenic, while the ApoA-containing lipoprotein (HDL particle) may exert anti-atherogenic effects as expanded below.

Classification of the HDL particle

The HDL particle is the densest and smallest of the lipoprotein particle family. They are a group of very heterogeneous particles, and their classification according to physicochemical properties is challenging due to the need for more consensus regarding the definitive categories of HDL subclasses and how to define them.²⁷ The commonly used subclasses are determined according to their separation method (Table 2).

HDL particle metabolism

The backbone of all HDL particles is their characteristic structural ApoA-I, synthesised in the liver and intestine. ApoA-I molecules are lipidated by the interaction with ABCA1, which forms nascent discoidal pre- β particles (Fig. 3). Further lipidation and transformation of free cholesterol into cholesteryl ester (by lecithin: cholesterol acyl transferase; LCAT) promote particle maturation to spherical α -HDLs, which primarily interact with ABCG1 and SR-B1 instead of ABCA1 (Fig. 3).

This, in combination with the enzymatic activity of CETP or the hepatic and endothelial lipase, as well as the phospholipid transfer activity of the phospholipid transfer protein (PLTP) leads to constant dynamic remodelling of the mature α -HDL particle, thus significantly modulating size and composition. The HDL particle's most important hallmark function is to clear excess cholesterol from peripheral organs and transport it to the liver for biliary excretion (reverse cholesterol transport). HDL-transported cholesteryl esters can either be delivered directly to the hepatocytes (SR-B1) or might be indirectly provided by ApoB-containing lipoproteins that receive HDL-transported cholesteryl esters in exchange for triglycerides. Cholesterol is an essential component of cell membranes and is required for the biosynthesis of vital, biologically active components such as vitamin D, lipid-digesting bile acids, and steroid hormones. Its levels are modulated by the absorption of exogenous. dietary cholesterol and endogenous biosynthesis of cholesterol, mainly performed by the liver and intestine. However, too high circulating cholesterol levels are the underlying cause and the first step towards ACVDs development.

HDL particle composition

HDL particles comprise a lipid core mainly containing the neutral lipids cholesteryl ester and triglycerides, which is protected by a surface layer of amphiphilic phospholipids, free cholesterol, and apolipoproteins. This composition ensures water solubility to all lipophilic components associated with HDL particles, enabling vital circulation transportation. In recent years, the list of HDL-bound components increased to proteins and enzymes, lipids, small non-coding RNAs (ncRNAs), hormones, and vitamins, making the HDL particle a very versatile but complex structure.

HDL proteome

Recent advances in proteomics and the "HDL Proteome Watch" project have increased our knowledge about HDL-bound protein species. While 85 different proteins were

Table 1 Classification and characteristics of lipoproteins.

Lipoprotein	Density (g/ml)	Size (nm)	Lipid/protein (%)	Main lipid	Main apolipoprotein
	, , , , , , , , , , , , , , , , , , ,		,	· · · · · · · · · · · · · · · · · · ·	<u> </u>
Chylomicrons	< 0.930	1000-70	98/2	TG	ApoB-48
VLDL	0.950-1.006	200–27	92/8	TG	ApoB-100
IDL	1.006-1.019	27-23	89/11	TG/CE	ApoB-100
LDL	1.019-1.063	23-18	79/21	CE	ApoB-100
HDL	1.063-1.210	13-7.3	50/50	CE	ApoA-I

VLDL: very low-density lipoprotein; IDL: intermediate-density lipoprotein; LDL: low-density lipoprotein; HDL: high-density lipoprotein; LD(a): lipoprotein(a); TG: triglycerides; CE: cholesteryl ester; ApoB: apolipoprotein B; ApoA: apolipoprotein A.

Classification	Method	HDL Subclass
Shape and charge	2D gel electrophoresis	Pre-ß HDL (discoidal)
		α -HDL 1/2/3/4 (spherical)
Size (nm)	Gradient gel electrophoresis	HDL2a (13.0-9.7), HDL2b (9.7-8.8)
		HDL3a (8.8-8.2), HDL3b (8.2-7.8), HDL3c (7.8-7.3)
Size (nm)	NMR spectroscopy	Large (13-8.8 nm)
		Medium (8.8-8.2 nm)
		Small (8.2-7.3 nm)
Density (g/ml)	Density gradient ultracentrifugation	HDL ₂ (1.063-1.125)
		HDL ₃ (1.125-1.210)

reported to associate with HDL particles a few years back, this number has risen to 251.²⁸ It is important, however, to mention the impact that HDL isolation has on the reported proteomic profile. While each of the common isolation techniques (ultracentrifugation, immunoaffinity, electrophoresis, and gel filtration) has its "flaws", cross-referencing proteomic profiles over studies and isolation techniques has enabled us to define a more robust selection of 91 "very likely" protein constituents of the HDL particle²⁸ (Table 3).

Recent and extensive work suggests HDL subclasses according to the main functions associated with respective protein clusters.²⁸ The biggest and most important subclasses include proteins involved in (1) lipid transportation (e.g., ApoA-I/-II, ApoC-I/-II/-III, ApoE, ApoM, CETP, LCAT) being ApoA-I and ApoA-II the most important structural pillars in all HDL particles; (2) homeostasis/protease inhibition (e.g., ITIH1/2/3, coagulation factors such as F5 and F13B); (3) inflammation/acute phase response (e.g., serum amyloid A1/2/4, ITIH4); (4) immunity/anti-microbial (e.g., immunoglobulins such as IGKV3-20, IGHV3-13, and complement pathway components such as C3, C4B, C6); and (5) cell/heparin-binding (e.g., SELL, DSG1, CD44, ANGPTL3). However, functions that are less represented, such as vitamin binding and transport, including the vitamin D- and retinol-binding proteins, or metal ion binding, need also to be considered. Nevertheless, whether strongly or weakly represented, the countless functions highlight important learning from the last years of research: HDL particles are much more than just lipid transporters.

HDL lipidome

The HDL lipidome is just as diverse as its proteomic counterpart, consisting of more than 400 HDL-bound lipid species identified so far.²⁹ Based on their chemical properties. amphiphilic lipids such as free cholesterol and phospholipids are carried on the surface of the HDL particle, protecting the neutral hydrophobic lipids such as cholesteryl esters and triglycerides in the HDL core. The major HDLbound lipid species are phospholipids with 37.4-49.3 mol%, cholesteryl esters with 35.0-37.0 mol%, free cholesterol with 8.7-13.5 mol%, sphingolipids with 5.7-6.9 mol%, and triglycerides with 2.8-3.2 mol% of total lipids. 30 Phospholipids and sphingolipids form a big part of the HDL lipidome, with phosphatidylcholine and sphingomyelin being the most abundant species, respectively.³⁰ The HDL lipidome constitutes many more lipids, though to a much lower abundance (Table 4).

Sphingomyelin and free cholesterol, two lipidic components that affect membrane fluidity, decrease with increasing density of HDL particles (HDL2 \rightarrow HDL3). ³¹ Hence, denser HDL3 particles exert better membrane fluidity, which may have functional implications for interactions with receptors and transporters. ³¹ Likewise, the abundance of sphingosine-1-phosphate (S1P) per HDL particle is significantly higher in dense HDL3 (40–50 mmol/mol HDL) particles compared with HDL2 particles (15–20 mmol/mol). ^{31–33} The enrichment of small, dense HDL3 with S1P is driven by HDL-bound ApoM, one of the main S1P carriers in circulation, ³⁴ which is predominantly associated with dense HDL3 particles ³⁵ and has functional implications. ³⁶

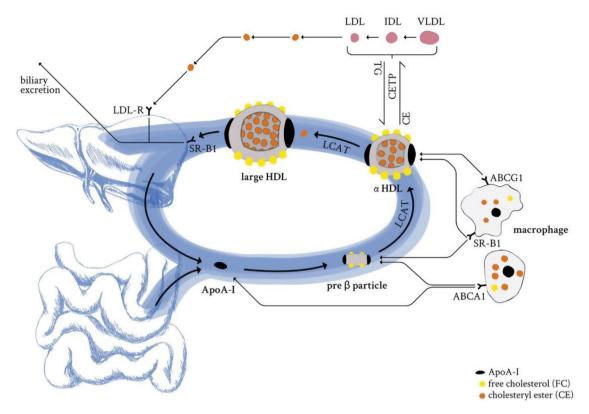


Figure 3 HDL life cycle in healthy conditions. ApoA-I from liver and the intestine is secreted into circulation and loaded with lipids by the interaction with the ABCA1 transporter (pre-β HDL). Further lipidation and conversion of free cholesterol to cholesteryl esters cause particle maturation which is accompanied with an increase in size and compositional complexity. LDL-R: low-density lipoprotein receptor; SR-B1: scavenger receptor B1; LCAT: lecithin cholesterol acyltransferase; CETP: cholesteryl ester transfer protein; TG: triglycerides; CE: cholesteryl ester; ABCG1: ATP binding cassette subfamily G member 1; ABCA1: ATP binding cassette subfamily A member 1; ApoA-I: apolipoprotein A-I; HDL: high-density lipoprotein; LDL: low-density lipoprotein; IDL: intermediate-density lipoprotein; VLDL: very low-density lipoprotein.

HDL-C measured in routine clinical analysis reflects the amount of cholesterol levels transported HDL particles but not whether they are functional.³⁷ As discussed below, the assessment of HDL function would better reflect HDL's capability to exert cardiovascular protective effects.

Other HDL constituents

Although HDL particles are primarily known for their lipidtransporting properties, they carry and deliver many other components, including hormones, carotenoids, vitamins, and ncRNAs.³⁸ The very first hormone to be identified on HDL particles was the thyroid hormone thyroxine (T4), which was shown to interact with apolipoprotein moieties (ApoA, ApoC) and to reduce binding upon increasing lipid content. 39,40 LCAT-mediated esterification of the hormone (derivates) oestrogen, pregnenolone, and dehydroepiandrosterone (fatty acid ester versions) have also been detected on HDL particles. 41-43 The physiological functions of these HDL-bound hormones remain largely unknown; they might, however, serve as precursors for cellular steroid synthesis or transcriptional regulators acting on nuclear receptor transcription factors once they have been delivered to the target cell by a potentially SR-B1-dependent mechanism.44

Furthermore, HDL has very recently been shown to be the primary transporter for carotenoids (β -carotene, zeaxanthin, and lutein in decreasing binding affinity to ApoA-I) from the liver to the retinal pigment epithelium where they protect against macular degeneration and blindness. ⁴⁵ In addition to these antioxidants, α -carotene, lycopene, and cryptoxanthin can also be transported by HDL particles. ⁴⁶ HDL particles transport more polar carotenoids (53% lutein, 39% cryptoxanthin) than nonpolar carotenoids (17% lycopene, 26% α -carotene, 22% β -carotene). ⁴⁴

Carotenes and cryptoxanthin are provitamin A carotenoids that can be converted into fat-soluble vitamin A, essential in conferring growth, vision, cell division, immunity, and reproduction. In addition, carotenes have also been demonstrated to decrease atherosclerotic risk in humans,⁴⁷ delay atherosclerosis progression,⁴⁸ and increase atherosclerosis resolution⁴⁹ in mice.

Nevertheless, vitamin A is not the only member of the fat-soluble vitamins that is transported in HDLs. Vitamin E (α -tocopherol) is probably the most important HDL-transported lipid-soluble antioxidant protecting cells and susceptible molecules from free radicals. ^{50,51} Mechanistically, vitamin E might interact with HDL-bound ApoA-I, as its transport is inhibited by ABC transporter inhibition. ⁵²

Protein	UniProt ID	Found in % of studies (HDL proteome watc project) ¹⁶¹
Actin	P60709	31
Afamin	P43652	41
Albumin	P02768	84
Alpha-1-antichymotrypsin	P01011	59
Alpha-1-antitrypsin	P01009	94
Alpha-1-microglobulin	P02760	75
Alpha-1B-glycoprotein	P04217	61
Alpha-2-antiplasmin	P08697	57
Alpha-2-HS-glycoprotein	P02765	88
Alpha-2-macroglobulin	P01023	61
Angiotensinogen	P01019	69
Antithrombin-III	P01008	55
Apolipoprotein A-I	P02647	100
Apolipoprotein A-II	P02652	96
Apolipoprotein A-IV	P06727	100
Apolipoprotein B-100	P04114	84
Apolipoprotein C-I	P02654	96
Apolipoprotein C-II	P02655	92
Apolipoprotein C-III	P02656	98
Apolipoprotein C-IV	P55056	43
Apolipoprotein D	P05090	82
	P02649	98
Apolipoprotein E		76
Apolipoprotein F	Q13790	
Apolipoprotein L1	014791	100
Apolipoprotein M	095445	92
Apolipoprotein(a)	P08519	63
C4b-binding protein alpha chain	P04003	47
Ceruloplasmin	P00450	39
Clusterin	P10909	90
Coagulation factor V	P12259	18
Complement C1r subcomponent	P00736	29
Complement C1s subcomponent	P09871	35
Complement C3	P01024	88
Complement C4-A	P0C0L4	63
Complement C4-B	P0C0L5	69
Complement C5	P01031	22
Complement component C9	P02748	59
Complement factor B	P00751	41
Complement factor H	P08603	33
Complement factor I	P05156	22
Fibrinogen alpha chain	P02671	78
Fibrinogen beta chain	P02675	55
Fibrinogen gamma chain	P02679	49
Fibronectin	P02751	41
Gelsolin	P06396	43
Haptoglobin	P00738	75
Haptoglobin-related protein	P00739	84
Hemoglobin subunit alpha	P69905	61
Hemoglobin subunit beta	P68871	65
Hemopexin	P02790	65
Heparin cofactor 2	P05546	49
Histidine-rich glycoprotein	P04196	39
Immunoglobulin heavy constant alpha 1	P04196 P01876	43

Protein	UniProt ID	Found in % of studies (HDL proteome watch project) ¹⁶¹
Immunoglobulin heavy constant gamma 1	P01857	55
Immunoglobulin heavy constant gamma 2	P01859	45
Immunoglobulin heavy constant gamma 3	P01860	29
Immunoglobulin heavy constant gamma 4	P01861	24
Immunoglobulin heavy constant mu	P01871	31
Immunoglobulin J chain	P01591	25
Immunoglobulin kappa constant	P01834	41
Immunoglobulin kappa variable 4-1	P06312	16
Immunoglobulin lambda constant 1	P0CG04	20
Immunoglobulin lambda constant 2	P0DOY2	20
Immunoglobulin lambda variable 3-21	P80748	18
Immunoglobulin lambda-like polypeptide 5	B9A064	25
Inter-alpha-trypsin inhibitor heavy chain H1	P19827	43
Inter-alpha-trypsin inhibitor heavy chain H2	P19823	49
Inter-alpha-trypsin inhibitor heavy chain H4	Q14624	65
Kallistatin	P29622	29
Kininogen-1	P01042	41
Lipopolysaccharide-binding protein	P18428	37
Phosphatidylinositol-glycan-specific phospholipase D	P80108	51
Phospholipid transfer protein	P55058	71
Plasma kallikrein	P03952	33
Plasma protease C1 inhibitor	P05155	55
Plasminogen	P00747	41
Pregnancy zone protein	P20742	24
Prothrombin	P00734	63
Retinol-binding protein 4	P02753	61
Serotransferrin	P02787	84
Serum amyloid A-1 protein	PODJI8	84
Serum amyloid A-2 protein	PODJI9	84
Serum amyloid A-4 protein	P35542	88
Serum amyloid P-component	P02743	24
Serum paraoxonase/arylesterase 1	P27169	94
Serum paraoxonase/lactonase 3	Q15166	59
Transthyretin	P02766	82
Vitamin D-binding protein	P02774	78
Vitamin K-dependent protein S	P07225	24
Vitronectin	P04004	73
Zinc-alpha-2-glycoprotein	P25311	35

Adjusted from Davidson et al.²⁸ and the HDL proteome watch project.¹⁶¹

However, while vitamin E delivery to epithelial cells has been reported to be SR-B1-independent,⁵³ delivery to endothelial cells was shown to be dependent on SR-B1.^{54,55}

On the other hand, vitamin D is not a typical HDL constituent since the serum protein vitamin D-binding protein mainly carries this vitamin. However, its carrier has been identified as part of the HDL proteome^{56,57} and can, therefore, be considered part of the HDL particle.

In recent years, HDL particles have also been shown to carry many types of small ncRNAs, amongst which microRNAs (miRNAs) are the second most abundant ncRNA class on HDL particles after ribosomal RNA-derived ncRNA. The combination and abundance of HDL-transported miRNAs is not just

a representation of the donor cells⁵⁸ but that they differ from cellular miRNAs in their post-transcriptional modification (non-templated 3'-uridylation versus -adenylation, respectively).⁵⁹ Although it is difficult to identify the cellular origin of HDL-transported miRNAs, the top-most abundant miRNAs on human HDL particles⁶⁰ have a restricted cell type-specific expression profile⁶¹: beta cells (miR-375-3p),⁶⁰ macrophages and neutrophils (miR-223-3p),^{60,62} and neurons (miR-124-3p, miR-9-5p).⁶³ So far, endothelial cells,⁶⁴⁻⁶⁶ hepatocytes,⁶⁰ and microglia⁶³ are cell types with confirmed uptake of HDL-transported miRNAs and subsequent downstream target inhibition. The mechanisms involved in miRNA delivery remain unclear, although SR-B1 has been suggested

⁹¹ most reproducible HDL-associated proteins identified with all four standard isolation methods including electrophoresis, filtration, immunoaffinity, and ultracentrifugation. Data represented in alphabetical order.

Lipid species	HDL content (mol% of total lipids)
Phospholipids	37.4-49.3
Phosphatidylcholine	32-35
PC-plasmalogen	2.2-3.5
LysoPC	1.4-8.1
Phosphatidylethanolamine	0.70-0.87
PE-plasmalogen	0.54-0.87
Phosphatidylinositol	0.47-0.76
Cardiolipin	0.077-0.201
Phosphatidylserine	0.016-0.030
Phosphatidylglycerol	0.004-0.006
Phosphatidic acid	0.006-0.009
Sphingolipids	5.7-6.9
Sphingomyelin	5.6-6.6
Ceramide	0.022-0.097
Hexosyl Cer	0.075-0.123
Lactosyl Cer	0.037-0.060
S1P d18:1	0.015-0.046
S1P d18:0	0.007
SPC d18:1	0.001
Neutral lipids	46.7-54.0
Cholesteryl esters	35-37
Free cholesterol	8.7-13.5
Triacylglycerides	2.8-3.2
Diacylglycerides	0.17-0.28
Minor lipids	
Free fatty acids	16:0, 18:0, 18:1
Isoprostane-containing PC	ND (IPGE2/D2-PC
	(36:4))

The lipids are ordered according to lipid class and abundance. PC: phosphatidylcholine; PE: phosphatidylethanolamine; S1P: sphingosine-1-phosphate; SPC: sphingosylphosphorylcholine.

to play a key role in miRNA-HDL loading. ^{60,64} As such, global SR-B1 deficiency has been shown to be associated with a lack of accumulation of miRNAs in circulating HDL particles. ^{59,60} Besides intensive research to identify the cross-talk between HDL-related miRNAs and cells efforts are being made to establish HDL-transported miRNAs as biomarkers for CVD. ⁶⁷

Cardiovascular protective functions of the HDL particle

Since the ground-breaking Framingham Heart Study in the 1960–1980s, ⁶⁸ several large-scale observational and experimental studies have accredited HDL particles with beneficial properties in the CV system. Although the hallmark protective of HDL particles is their ability to efflux cholesterol and mediate reverse cholesterol transport (RCT), ^{69,70} they possess further athero- and cardio-protective functions, including protection against oxidative and inflammatory damage, ^{71–74} inhibition of thrombosis, ^{75,76} cardioprotection, ⁷⁷ as well as induction of NO synthesis ^{72,78} and endothelial cell renewal. ^{72,79}

Reverse cholesterol transport

HDL particles are the key molecules in reverse cholesterol transport, as they function as cholesterol acceptors, carriers for circulation, and deliverers to the liver for biliary excretion. As such, HDL particles facilitate cholesterol efflux from peripheral tissues, including atherosclerotic foam cells, in various ways. Upon normal cholesterol levels, passive agueous diffusion of free cholesterol between membranes and HDL particles occurs following the concentration gradient. 80 Free cholesterol is translocated into the particle core by LCAT-dependent transformation into cholesteryl esters to maintain the efflux from the cells to the HDL particle. In addition, SR-B1-dependent non-aqueous efflux may occur, a pathway promoted by low total HDL levels and large HDL particles.⁸¹ In conditions with elevated cellular cholesterol concentrations (e.g., foam cells), active cholesterol efflux pathways dependent on the ABC transporter family members A1 and G1 are activated. 80 Further LCAT activity leads to particle maturation and cholesteryl ester enrichment in the lipid core.

Antioxidant and anti-inflammatory capacity

Oxidative stress (ROS, oxLDL) and a pro-inflammatory environment (cytokines, chemokines, and endothelial adhesion molecules) drive atherosclerosis initiation and progression. HDL particles provide potent atheroprotection and prevent LDLs lipid oxidation (HDL3 > HDL2) by transferring those oxidised lipids to the HDL particle, where redox-active ApoA-I residues can inactivate them. In addition, HDL particles carry multiple antioxidant enzymes such as serum paraoxonase 1 (PON1), lipoprotein-associated phospholipid A2, and LCAT. PON1 is the most investigated HDL-associated enzyme and equally protects HDL and LDL particles by hydrolysing lactones, phosphate esters, and lipid peroxide derivates. The presence or interaction with ApoA-I seems crucial since particles containing ApoA-II or -IV have shown significantly less antioxidative protection. Sc. 87

Interestingly, PON1 has also been shown to play a role in the classical activation of eNOS through the ApoA-I/SR-B1 axis. Representation of eNOS activation by S1P3 signalling. Representation by S1P3 signalling. Representation by converging the non-receptor tyrosine kinase Src-activated Akt and MAP kinases.

HDL particles also have a crucial role in inhibiting the expression of the inflammatory adhesion molecules VCAM-1 and ICAM-1,90 E-selectin,91 and MCP-192 on activated endothelial cells thereby preventing the recruiting and extravasation of immune cells into the intimal space. The suggested molecular mechanisms behind are HDL-S1P-dependent and involve the interference with NFkB signalling $(S1P_1/\beta$ -arrestin2 axis⁹³) and a reduction in endothelial apoptosis and TNF-mediated inflammatory signalling (P13K/AKT/eNOS axis^{94,95}). HDL-S1P has also been reported to inhibit macrophage apoptosis through JAK2/STAT3 signalling.96 Whereas HDL-related inhibition of endothelial inflammatory surface molecules seems independent of variations in HDL size and composition of apolipoproteins, cholesteryl ester, and triglycerides, 97 phospholipid subspecies have shown to exert antiinflammatory

effects⁹⁸ when protected from oxidation by either ApoA-I or other antioxidants.⁹⁹

Antithrombotic effects

HDL particles exert several antithrombotic effects that may protect against atherothrombotic clinical events. As such, HDL particles promote blood flow by increased NO and prostacyclin synthesis with subsequent vasodilation. HDL also prevent endothelial cell activation by inhibiting endothelial cell apoptosis⁸⁹ and the expression of the prothrombotic factors P-selectin,¹⁰⁰ E-selectin,¹⁰⁰ and tissue factor.¹⁰¹ NO and prostacyclin also inhibit platelet activation and aggregation,¹⁰² which, together with their vasodilator properties, counterbalance restricted blood flow. In addition, HDL particles have been reported to upregulate endothelial thrombomodulin,¹⁰³ an anticoagulant factor that inhibits thrombin formation.

Cardioprotection: protection against ischemia-reperfusion injury

While cardiac tissue and performance are more imminently threatened during ischemia, 104 reperfusion – although indispensable for saving ischemic heart tissue from cell death - further damages the injured heart tissue. 105 Consequently, the total damage of ischemia-reperfusion injury (IRI) is the sum of the ischemic insult (restricted or interrupted blood flow) and reperfusion (restored blood flow and re-oxygenation). 106 Early experimental studies in rodents reported that intravenous infusions of HDL particles strongly reduced infarct size^{77,107} and improved post-ischemic cardiac function. 108 The mechanisms behind involve the reduction in apoptosis execution through activation of the reperfusion injury signalling kinase (RISK) and survivor activating factor enhancement (SAFE) signalling pathways¹⁰⁹ (Fig. 4). The RISK pathway (P13K/AKT/eNOS axis) protects endothelial cells from pro-inflammatory TNF α signalling, whereas the SAFE pathway is likely to be activated by HDL-bound S1P¹¹⁰ with TNF α and STAT3 downstream signalling effectors. 111 STAT3 activation is involved in the early protection phase by preventing the opening of the mitochondrial permeability transition pore and subsequent apoptosis execution, and in the late protective phase, in which STAT3 phosphorylation leads to expression of target genes, eventually reducing oxidative stress, apoptosis, and IRI¹¹⁰ (Fig. 4). Furthermore, HDL-S1P signalling also prevents cardiomyocyte cell death and significantly reduces infarct size by phosphorylating connexin 43 (Cx43), the main protein of cardiomyocyte Cx43 gap junction channels¹¹² (Fig. 4).

Evidence also suggests that HDL-transported miRNAs are involved in protection against IRI. As such, inhibition of miR-92a, a miRNA reported to be carried by HDL particles, has been shown in small and large animal models to reduce infarct size and improve post-ischemic cardiac function. 113-116 Likewise, miR-486 has also been shown to attenuate IRI by reducing cardiomyocyte apoptosis *in vitro* 117,118 and *in vivo* 118 and also mediate beneficial effects on exercise-induced myocardial protection. 118 Furthermore, recent advances also suggest protective effects for the presence of miR-146a 119,120 and miR-125, 121,122 both part of an HDL-miRNA profile identified in coronary artery disease patients. 67

Endothelial protection

The endothelium forms a semipermeable barrier that separates blood-carried factors and cells from the surrounding tissue¹²³ and its integrity is essential for sustaining vascular homeostasis, including the modulation of vascular tone and trafficking of substances between systemic circulation and adjacent tissues. 124 HDL particles support endothelial monolayer integrity by promoting junction closure, 125 preventing loss of glycocalyx (proteoglycan shedding), 126 reducing endothelial cell apoptosis, 110 and stimulating endothelial cell proliferation and migration of both mature endothelial cells¹²⁷ as well as endothelial progenitor cells. 128 As part of their antiapoptotic properties, HDL particles prevent sustained increases of intracellular calcium, 89 the activation of caspases 3 and $9^{129,130}$ (TNF α - and growth factor deprivation-mediated apoptosis), and modulate S1P-dependent activation of PI3K/AKT^{94,110,131} and ERK1/2.¹³² On the other hand, endothelial cell proliferation and migration are crucial for neovascularisation and reendothelisation after vascular injury. 127 HDL stimulation of endothelial cell migration and proliferation is mediated by multiple kinase cascades involving SR-B1-dependent activation of MAPK/Rac1¹³³ (migration and re-endothelialisation), PI3K/AKT¹³⁴ (HIF1 α mediated angiogenesis), as well as S1P1/3-dependent activation of ERK¹³⁵ (endothelial cell survival), PI3K/AKT, 135 p38 MAPK, 135 Rho/Rho kinase 135 (endothelial cell migration), p42/44 MAPK/Ras¹³⁶ (tube formation), AKT¹²⁵ and AMPK¹³⁷ (endothelial barrier integrity), NFκB^{90,95} PI3K/AKT/eNOS¹¹⁰ (anti-inflammatory, decrease in surface expression of VCAM-1, ICAM-1, E-selectin, MCP-1), and VEGFR2¹³⁸ (angiogenesis) (Fig. 5). As interference with the S1P/S1P_{1/3} axis mimics effects seen by interference with SR-B1, HDL particles were suggested to primarily interact with the SR-B1 receptor on endothelial cells, which secondarily leads to spatial proximity between HDL-bound S1P and endothelial S1P_{1/3} receptors. As stated above, endothelial cells are important SR-B1-dependent inducers of NO and prostacyclin synthesis, which strongly contribute to PDZK1/SRC/AKT/MAPK/ eNOS-139 and PDZK1/AMPK/PI3K/AKT/COX-2/prostacyclin-140 as well as SphK2/COX-2/prostacyclin¹⁴¹-mediated vasodilation (Fig. 5).

The impact of comorbidities on HDL-related cardiovascular protection

The concept that raising HDL-C would promote protection against CVD was developed from observing several large epidemiological studies that evidenced that low HDL-C levels strongly correlated with future CV events in healthy individuals without baseline CVD.^{68,142–144} However, doubts arose when extremely high HDL-C levels were paradoxically shown to increase all-cause and CV mortality in both men and women¹⁴⁵ and Mendelian randomisation studies failed to confirm a causative interaction between increased CVD risk and HDL-C.¹⁴⁶ Furthermore, extensive efforts to pharmacologically raise HDL-C levels through several randomized clinical trials (i.e., niacin, CETP inhibitors) failed to reduce CV events in secondary prevention.^{147–150}

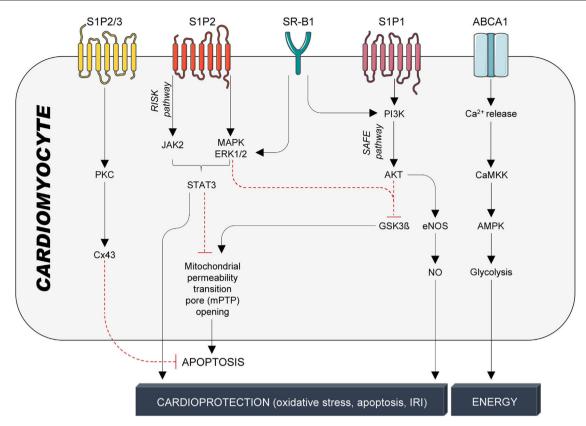


Figure 4 HDL-mediated protective signalling in cardiomyocytes. Protective signalling in cardiomyocytes is mediated through the G-protein coupled receptors S1P1-3 and the cholesterol transporters SR-B1 and ABCA1. S1P1-3 are activated by the interaction with S1P, which is bound to HDL particles by its interaction with ApoM. SR-B1 and ABCA1, on the other hand, are activated by binding to ApoA-I, the main structural protein of HDL particles and the main inducer of active cholesterol efflux. HDL particles confer cardioprotection in cardiomyocytes through a multitude of kinase cascades that reduce stressors, prevent cardiomyocyte apoptosis, and, therefore, improve infarct size, cardiac function, and overall outcome. S1P1-3: Sphingosine-1-phosphate receptors 1-3; SR-B1: scavenger receptor B1; ABCA1: ATP-binding cassette transporter A1; PKC: protein kinase C; Cx43: connexin 43; JAK2; Janus kinase 2; MAPK: mitogen-activated protein kinase; ERK1/2: extracellular signal-regulated kinase 1/2; STAT3: signal transducer and activator of transcription 3; Pl3K: phosphatidylinositol-4,5-bisphosphate 3-kinase; AKT: serine/threonine kinase 1; GSK3β: inosine/guanosine kinase 3β; eNOS: endothelial nitric oxide synthase; NO: nitric oxide; CaMKK: calcium/calmodulin-activated protein kinase; AMPK: adenosine monophosphate-activated protein kinase.

The failure of the cetrapib trials

The efficacy of several CETP inhibitors (torcetrapib, dalcetrapib, evacetrapib, and anacetrapib) was tested in four large-scale phase-III clinical trials^{148–151} on top of statin treatment. The hypothesis was that while statins would limit atherosclerotic disease progression, HDL-C-raising treatment would simultaneously work on reversing the existing atherosclerotic plaques. Besides, CETP inhibitors would also minimise the residual CV risk in high-risk patients on statins with successfully lowered LDL-C levels but low HDL-C levels.

The first trial was the ILLUMINATE (Investigation of Lipid Level Management to Understand its Impact in Atherosclerotic Events) trial, which administered torcetrapib in combination with atorvastatin to 15,067 patients for 12 months. ¹⁵⁰ Despite significantly increased HDL-C (70%) and decreased LDL-C (25%), torcetrapib treatment elevated the risk for major adverse CV events (MACE) and CVD-related as well as unrelated deaths compared to the placebo group. ¹⁵⁰ As a result, the trial was terminated prematurely.

In the dal-OUTCOME (Study of RO4607381 in Stable Coronary Heart Disease Patients With Recent Acute Coronary

Syndrome) trial, dalcetrapib or placebo was administered to patients on statins and with a recent acute coronary syndrome for a mean of 31 months. 149 Despite a significant increase in HDL-C levels by 31–40% and fewer adverse effects than torcetrapib, the trial was terminated due to futility as no significant reduction in hard primary endpoints was detected. Nevertheless, later studies found that a minority of 17% of the cohort significantly benefited from dalcetrapib treatment (39% reduction in CV endpoints) due to a homozygous single nucleotide morphism in the adenylate cyclase nine gene. 152

The ACCELERATE (Assessment of Clinical Effects of Cholesteryl Ester Transfer Protein Inhibition with evacetrapib in Patients at a High Risk for Vascular Outcomes) trial administered anacetrapib or placebo to 12,092 patients on statins and with high-risk vascular disease (over 63% of the patients presented with diabetes) for a mean period of 26 months. 148 Despite the massive change in lipoprotein cholesterol levels (HDL-C +133%; LDL-C -31%) and significantly lower incidence rates of death from any cause in anacetrapib-treated patients, the ACCELERATE trial did not

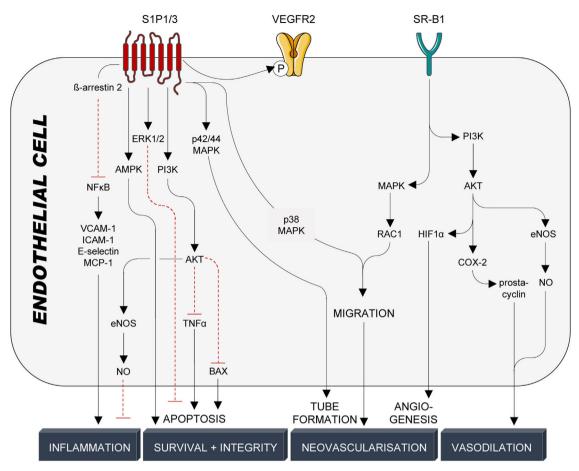


Figure 5 HDL-mediated protective signalling in endothelial cells. Protective signalling in endothelial cells is mediated through the G-protein coupled receptors S1P1/3 and the cholesterol transporter SR-B1. S1P1/3 are activated by the interaction with S1P, which is bound to HDL particles by its interaction with ApoM. SR-B1, on the other hand, is activated by binding to ApoA-I, the main structural protein of HDL particles and the main inducer of active cholesterol efflux. HDL particles confer cardioprotection in endothelial cells through a multitude of kinase cascades, NF κ B and VEGFR2, that prevent endothelial cell activation and inflammation, promote endothelial cell survival and barrier integrity and induce tube formation, migration, and angiogenesis. Production and release of NO and prostacyclin from endothelial cells also trigger vascular smooth muscle cell relaxation and subsequent vasodilation. S1P1/3: sphingosine-1-phosphate receptors 1/3; VEGFR2: vascular endothelial growth factor receptor 2; SR-B1: scavenger receptor B1; NF κ B: nuclear factor κ B; VCAM-1: vascular cell adhesion molecule 1; ICAM-1: intracellular cell adhesion molecule 1; MCP-1: monocyte chemoattractant protein 1; AMPK: adenosine monophosphate-activated protein kinase; ERK1/2: extracellular signal-regulated kinase 1/2; P13K: phosphatidylinositol-4,5-bisphosphate 3-kinase; AKT: serine/threonine kinase 1; TNF κ : tumor necrosis factor κ ; BAX: Bcl-2-associated X protein; eNOS: endothelial nitric oxide synthase; NO: nitric oxide; MAPK: mitogen-activated protein kinase; RAC1: Ras-related C3 botulinum toxin substrate 1; HIF1 κ : hypoxia-inducible factor 1 κ ; COX-2: cyclooxygenase-2.

lead to reduced primary CV endpoints. It was terminated after two years of follow-up due to a lack of efficacy.

Finally, the REVEAL (Randomized Evaluation of the Effects of Anacetrapib through Lipid-modification) trial was the longest and largest trial, with 30,449 patients on atorvastatin and with established atherosclerotic CVD receiving anacetrapib or placebo for a median period of over four years. ¹⁵¹ Anacetrapib strongly modified the lipid profile (HDL-C +104%, LDL-C -41%) and was the first to report modest but significantly reduced primary endpoint rates, including coronary death, myocardial infarction, and coronary revascularisation in anacetrapib-treated patients (10.8% vs 11.8% in placebo, p = 0.004). However, the benefit was small and only significant when combining data from all years or years > 1, which questioned the usefulness for

high-risk patients needing short-term solutions. In the end, the reduction in the primary CV endpoint was attributed to the drop in LDL-C levels rather than the rise of HDL-C.

The results of the trials above failed to meet the expectations for therapeutically raising HDL-C levels in high-risk patients on statin treatment. Furthermore, a meta-analysis of over 113,000 statin-treated patients in 39 trials involving the HDL-C-raising therapies niacin, fibrates, and CETP inhibitors demonstrated no difference in CV events between those patients on these HDL-C-related therapies and placebo controls. ¹⁴⁷ The beneficial effect that was provided by lowering LDL-C (by statin or otherwise) was strongly diminished by very high HDL-C (in putatively dysfunctional HDL), as seen in the REVEAL trial and suggested for both the ACCELERATE and dal-OUTCOME trials. It is essential to consider patients'

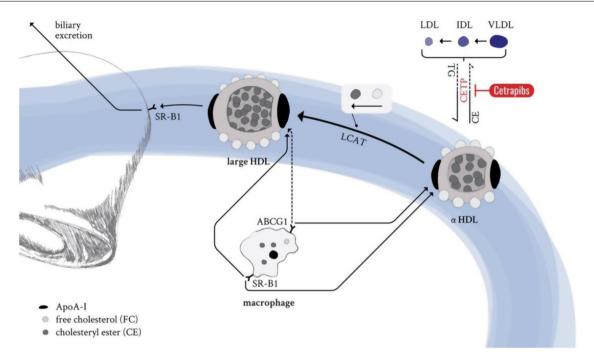


Figure 6 Mechanism of CETP inhibitors (cetrapibs). The cholesteryl ester transfer protein (CETP) is an enzyme that catalyzes the transport of cholesteryl esters and triglycerides between lipoproteins. Delivery of HDL-transported cholesteryl ester to ApoB-containing particles (LDLs/IDLs/VLDLs) is mediated by CETP in exchange for triglycerides. This process is selectively blocked by the drug family of CETP inhibitors, also known as cetrapibs, and leads to the formation of large, cholesteryl ester-enriched HDL particles. These particles lack efficient interaction with the ABC transporter family, the major mediators of cholesterol efflux from macrophages to HDL particles, subsequently reducing (cardio-) vascular protection via reverse cholesterol transport. ABCG1: ATP-binding cassette transporter G1; SR-B1: scavenger receptor B1; LCAT: lecithin cholesteryl acetyltransferase; CE: cholesteryl ester; HDL: high-density lipoprotein; LDL: low-density lipoprotein; IDL: intermediate-density lipoprotein; VLDL: very low-density lipoprotein; LDL-R: LDL receptor.

additional CV risk factors and comorbidities on top of dyslipidaemia. In this regard, besides hypertension and chronic kidney disease, two out of four CETP inhibitors promoted *de novo* onset of diabetes, likely increasing the patient's risk.

In summary, cetrapibs were developed assuming that increasing HDL-C would translate into enhanced hepatic cholesterol excretion and subsequent cardio/vascular protection. CETP inhibitors block cholesteryl ester transport from HDL to VLDL, IDL, and LDL particles to enhance the amount of cholesterol reaching the liver (Fig. 6). However, in turn, CETP inhibition leads to the formation of large cholesteryl ester-enriched particles that, in contrast to the smaller and denser HDL subfractions (i.e., HDL3), have been shown to not interact efficiently with ABCA1, the major mediator of cholesterol efflux from macrophages to HDLs¹⁵³ (Fig. 6).

Pioneering studies from our group demonstrated that administration of cholesterol-poor HDL particles (equivalent to HDL3 particles) isolated from healthy animals limited further lipid deposition and even regressed atherosclerotic lesions. These and other experimental studies have supported the 'HDL quality over quantity hypothesis'. The scientific field has accordingly redirected its efforts towards deciphering HDL structure and function to restore HDL-related CV protection, particularly since multiple studies have suggested that pathophysiologic conditions adversely remodel HDL particles [reviewed in 30,82,156], thus impairing HDL protective effects. 82,156 In this regard, we

have demonstrated in a highly translatable pig model that diet-induced hypercholesterolemia, one of the most prevalent CV risk factors, render HDL particles dysfunctional and induce compositional changes at a proteomic, lipidomic and miRNA level. ¹⁰⁸ Interestingly, we have also evidenced that such adverse HDL remodelling can be reversed and cardio-vascular protection restored by feeding a low-fat diet and decreasing LDL-C levels back to the physiological state. ¹⁵⁷

Conclusions

The landmark Framingham Heart Study evidenced an inverse association between CV risk and HDL-C plasma levels, fostering experimental research towards deciphering the beneficial properties of HDL particles. However, excitement towards the therapeutic potential of HDL-C-raising drugs faded with the disappointing outcomes of the cetrapib trials. These data evidenced no clear benefit of increasing HDL-C in patients with established CVD in secondary prevention and, most importantly, questioned whether HDL-C was the adequate parameter to reflect HDL-related protection. 158 HDL is a complex and heterogeneous particle with many components that exert various cellular and molecular functions beyond cholesterol transport and removal. 82,156 The presence of CV risk factors and comorbidities have been shown to remodel HDL particles towards a dysfunctional state, yet, experimental evidence supports the ability to

restore HDL protective function by implementing healthy habits re-opening the door for new considerations in CVD prevention. ¹⁵⁷ Further research is warranted to identify how these particles can rise to their former glory and maintain their protective phenotype in primary and secondary prevention.

Funding

This work was supported by the Spanish Society of Atherosclerosis/Atherosclerosis Spanish Foundation (Research Grant SEA/FEA 2019), and PID2021-1288910B-(to GV), PID2019-107160RB-I00 (to LB), PLEC2021-007664 NextGenerationEU (to GV) funded by MCIN/AEI/10.13039/501100011033 and Fondo Europeo de Desarrollo Regional (FEDER) A way of making Europe; the Instituto de Salud Carlos III [CIBERCV CB16/11/00411 to LB]; the Generalitat of Catalunya-Secretaria d'Universitats i Recerca del Departament d'Economia i Coneixement de la Generalitat [2017SGR1480 to LB] and 2016PROD00043 (Agencia Gestión Ayudas Universitarias Investigación: AGAUR), CERCA programme/Generalitat de Cataluña, and Fundación Investigación Cardiovascular - Fundación Jesús Serra for their continuous support.

Conflict of interest

The authors have no conflict of interest to declare related to this manuscript.

References

- Roth GA, Mensah GA, Johnson CO, Addolorato G, Ammirati E, Baddour LM, et al. Global burden of cardiovascular diseases and risk factors, 1990–2019: update from the GBD 2019 study. J Am Coll Cardiol. 2020;76:2982–3021.
- 2. Fuster V, Narula J, Vaishnava P, Leon BM, Callans DJ, Rumsfeld JS, et al. In: Fuster V, Narula J, Vaishnava P, Leon BM, Callans DJ, Rumsfeld JS, et al., editors. Fuster and Hurst's the heart. Mc Graw Hill: 2022. p. 1104.
- 3. Vos T, Lim SS, Abbafati C, Abbas KM, Abbasi M, Abbasifard M, et al. Global burden of 369 diseases and injuries in 204 countries and territories, 1990–2019: a systematic analysis for the Global Burden of Disease Study 2019. Lancet. 2020;396:1204–22.
- Schwingshackl L, Knüppel S, Michels N, Schwedhelm C, Hoffmann G, Iqbal K, et al. Intake of 12 food groups and disability-adjusted life years from coronary heart disease, stroke, type 2 diabetes, and colorectal cancer in 16 European countries. Eur J Epidemiol. 2019;34:765–75.
- 5. Freedman DS, Srinivasan SR, Shear CL, Hunter SM, Croft JB, Webber LS, et al. Cigarette smoking initiation and longitudinal changes in serum lipids and lipoproteins in early adulthood: the Bogalusa Heart Study. Am J Epidemiol. 1986;124:207–19.
- Nielsen VG, Hafner DT, Steinbrenner EB. Tobacco smokeinduced hypercoagulation in human plasma: role of carbon monoxide. Blood Coagul Fibrinolysis Int J Haemost Thromb. 2013;24:405–10.
- 7. GBD 2019 Demographics Collaborators. Global age-sex-specific fertility, mortality, healthy life expectancy (HALE), and population estimates in 204 countries and territories, 1950–2019: a comprehensive demographic analysis for the Global Burden of Disease Study 2019. Lancet Lond Engl. 2020;396:1160–203.

- 8. Danaei G, Finucane MM, Lu Y, Singh GM, Cowan MJ, Paciorek CJ, et al. National, regional, and global trends in fasting plasma glucose and diabetes prevalence since 1980: systematic analysis of health examination surveys and epidemiological studies with 370 country-years and 2.7 million participants. Lancet. 2011;378:31–40.
- Gallucci G, Tartarone A, Lerose R, Lalinga AV, Capobianco AM. Cardiovascular risk of smoking and benefits of smoking cessation. J Thorac Dis. 2020;12:3866-76.
- Jebari-Benslaiman S, Galicia-García U, Larrea-Sebal A, Olaetxea JR, Alloza I, Vandenbroeck K, et al. Pathophysiology of atherosclerosis. Int J Mol Sci. 2022;23. Available from: http://www.ncbi.nlm.nih.gov/pubmed/35328769 [cited 2.8.22].
- 11. Chiu JJ, Chien S. Effects of disturbed flow on vascular endothelium: pathophysiological basis and clinical perspectives. Physiol Rev. 2011;91:327–87.
- Dessalles CA, Leclech C, Castagnino A, Barakat AI. Integration of substrate- and flow-derived stresses in endothelial cell mechanobiology. Commun Biol. 2021;4:764.
- 13. Lemaster E, Huang RT, Zhang C, Bogachkov Y, Coles C, Shentu TP, et al. Pro-atherogenic flow increases endothelial stiffness via enhanced CD36-mediated oxLDL uptake HHS Public Access. Arter Thromb Vasc Biol. 2018;38:64–75.
- 14. Borén J, Chapman MJ, Krauss RM, Packard CJ, Bentzon JF, Binder CJ, et al. Low-density lipoproteins cause atherosclerotic cardiovascular disease: pathophysiological, genetic, and therapeutic insights: a consensus statement from the European Atherosclerosis Society Consensus Panel. Eur Heart J. 2020;41:2313–30.
- **15.** Chiu JJ, Usami S, Chien S. Vascular endothelial responses to altered shear stress: pathologic implications for atherosclerosis. Ann Med. 2009;41:19–28.
- 16. Zhao Y, Ren P, Li Q, Umar SA, Yang T, Dong Y, et al. Low shear stress upregulates CX3CR1 expression by inducing VCAM-1 via the NF-κB pathway in vascular endothelial cells. Cell Biochem Biophys. 2020;78:383-9.
- 17. Chiu JJ, Chen CN, Lee PL, Yang CT, Chuang HS, Chien S, et al. Analysis of the effect of disturbed flow on monocytic adhesion to endothelial cells. J Biomech. 2003;36:1883–95.
- Chen Z, Tzima E. PECAM-1 is necessary for flowinduced vascular remodeling. Available from: http://www.transonic.com/workbook.shtml [cited 2.8.22].
- Gimbrone MA, García-Cardeña G. Endothelial cell dysfunction and the pathobiology of atherosclerosis. Circ Res. 2016;118:620–36.
- Jang E, Robert J, Rohrer L, von Eckardstein A, Lee WL. Transendothelial transport of lipoproteins. Atherosclerosis. 2020;315:111–25.
- 21. Colin S, Chinetti-Gbaguidi G, Staels B. Macrophage phenotypes in atherosclerosis. Immunol Rev. 2014;262:153–66.
- 22. Grootaert MOJ, Bennett MR. Vascular smooth muscle cells in atherosclerosis: time for a re-assessment. Cardiovasc Res. 2021:117:2326–39.
- 23. Kockx MM. Apoptosis in the atherosclerotic plaque: quantitative and qualitative aspects. Arterioscler Thromb Vasc Biol. 1998;18:1519–22.
- 24. Chen Y, Zhao X, Wu H. Arterial stiffness: a focus on vascular calcification and its link to bone mineralization. Arterioscler Thromb Vasc Biol. 2020;40:1078–93.
- 25. Lakatta EG. Arterial and cardiac aging: major shareholders in cardiovascular disease enterprises: Part III: Cellular and molecular clues to heart and arterial aging. Circulation. 2003;107:490–7.
- Vergallo R, Crea F. Atherosclerotic plaque healing. N Engl J Med. 2020;383:846–57.
- 27. Zhang Y, Gordon SM, Xi H, Choi S, Paz MA, Sun R, et al. HDL subclass proteomic analysis and functional implication of

- protein dynamic change during HDL maturation. Redox Biol. 2019;24:101222.
- 28. Davidson WS, Shah AS, Sexmith H, Gordon SM, The HDL. Proteome watch: compilation of studies leads to new insights on HDL function. Biochim Biophys Acta Mol Cell Biol Lipids. 2022;1867:159072.
- 29. Chapman MJ. HDL functionality in type 1 and type 2 diabetes: new insights. Curr Opin Endocrinol Diabetes Obes. 2022;29:112–23.
- 30. von Eckardstein A, Kardassis D. In: von Eckardstein A, Kardassis D, editors. Handbook of experimental pharmacology high density lipoprotein from biological understanding to clinical exploitation. Springer Open; 2015. p. 694.
- 31. Kontush A, Therond P, Zerrad A, Couturier M, Négre-Salvayre A, de Souza JA, et al. Preferential sphingosine-1-phosphate enrichment and sphingomyelin depletion are key features of small dense HDL3 particles: relevance to antiapoptotic and antioxidative activities. Arterioscler Thromb Vasc Biol. 2007;27:1843-9.
- 32. de Souza JA, Vindis C, Nègre-Salvayre A, Rye KA, Couturier M, Therond P, et al. Small, dense HDL 3 particles attenuate apoptosis in endothelial cells: pivotal role of apolipoprotein A-I. J Cell Mol Med. 2010;14:608–20.
- 33. de Souza JA, Vindis C, Hansel B, Nègre-Salvayre A, Therond P, Serrano CV, et al. Metabolic syndrome features small, apolipoprotein A-I-poor, triglyceride-rich HDL3 particles with defective anti-apoptotic activity. Atherosclerosis. 2008;197:84–94.
- 34. Christoffersen C, Obinata H, Kumaraswamy SB, Galvani S, Ahnström J, Sevvana M, et al. Endothelium-protective sphingosine-1-phosphate provided by HDL-associated apolipoprotein M. Proc Natl Acad Sci USA. 2011;108:9613–8.
- 35. Davidson WS, Silva RAGD, Chantepie S, Lagor WR, Chapman MJ, Kontush A. Proteomic analysis of defined HDL subpopulations reveals particle-specific protein clusters: relevance to antioxidative function. Arterioscler Thromb Vasc Biol. 2009;29:870–6.
- **36.** Levkau B. HDL-S1P: cardiovascular functions, disease-associated alterations, and therapeutic applications. Front Pharmacol. 2015;6:243.
- 37. Kontush A, Lhomme M, Chapman MJ. Unraveling the complexities of the HDL lipidome. J Lipid Res. 2013;54:2950–63.
- Ben-Aicha S, Badimon L, Vilahur G. Advances in HDL: much more than lipid transporters. Int J Mol Sci. 2020;21. Available from: http://www.ncbi.nlm.nih.gov/pubmed/31979129 [cited 22.9.20].
- 39. Benvenga S, Cahnmann HJ, Gregg RE, Robbins J. Characterization of the binding of thyroxine to high density lipoproteins and apolipoproteins A-I. J Clin Endocrinol Metab. 1989;68:1067–72.
- 40. Benvenga S, Cahnmann HJ, Rader D, Kindt M, Facchiano A, Robbins J. Thyroid hormone binding to isolated human apolipoproteins A-II, C-I, C-II, and C-III: homology in thyroxine binding sites. Thyroid Off J Am Thyroid Assoc. 1994;4:261–7.
- 41. Höckerstedt A, Jauhiainen M, Tikkanen MJ. Estradiol fatty acid esterification is increased in high density lipoprotein subclass 3 isolated from hypertriglyceridemic subjects. Atherosclerosis. 2006;185:264–70.
- **42.** Leszczynski DE, Schafer RM, Perkins EG, Jerrell JP, Kummerow FA. Esterification of dehydroepiandrosterone by human plasma HDL. Biochim Biophys Acta. 1989;1014:90–7.
- Provost PR, Lavallée B, Bélanger A. Transfer of dehydroepiandrosterone- and pregnenolone-fatty acid esters between human lipoproteins. J Clin Endocrinol Metab. 1997;82:182-7.
- **44.** Vickers KC, Remaley AT. HDL and cholesterol: life after the divorce? J Lipid Res. 2014;55:4–12.

- **45.** Li B, Vachali P, Chang FY, Gorusupudi A, Arunkumar R, Shi L, et al. HDL is the primary transporter for carotenoids from liver to retinal pigment epithelium in transgenic ApoA-I—/—/Bco2—/— mice. Arch Biochem Biophys. 2022;716:109111.
- **46.** Cardinault N, Abalain JH, Sairafi B, Coudray C, Grolier P, Rambeau M, et al. Lycopene but not lutein nor zeaxanthin decreases in serum and lipoproteins in age-related macular degeneration patients. Clin Chim Acta Int J Clin Chem. 2005;357:34–42.
- 47. D'Odorico A, Martines D, Kiechl S, Egger G, Oberhollenzer F, Bonvicini P, et al. High plasma levels of α and β -carotene are associated with a lower risk of atherosclerosis: Results from the Bruneck study. Atherosclerosis. 2000;153:231–9.
- **48.** Zhou F, Wu X, Pinos I, Abraham BM, Barrett TJ, von Lintig J, et al. β-Carotene conversion to vitamin A delays atherosclerosis progression by decreasing hepatic lipid secretion in mice. J Lipid Res. 2020;61:1491–503.
- Albakri A, Coronel J, Tamane S, Black M, Fisher E, Amengual J. β-Carotene enhances atherosclerosis resolution in a reversible murine model of atherosclerosis. Curr Dev Nutr. 2021;5 Suppl. 2:68
- Qian J, Morley S, Wilson K, Nava P, Atkinson J, Manor D. Intracellular trafficking of vitamin E in hepatocytes: the role of tocopherol transfer protein. J Lipid Res. 2005;46:2072–82.
- 51. Carr T, Traber M, Haines J, Kayden H, Parks J, Rudel L. Interrelationships of alpha-tocopherol with plasma lipoproteins in African green monkeys: effects of dietary fats. J Lipid Res. 1993;34:1863–71.
- Anwar K, Iqbal J, Hussain MM. Mechanisms involved in vitamin E transport by primary enterocytes and in vivo absorption. J Lipid Res. 2007:48:2028–38.
- 53. Akanuma S, Yamamoto A, Okayasu S, Tachikawa M, Hosoya K. High-density lipoprotein-associated alpha-tocopherol uptake by human retinal pigment epithelial cells (ARPE-19 Cells): the irrelevance of scavenger receptor class B, type I. Biol Pharm Bull. 2009;32:1131–4.
- 54. Tachikawa M, Okayasu S, Hosoya K. Functional involvement of scavenger receptor class B, type I, in the uptake of alphatocopherol using cultured rat retinal capillary endothelial cells. Mol Vis. 2007;13:2041–7.
- 55. Balazs Z, Panzenboeck U, Hammer A, Sovic A, Quehenberger O, Malle E, et al. Uptake and transport of high-density lipoprotein (HDL) and HDL-associated alpha-tocopherol by an in vitro blood-brain barrier model. J Neurochem. 2004;89:939–50.
- **56.** Vaisar T, Pennathur S, Green PS, Gharib SA, Hoofnagle AN, Cheung MC, et al. Shotgun proteomics implicates protease inhibition and complement activation in the antiinflammatory properties of HDL. J Clin Invest. 2007;117:746–56.
- 57. Alwaili K, Bailey D, Awan Z, Bailey SD, Ruel I, Hafiane A, et al. The HDL proteome in acute coronary syndromes shifts to an inflammatory profile. Biochim Biophys Acta. 2012;1821:405–15.
- 58. Sedgeman LR, Beysen C, Ramirez Solano MA, Michell DL, Sheng Q, Zhao S, et al. Beta cell secretion of miR-375 to HDL is inversely associated with insulin secretion. Sci Rep. 2019;9:3803.
- Allen RM, Zhao S, Ramirez Solano MA, Zhu W, Michell DL, Wang Y, et al. Bioinformatic analysis of endogenous and exogenous small RNAs on lipoproteins. J Extracell Ves. 2018;7:1506198.
- **60.** Vickers KC, Palmisano BT, Shoucri BM, Shamburek RD, Remaley AT. MicroRNAs are transported in plasma and delivered to recipient cells by high-density lipoproteins. Nat Cell Biol. 2011;13:423–33.
- **61.** Vickers KC, Michell DL. HDL-small RNA export, transport, and functional delivery in atherosclerosis. Curr Atheroscler Rep. 2021;23:38.

- 62. Cuesta Torres LF, Zhu W, Öhrling G, Larsson R, Patel M, Wiese CB, et al. High-density lipoproteins induce miR-223-3p biogenesis and export from myeloid cells: role of scavenger receptor BI-mediated lipid transfer. Atherosclerosis. 2019;286:20–9.
- 63. Veremeyko T, Kuznetsova IS, Dukhinova MWY, Yung A, Kopeikina E, Barteneva NS, et al. Neuronal extracellular microRNAs miR-124 and miR-9 mediate cell-cell communication between neurons and microglia. J Neurosci Res. 2019;97:162–84.
- 64. Ben-Aicha S, Escate R, Casaní L, Padró T, Peña E, Arderiu G, et al. High-density lipoprotein remodelled in hypercholestero-laemic blood induce epigenetically driven down-regulation of endothelial HIF-1α expression in a preclinical animal model. Cardiovasc Res. 2020;116:1288–99.
- 65. Tabet F, Vickers KC, Cuesta Torres LF, Wiese CB, Shoucri BM, Lambert G, et al. HDL-transferred microRNA-223 regulates ICAM-1 expression in endothelial cells. Nat Commun. 2014;5:3292.
- 66. Li HM, Mo ZW, Peng YM, Li Y, Dai WP, Yuan HY, et al. Angiogenic and antiangiogenic mechanisms of high density lipoprotein from healthy subjects and coronary artery diseases patients. Redox Biol. 2020;36:101642.
- **67.** Schoch L, Badimon L, Vilahur G. Unraveling the complexity of HDL remodeling: on the hunt to restore HDL quality. Biomedicines. 2021;9:805.
- **68.** Gordon T, Castelli WP, Hjortland MC, Kannel WB, Dawber TR. High density lipoprotein as a protective factor against coronary heart disease: the Framingham study. Am J Med. 1977:62:707–14.
- **69.** Ouimet M, Barrett TJ, Fisher EA. HDL and reverse cholesterol transport. Circ Res. 2019;124:1505–18.
- Badimon J, Badimon L, Galvez A, Dische R, Fuster V. High density lipoprotein plasma fractions inhibit aortic fatty streaks in cholesterol-fed rabbits. Lab Invest. 1989;60:455–61.
- 71. Brites F, Martin M, Guillas I, Kontush A. Antioxidative activity of high-density lipoprotein (HDL): mechanistic insights into potential clinical benefit. BBA Clin. 2017;8:66–77.
- **72.** Riwanto M, Landmesser U. High density lipoproteins and endothelial functions: mechanistic insights and alterations in cardiovascular disease. J Lipid Res. 2013;54:3227–43.
- Moya C, Máñez S. Paraoxonases: metabolic role and pharmacological projection. Naunyn Schmiedebergs Arch Pharmacol. 2018;391:349-59.
- 74. Keul P, Polzin A, Kaiser K, Gräler M, Dannenberg L, Daum G, et al. Potent anti-inflammatory properties of HDL in vascular smooth muscle cells mediated by HDL-S1P and their impairment in coronary artery disease due to lower HDL-S1P: a new aspect of HDL dysfunction and its therapy. FASEB J Off Publ Fed Am Soc Exp Biol. 2019;33:1482–95.
- **75.** van der Stoep M, Korporaal SJA, Van Eck M. High-density lipoprotein as a modulator of platelet and coagulation responses. Cardiovasc Res. 2014;103:362–71.
- Chung DW, Chen J, Ling M, Fu X, Blevins T, Parsons S, et al. High-density lipoprotein modulates thrombosis by preventing von Willebrand factor self-association and subsequent platelet adhesion. Blood. 2016;127:637–45.
- Gomaraschi M, Calabresi L, Franceschini G. Protective effects of HDL against ischemia/reperfusion injury. Front Pharmacol. 2016:7:2.
- **78.** Schwertani A, Choi HY, Genest J. HDLs and the pathogenesis of atherosclerosis. Curr Opin Cardiol. 2018;33:311–6.
- Robert J, Osto E, von Eckardstein A. The endothelium is both a target and a barrier of HDL's protective functions. Cells. 2021;10. Available from: http://www.ncbi.nlm. nih.gov/pubmed/33924941 [cited 7.9.22].
- Adorni MP, Zimetti F, Billheimer JT, Wang N, Rader DJ, Phillips MC, et al. The roles of different pathways in the release of cholesterol from macrophages. J Lipid Res. 2007;48:2453–62.

- 81. Thuahnai ST, Lund-Katz S, Dhanasekaran P, de la Llera-Moya M, Connelly MA, Williams DL, et al. Scavenger receptor class B type I-mediated cholesteryl ester-selective uptake and efflux of unesterified cholesterol. Influence of high density lipoprotein size and structure. J Biol Chem. 2004;279:12448-55.
- **82.** Chiesa ST, Charakida M. High-density lipoprotein function and dysfunction in health and disease. Cardiovasc Drugs Ther. 2019;33:207–19.
- Miller NE, Cenini G, Upadhyay G, Durrington PN, Soran H, Schofield JD. Antioxidant properties of HDL. Front Pharmacol. 2015;6:222.
- **84.** Silva IT, Mello APQ, Damasceno NRT. Antioxidant and inflammatory aspects of lipoprotein-associated phospholipase A₂ (Lp-PLA₂): a review. Lipids Health Dis. 2011;10:170.
- **85.** Vohl MC, Neville TA, Kumarathasan R, Braschi S, Sparks DL. A novel lecithin-cholesterol acyltransferase antioxidant activity prevents the formation of oxidized lipids during lipoprotein oxidation. Biochemistry. 1999;38:5976–81.
- 86. Marín M, Moya C, Máñez S. Mutual influences between nitric oxide and paraoxonase 1. Antioxid Basel Switz. 2019;8. Available from: http://www.ncbi.nlm.nih.gov/pubmed/31817387 [cited 8.9.22].
- 87. Gaidukov L, Tawfik DS. High affinity, stability, and lactonase activity of serum paraoxonase PON1 anchored on HDL with ApoA-I. Biochemistry. 2005;44:11843–54.
- 88. Nofer JR, van der Giet M, Tölle M, Wolinska I, von Wnuck Lipinski K, Baba HA, et al. HDL induces NO-dependent vasore-laxation via the lysophospholipid receptor S1P3. J Clin Invest. 2004;113:569–81.
- 89. Mineo C, Deguchi H, Griffin JH, Shaul PW. Endothelial and antithrombotic actions of HDL. Circ Res. 2006;98:1352-64.
- 90. Kimura T, Tomura H, Mogi C, Kuwabara A, Damirin A, Ishizuka T, et al. Role of scavenger receptor class B type I and sphingosine 1-phosphate receptors in high density lipoprotein-induced inhibition of adhesion molecule expression in endothelial cells. J Biol Chem. 2006;281:37457–67.
- 91. Nofer JR, Geigenmüller S, Göpfert C, Assmann G, Buddecke E, Schmidt A. High density lipoprotein-associated lysosphingolipids reduce E-selectin expression in human endothelial cells. Biochem Biophys Res Commun. 2003;310:98–103.
- 92. Tölle M, Pawlak A, Schuchardt M, Kawamura A, Tietge UJ, Lorkowski S, et al. HDL-associated lysosphingolipids inhibit NAD(P)H oxidase-dependent monocyte chemoattractant protein-1 production. Arterioscler Thromb Vasc Biol. 2008;28:1542–8.
- **93.** Galvani S, Sanson M, Blaho VA, Swendeman SL, Obinata H, Conger H, et al. HDL-bound sphingosine 1-phosphate acts as a biased agonist for the endothelial cell receptor S1P1 to limit vascular inflammation. Sci Signal. 2015;8:ra79.
- 94. Wilkerson BA, Grass GD, Wing SB, Argraves WS, Argraves KM. Sphingosine 1-phosphate (S1P) carrier-dependent regulation of endothelial barrier: high density lipoprotein (HDL)-S1P prolongs endothelial barrier enhancement as compared with albumin-S1P via effects on levels, trafficking, and signaling of S1P1. J Biol Chem. 2012;287:44645–53.
- 95. Ruiz M, Frej C, Holmér A, Guo LJ, Tran S, Dahlbäck B. High-density lipoprotein-associated apolipoprotein m limits endothelial inflammation by delivering sphingosine-1-phosphate to the sphingosine-1-phosphate receptor 1. Arterioscler Thromb Vasc Biol. 2017;37:118–29.
- 96. Feuerborn R, Becker S, Potì F, Nagel P, Brodde M, Schmidt H, et al. High density lipoprotein (HDL)-associated sphingosine 1-phosphate (S1P) inhibits macrophage apoptosis by stimulating STAT3 activity and survivin expression. Atherosclerosis. 2017;257:29–37.
- 97. Baker PW, Rye KA, Gamble JR, Vadas MA, Barter PJ. Ability of reconstituted high density lipoproteins to inhibit cytokine-

- induced expression of vascular cell adhesion molecule-1 in human umbilical vein endothelial cells. J Lipid Res. 1999;40:345–53.
- **98.** Baker PW, Rye KA, Gamble JR, Vadas MA, Barter PJ. Phospholipid composition of reconstituted high density lipoproteins influences their ability to inhibit endothelial cell adhesion molecule expression. J Lipid Res. 2000;41:1261–7.
- 99. Barter PJ, Nicholls S, Rye KA, Anantharamaiah GM, Navab M, Fogelman AM. Antiinflammatory properties of HDL. Circ Res. 2004;95:764–72.
- 100. Hossain M, Qadri SM, Liu L. Inhibition of nitric oxide synthesis enhances leukocyte rolling and adhesion in human microvasculature. J Inflamm. 2012;9:28.
- 101. Yang Y, Loscalzo J. Regulation of tissue factor expression in human microvascular endothelial cells by nitric oxide. Circulation. 2000;101:2144–8.
- 102. Mitchell JA, Ali F, Bailey L, Moreno L, Harrington LS. Role of nitric oxide and prostacyclin as vasoactive hormones released by the endothelium. Exp Physiol. 2008;93:141–7.
- 103. Nicholls SJ, Cutri B, Worthley SG, Kee P, Rye KA, Bao S, et al. Impact of short-term administration of high-density lipoproteins and atorvastatin on atherosclerosis in rabbits. Arterioscler Thromb Vasc Biol. 2005;25:2416–21.
- 104. Vilahur G, Juan-Babot O, Peña E, Oñate B, Casaní L, Badimon L. Molecular and cellular mechanisms involved in cardiac remodeling after acute myocardial infarction. J Mol Cell Cardiol. 2011;50:522–33.
- Carden DL, Granger DN. Pathophysiology o ischaemia-reperfusion injury. J Pathol. 2000;190:255-66.
- **106.** Gunata M, Parlakpinar H. A review of myocardial ischaemia/reperfusion injury: pathophysiology, experimental models, biomarkers, genetics and pharmacological treatment. Cell Biochem Funct. 2021;39:190–217.
- 107. Theilmeier G, Schmidt C, Herrmann J, Keul P, Schäfers M, Herrgott I, et al. High-density lipoproteins and their constituent, sphingosine-1-phosphate, directly protect the heart against ischemia/reperfusion injury in vivo via the S1P3 lysophospholipid receptor. Circulation. 2006;114:1403–9.
- 108. Vilahur G, Gutiérrez M, Casaní L, Cubedo J, Capdevila A, Pons-Llado G, et al. Hypercholesterolemia abolishes high-density lipoprotein-related cardioprotective effects in the setting of myocardial infarction. J Am Coll Cardiol. 2015;66:2469–70.
- 109. Kalakech H, Hibert P, Prunier-Mirebeau D, Tamareille S, Letournel F, Macchi L, et al. RISK and SAFE signaling pathway involvement in apolipoprotein A-I-induced cardioprotection. PLoS ONE. 2014;9:e107950.
- 110. Sposito AC, de Lima-Junior JC, Moura FA, Barreto J, Bonilha I, Santana M, et al. Reciprocal multifaceted interaction between HDL (high-density lipoprotein) and myocardial infarction. Arterioscler Thromb Vasc Biol. 2019;39:1550–64.
- 111. Somers SJ, Frias M, Lacerda L, Opie LH, Lecour S. Interplay between SAFE and RISK pathways in sphingosine-1-phosphate-induced cardioprotection. Cardiovasc Drugs Ther. 2012;26:227–37.
- 112. Morel S, Frias MA, Rosker C, James RW, Rohr S, Kwak BR. The natural cardioprotective particle HDL modulates connexin43 gap junction channels. Cardiovasc Res. 2012;93:41–9.
- 113. Wang J, Wang W, Yan C, Wang T. Ischemic postconditioning protects nonculprit coronary arteries against ischemia-reperfusion injury via downregulating miR-92a, miR-328 and miR-494. Aging. 2022;14:2748-57.
- 114. Wu Q, Wang H, He F, Zheng J, Zhang H, Cheng C, et al. Depletion of microRNA-92a enhances the role of sevoflurane treatment in reducing myocardial ischemia-reperfusion injury by upregulating KLF4. Cardiovasc Drugs Ther. 2022. Available from: http://www.ncbi.nlm.nih.gov/pubmed/35171385 [cited 12.9.22].

- 115. Hinkel R, Penzkofer D, Zühlke S, Fischer A, Husada W, Xu QF, et al. Inhibition of microRNA-92a protects against ischemia/reperfusion injury in a large-animal model. Circulation. 2013;128:1066-75.
- 116. Zhang B, Zhou M, Li C, Zhou J, Li H, Zhu D, et al. MicroRNA-92a inhibition attenuates hypoxia/reoxygenationinduced myocardiocyte apoptosis by targeting Smad7. PLoS ONE. 2014;9:e100298.
- 117. Wang N, Yu YB. MiR-486 alleviates hypoxia/reoxygenation-induced H9c2 cell injury by regulating forkhead box D3. Eur Rev Med Pharmacol Sci. 2022;26:422-31.
- 118. Bei Y, Lu D, Bär C, Chatterjee S, Costa A, Riedel I, et al. miR-486 attenuates cardiac ischemia/reperfusion injury and mediates the beneficial effect of exercise for myocardial protection. Mol Ther J Am Soc Gene Ther. 2022;30:1675–91.
- **119.** Su Q, Xu Y, Cai R, Dai R, Yang X, Liu Y, et al. miR-146a inhibits mitochondrial dysfunction and myocardial infarction by targeting cyclophilin D. Mol Ther Nucleic Acids. 2021;23:1258–71.
- 120. Li G, Xu M, Wang H, Qi X, Wang X, Li Y, et al. MicroRNA-146a overexpression alleviates intestinal ischemia/reperfusioninduced acute lung injury in mice. Exp Ther Med. 2021;22: 937
- 121. Díaz I, Calderón-Sánchez E, Toro RD, Ávila-Médina J, de Rojas-de Pedro ES, Domínguez-Rodríguez A, et al. miR-125a, miR-139 and miR-324 contribute to Urocortin protection against myocardial ischemia-reperfusion injury. Sci Rep. 2017;7:8898.
- **122.** Wu Q, Shang Y, Bai Y, Wu Y, Wang H, Shen T. Sufentanil preconditioning protects against myocardial ischemia/reperfusion injury via miR-125a/DRAM2 axis. Cell Cycle Georget Tex. 2021;20:383–91.
- **123.** Baeyens N, Bandyopadhyay C, Coon BG, Yun S, Schwartz MA. Endothelial fluid shear stress sensing in vascular health and disease. J Clin Invest. 2016;126:821–8.
- **124.** McCarron JG, Lee MD, Wilson C. The endothelium solves problems that endothelial cells do not know exist. Trends Pharmacol Sci. 2017;38:322–38.
- 125. Argraves KM, Gazzolo PJ, Groh EM, Wilkerson BA, Matsuura BS, Twal WO, et al. High density lipoprotein-associated sphingosine 1-phosphate promotes endothelial barrier function. J Biol Chem. 2008;283:25074–81.
- 126. Hesse B, Rovas A, Buscher K, Kusche-Vihrog K, Brand M, Di Marco GS, et al. Symmetric dimethylarginine in dysfunctional high-density lipoprotein mediates endothelial glycocalyx breakdown in chronic kidney disease. Kidney Int. 2020;97:502–15.
- **127.** Tamagaki T, Sawada S, Imamura H, Tada Y, Yamasaki S, Toratani A, et al. Effects of high-density lipoproteins on intracellular pH and proliferation of human vascular endothelial cells. Atherosclerosis. 1996;123:73–82.
- 128. Zhang Q, Yin H, Liu P, Zhang H, She M. Essential role of HDL on endothelial progenitor cell proliferation with PI3K/Akt/cyclin D1 as the signal pathway. Exp Biol Med (Maywood NJ). 2010;235:1082-92.
- 129. Sugano M, Tsuchida K, Makino N. High-density lipoproteins protect endothelial cells from tumor necrosis factor-alpha-induced apoptosis. Biochem Biophys Res Commun. 2000;272:872–6.
- 130. Nofer JR, Levkau B, Wolinska I, Junker R, Fobker M, von Eckardstein A, et al. Suppression of endothelial cell apoptosis by high density lipoproteins (HDL) and HDL-associated lysosphingolipids. J Biol Chem. 2001;276:34480-5.
- 131. Takino J, Sato T, Nagamine K, Hori T. The inhibition of Bax activation-induced apoptosis by RasGRP2 via R-Ras-PI3K-Akt signaling pathway in the endothelial cells. Sci Rep. 2019;9:16717.
- 132. Michaelis M, Suhan T, Michaelis UR, Beek K, Rothweiler F, Tausch L, et al. Valproic acid induces extracellular signal-

- regulated kinase 1/2 activation and inhibits apoptosis in endothelial cells. Cell Death Differ. 2006;13:446–53.
- 133. Seetharam D, Mineo C, Gormley AK, Gibson LL, Vongpatanasin W, Chambliss KL, et al. High-density lipoprotein promotes endothelial cell migration and reendothelialization via scavenger receptor-B type I. Circ Res. 2006;98:63–72.
- 134. Tan JTM, Prosser HCG, Vanags LZ, Monger SA, Ng MKC, Bursill CA. High-density lipoproteins augment hypoxia-induced angiogenesis via regulation of post-translational modulation of hypoxia-inducible factor 1α . FASEB J Off Publ Fed Am Soc Exp Biol. 2014;28:206–17.
- 135. Kimura T, Sato K, Malchinkhuu E, Tomura H, Tamama K, Kuwabara A, et al. High-density lipoprotein stimulates endothelial cell migration and survival through sphingosine 1-phosphate and its receptors. Arterioscler Thromb Vasc Biol. 2003;23:1283–8.
- 136. Miura S, ichiro, Fujino M, Matsuo Y, Kawamura A, Tanigawa H, et al. High density lipoprotein-induced angiogenesis requires the activation of Ras/MAP kinase in human coronary artery endothelial cells. Arterioscler Thromb Vasc Biol. 2003;23:802–8.
- 137. Dennhardt S, Finke KR, Huwiler A, Coldewey SM. Sphingosine-1-phosphate promotes barrier-stabilizing effects in human microvascular endothelial cells via AMPK-dependent mechanisms. Biochim Biophys Acta Mol Basis Dis. 2019;1865:774–81.
- 138. Jin F, Hagemann N, Sun L, Wu J, Doeppner TR, Dai Y, et al. High-density lipoprotein (HDL) promotes angiogenesis via S1P3-dependent VEGFR2 activation. Angiogenesis. 2018;21:381–94.
- 139. Mineo C, Yuhanna IS, Quon MJ, Shaul PW. High density lipoprotein-induced endothelial nitric-oxide synthase activation is mediated by Akt and MAP kinases. J Biol Chem. 2003;278:9142-9.
- 140. Zhang QH, Zu XY, Cao RX, Liu JH, Mo ZC, Zeng Y, et al. An involvement of SR-B1 mediated PI3K-Akt-eNOS signaling in HDL-induced cyclooxygenase 2 expression and prostacyclin production in endothelial cells. Biochem Biophys Res Commun. 2012;420:17–23.
- 141. Xiong SL, Liu X, Yi GH. High-density lipoprotein induces cyclooxygenase-2 expression and prostaglandin I-2 release in endothelial cells through sphingosine kinase-2. Mol Cell Biochem. 2014;389:197–207.
- **142.** Castelli WP, Garrison RJ, Wilson PWF, Abbott RD, Kalousdian S, Kannel WB. Incidence of coronary heart disease and lipoprotein cholesterol levels. JAMA. 1986;256:2835.
- 143. Stampfer MJ, Sacks FM, Salvini S, Willett WC, Hennekens CH. A prospective study of cholesterol, apolipoproteins, and the risk of myocardial infarction. N Engl J Med. 1991;325:373–81.
- 144. Emerging Risk Factors Collaboration DA, Di Angelantonio E, Sarwar N, Perry P, Kaptoge S, Ray KK, et al. Major lipids, apolipoproteins, and risk of vascular disease. JAMA. 2009;302:1993–2000.
- **145.** Madsen CM, Varbo A, Nordestgaard BG. Extreme high high-density lipoprotein cholesterol is paradoxically associated with high mortality in men and women: two prospective cohort studies. Eur Heart J. 2017;38:2478–86.

- **146.** Voight BF, Peloso GM, Orho-Melander M, Frikke-Schmidt R, Barbalic M, Jensen MK, et al. cholesterol and risk of myocardial infarction: a mendelian randomisation study. Lancet Lond Engl. 2012;380:572–80.
- 147. Keene D, Price C, Shun-Shin MJ, Francis DP. Effect on cardiovascular risk of high density lipoprotein targeted drug treatments niacin, fibrates, and CETP inhibitors: meta-analysis of randomised controlled trials including 117,411 patients. BMJ. 2014;349:g4379.
- 148. Lincoff AM, Nicholls SJ, Riesmeyer JS, Barter PJ, Brewer HB, Fox KAA, et al. Evacetrapib and cardiovascular outcomes in high-risk vascular disease. N Engl J Med. 2017;376:1933–42.
- **149.** Schwartz GG, Olsson AG, Abt M, Ballantyne CM, Barter PJ, Brumm J, et al. Effects of dalcetrapib in patients with a recent acute coronary syndrome. N Engl J Med. 2012;367:2089–99.
- **150.** Barter PJ, Caulfield M, Eriksson M, Grundy SM, Kastelein JJP, Komajda M, et al. Effects of torcetrapib in patients at high risk for coronary events. N Engl J Med. 2007;357:2109–22.
- **151.** Group THC. Effects of anacetrapib in patients with atherosclerotic vascular disease. N Engl J Med. 2017;377:1217–27.
- **152.** Tardif JC, Rhéaume E, Lemieux Perreault LP, Grégoire JC, Feroz Zada Y, Asselin G, et al. Pharmacogenomic determinants of the cardiovascular effects of dalcetrapib. Circ Cardiovasc Genet. 2015;8:372–82.
- 153. Du XM, Kim MJ, Hou L, Le Goff W, Chapman MJ, Van Eck M, et al. HDL particle size is a critical determinant of ABCA1-mediated macrophage cellular cholesterol export. Circ Res. 2015;116:1133–42.
- **154.** Badimon JJ, Badimon L, Fuster V. Regression of atherosclerotic lesions by high density lipoprotein plasma fraction in the cholesterol-fed rabbit. J Clin Invest. 1990;85:1234–41.
- **155.** Jomard A, Osto E. High density lipoproteins: metabolism, function, and therapeutic potential. Front Cardiovasc Med. 2020;7:39.
- 156. Bonizzi A, Piuri G, Corsi F, Cazzola R, Mazzucchelli S. HDL dysfunctionality: clinical relevance of quality rather than quantity. Biomedicines. 2021;9. Available from: http://www.ncbi.nlm.nih.gov/pubmed/34202201
- **157.** Schoch L, Sutelman P, Suades R, Casani L, Padro T, Badimon L, et al. Hypercholesterolemia-induced HDL dysfunction can be reversed: the impact of diet and statin treatment in a preclinical animal model. Int J Mol Sci. 2022;23:8596.
- **158.** Badimon L, Vilahur G. HDL particles more complex than we thought. Thromb Haemost. 2014;112:857.
- 159. Feingold KR. Introduction to lipids and lipoproteins. Endotext. 2000. Available from: http://www.ncbi.nlm.nih.gov/pubmed/26247089 [cited 22.8.22].
- 160. Thakkar H, Vincent V, Sen A, Singh A, Roy A. Changing perspectives on HDL: from simple quantity measurements to functional quality assessment. J Lipids. 2021;2021:5585521.
- 161. HDL Proteome Watch page. Available from: https:// homepages.uc.edu/%7Edavidswm/HDLproteome.html [cited 11.10.23].