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#### Concise review

# Mechanisms and therapeutic targets of mitochondria in the progression of metabolic dysfunction-associated steatotic liver disease



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#### ABSTRACT

Metabolic Dysfunction-Associated Steatotic Liver Disease (MASLD) includes liver disease processes from simple fatty liver to nonalcoholic steatohepatitis, which may progress to liver fibrosis, cirrhosis, and even hepatocellular carcinoma (HCC). As the incidence of HCC derived from viral hepatitis decreases, MASLD has emerged as a significant health threat, driven by lifestyle changes and rising obesity rates among patients. The pathogenesis of MASLD is complex, involving factors such as insulin resistance, gut microbiota imbalance, and genetic and epigenetic factors. In recent years, the role of mitochondrial dysfunction in MASLD has gained significant attention, involving  $\beta$ -oxidation imbalance, oxidative stress increase, mitophagy defects, and mitochondrial DNA (mtDNA) mutations. This article reviews the pathophysiological mechanisms of mitochondrial dysfunction in MASLD, diagnostic methods, and potential therapeutic strategies. By synthesizing current research findings, the review aims to highlight the critical role of mitochondrial dysfunction as a target for future diagnostic and therapeutic interventions. This focus could pave the way for innovative clinical strategies, ultimately improving treatment options and patient prognosis in MASLD.

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# 1. Introduction

Recently, "Non-Alcoholic Fatty Liver Disease" (NAFLD) has been renamed to "Metabolic Dysfunction-Associated Steatotic Liver Disease" (MASLD). This new term, "Metabolic Associated Steatotic Liver Disease" (MASLD), underscores the critical role of metabolic factors

Abbreviations: 17-AAG, 17-Allylamino-17-demethoxygeldanamycin; ALT, Alanine aminotransferase; AMPK, AMP-activated protein kinase; BMI, Body mass index; CD36, Cluster of differentiation 36; CDS2, CDP-diacylglycerol synthase 2; CPT2, Carnitine palmitoyltransferase 2; DNL, De novo lipogenesis; DNM2, Dynamin 2; DRP1, Dynamin-related protein 1; ETC, Electron transport chain; FFA, Free fatty acids; GCGR, Glucagon receptor; HCC, Hepatocellular carcinoma; HFD, High-fat diet; IP3R, Inositol 1,4,5-triphosphate receptor; JNK, c-Jun N-terminal kinase; LC3, Microtubule-associated protein 1 light chain 3; MAM, Mitochondria-associated endoplasmic reticulum membranes; MASLD, Metabolic dysfunction-associated steatotic liver disease; MCJ, Methylation-controlled J protein; MFN2, Mitofusin 2; mtDNA, Mitochondrial DNA; NASH, Nonalcoholic steatohepatitis; NP, Nanoplastic; OPA1, Optic atrophy 1; OXPHOS, Oxidative phosphorylation; PGC-1α, Peroxisome proliferator-activated receptor gamma coactivator 1 alpha; PUFAs, Polyunsaturated fatty acids; ROS, Reactive oxygen species; VDAC, Voltage-dependent anion channel

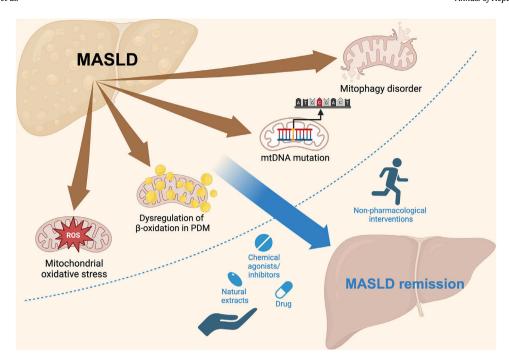
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in the disease's progression, such as obesity, diabetes, and hyperlipidemia [1]. With the increasing prevalence of metabolic syndrome, MASLD has become a common liver condition [2]. As of 2019, MASLD affected 38 % of the global population [3,4]. Chris Estes and his team utilized mathematical models to study the status of MASLD in countries including China, the United States, France, and Japan from 2016 to 2030. Their findings indicated that China had the highest number of MASLD cases worldwide, with the patient population expected to grow from 246.33 million in 2016 to 314.58 million by 2030, an increase of approximately 29.1 % [5]. MASLD exhibits a strong association with hepatocellular carcinoma (HCC), with its microenvironment fostering HCC cell proliferation and migration [6]. In recent years, the incidence of MASLD-driven HCC has been increasing [7]. MASLD has become a global health problem that brought huge medical and economic burden [8].

Increasing evidence indicated that mitochondrial dysfunction is associated with the progression of MASLD. During oxidative stress, the activity of the mitochondrial electron transport chain (ETC) is inhibited, leading to increased generation of reactive oxygen species (ROS), which plays an important role in the pathogenesis of MASLD [9]. Blocked mitochondrial  $\beta$ -oxidation in the liver leads to the



**Fig. 1.** Schematic representation of the mechanisms of MASLD and potential therapeutic interventions. MASLD progression is linked to mitochondrial ROS, dysregulation of β-oxidation in PDM, mtDNA mutations, and mitophagy disorders. These mitochondrial dysfunctions contribute to disease progression but can be targeted by various interventions. Non-pharmacological approaches, such as physical activity, and pharmacological strategies, including drugs, chemical agonists/inhibitors, and natural extracts, can promote MASLD remission by addressing mitochondrial dysfunction. MASLD, metabolic dysfunction-associated steatotic liver disease.

accumulation of fatty acid, the generation of lipotoxicity, and the activation of pro-inflammatory signaling pathways. In addition, excessive reactive oxygen species can lead to liver cell damage and fibrosis, which are generated by oxidative stress. Furthermore, mitophagy defects hinder the timely clearance of defective mitochondria and aggravate cell damage. Mutations and damage of mitochondrial DNA further impair mitochondrial function and are one of the causes of energy metabolism disorders in MASLD [10–14].

We review the mechanisms of MASLD development and the structure and function of mitochondria, highlighting mitochondrial dysfunction as a key risk factor for MASLD. Finally, we discussed the significance of targeting mitochondrial dysfunction for diagnosis and treatment, aiming to improve therapy and patient outcomes (Fig. 1).

# 2. Pathogenesis of MASLD

In 1998, Day and James proposed the "two-hit" hypothesis to explain how simple steatosis progresses to nonalcoholic steatohepatitis (NASH) in MASLD. The first hit is the accumulation of fat in the liver, which is driven by lifestyle, dietary habits, obesity, and insulin resistance. The second hit involves inflammation and oxidative stress, leading to hepatocyte damage, inflammation, and fibrosis, which promote the progression of MASLD [15]. Although significant hepatic lipid accumulation is observed in the genetic obesity ob/ob mouse model of leptin deficiency, a second hit is required to induce inflammation and fibrosis [16]. As research on MASLD has advanced, it has become clear that the "two-hit" model is too simplistic to fully capture the complexity of the disease. The current trend in MASLD research favors the "multiplehit", which offers a more comprehensive explanation of the disease's development by suggesting that various factors-including insulin resistance, immune dysregulation, hormones derived from adipose tissue, nutritional components, gut microbiota, and both genetic and epigenetic factors-play integral roles in its pathogenesis (Fig. 2) [17].

# 2.1. Insulin resistance

Insulin resistance is a physiological state that refers to the disorder of molecular signaling pathways when insulin acts on target tissues, resulting in a decrease in the ability of cells to respond to insulin. Insulin resistance plays a key role in the development of MASLD. The reduced expression of insulin receptor substrate 2 during insulin resistance leads to the overexpression of sterol regulatory element-binding protein 1c, a transcription factor that promotes hepatic de novo lipogenesis (DNL), thereby upregulating DNL [18,19]. Moreover, the inhibited  $\beta$ -oxidation of free fatty acids (FFA) in liver cells causes further lipid accumulation [20]. Elevated intracellular FFA concentrations activate multiple serine kinases, including c-Jun N-terminal kinase (INK), an inhibitor of nuclear factor  $\kappa$ -B kinase, and protein kinase  $C-\theta$  [21], which phosphorylate serine residues of insulin receptor substrate 1, thereby reducing its activity and exacerbating insulin resistance [22]. Insulin resistance is the main feature of MASLD, and the two are in a mutually promoting bidirectional relationship.

# 2.2. Hepatic lipid accumulation

In MASLD, lipid accumulation significantly affects liver health. In the liver, fatty acids are metabolized by mitochondrial  $\beta$ -oxidation or esterified to form triglycerides. Research indicates that as MASLD severity increases, there is a significant downregulation of both complete oxidation of long-chain fatty acids and the rate-limiting steps of  $\beta$ -oxidation in the liver. This leads to the excessive formation of triglyceride-laden lipid droplets, which is a hallmark feature of MASLD [23]. At the same time, the breakdown of these lipid droplets increases intracellular fatty acid levels. Specific lipotoxic lipids such as diacylglycerols, ceramides, and lysophosphatidylcholine may cause cell damage and trigger NASH. Lipotoxic lipids can induce pathological changes such as endoplasmic reticulum stress, inflammation, apoptosis, and injury responses. External factors like cytokine imbalance, ATP depletion, and periodic hypoxia further exacerbate hepatocyte lipotoxic stress and inflammation [24].

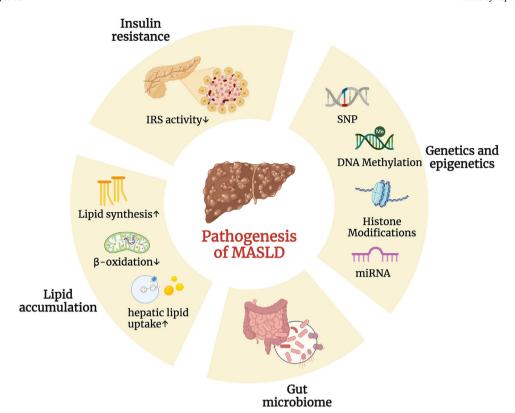


Fig. 2. Various factors play integral roles in the pathogenesis of MASLD. Differential changes in multiple areas (including insulin resistance, genetics and epigenetics, gut microbiome, and lipid accumulation) have a profound impact on the pathogenesis of MASLD. (Created with BioRender.com). DNA, deoxyribonucleic acid; IRS, insulin receptor substrate; miRNA, microRNA; SNP, single nucleotide polymorphism.

#### 2.3. Immune dysregulation

The liver, as a complex immunological organ, contains a variety of immune cell types, including Kupffer cells, non-Kupffer macrophages, natural killer (NK) cells, natural killer T (NKT) cells, dendritic cells (DCs), and hepatic stellate cells (HSCs), among others. Through their interactions, these cells play a critical role in maintaining hepatic immune homeostasis, defending against pathogen invasion, and regulating the progression of liver damage [25].

In MASLD, the polarization mechanisms of M1 and M2 macrophages are mediated through multiple signaling pathways. The polarization of M1 macrophages is primarily induced by pro-inflammatory factors such as saturated fatty acids (SFA) and lipopolysaccharides (LPS), which activate the NF- $\kappa$ B signaling pathway. This activation leads to the upregulation of pro-inflammatory cytokines, such as TNF- $\alpha$  and IL-6, and is mainly dependent on the activation of the TLR4 receptor [26,27]. In contrast, the polarization of M2 macrophages is mediated by anti-inflammatory factors such as n-3 polyunsaturated fatty acids (PUFA) and IL-4, which activate the PPAR- $\gamma$ signaling pathway. This activation promotes the expression of antiinflammatory cytokines, such as IL-10, and effectively inhibits the activity of the NF- $\kappa$ B signaling pathway [28,29]. Moreover, the activation of PPAR- $\gamma$  can inhibit M1 polarization by directly binding to the NF-κB p65 subunit, thereby promoting M2 polarization. These mechanisms reveal the dynamic changes in M1 and M2 macrophage polarization and their roles in the progression of NAFLD [29].

#### 2.4. Gut microbiota

In MASLD, poor dietary habits can induce a series of chain reactions by disrupting the gut microbiota, leading to dysbiosis, intestinal barrier disruption, and increased intestinal permeability, thereby allowing more pathogen-associated molecular patterns and bacteria

to translocate through the portal vein into the liver, resulting in sustained immune activation and inflammation, which ultimately promotes the progression of MASLD [30]. Studies have shown that the gut microbiota can produce substantial amounts of ethanol in MASLD patients. This ethanol enters the liver through the portal circulation, bypassing the first-pass effect, thereby inducing hepatic steatosis and inflammation [31].

# 2.5. Genetic and epigenetic factors

MASLD exhibits familial aggregation. In one study, 16 out of 90 MASLD patients had first-degree relatives who were also affected by the disease [32]. Furthermore, parents with type 2 diabetes have an increased risk of MASLD in their offspring [33]. Research on obese children with MASLD and their family members revealed that siblings and parents of MASLD children have a higher probability of developing fatty liver, with a stronger correlation between liver fat fraction and body mass index (BMI) [34]. Twin studies have shown that after adjusting for gender and BMI, 60 % of the variation in serum alanine aminotransferase (ALT), a marker of liver damage (hepatitis, cirrhosis, fatty liver), is determined by genetic factors [35].

Although MASLD is defined by its histological phenotype, genome-wide association studies have identified several strongly associated genes. The PNPLA3 gene plays a pivotal role in various aspects of MASLD. Studies indicate that the PNPLA3 rs738409 (Ile148Met) variant is associated with increased inflammation and fibrosis in MASLD [33,36,37]. This variant also elevates the risk of MASLD-associated HCC [37]. The PNPLA3 gene modulates hepatic steatosis by enhancing the liver's response to environmental stressors [37,38]. The GCKR rs1260326 (Pro446Leu) variant leads to increased hepatic glucokinase activity, perpetuating hepatic glycolysis, downregulating glucose and insulin levels, increasing malonyl-CoA content in hepatocytes, inhibiting fatty acid  $\beta$ -oxidation, and

promoting lipid accumulation [38]. The TM6SF2 rs58542926 (Glu167Lys) variant results in loss of protein function, leading to decreased VLDL levels, elevated ALT levels, and hepatic steatosis [39]. Genes such as superoxide dismutase 2, phosphatidylethanolamine N-methyltransferase, angiotensin II receptor type 1, and kruppel-like factor 6 have been found to be independently associated with MASLD progression [40–43].

During the progression from simple fatty liver to NASH, epigenetic modifications such as DNA methylation, histone acetylation, and miRNAs play crucial roles [44]. The DNA methylation levels of apolipoprotein B and nuclear factor erythroid 2-related factor 2 are positively correlated with lipid accumulation in MASLD, whereas the methylation level of dipeptidyl peptidase 4 is negatively correlated with NASH staging [45-47]. The SIRT protein family and specific miR-NAs (such as miR-122) significantly influence the development of NASH by regulating key genes involved in lipid and glucose metabolism [41,48]. Furthermore, these genetic modifications can be inherited by offspring, increasing the risk of MASLD [49]. Small RNAs derived from the 5' end of tRNA, known as tsRNA, have been found to be significantly upregulated in the sperm tsRNA of high-fat diet (HFD) mice, with increased m5C and m2G modifications. Changes in the paternal RNA modification profile can be transmitted to offspring, thereby perpetuating metabolic disorder traits [50].

#### 3. Overview of mitochondria

#### 3.1. Structure and dynamics of mitochondria

Mitochondria are double-membrane organelles found in most eukaryotic cells, consisting of several functional regions including the outer membrane, intermembrane space, inner membrane, and matrix [51]. They have their own double-stranded circular DNA, independent of nuclear DNA, and are able to replicate, transcribe, and translate mitochondrial-specific proteins. These proteins include 13 polypeptides of the mitochondrial respiratory chain, 22 transfer RNAs, and 2 ribosomal RNAs, all of which are essential for mitochondrial function and self-replication [52].

Mitochondrial dynamics refer to the continuous processes of mitochondrial fusion and fission, regulated by specific proteins, which maintain mitochondrial homeostasis [53]. Mitochondrial fission involves multiple steps: first, actin filaments assist the endoplasmic reticulum in tightly wrapping around the mitochondrion, leading to local constriction of the mitochondrial surface and providing a platform for the assembly of dynamin-related protein 1 (DRP1) [54,55]; Next, DRP1 mediates further constriction, promoting the recruitment of dynamin 2 (DNM2); finally, mitochondrial division is completed through membrane scission induced by DNM2 [56].

Oxidative phosphorylation (OXPHOS) is a vital mitochondrial function involving multiple enzyme complexes in the inner mitochondrial membrane, which generate ATP by transferring electrons from NADH and FADH2 [57,58]. During this process, electrons sequentially pass through complexes I, III, and IV, ultimately combining with oxygen and yielding H2O [59]. This electron transfer drives the translocation of protons from the mitochondrial matrix across the membrane, creating a proton motive force. This force, consisting of an electrochemical gradient, powers ATP synthase to synthesize ATP [60]. However, electron leakage in the electron transport chain can lead to the generation of ROS, which serves as signaling molecules in cells, though excessive ROS can cause oxidative damage.

Mitochondrial redox homeostasis affects mitochondrial dynamics through redox-sensitive post-translational modifications of specific proteins. An illustration of this is the enhancement of Drp1's GTPase activity via S-nitrosylation at Cys644, thereby promoting mitochondrial fission [61]. S-nitrosylation of optic atrophy 1 (OPA1) may be involved in mitochondrial fusion, although the specific function is still unclear [62]. Mitofusin 2(Mfn2) is susceptible to ubiquitination

and degradation after JNK-mediated phosphorylation, leading to mitochondrial fission [63]. Additionally, S-nitrosylation of Parkin inhibits its E3 ubiquitin ligase activity, affecting the degradation of Mfn1 and possibly promoting mitochondrial elongation [64]. These proteins interact with each other and, in conjunction with changes in redox status, dynamically regulate mitochondrial morphology.

# 3.2. Mitochondrial dysfunction

Mitochondrial dysfunction includes a variety of damages, such as reduced ATP production due to ETC damage, ROS generation, calcium ion dysregulation, increased mitochondrial membrane permeability due to the opening of the permeability transition pore (PTP), and dysregulation of cell apoptosis. mtDNA, as the energy metabolism center of the cell, is more susceptible to damage by toxic metabolites. Compared with nuclear DNA, mtDNA lacks effective protection and repair mechanisms and is more susceptible to damage and mutation [65]. Damage to mtDNA impairs mitochondrial function, reduces energy production, and leads to a variety of diseases, including mtDNA depletion syndrome, multiple cancers, and age-related diseases [66 -71]. PTP is a voltage-dependent channel protein complex located in the inner mitochondrial membrane. When PTP is open, mitochondrial membrane permeability increases significantly, allowing small molecules such as water, inorganic ions, metabolites, and proteins to pass through. High levels of ROS, Ca<sup>2+</sup> overload, changes in the lipid microenvironment, inactivation of B-cell lymphoma 2 protein, and high pH environment can trigger the opening of PTP [72,73], ultimately leading to mitochondrial swelling and membrane rupture. This process releases various apoptotic factors, such as cytochrome c, which activate downstream effectors like caspases, resulting in abnormal cell apoptosis [74].

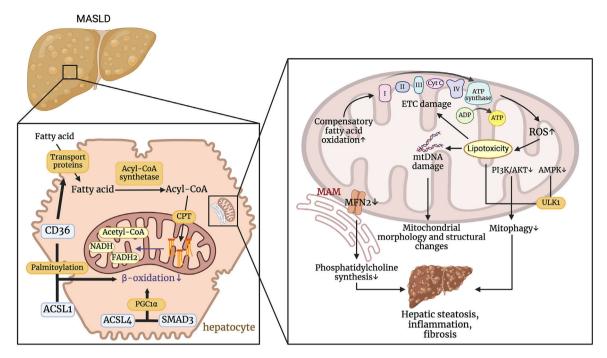
# 4. Mitochondrial dysfunction and MASLD

Mitochondria play a crucial role in the development of MASLD by regulating fatty acid  $\beta$ -oxidation, managing oxidative stress, and interacting with cellular structures such as lipid droplets and the endoplasmic reticulum. Dysfunction in these processes leads to increased ROS production, mtDNA damage, and impaired mitophagy, which contribute to hepatic steatosis, inflammation, and fibrosis. Understanding these mitochondrial mechanisms is essential for developing targeted therapeutic strategies to mitigate the progression of MASLD (Fig. 3).

# 4.1. Mitochondrial $\beta$ -oxidation and MASLD

Mitochondrial fatty acid  $\beta$ -oxidation is the primary pathway for fatty acid degradation and is crucial for maintaining energy homeostasis in the human body. During the post-absorptive state and fasting, fatty acids become a key energy source due to limited glucose availability. However, even in glucose-rich conditions, fatty acid  $\beta$ -oxidation remains a major energy source for the heart, skeletal muscles, and kidneys. In the cytoplasm, fatty acids are activated into acyl-CoA by acyl-CoA synthetase and then transported into the mitochondria via the carnitine shuttle. Subsequently, acyl-CoA undergoes four enzymatic steps: dehydrogenation, hydration, re-dehydrogenation, and thiolysis, being broken down into acetyl-CoA, with the generated FADH2 and NADH entering the ETC [75].

In the early stages of insulin resistance-related NASH, the liver responds to lipid overload by increasing  $\beta$ -oxidation activity and VLDL secretion [76]. Insulin resistance enhances lipolysis in adipose tissue, releasing a large amount of FFA into the bloodstream, which are subsequently absorbed by the liver. As  $\beta$ -oxidation increases, a substantial amount of ROS is generated, leading to oxidative stress and mitochondrial damage. Persistent oxidative stress and mitochondrial damage gradually impair  $\beta$ -oxidation function, reducing the



**Fig. 3.** Pathophysiological mechanisms by which Mitochondrial Dysfunction Affects MASLD. CD36 regulates transport proteins that transport fatty acids into cells. CD36 can promote the transport of long-chain fatty acids into the cell via transport proteins, and palmitoylated CD36 interacts with ACSL1 to inhibit β-oxidation. ACSL4 can also regulate PGC1α in a smad3-dependent manner to affect β-oxidation. These processes result in the accumulation of fatty acids within the hepatocyte. Impaired ETC and increased ROS production lead to lipotoxicity, which in turn damages mtDNA and exacerbates changes in mitochondrial structure and function. The accumulation of lipotoxicity and the inhibition of the PI3K/AKT and AMPK pathways impair mitochondrial autophagy. In addition, decreased MFN2 expression in MAMs leads to reduced phosphatidylcholine synthesis. This series of events ultimately promotes hepatic steatosis, inflammation, and fibrosis. (Created with BioRender.com). ACSL1, acyl-CoA synthetase long-chain family member 1; ACSL4, acyl-CoA synthetase long-chain family member 4; ADP, adenosine diphosphate; AMPK, AMP-activated protein kinase; ATP, adenosine triphosphate; ATP synthase, adenosine triphosphate synthase; CD36, cluster of differentiation 36; CPT, carnitine palmitoyltransferase; ETC damage, electron transport chain damage; FADH, flavin adenine dinucleotide (Reduced Form); MASLD, metabolic dysfunction-associated steatotic liver disease; MAM, mitochondria-associated membranes; MFN2, mitofusin 2; mtDNA, mitochondrial DNA; NADH, nicotinamide adenine dinucleotide (Reduced Form); PI3K/AKT, phosphoinositide 3-kinase / protein kinase B pathway; ROS, reactive oxygen species; SMAD3, SMAD family member 3; ULK1, unc-51 like autophagy activating kinase 1.

liver's efficiency in processing FFA, resulting in further lipid accumulation and forming a positive feedback loop of hepatocellular damage in MASLD [77]. When hepatic  $\beta$ -oxidation is obstructed, fatty acids produce lipotoxic substances that induce pro-inflammatory and profibrotic signaling pathways, promoting steatohepatitis and fibrosis. Reversing the inhibition of  $\beta$ -oxidation can slow the progression of MASLD [78]. Cluster of differentiation 36 (CD36) facilitates the uptake of long-chain fatty acids by hepatocytes. In MASLD, palmitoylation of CD36 is significantly increased. Depalmitoylation of CD36 increases its mitochondrial distribution and, through interaction with longchain acyl-CoA synthetase 1, upregulates mitochondrial  $\beta$ -oxidation, thereby reducing lipid accumulation in hepatocytes [79]. The longchain acyl-CoA synthetase family member ACSL4 is highly expressed in the liver tissues of MASLD patients and is positively correlated with hepatocellular steatosis and fibrosis. Although ACSL4 does not directly participate in  $\beta$ -oxidation, its silencing upregulates  $\beta$ -oxidation-related enzymes peroxisome proliferator-activated receptor  $\alpha$ and its coactivator peroxisome proliferator-activated receptor gamma coactivator 1 alpha (PGC- $1\alpha$ ) in an SMAD family member 3dependent manner, promoting mitochondrial  $\beta$ -oxidation and thereby protecting cells from death [80].

# 4.2. Mitochondrial oxidative stress and MASLD

Mitochondrial oxidative stress is characterized by excessive production and accumulation of ROS due to mitochondrial dysfunction, leading to cellular oxidative damage. Under normal metabolic conditions, ROS are produced by the ETC, but excessive ROS can oxidize proteins, lipids, and DNA, resulting in cellular dysfunction and death [81]. Enzymatic antioxidants such as superoxide dismutase and

catalase, along with non-enzymatic antioxidants like glutathione and ascorbic acid, work together to eliminate ROS. However, when this balance is disrupted, oxidative stress intensifies, influencing the onset and progression of MASLD [10,11]. Mitochondrial antioxidant capacity is influenced by several redox pairs, including NADH/NAD+ and reduced glutathione /oxidized glutathione. Changes in the mitochondrial redox state lead to an increased NADH/NAD+ ratio, which activates fatty acid synthases (such as fatty acid synthase and glycerol-3phosphate acyltransferase) and inhibits fatty acid oxidases (such as acyl-CoA dehydrogenase and  $\beta$ -hydroxyacyl-CoA dehydrogenase) [82]. As MASLD progresses, fatty acids accumulate in the liver. To control excess lipids, hepatocytes enhance fatty acid oxidation and the tricarboxylic acid (TCA) cycle [83]. However, lipid oxidation produces a large amount of reducing equivalents, leading to electron overload of the mitochondrial respiratory chain, thereby increasing the production of ROS [84].

ROS not only induce the directed migration of pro-fibrotic cells in the liver, promoting hepatic fibrosis, but also trigger lipid peroxidation of PUFAs, producing harmful aldehyde byproducts such as 4-hydroxy-2-nonenal and malondialdehyde. This exacerbates cellular damage and leads to cell death [85]. Furthermore, due to insufficient mitochondrial antioxidant defense mechanisms to cope with sustained oxidative stress induced by long-chain free fatty acids, the mitochondrial ETC and mtDNA suffer oxidative damage. This damage not only impairs normal mitochondrial function but also promotes the generation of more ROS, creating a vicious cycle that accelerates the progression of MASLD [86–88]. The methylation-controlled J protein (MCJ) is a transmembrane protein located in the mitochondrial inner membrane, possessing a unique regulatory mechanism for fatty acid metabolism. As a negative regulator of ETC complex I, MCJ

inhibits its expression, which can enhance the activity of mitochondrial complex I, thereby increasing the efficiency of fatty acid  $\beta$ -oxidation. Inhibiting MCJ not only boosts the activity of complex I but also promotes the formation of respiratory chain complexes I, III, and IV, reducing electron leakage, thereby preventing ROS production and the downstream activation of the JNK pathway, ultimately alleviating MASLD [89].

# 4.3. Peridroplet mitochondria and MASLD

Peridroplet Mitochondria (PDM) are mitochondria closely arranged around lipid droplets, playing an important role in cellular energy and lipid metabolism. PDM enhance fatty acid  $\beta$ -oxidation and ATP synthesis, providing necessary energy for cellular metabolic activities, and support triglyceride synthesis and the maintenance of lipid homeostasis. Additionally, PDM capture free fatty acids, reducing their accumulation within cells, thereby decreasing lipotoxicity and protecting cells from damage caused by lipid overload. These functions make PDM crucial in regulating cellular metabolism [90]. In contrast, cytosolic mitochondria (CM) mainly regulate cellular energy metabolism through the transport and oxidation of fatty acids, complementing the role of PDM in lipid metabolism.

In the pathological mechanisms of MASLD, abnormal accumulation of hepatic lipid droplets and mitochondrial dysfunction are key features. PLIN5, as a lipid droplet-associated protein, regulates PDM, promoting fatty acid release and oxidative metabolism, thereby reducing lipid droplet accumulation and lipotoxicity, and maintaining the balance of hepatic lipid metabolism [91,92]. Additionally, PLIN5 plays a dual role in lipolysis and lipid metabolism, depending on the cell's energy status and hormonal environment. After catecholamine stimulation, PLIN5 is phosphorylated and translocates to the nucleus, where it forms a complex with SIRT1 and PGC-1 $\alpha$ . By activating the deacetylase activity of SIRT1, PLIN5 enhances the deacetylation of PGC-1 $\alpha$ , thereby promoting the expression of PGC- $1\alpha$ -regulated fatty acid oxidation genes. This mechanism ensures that the fatty acids released during lipolysis are efficiently utilized by mitochondria, avoiding mitochondrial overload and maintaining their function and metabolic efficiency [93]. Recent research by Xiangyun Sun et al. has shown that diethyldithiocarbamate (DDC) can upregulate the expression of PLIN5, thereby promoting the formation of PDM. DDC improves mitochondrial function by enhancing mitochondrial OXPHOS levels and increasing ATP production. This mechanism not only promotes the synthesis of triglycerides (TG) but also significantly alleviates mitochondrial dysfunction caused by MASLD, reducing liver inflammation and fibrosis. Consequently, it inhibits the progression of MASLD to more severe stages, such as metabolic-associated steatohepatitis (MASH) [94].

# 4.4. Mitophagy and MASLD

Mitophagy refers to the selective degradation of damaged and dysfunctional mitochondria by the cell, which helps maintain the normal structure, quantity, and function of mitochondria [95]. In the liver, there are two main pathways of mitochondrial autophagy: ubiquitindependent pathway and ubiquitin-independent pathway. In the ubiquitin-dependent pathway, E3 ubiquitin ligases such as Parkin bind ubiquitin to mitochondrial outer membrane proteins and recruit autophagy receptors such as p62, OPTN and NDP52. These autophagy receptors contain Microtubule-associated protein 1 light chain 3 (LC3) interacting regions (LIRs), which can bind to LC3 and anchor ubiquitinated mitochondria to the autophagosome membrane, leading to the degradation of damaged mitochondria. In the ubiquitin-independent pathway, receptors on the mitochondrial outer membrane, such as FUN14 domain-containing protein 1, contain LIR sequences and can directly bind to LC3 under stress conditions to form autophagosome membranes that encapsulate damaged mitochondria and complete

mitophagy [96]. Studies have shown that upregulation of macrophage stimulating factor 1 (Mst1) leads to liver vacuolation, steatosis, fibrosis, oxidative stress, and inflammation, and plays a key role in HFD-induced MASLD. Mst1 gene knockout alleviates HFD-induced liver damage by activating the AMP-activated protein kinase (AMPK) signaling pathway, regulating Parkin-related mitophagy [97]. Additionally, as a key protein in glucose regulation, AMPK also regulates mitophagy through the ubiquitin-independent pathway by activating the autophagy-related protein Unc-51-like kinase 1 [98].

In MASLD, an imbalance between mitochondrial fusion and fission leads to abnormally enlarged mitochondria, affecting the mitophagy process. DRP1 and OPA1 respectively regulate mitochondrial fission and fusion in hepatocytes, with their actions being antagonistic to each other. Research has found that simultaneous knockout of DRP1 and OPA1 in mice maintains mitochondrial morphology stability similar to that of the control group. In MASLD model mice, OPA1 knockout significantly reduces mitochondrial size, thereby promoting mitophagy and preventing the progression of MASLD [99].

# 4.5. mt DNA and MASLD

Patients with MASLD exhibit a higher mutation rate in mitochondrial DNA (mt DNA) compared to healthy individuals [14]. Studies have identified that the progression of MASLD is closely related to nucleotide variations in the D-loop region of mt DNA. Specifically, the m.16318C>A mutation is associated with NASH, while the Mt16129AA homozygous mutation genotype correlates with a higher degree of liver fibrosis [100]. In another cohort of MASLD patients, the mtDNA mutation profile in the liver includes several point mutations related to the disease state. These mutations primarily target the OXPHOS system and affect phenotypes. Notably, the m.14766 C > T missense mutation in the mitochondrial cytochrome b (MT-CYB) gene is predicted to be a deleterious mutation. MASLD patients carrying this mutation exhibit significant mitochondrial morphological changes under electron microscopy, including increased mitochondrial volume, condensed mitochondrial matrix, loss of mitochondrial membranes and cristae, and peroxisome proliferation [14]. Additionally, studies have indicated that MT-CYB mutations are associated with histological features such as steatosis, inflammation, and fibrosis. Lipid peroxidation and ROS are suggested as the causes of MT-CYB mutations [101].

# 4.6. Interactions between mitochondria and endoplasmic reticulum in MASLD

In 1971, Franke et al. observed a close association between the mitochondrial outer membrane and the endoplasmic reticulum (ER) in vivo [102]. Mitochondria-associated endoplasmic reticulum membranes (MAM) are structures rich in phospholipid-synthesizing enzymes such as diacylglycerol acyltransferase, phosphatidylserine synthase, and cholesterol acyltransferase, primarily responsible for lipid transport between the ER and mitochondria [103]. MAMs contain many proteins, including inositol 1,4,5-triphosphate receptor (IP3R), voltage-dependent anion channel (VDAC), Sigma-1 receptor, glucose-regulated protein 75, and various calcium-binding partners. These proteins mediate efficient calcium ion transfer from the ER to mitochondria, participating in the regulation of cellular energy metabolism, survival, and death processes [104].

In MASLD, loss of CDP-diacylglycerol synthase 2 (CDS2) results in reduced mitochondrial content in the liver and reduced activity of ETC complexes I and II, along with triglyceride Ester, cholesterol, ALT and AST levels are increased. Furthermore, reduced CDS2 expression altered the protein composition of MAMs, including significant downregulation of the mitochondrial protein Mitofusin 2 (MFN2). [105]. MFN2 is a mitochondrial membrane protein that connects mitochondria and the ER membrane, selectively binding and

**Table 1**Therapeutic intervention for mitochondria in MASLD.

Categories	Therapeutics	Main effects	Reference
Non-pharmacological interventions	Physical activity	Enhances hepatic mitochondrial fatty acid oxidation and modulates the structure of mitochondrial permeability transition pores, thus reducing mitochondrial damage	[109]
	Ketogenic diet	Markedly altered hepatic mitochondrial fluxes and redox state	[110]
	Metabolic surgery	Increases oxidative phosphorylation, enhances citrate synthase activity, promotes ETC gene expression, and reduces oxidative stress.	[111]
Chemical agonists/inhibitors	17-AAG	Suppresses lipid synthesis via inhibiting Heat shock protein 90; prevents excessive oxidation and damage of mitochondria by promoting the formation of albumosomes that capture CPT2	<u>[126]</u>
	A54556A (caseinolytic proteaseP activator)	Reduces PA/OA-induced cytokine expression and triglyceride Accumulation; protects the liver damage and fat deposition	<u>[</u> 127]
	JT003 (adiponectin-based agonist)	Enhances fatty acid uptake, $\beta$ -oxidation, and mitochondrial utilization by activating AMPK and PPAR $\gamma$ ; inhibits lipogenesis by activating PI3K-Akt; alleviates liver fibrosis by suppressing the activation of hepatic stellate cells	[128]
Natural extracts (The following substances have not been found to be hepatotoxic.)	Spermidine	Improves of mitochondrial fatty acid oxidation by maintaining hypusination of Eukaryotic Initiation Factor 5A	<u>[129]</u>
	Uroguanylin	Activates AMPK-induced mitochondrial fatty acid $\beta$ -oxidation; augments mitochondrial oxidative phosphorylation and preserves mtDNA content.	<u>[</u> 130]
	Silybin	Reduces endoplasmic reticulum stress and improves mitochondrial respiratory chain complex activity, thereby enhances oxidative phosphorylation; inhibits the release of cytochrome c in mitochondria, prevent mitochondrial fragmentation,	[131]
	Astaxanthin	Activates PPAR $-\alpha$ while inhibiting PPAR $-\gamma$ , thus reducing lipid accumulation in hepatocytes; regulates fatty acid oxidation and mitochondrial biogenesis; regulates fatty acid oxidation and mitochondrial biogenesis; attenuates hepatic damage and mitochondrial dysfunction in non—alcoholic fatty liver disease by up—regulating the FGF21/PGC $-1\alpha$	[132–134]
Drugs	Pioglitazone	Alleviate mitochondrial energy deficit by slowing down the TCA cycle and cardiolipin remodeling	[112]
	Sorafenib	Increased mitochondrial uncoupling to activate AMPK activity	[135]
	Survodutide	Drives increased gluconeogenesis and glycogenolysis, reduced hepatic lipid accumulation, increased mitochondrial turnover, and reduced oxidative stress	<u>[125]</u>
	Cotadutide	Inhibits hepatic lipogenesis, enhances mitochondrial turnover and oxida- tive capacity by activating GCGR	<u>[124]</u>
	Semaglutide	Reduces hepatic lipid deposition, inflammation, and fibrosis; improves mitochondrial structure.	[136]
	Resmetirom	Restores $\beta$ -oxidation function of mitochondrial	[120]

separating phosphatidylserine (PS) from the membrane, forming PS-rich rigid domains. Reduced MFN2 expression in MASLD leads to decreased PS transport from the ER to mitochondria, thereby inhibiting the synthesis of phosphatidylcholine in mitochondria. Disruption of phospholipid metabolism-induced hepatic MAM remodeling has been shown to result in triglyceride accumulation and insulin resistance, thereby promoting the progression of MASLD [106].

With industrial development, the impact of environmental factors on the progression of MASLD has become increasingly significant. In mice exposed to Nanoplastic (NP), a common environmental pollutant, mitochondrial swelling, cristae disappearance, endoplasmic reticulum dilation, and more frequent contacts between mitochondria and the endoplasmic reticulum were observed, resulting in the formation of more MAMs. Additionally, NP increases the levels of IP3R1, GRP75, and VDAC1 in MAMs by binding to IP3R1, enhancing the stability of these proteins and promoting MAM formation. The increased MAMs enhance the uptake of Ca2+ by mitochondria, leading to mitochondrial Ca2+ overload and excessive production of ROS, which suppresses downstream NRF2. By binding to the promoter region of the microRNA-26a (miR26a) gene, downregulated NRF2 inhibits the expression of miR26a, which promotes the expression of the target gene VDAC1 and facilitates the nuclear translocation of nuclear factor p65 (p65) and kelch-like ECH-associated protein 1, leading to the inactivation of NRF2. Downregulation of miR26a forms a positive feedback loop that exacerbates oxidative stress, steatosis, inflammation, and fibrosis in the liver, thereby promoting the progression of MASLD [107].

# 5. Mitochondrial dysfunction: a therapeutic target for MASLD

Recent clinical studies have evaluated the impact of dietary interventions and bariatric surgery on hepatic mitochondria, which are emerging as a promising therapeutic target for MASLD.

Numerous studies have demonstrated that enhancing hepatic mitochondrial  $\beta$ -oxidation through non-pharmacological treatments can reduce the accumulation of free fatty acids, thereby alleviating hepatic steatosis and inflammation. For instance, glycine treatment has been shown to enhance mitochondrial activity and reduce lipid accumulation [108]. Regular physical activity is another effective non-pharmacological intervention, as it improves the pathological characteristics of NAFLD by enhancing hepatic mitochondrial fatty acid oxidation and modulating the structure of PTP, thus reducing mitochondrial damage [109]. Caloric restriction is an effective strategy for treating metabolic disorders. The ketogenic diet can significantly improve liver fatty acid content, and metabolic surgery has been shown to significantly affect patient weight, increase OXPHOS capacity, enhance citrate synthase activity, promote ETC gene expression, and reduce oxidative stress [110,111].

In the pharmacological treatment of MASLD, pioglitazone ameliorates insulin resistance by increasing mitochondrial activity and reducing lipid accumulation [112]. Vitamin E alleviates mitochondrial dysfunction through its powerful antioxidant capacity. It can scavenge ROS and reactive nitrogen species (RNS) to protect mitochondria from oxidative damage [113]. In addition, vitamin E enhances the activities of antioxidant enzymes such as superoxide dismutase,

catalase, and glutathione peroxidase, enhancing cellular antioxidant defense [114-116]. Vitamin E also improves mitochondrial membrane potential by increasing the level of anti-apoptotic protein BCL-2, reducing the level of pro-apoptotic proteins BAX and p53, and inhibiting caspase activity, thereby reducing cell apoptosis [117]. In March 2024, the U.S. Food and Drug Administration (FDA) approved resmetirom to treat MASH with moderate to advanced fibrosis for the first time. Its derivative, HSK31679, is currently undergoing Phase II clinical trials [118,119]. Resmetirom alleviates MASH by targeting the liver-specific thyroid hormone receptor  $\beta$ , stimulating hepatic fatty acid  $\beta$ -oxidation, and thereby reducing the burden of lipotoxic lipids [120]. Additionally, Resmetirom decreases oxidative stress and inflammation, lowering the risk of fibrosis, and promotes mitochondrial biogenesis to maintain mitochondrial health. For example, in cultured HepG2–THR- $\beta$  cells from mouse models, thyroid hormone treatment increased mitochondrial respiration and fatty acid oxidation under both basal and palmitic acid-treated conditions, while reducing inflammation and fibrotic responses induced by lipopolysaccharides and palmitic acid stimulation [121–123]. Cotadutide is a novel dual agonist that activates both the glucagon-like peptide-1 receptor and the glucagon receptor (GCGR), exerting its therapeutic effects by inhibiting fatty acid synthesis through GCGR-mediated pathways. In addition, it promotes mitochondrial turnover, facilitates the maintenance of mitochondrial function, and alleviates hepatic steatosis, phosphorylation, and inflammation [124]. Another dual agonist, survodutide, enhances gluconeogenesis and glycogenolysis while reducing hepatic lipid accumulation. It also supports mitochondrial renewal and reduces oxidative stress, further demonstrating its potential in metabolic and liver disease interventions [125]. Recent study has found that endogenous unfolded albumin in the cytoplasm undergoes phase separation to form membrane-less, shell-like organelles called proteosomes. Carnitine palmitoyltransferase 2 (CPT2) transports fatty acids from the mitochondrial outer membrane into the matrix for  $\beta$ -oxidation, while proteosomes capture CPT2 and regulate its entry into mitochondria, thereby modulating mitochondrial metabolism and respiration. The Hsp90 inhibitor 17-Allylamino-17demethoxygeldanamycin (17-AAG) can promote the accumulation of hepatic proteosomes, thereby inhibiting the progression of NAFLD, reducing mitochondrial oxidative stress in the early stages of MASLD, and decreasing ROS production [126].

In summary, mitochondrial dysfunction represents a promising target for the treatment of MASLD. The combination of non-pharmacological and pharmacological interventions, including increased physical activity, dietary modifications, anti-diabetic medications, and thyroxine analogs, has been shown to significantly enhance mitochondrial function, thereby alleviating the symptoms of MASLD and slowing its progression (Table 1).

# 6. Conclusions

MASLD, a highly prevalent liver disease, has a complex and multifaceted pathogenesis involving insulin resistance, immune response, gut microbiota, and genetic and epigenetic factors, among others. Mitochondrial dysfunction plays a crucial role in the occurrence and development of MASLD, including impaired  $\beta$ -oxidation, PDM, oxidative stress, defective mitophagy, and mtDNA mutations (Fig. 3). The reduction in fatty acid oxidation and mitochondrial enzyme activity precedes the onset of insulin resistance and MASLD, with decreased insulin sensitivity and diminished levels of free radical scavenging enzymes such as superoxide dismutase exacerbating the loss of mitochondrial function [137]. In recent years, research into the relationship between mitochondria and MASLD has deepened, revealing how mitochondrial dysfunction accelerates MASLD progression by affecting hepatocyte metabolism, oxidative stress, and apoptosis. Particularly, the interaction between mitochondria and the endoplasmic reticulum in lipid metabolism regulation underscores the central role

of mitochondria in the pathogenesis of MASLD. In the future, diagnostic and therapeutic strategies targeting mitochondrial dysfunction are expected to become new directions for clinical intervention in MASLD, providing more treatment options and better prognoses for patients.

#### **Authors contributions**

Chenyang Mu conceived and wrote the manuscript. Sijie Wang and Fu Yang conceived and revised the manuscript. All authors collected and analyzed the data.

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# Availability of data and materials

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# **Declaration of interests**

The authors declare no competing interests.

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#### References

- Chan W-K, Chuah K-H, Rajaram RB, Lim L-L, Ratnasingam J, Vethakkan SR. Metabolic dysfunction-associated steatotic liver disease (MASLD): a state-of-the-art review. J Obes Metab Syndr 2023;32:197–213. https://doi.org/10.7570/ iomzc20252
- [2] Eslam M, Newsome PN, Sarin SK, Anstee QM, Targher G, Romero-Gomez M, et al. A new definition for metabolic dysfunction-associated fatty liver disease: an international expert consensus statement. J Hepatol 2020;73:202–9. https://doi. org/10.1016/j.jhep.2020.03.039.
- [3] Wong VW-S, Ekstedt M, Wong GL-H, Hagström H. Changing epidemiology, global trends and implications for outcomes of NAFLD. J Hepatol 2023;79:842– 52. https://doi.org/10.1016/j.jhep.2023.04.036.
- [4] Lee BP, Dodge JL, Terrault NA. National prevalence estimates for steatotic liver disease and subclassifications using consensus nomenclature. Hepatology 2024;79:666–73. https://doi.org/10.1097/HEP.00000000000000604.
- [5] Estes C, Anstee QM, Arias-Loste MT, Bantel H, Bellentani S, Caballeria J, et al. Modeling NAFLD disease burden in China, France, Germany, Italy, Japan, Spain, United Kingdom, and United States for the period 2016-2030. J Hepatol 2018;69:896–904. https://doi.org/10.1016/j.jhep.2018.05.036.
- [6] Korhan Peyda, Bağırsakçı Ezgi, Öztemur Islakoğlu Yasemin, Solmaz Gülhas, Sarı-kaya Burcu, Nart Deniz, Yılmaz Funda, Atabey Neşe. MASLD-mimicking micro-environment drives an aggressive phenotype and represses IDH2 expression in hepatocellular carcinoma. Hepatoma Res 2024;10:22. https://doi.org/10.20517/2394-5079.2023.140.
- [7] Ertle J, Dechêne A, Sowa J-P, Penndorf V, Herzer K, Kaiser G, et al. Non-alcoholic fatty liver disease progresses to hepatocellular carcinoma in the absence of apparent cirrhosis. Int J Cancer 2011;128:2436–43. https://doi.org/10.1002/ iic.25797.
- [8] Miao L, Targher G, Byrne CD, Cao Y-Y, Zheng M-H. Current status and future trends of the global burden of MASLD. Trends Endocrinol Metab 2024;35:697– 707. https://doi.org/10.1016/j.tem.2024.02.007.
- [9] Chen Z, Tian R, She Z, Cai J, Li H. Role of oxidative stress in the pathogenesis of nonalcoholic fatty liver disease. Free Radic Biol Med 2020;152:116–41. https:// doi.org/10.1016/i.freeradbiomed.2020.02.025.
- [10] Filomeni G, Rotilio G, Ciriolo MR. Cell signalling and the glutathione redox system. Biochem Pharmacol 2002;64:1057–64. https://doi.org/10.1016/s0006-2952(02)01176-0.
- [11] Flohé L. Changing paradigms in thiology from antioxidant defense toward redox regulation. Methods Enzymol 2010;473:1–39. https://doi.org/10.1016/S0076-6879(10)73001-9.
- [12] Greenberg AS, Coleman RA, Kraemer FB, McManaman JL, Obin MS, Puri V, et al. The role of lipid droplets in metabolic disease in rodents and humans. J Clin Invest 2011;121:2102–10. https://doi.org/10.1172/JCI46069.

[13] Rashid H-O, Yadav RK, Kim H-R, Chae H-J. ER stress: autophagy induction, inhibition and selection. Autophagy 2015;11:1956-77. https://doi.org/10.1080/ 5548627 2015 1091141

- [14] Sookoian S, Flichman D, Scian R, Rohr C, Dopazo H, Gianotti TF, et al. Mitochondrial genome architecture in non-alcoholic fatty liver disease. I Pathol 2016;240:437-49. https://doi.org/10.1002/path.4803
- [15] Day CP, James OF. Steatohepatitis: a tale of two "hits"? Gastroenterology 1998;114:842-5. https://doi.org/10.1016/s0016-5085(98)70599-2.
- Leamy AK, Egnatchik RA, Young JD. Molecular mechanisms and the role of saturated fatty acids in the progression of non-alcoholic fatty liver disease. Prog Lipid Res 2013;52:165-74. https://doi.org/10.1016/j.plipres.2012.10.004.
- Buzzetti E, Pinzani M, Tsochatzis EA. The multiple-hit pathogenesis of non-alcoholic fatty liver disease (NAFLD). Metabolism 2016;65:1038-48. https://doi.org/ 10.1016/j.metabol.2015.12.012.
- [18] Schreuder TCMA, Verwer BJ, van Nieuwkerk CMJ, Mulder CJJ. Nonalcoholic fatty liver disease: an overview of current insights in pathogenesis, diagnosis and treatment. World J Gastroenterol 2008;14:2474-86. https://doi.org/10.3748/ wjg.14.2474.
- [19] Stefan N, Kantartzis K, Häring H-U. Causes and metabolic consequences of Fatty liver. Endocr Rev 2008;29:939-60 2019041121355393300.
- Postic C, Girard J. Contribution of de novo fatty acid synthesis to hepatic steatosis and insulin resistance: lessons from genetically engineered mice. J Clin Invest 2008;118:829-38. https://doi.org/10.1172/JCI34275.
- Wellen KE, Hotamisligil GS. Inflammation, stress, and diabetes. J Clin Invest 2005:115:1111-9.
- [22] Hotamisligil GS. Role of endoplasmic reticulum stress and c-Jun NH2-terminal kinase pathways in inflammation and origin of obesity and diabetes. Diabetes 2005;54(Suppl 2):S73-8. https://doi.org/10.2337/diabetes.54.suppl\_2.s73
- Moore MP, Cunningham RP, Meers GM, Johnson SA, Wheeler AA, Ganga RR, et al. Compromised hepatic mitochondrial fatty acid oxidation and reduced markers of mitochondrial turnover in human NAFLD. Hepatology 2022;76:1452-65. https://doi.org/10.1002/hep.32324.
- Friedman SL, Neuschwander-Tetri BA, Rinella M, Sanyal AJ. Mechanisms of NAFLD development and therapeutic strategies. Nat Med 2018;24:908-22. https://doi.org/10.1038/s41591-018-0104-9.
- [25] Kubes P, Jenne C. Immune Responses in the Liver. Annu Rev Immunol 2018;36:247-77. https://doi.org/10.1146/annurev-immunol-051116-052415.
- Baffy G. Kupffer cells in non-alcoholic fatty liver disease: the emerging view. J Hepatol 2009;51:212-23. https://doi.org/10.1016/j.jhep.2009.03.008.
- Sarkar A, Mitra P, Lahiri A, Das T, Sarkar J, Paul S, et al. Butyrate limits inflammatory macrophage niche in NASH. Cell Death Dis 2023;14:332. https://doi.org/ 10.1038/s41419-023-05853-6.
- [28] Neri T, Armani C, Pegoli A, Cordazzo C, Carmazzi Y, Brunelleschi S, et al. Role of NF-kappaB and PPAR-gamma in lung inflammation induced by monocytederived microparticles. Eur Respir J 2011;37:1494-502, https://doi.org/10.1183/ 09031936 00023310
- [29] Luo W, Xu Q, Wang Q, Wu H, Hua J. Effect of modulation of PPAR- $\gamma$  activity on Kupffer cells M1/M2 polarization in the development of non-alcoholic fatty liver disease. Sci Rep 2017;7:44612. https://doi.org/10.1038/srep44612.
- Martín-Mateos R, Albillos A. The role of the gut-liver axis in metabolic dysfunction-associated fatty liver disease. Front Immunol 2021;12:660179. https://doi. org/10.3389/fimmu.2021.660179.
- [31] Meijnikman AS, Davids M, Herrema H, Aydin O, Tremaroli V, Rios-Morales M, et al. Microbiome-derived ethanol in nonalcoholic fatty liver disease. Nat Med 2022;28:2100-6. https://doi.org/10.1038/s41591-022-02016-6.
- [32] Willner IR, Waters B, Patil SR, Reuben A, Morelli J, Riely CA. Ninety patients with nonalcoholic steatohepatitis: insulin resistance, familial tendency, and severity of disease. Am J Gastroenterol 2001;96:2957-61. https://doi.org/10.111 .1572-0241.2001.04667.x.
- [33] Wajsbrot NB, Leite NC, Franca PHC, Cardoso CRL, Salles GF, Villela-Nogueira CA. Parental history of Type 2 diabetes mellitus and PNPLA3 polymorphism increase the risk of severe stages of nonalcoholic fatty liver disease. Dig Dis Sci 2024;69:634–42. https://doi.org/10.1007/s10620-023-08214-7.
- [34] Schwimmer JB, Celedon MA, Lavine JE, Salem R, Campbell N, Schork NJ, et al. Heritability of nonalcoholic fatty liver disease. Ga 2009;136:1585–92. https://doi.org/10.1053/j.gastro.2009.01.050. Gastroenterology
- Makkonen J, Pietiläinen KH, Rissanen A, Kaprio J, Yki-Järvinen H. Genetic factors contribute to variation in serum alanine aminotransferase activity independent of obesity and alcohol: a study in monozygotic and dizygotic twins. J Hepatol 2009;50:1035-42. https://doi.org/10.1016/j.jhep.2008.12.025
- [36] Rotman Y, Koh C, Zmuda JM, Kleiner DE, Liang TJ, NASH CRN. The association of genetic variability in patatin-like phospholipase domain-containing protein 3 (PNPLA3) with histological severity of nonalcoholic fatty liver disease. Hepatology 2010;52:894–903. https://doi.org/10.1002/hep.23759
- Valenti L, Al-Serri A, Daly AK, Galmozzi E, Rametta R, Dongiovanni P, et al. Homozygosity for the patatin-like phospholipase-3/adiponutrin I148M polymorphism influences liver fibrosis in patients with nonalcoholic fatty liver disease. Hepatology 2010;51:1209-17. https://doi.org/10.1002/hep.23622
- [38] Beer NL, Tribble ND, McCulloch LJ, Roos C, Johnson PRV, Orho-Melander M, et al. The P446L variant in GCKR associated with fasting plasma glucose and triglyceride levels exerts its effect through increased glucokinase activity in liver. Hum Mol Genet 2009;18:4081-8. https://doi.org/10.1093/hmg/ddp353
- [39] Kozlitina J, Smagris E, Stender S, Nordestgaard BG, Zhou HH, Tybjærg-Hansen A, et al. Exome-wide association study identifies a TM6SF2 variant that confers susceptibility to nonalcoholic fatty liver disease. Nat Genet 2014;46:352-6. https://doi.org/10.1038/ng.2901.

[40] Al-Serri A, Anstee QM, Valenti L, Nobili V, Leathart JBS, Dongiovanni P, et al. The SOD2 C47T polymorphism influences NAFLD fibrosis severity: evidence from case-control and intra-familial allele association studies. J Hepatol 2012;56:448-54. https://doi.org/10.1016/j.jhep.2011.05.029.

- [41] Dong H, Wang J, Li C, Hirose A, Nozaki Y, Takahashi M, et al. The phosphatidylethanolamine N-methyltransferase gene V175M single nucleotide polymorphism confers the susceptibility to NASH in Japanese population. J Hepatol 2007;46:915-20. https://doi.org/10.1016/j.jhep.2006.12.012
- [42] Yoneda M, Hotta K, Nozaki Y, Endo H, Uchiyama T, Mawatari H, et al. Association between angiotensin II type 1 receptor polymorphisms and the occurrence of nonalcoholic fatty liver disease. Liver Int 2009;29:1078-85. https://doi.org/ 10.1111/j.1478-3231.2009.01988.x.
- [43] Narla G, Difeo A, Reeves HL, Schaid DJ, Hirshfeld J, Hod E, et al. A germline DNA polymorphism enhances alternative splicing of the KLF6 tumor suppressor gene and is associated with increased prostate cancer risk. Cancer Res 2005;65:1213-22. https://doi.org/10.1158/0008-5472.CAN-04-4249.
- [44] Wree A, Broderick L, Canbay A, Hoffman HM, Feldstein AE. From NAFLD to NASH to cirrhosis-new insights into disease mechanisms. Nat Rev Gastroenterol Hepatol 2013;10:627-36. https://doi.org/10.1038/nrgastro.2013.149.
- [45] Chen H-C, Chen Y-Z, Wang C-H, Lin F-J. The nonalcoholic fatty liver disease-like phenotype and lowered serum VLDL are associated with decreased expression and DNA hypermethylation of hepatic ApoB in male offspring of ApoE deficient mothers fed a with Western diet. J Nutr Biochem 2020;77:108319. https://doi. org/10.1016/j.jnutbio.2019.108319.
- [46] Baumeier C, Saussenthaler S, Kammel A, Jähnert M, Schlüter L, Hesse D, et al. Hepatic DPP4 DNA methylation associates with fatty liver. Diabetes 2017;66:25-35. https://doi.org/10.2337/db15-1716.
- [47] Hosseini H, Teimouri M, Shabani M, Koushki M, Babaei Khorzoughi R, Namvarjah F, et al. Resveratrol alleviates non-alcoholic fatty liver disease through epigenetic modification of the Nrf2 signaling pathway. Int J Biochem Cell Biol 2020;119:105667. https://doi.org/10.1016/j.biocel.2019.105667.
- [48] Anstee QM, Concas D, Kudo H, Levene A, Pollard J, Charlton P, et al. Impact of pan-caspase inhibition in animal models of established steatosis and non-alcoholic steatohepatitis. J Hepatol 2010;53:542–50. https://doi.org/10.1016/j. ihep.2010.03.016.
- [49] Iredale JP. Models of liver fibrosis: exploring the dynamic nature of inflammation and repair in a solid organ. J Clin Invest 2007;117:539-48. https://doi.org/ 10.1172/ICI30542
- [50] Chen Q, Yan M, Cao Z, Li X, Zhang Y, Shi J, et al. Sperm tsRNAs contribute to intergenerational inheritance of an acquired metabolic disorder. Science 2016;351:397-400. https://doi.org/10.1126/science.aad797
- [51] Frey TG, Mannella CA. The internal structure of mitochondria. Trends Biochem Sci 2000;25:319-24. https://doi.org/10.1016/s0968-0004(00)01609-1.
- [52] Mansouri A, Gattolliat C-H, Asselah T. Mitochondrial dysfunction and signaling in chronic liver diseases. Gastroenterology 2018;155:629-47, https://doi.org/ 10.1053/j.gastro.2018.06.083.
- Chan DC. Mitochondrial dynamics and its involvement in disease. Annu Rev Pathol 2020;15;235-59, https://doi.org/10.1146/annurev-pathmechdis-012419-032711.
- [54] Liu X, Weaver D, Shirihai O, Hajnóczky G. Mitochondrial "kiss-and-run": interplay between mitochondrial motility and fusion-fission dynamics. EMBO J 2009;28:3074–89. https://doi.org/10.1038/emboj.2009.255.
- [55] Prudent J, McBride HM. Mitochondrial dynamics: ER actin tightens the Drp1 noose, Curr Biol 2016;26:R207-9, https://doi.org/10.1016/j.cub.2016.01.009.
- [56] Lee JE, Westrate LM, Wu H, Page C, Voeltz GK. Multiple dynamin family members collaborate to drive mitochondrial division. Nature 2016;540:139-43. https://doi.org/10.1038/nature20555.
- Saraste M. Oxidative phosphorylation at the fin de siècle. Science 1999;283:1488–93. Smeitink I, van den Heuvel L, DiMauro S. The genetics and pathology of oxidative
- [59] Mitchell P. Coupling of phosphorylation to electron and hydrogen transfer by a chemi-osmotic type of mechanism. Nature 1961;191:144-8.

phosphorylation. Nat Rev Genet 2001;2:342-52.

- [60] Szabo I, Zoratti M. Mitochondrial channels: ion fluxes and more. Physiol Rev 2014;94:519-608. https://doi.org/10.1152/physrev.00021.2013.
- [61] Cho D-H, Nakamura T, Fang J, Cieplak P, Godzik A, Gu Z, Lipton SA. S-nitrosylation of Drp1 mediates beta-amyloid-related mitochondrial fission and neuronal injury. Science 2009;324:102–5. https://doi.org/10.1126/science.1171091
- Bossy B, Petrilli A, Klinglmayr E, Chen J, Lütz-Meindl U, Knott AB, et al. S-Nitrosylation of DRP1 does not affect enzymatic activity and is not specific to Alzheimer's disease. J Alzheimers Dis 2010;20(Suppl 2):S513-26. https://doi.org/ 10.3233/JAD-2010-100552.
- [63] Leboucher GP, Tsai YC, Yang M, Shaw KC, Zhou M, Veenstra TD, et al. Stressinduced phosphorylation and proteasomal degradation of mitofusin 2 facilitates mitochondrial fragmentation and apoptosis. Mol Cell 2012;47:547-57. https:// doi.org/10.1016/j.molcel.2012.05.041.
- [64] Chung KKK, Thomas B, Li X, Pletnikova O, Troncoso JC, Marsh L, Dawson VL, Dawson TM. S-nitrosylation of parkin regulates ubiquitination and compromises parkin's protective function. Science 2004;304:1328-31.
- [65] Mandavilli BS, Santos JH, Van Houten B. Mitochondrial DNA repair and aging. Mutat Res 2002;509:127-51. https://doi.org/10.1016/s0027-5107(02)00220-8.
- [66] Shidara Y, Yamagata K, Kanamori T, Nakano K, Kwong JQ, Manfredi G, et al. Positive contribution of pathogenic mutations in the mitochondrial genome to the promotion of cancer by prevention from apoptosis. Cancer Res 2005;65:1655-63. https://doi.org/10.1158/0008-5472.CAN-04-2012.
- [67] Petros JA, Baumann AK, Ruiz-Pesini E, Amin MB, Sun CQ, Hall J, et al. mtDNA mutations increase tumorigenicity in prostate cancer. Proc Natl Acad Sci U S A 2005;102:719-24. https://doi.org/10.1073/pnas.0408894102.

[68] Park JS, Sharma LK, Li H, Xiang R, Holstein D, Wu J, et al. A heteroplasmic, not homoplasmic, mitochondrial DNA mutation promotes tumorigenesis via alteration in reactive oxygen species generation and apoptosis. Hum Mol Genet 2009;18:1578–89. https://doi.org/10.1093/hmg/ddp069.

- [69] Imanishi H, Hattori K, Wada R, Ishikawa K, Fukuda S, Takenaga K, et al. Mito-chondrial DNA mutations regulate metastasis of human breast cancer cells. PLoS One 2011;6:e23401. https://doi.org/10.1371/journal.pone.0023401.
- [70] Horton TM, Petros JA, Heddi A, Shoffner J, Kaufman AE, Graham SD, et al. Novel mitochondrial DNA deletion found in a renal cell carcinoma. Genes Chromosomes Cancer 1996;15:95–101. https://doi.org/10.1002/(SICI)1098-2264 (199602)15:2<95::AID-GCC3>3.0.CO:2-Z.
- [71] Kujoth GC, Hiona A, Pugh TD, Someya S, Panzer K, Wohlgemuth SE, et al. Mito-chondrial DNA mutations, oxidative stress, and apoptosis in mammalian aging. Science 2005;309:481–4. https://doi.org/10.1126/science.1112125.
- [72] Roy MJ, Vom A, Czabotar PE, Lessene G. Cell death and the mitochondria: therapeutic targeting of the BCL-2 family-driven pathway. Br J Pharmacol 2014;171:1973–87. https://doi.org/10.1111/bph.12431.
- [73] Weiss JN, Korge P, Honda HM, Ping P. Role of the mitochondrial permeability transition in myocardial disease. Circ Res 2003;93:292–301. https://doi.org/ 10.1161/01.RES.0000087542.26971.D4.
- [74] Bock FJ, Tait SWG. Mitochondria as multifaceted regulators of cell death. Nat Rev Mol Cell Biol 2020;21:85–100. https://doi.org/10.1038/s41580-019-0173-8.
- [75] Houten SM, Violante S, Ventura FV, Wanders RJA. The biochemistry and physiology of mitochondrial fatty acid β-oxidation and its genetic disorders. Annu Rev Physiol 2016;78:23–44. https://doi.org/10.1146/annurev-physiol-021115-105045
- [76] Lee J, Choi J, Scafidi S, Wolfgang MJ. Hepatic fatty acid oxidation restrains systemic catabolism during starvation. Cell Rep 2016;16:201–12. https://doi.org/10.1016/j.celrep.2016.05.062.
- [77] Begriche K, Igoudjil A, Pessayre D, Fromenty B. Mitochondrial dysfunction in NASH: causes, consequences and possible means to prevent it. Mitochondrion 2006;6:1–28. https://doi.org/10.1016/j.mito.2005.10.004.
- [78] Huh JY, Reilly SM, Abu-Odeh M, Murphy AN, Mahata SK, Zhang J, et al. TANK-binding kinase 1 regulates the localization of acyl-CoA synthetase ACSL1 to control hepatic fatty acid oxidation. Cell Metab 2020;32:1012–27 e7. https://doi.org/10.1016/j.cmet.2020.10.010.
- [79] Zeng S, Wu F, Chen M, Li Y, You M, Zhang Y, et al. Inhibition of fatty acid translocase (FAT/CD36) palmitoylation enhances hepatic fatty acid β-oxidation by increasing its localization to mitochondria and interaction with long-chain acylcoa synthetase 1. Antioxid Redox Signal 2022;36:1081–100. https://doi.org/10.1089/ars.2021.0157.
- [80] Duan J, Wang Z, Duan R, Yang C, Zhao R, Feng Q, et al. Therapeutic targeting of hepatic ACSL4 ameliorates NASH in mice. Hepatology 2022;75:140–53. https:// doi.org/10.1002/hep.32148.
- [81] Filomeni G, De Zio D, Cecconi F. Oxidative stress and autophagy: the clash between damage and metabolic needs. Cell Death Differ 2015;22:377–88. https://doi.org/10.1038/cdd.2014.150.
- [82] Serviddio G, Bellanti F, Vendemiale G. Free radical biology for medicine: learning from nonalcoholic fatty liver disease. Free Radic Biol Med 2013;65:952–68. https://doi.org/10.1016/j.freeradbiomed.2013.08.174.
- [83] Sunny NE, Parks EJ, Browning JD, Burgess SC. Excessive hepatic mitochondrial TCA cycle and gluconeogenesis in humans with nonalcoholic fatty liver disease. Cell Metab 2011;14:804–10. https://doi.org/10.1016/j.cmet.2011.11.004.
- [84] Serviddio G, Bellanti F, Tamborra R, Rollo T, Capitanio N, Romano AD, et al. Uncoupling protein-2 (UCP2) induces mitochondrial proton leak and increases susceptibility of non-alcoholic steatohepatitis (NASH) liver to ischaemia-reperfusion injury. Gut 2008;57:957-65. https://doi.org/10.1136/gut.2007.147496.
- [85] Yang J, Fernández-Galilea M, Martínez-Fernández L, González-Muniesa P, Pérez-Chávez A, Martínez JA, et al. Oxidative stress and non-alcoholic fatty liver disease: effects of omega-3 fatty acid supplementation. Nutrients 2019;11:872. https://doi.org/10.3390/nu11040872.
- [86] Novo E, Busletta C, di Bonzo LV, Povero D, Paternostro C, Mareschi K, et al. Intracellular reactive oxygen species are required for directional migration of resident and bone marrow-derived hepatic pro-fibrogenic cells. J Hepatol 2011;54:964–74. https://doi.org/10.1016/j.jhep.2010.09.022.
- [87] Rachek LI, Yuzefovych LV, Ledoux SP, Julie NL, Wilson GL. Troglitazone, but not rosiglitazone, damages mitochondrial DNA and induces mitochondrial dysfunction and cell death in human hepatocytes. Toxicol Appl Pharmacol 2009;240:348–54. https://doi.org/10.1016/j.taap.2009.07.021.
- [88] Ricci C, Pastukh V, Leonard J, Turrens J, Wilson G, Schaffer D, et al. Mitochondrial DNA damage triggers mitochondrial-superoxide generation and apoptosis. Am J Physiol Cell Physiol 2008;294:C413–22. https://doi.org/10.1152/ajpcell.00362.2007.
- [89] Barbier-Torres L, Fortner KA, Iruzubieta P, Delgado TC, Giddings E, Chen Y, al Cet. Silencing hepatic MCJ attenuates non-alcoholic fatty liver disease (NAFLD) by increasing mitochondrial fatty acid oxidation. Nat Commun 2020;11:3360. https://doi.org/10.1038/s41467-020-16991-2.
- [90] Najt CP, Adhikari S, Heden TD, Cui W, Gansemer ER, Rauckhorst AJ, et al. Organelle interactions compartmentalize hepatic fatty acid trafficking and metabolism. Cell Rep 2023:42:112435. https://doi.org/10.1016/j.celrep.2023.112435.
- lism. Cell Rep 2023;42:112435. https://doi.org/10.1016/j.celrep.2023.112435.
  [91] Benador IV, Veliova M, Mahdaviani K, Petcherski A, Wikstrom JD, Assali E, Acín-Peréz R, et al. Mitochondria bound to lipid droplets have unique bioenergetics, composition, and dynamics that support lipid droplet expansion. Cell Metab 2018;27:869–85 e6. https://doi.org/10.1016/j.cmet.2018.03.003.
- [92] Wang H, Bell M, Sreenevasan U, Hu H, Liu J, Dalen K, et al. Unique regulation of adipose triglyceride lipase (ATGL) by perilipin 5, a lipid droplet-associated protein. J Biol Chem 2011;286:15707–15. https://doi.org/10.1074/jbc.M110.207779.

[93] Gallardo-Montejano VI, Saxena G, Kusminski CM, Yang C, McAfee JL, Hahner L, et al. Nuclear Perilipin 5 integrates lipid droplet lipolysis with PGC-1α/SIRT1-dependent transcriptional regulation of mitochondrial function. Nat Commun 2016;7:12723. https://doi.org/10.1038/ncomms12723.

- [94] Sun X, Yu Q, Qi Y, Kang B, Zhao X, Liu L, et al. Peridroplet mitochondria are associated with the severity of MASLD and the prevention of MASLD by diethyldithiocarbamate. J Lipid Res 2024;65:100590. https://doi.org/10.1016/j.iir.2024.100590.
- [95] Pickles S, Vigié P, Youle RJ. Mitophagy and quality control mechanisms in mitochondrial maintenance. Curr Biol 2018;28:R170–85. https://doi.org/10.1016/j. cub.2018.01.004.
- [96] Lu Y, Li Z, Zhang S, Zhang T, Liu Y, Zhang L. Cellular mitophagy: mechanism, roles in diseases and small molecule pharmacological regulation. Theranostics 2023;13:736–66. https://doi.org/10.7150/thno.79876.
- [97] Zhou T, Chang L, Luo Y, Zhou Y, Zhang J. Mst1 inhibition attenuates non-alco-holic fatty liver disease via reversing Parkin-related mitophagy. Redox Biol 2019;21:101120. https://doi.org/10.1016/j.redox.2019.101120.
- [98] Egan DF, Shackelford DB, Mihaylova MM, Gelino S, Kohnz RA, Mair W, et al. Phosphorylation of ULK1 (hATG1) by AMP-activated protein kinase connects energy sensing to mitophagy. Science 2011;331:456-61. https://doi.org/ 10.1126/science.1196371.
- [99] Yamada T, Murata D, Adachi Y, Itoh K, Kameoka S, Igarashi A, et al. Mitochondrial stasis reveals p62-mediated ubiquitination in parkin-independent mitophagy and mitigates nonalcoholic fatty liver disease. Cell Metab 2018;28:588–604 e5. https://doi.org/10.1016/j.cmet.2018.06.014.
- [100] Hasturk B, Yilmaz Y, Eren F. Potential clinical variants detected in mitochondrial DNA p-loop hypervariable region I of patients with non-alcoholic steatohepatitis. Hor-mones (Athens) 2019;18:463–75. https://doi.org/10.1007/s42000-019-00137-1.
- [101] Pirola CJ, Garaycoechea M, Flichman D, Castaño GO, Sookoian S. Liver mitochondrial DNA damage and genetic variability of Cytochrome b a key component of the respirasome drive the severity of fatty liver disease. J Intern Med 2021;289:84–96. https://doi.org/10.1111/joim.13147.
- [102] Franke WW, Kartenbeck J. Outer mitochondrial membrane continuous with endoplasmic reticulum. Protoplasma 1971;73:35–41. https://doi.org/10.1007/BF01286409.
- [103] Vance JE, Stone SJ, Faust JR. Abnormalities in mitochondria-associated membranes and phospholipid biosynthetic enzymes in the mnd/mnd mouse model of neuronal ceroid lipofuscinosis. Biochim Biophys Acta 1997;1344:286–99.
- [104] Wieckowski MR, Giorgi C, Lebiedzinska M, Duszynski J, Pinton P. Isolation of mitochondria-associated membranes and mitochondria from animal tissues and cells. Nat Protoc 2009;4:1582–90. https://doi.org/10.1038/nprot.2009.151.
- [105] Xu J, Chen S, Wang W, Man Lam S, Xu Y, Zhang S, Pan H, Liang J, Huang X, Wang Y, Li T, Jiang Y, Wang Y, Ding M, Shui G, et al. Hepatic CDP-diacylglycerol synthase 2 deficiency causes mitochondrial dysfunction and promotes rapid progression of NASH and fibrosis. Sci Bull (Beijing) 2022;67:299–314. https://doi.org/10.1016/j.scib.2021.10.014.
- [106] Hernández-Alvarez MI, Sebastián D, Vives S, Ivanova S, Bartoccioni P, Kakimoto P, et al. Deficient endoplasmic reticulum-mitochondrial phosphatidylserine transfer causes liver disease. Cell 2019;177:881–95 e17. https://doi.org/10.1016/j.cell.2019.04.010.
- [107] Wei J, Liu J, Wang H, Wen K, Ni X, Lin Y, et al. Nanoplastic propels diet-induced NAFL to NASH via ER-mitochondrial tether-controlled redox switch. J Hazard Mater 2024;465:133142. https://doi.org/10.1016/j.jhazmat.2023.133142.
- [108] Rom O, Liu Y, Liu Z, Zhao Y, Wu J, Ghrayeb A, Villacorta L, et al. Glycine-based treatment ameliorates NAFLD by modulating fatty acid oxidation, glutathione synthesis, and the gut microbiome. Sci Transl Med 2020;12:eaaz2841. https:// doi.org/10.1126/scitranslmed.aaz2841.
- [109] Gonçalves IO, Passos E, Diogo CV, Rocha-Rodrigues S, Santos-Alves E, Oliveira PJ, et al. Exercise mitigates mitochondrial permeability transition pore and quality control mechanisms alterations in nonalcoholic steatohepatitis. Appl Physiol Nutr Metab 2016;41:298–306. https://doi.org/10.1139/apnm-2015-0470.
- [110] Luukkonen PK, Dufour S, Lyu K, Zhang X-M, Hakkarainen A, Lehtimäki TE, et al. Effect of a ketogenic diet on hepatic steatosis and hepatic mitochondrial metabolism in nonalcoholic fatty liver disease. Proc Natl Acad Sci USA 2020;117:7347–54. https://doi.org/10.1073/pnas.1922344117.
- [111] Verbeek J, Lannoo M, Pirinen E, Ryu D, Spincemaille P, Vander Elst I, et al. Rouxen-y gastric bypass attenuates hepatic mitochondrial dysfunction in mice with non-alcoholic steatohepatitis. Gut 2015;64:673–83. https://doi.org/10.1136/ gutinl-2014-306748
- [112] Shannon CE, Ragavan M, Palavicini JP, Fourcaudot M, Bakewell TM, Valdez IA, et al. Insulin resistance is mechanistically linked to hepatic mitochondrial remodeling in non-alcoholic fatty liver disease. Mol Metab 2021;45:101154. https://doi.org/10.1016/j.molmet.2020.101154.
- [113] Niki E. Role of vitamin E as a lipid-soluble peroxyl radical scavenger: in vitro and in vivo evidence. Free Radic Biol Med 2014;66. https://doi.org/10.1016/j.freeradhiomed 2013 03 022
- [114] Debbabi M, Nury T, Zarrouk A, Mekahli N, Bezine M, Sghaier R, et al. Protective effects of α-tocopherol, γ-tocopherol and oleic acid, three compounds of olive oils, and no effect of trolox, on 7-ketocholesterol-induced mitochondrial and peroxisomal dysfunction in microglial BV-2 cells. Int J Mol Sci 2016;17:1973. https://doi.org/10.3390/lijms17121973.
- [115] Singh N, Chander Narula S, Kumar Sharma R, Tewari S, Kumar Sehgal P. Vitamin E supplementation, superoxide dismutase status, and outcome of scaling and root planing in patients with chronic periodontitis: a randomized clinical trial. J Periodontol 2014;85:242–9. https://doi.org/10.1902/jop.2013.120727.
- [116] Nor Azman NHE, Goon JA, Abdul Ghani SM, Hamid Z, Wan Ngah WZ. Comparing palm oil, tocotrienol-rich fraction and α-tocopherol supplementation on the

- antioxidant levels of older adults. Antioxidants (Basel) 2018;7. https://doi.org/10.3390/antiox7060074.
- [117] Jin X, Song L, Liu X, Chen M, Li Z, Cheng L, et al. Protective efficacy of vitamins C and E on p,p'-DDT-induced cytotoxicity via the ROS-mediated mitochondrial pathway and NF-κB/FasL pathway. PLoS One 2014;9:e113257. https://doi.org/10.1371/journal.pone.0113257.
- [118] Radosavljevic T, Brankovic M, Samardzic J, Djuretić J, Vukicevic D, Vucevic D, et al. Altered mitochondrial function in masld: key features and promising therapeutic approaches. Antioxidants (Basel) 2024;13. https://doi.org/10.3390/antiox13080906
- [119] Zhang Y-H, Xie R, Dai C-S, Gao H-W, Zhou G, Qi T-T, et al. Thyroid hormone receptor-beta agonist HSK31679 alleviates MASLD by modulating gut microbial sphingolipids. J Hepatol 2024. https://doi.org/10.1016/j.jhep.2024.08.008.
- [120] Harrison SA, Bashir MR, Guy CD, Zhou R, Moylan CA, Frias JP, et al. Resmetirom (MGL-3196) for the treatment of non-alcoholic steatohepatitis: a multicentre, randomised, double-blind, placebo-controlled, phase 2 trial. The Lancet 2019;394:2012-24. https://doi.org/10.1016/S0140-6736(19)32517-6.
- [121] Sookoian S, Pirola CJ. Resmetirom for treatment of MASH. Cell 2024;187. https://doi.org/10.1016/j.cell.2024.05.009.
- [122] Karim G, Bansal MB. Resmetirom: an orally administered, smallmolecule, liver-directed, β-selective THR agonist for the treatment of non-alcoholic fatty liver disease and non-alcoholic steatohepatitis, touchREV Endocrinol 2023;19:60–70. https://doi.org/10.17925/EE.2023.19.1.60.
- [123] Kihara S, Wölle J, Ehnholm C, Chan L, Oka K. Regulation of hepatic triglyceride lipase by thyroid hormone in HepG2 cells. J Lipid Res 1993;34:961–70.
- [124] Boland ML, Laker RC, Mather K, Nawrocki A, Oldham S, Boland BB, et al. Resolution of NASH and hepatic fibrosis by the GLP-1R/GcgR dual-agonist Cotadutide via modulating mitochondrial function and lipogenesis. Nat Metab 2020;2:413–31. https://doi.org/10.1038/s42255-020-0209-6.
- [125] Newsome PN, Ambery P. Incretins (GLP-1 receptor agonists and dual/triple agonists) and the liver. J Hepatol 2023;79:1557-65. https://doi.org/10.1016/j.jhep.2023.07.033.
- [126] Ma B, Ju A, Zhang S, An Q, Xu S, Liu J, Yu L, Fu Y, Luo Y. Albumosomes formed by cytoplasmic pre-folding albumin maintain mitochondrial homeostasis and inhibit nonalcoholic fatty liver disease. Signal Transduct Target Ther 2023;8:229. https://doi.org/10.1038/s41392-023-01437-0.
- [127] Choi S-E, Hwang Y, Lee S-J, Jung H, Shin TH, Son Y, et al. Mitochondrial protease ClpP supplementation ameliorates diet-induced NASH in mice. J Hepatol 2022;77:735–47. https://doi.org/10.1016/j.jhep.2022.03.034.

- [128] Xu H, Zhao Q, Song N, Yan Z, Lin R, Wu S, et al. AdipoR1/AdipoR2 dual agonist recovers nonalcoholic steatohepatitis and related fibrosis via endoplasmic reticulum-mitochondria axis. Nat Commun 2020;11:5807. https://doi.org/10.1038/ s41467-020-19668-v.
- [129] Zhou J, Pang J, Tripathi M, Ho JP, Widjaja AA, Shekeran SG, et al. Spermidine-mediated hypusination of translation factor EIF5A improves mitochondrial fatty acid oxidation and prevents non-alcoholic steatohepatitis progression. Nat Commun 2022;13:5202. https://doi.org/10.1038/s41467-022-32788-x.
- [130] Fernández-Sáez EM, Losarcos M, Becerril S, Valentí V, Moncada R, Martín M, et al. Uroguanylin prevents hepatic steatosis, mitochondrial dysfunction and fibrosis in obesity-associated NAFLD. Metabolism 2023;147:155663. https://doi.org/10.1016/j.metabol.2023.155663.
- [131] Wu J, Lou Y, Yang X, Wang R, Zhang R, Aa J, Wang G, Xie Y. Silybin regulates P450s activity by attenuating endoplasmic reticulum stress in mouse nonalcoholic fatty liver disease. Acta Pharmacol Sin 2023;44:133–44. https://doi.org/ 10.1038/s41401-022-00924-4.
- [132] Jia Y, Wu C, Kim J, Kim B, Lee S-J. Astaxanthin reduces hepatic lipid accumulations in high-fat-fed C57BL/6J mice via activation of peroxisome proliferator-activated receptor (PPAR) alpha and inhibition of PPAR gamma and Akt. J Nutr Biochem 2016;28:9–18. https://doi.org/10.1016/j.jnutbio.2015.09.015.
- [133] Kim B, Farruggia C, Ku CS, Pham TX, Yang Y, Bae M, et al. Astaxanthin inhibits inflammation and fibrosis in the liver and adipose tissue of mouse models of diet-induced obesity and nonalcoholic steatohepatitis. J Nutr Biochem 2017;43:27–35. https://doi.org/10.1016/j.jnutbio.2016.01.006.
- [134] Wu L, Mo W, Feng J, Li J, Yu Q. Li S, et al. Astaxanthin attenuates hepatic damage and mitochondrial dysfunction in non-alcoholic fatty liver disease by up-regulating the FGF21/PGC-1α pathway. Br J Pharmacol 2020;177:3760–77. https:// doi.org/10.1111/bph.15099.
- [135] Jian C, Fu J, Cheng X, Shen L-J, Ji Y-X, Wang X, Pan S, et al. Low-dose sorafenib acts as a mitochondrial uncoupler and ameliorates nonalcoholic steatohepatitis. Cell Metab 2020;31:892–908 e11. https://doi.org/10.1016/j.cmet.2020.04.011.
- [136] Niu S, Chen S, Chen X, Ren Q, Yue L, Pan X, et al. Semaglutide ameliorates metabolism and hepatic outcomes in an NAFLD mouse model. Front Endocrinol (Lausanne) 2022;13:1046130. https://doi.org/10.3389/fendo.2022.1046130.
- [137] Rector RS, Thyfault JP, Uptergrove GM, Morris EM, Naples SP, Borengasser SJ, et al. Mitochondrial dysfunction precedes insulin resistance and hepatic steatosis and contributes to the natural history of non-alcoholic fatty liver disease in an obese rodent model. J Hepatol 2010;52:727–36. https://doi.org/10.1016/j.ihep.2009.11.030.