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Original article

A comprehensive meta-analysis of stem cell therapy for liver failure: Assessing treatment efficacy and modality



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ABSTRACT

Introduction and Objectives: This meta-analysis aims to evaluate the efficacy of stem cell therapy (SCT) for liver failure

Materials and Methods: The study adhered to the recommended guidelines of the PRISMA statement. Eligible studies published prior to May 13, 2023, were comprehensively searched in databases including PubMed, Web of Science, and Embase. Quality assessment was conducted using the Cochrane risk-of-bias tool, and the standard mean differences were calculated for the clinical parameters. The hazard ratios were determined by extracting individual patient data from the Kaplan-Meier curve.

Results: A total of 2,937 articles were retrieved, and eight studies were included in the final analysis. Most of the studies focused on HBV-related liver failure and were randomized controlled trials. All studies utilized mesenchymal stem cells (MSCs), with the majority (62.5%) being allogeneic. The analysis revealed that combining stem cell therapy with standard medical treatment or plasma exchange significantly enhanced patient survival and reduced MELD scores. Specifically, allogeneic stem cells showed superior efficacy in improving survival outcomes compared to autologous stem cells. Furthermore, deep vessel injection plus a single injection demonstrated better effectiveness than peripheral vessel injection plus multiple injections in reducing MELD scores.

Conclusions: This comprehensive analysis underscores the potential of MSC therapy in significantly improving survival and clinical outcomes in patients with liver failure, highlighting the superior benefits of allogeneic MSCs and deep vessel plus single injection administration.

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Abbreviations: PRISMA, Preferred Reporting Items for Systematic Reviews and Meta-Analyses; SMD, standard mean differences; HR, hazard ratios; HBV, hepatitis B virus; CLF, chronic liver failure; ACLF, acute-on-chronic liver failure; MSCs, mesenchymal stem cells; SCT, stem cell therapy; SMT, standard medicine treatment; CT, cell transplantation; PE, plasma exchange; MELD, model for end-stage liver disease; iPSCs, induced pluripotent stem cells; BM-MSCs, bone marrow MSCs; AD-MSCs, adipose MSCs; UC-MSCs, umbilical cord MSCs; PB-MSCs, peripheral blood derived MSCs; HSCs, hepatic stellate cells; NO, nitric oxide; PGE2, prostaglandin E2; IDO, indoleamine 2,3—dioxygenase; IL-6, interleukin-6; SDF-1, stromal cell-derived factor 1; ALB, albumin; TBIL, total bilirubin; ALT, alanine aminotransferase; INR, international normalized ratio; K-M, Kaplan-Meier; logHR, logarithm of the HR; SE, standard error; HCV, hepati-

tis C virus

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1. Introduction

Liver failure is a severe condition characterized by impaired liver function. It can manifest itself suddenly as acute liver failure or emerge gradually over a period of time as chronic liver failure (CLF). Acute-on-chronic liver failure (ACLF) has a worldwide prevalence of 35% among patients with decompensated cirrhosis, with the highest occurrence in South Asia at 65%. At present, the global 90-day mortality rate stands at 58%, with the highest rate found in South America at 73% [1]. The primary treatment approach for liver failure is centered on addressing the underlying causes, including antiviral therapy for viral hepatitis, discontinuation of hepatotoxic drugs, promoting alcohol cessation and avoiding hepatotoxic substances. Additionally, comprehensive medical management, artificial liver support therapy and liver transplantation (LT) surgery, are employed. LT is currently the only curative approach available for liver failure [2]. In recent years, an improvement in post-LT survival rates has been observed

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(current 5-year survival rates: 75% in Asia, 74% in Europe and 78.1% in the United States). However, the high cost of the surgery, coupled with the scarcity of liver donors, leads to delayed treatment for numerous patients on LT waiting list [3]. Moreover, the high mortality of acute graft-versus-host disease following LT contributes to treatment failure [4]. Therefore, the development of novel approaches for the treatment of liver failure is of utmost importance in terms of clinical practice.

Human stem cells may be essentially categorized into embryonic stem cells (ESCs), mesenchymal stem cells (MSCs) and somatic stem cells. Among these categories, ESCs represent the most proliferative type of pluripotent stem cells, although ethical issues arise from their usage. Recent times have seen the development of somatic stem cells (also termed induced pluripotent stem cells, or iPSCs), which are obtained by introducing specific genes, such as Myc, Oct3/4 and Klf4, into the somatic cells, thereby converting them into pluripotent stem cells. Although iPSCs show great promise, the current lack of a standardized generation procedure limits their applicability at present [5,6]. MSCs are pluripotent stem cells derived from various tissues, such as bone marrow, adipose tissue, umbilical cord blood, placenta and peripheral blood. They have low immunogenicity, and possess the ability to self-proliferate and differentiate. MSCs, including bone marrow MSCs (BM-MSCs), adipose MSCs (AD-MSCs), umbilical cord MSCs (UC-MSCs) and peripheral blood derived MSCs (PB-MSCs), have shown potential in treating liver diseases via their differentiation into hepatocyte-like cells, with their subsequent participation in immune regulation, cell proliferation and injury repair [7,8]. Due to their abundant sources, easy obtainability, low immunogenicity and compatibility with different delivery methods, MSCs hold promise as an ideal source for stem cell research in liver diseases. The mechanism associated with MSCs function comprises their differentiation into hepatocyte-like cells and the paracrine secretion of numerous signaling pathway factors. The immunomodulatory activities and antifibrotic properties of MSCs are important in terms of the treatment of liver failure. MSCs indirectly regulate the activity of hepatic stellate cells (HSCs) through their influence of immune cells. In an inflammatory environment, MSCs migrate to the liver injury site, where they secrete various factors [e.g., nitric oxide (NO), prostaglandin E2 (PGE2), indoleamine 2,3-dioxygenase (IDO), interleukin (IL)-6, IL-10 and stromal cell-derived factor 1 (SDF-1)] to inhibit immune cell proliferation and activation, thereby reducing both fibrosis and the accumulation of extracellular matrix (ECM) [9]. Moreover, MSCs exert direct anti-fibrotic effects by inhibiting the proliferation of HSCs and ECM synthesis [10]. They also reduce gene expression of collagen and inhibit the notch pathway [11].

In recent years, numerous studies have reported on the safety of stem cell therapy (SCT) in liver failure [12]; however, the therapeutic effects have been found to be inconsistent. Although systematic reviews and meta-analyses on the use of stem cells in the treatment of liver failure have been published, the majority of them have included studies on liver cirrhosis due to an insufficient number of clinical trials having been performed. It is important to note that liver failure and cirrhosis proceed via distinct pathogenic mechanisms. Additionally, previously published meta-analyses have primarily pooled the overall survival (OS) rate to address the impact of SCT on the prognosis of patients with liver failure [12-15]. Moreover, the inclusion of studies with varying follow-up durations in these metaanalyses may have had the effect of limiting the accuracy and comprehensiveness of the OS rate in reflecting the true survival status of patients [16]. Therefore, the present study aimed to systematically include and analyze the clinical trials associated with SCT for liver failure, with a comprehensive utilization of the reported survival curves, in order to evaluate and summarize the therapeutic efficacy of stem cells in the treatment of liver failure; in addition, we further discuss the limitations present in existing research in this field.

2. Patients and methods

This meta-analysis study was conducted according to the recommended guidelines of the Preferred Reporting Items for Systematic Reviews and Meta-Analyses (PRISMA) statement [17].

2.1. Eligibility criteria for selection

Previous studies of human stem cell-based treatment for liver failure were collected and collated for this meta-analysis. To ensure the inclusion of high-quality clinical investigations, the present study only considered publications in English. Specifically, our focus was on formally published randomized controlled trials (RCTs) that examined the use of stem cell therapy for liver failure. Conference abstracts were excluded in the present study. The selected studies were required to meticulously report detailed information for further comprehensive re-investigation. The details sought after included the number of participants, diagnostic criteria for liver failure, clear definition of groups, treatment methods, durations of follow-up, clinical parameters and survival outcome.

2.2. Search strategy using various databases

A thorough investigation was conducted in the PubMed, Web of Science (WOS) and Embase databases to identify relevant studies published prior to May 13th, 2023. Our search queries were constructed in accordance with the retrieval guidelines of these databases, using a combination of keywords, fields and Boolean logical operators. For PubMed, the search terms employed were "{(stem cell transplantation[Title/Abstract]) OR (stem cells transplantation[Title/ Abstract]) OR (stem cells infusion[Title/Abstract]) OR (stem cell infusion[Title/Abstract]) OR (stem cells therapy[Title/Abstract]) OR (stem cell therapy[Title/Abstract]) OR (umbilical cord[Title/Abstract]) OR (bone marrow[Title/Abstract]) OR (mesenchymal stromal[Title/ Abstract])) AND (liver failure [Title/Abstract])}". In the case of WOS, our search term was "TI (limit to title) = [(stem cell* transplantation) OR (stem cell* infusion) OR (stem cell* therapy) OR (umbilical cord) OR (mesenchymal stromal) OR (bone marrow)] AND (liver failure) OR AB (abstract) = [(stem cell* transplantation) OR (stem cell* infusion) OR (stem cell* therapy) OR (umbilical cord) OR (mesenchymal stromal) OR (bone marrow)] AND (liver failure)". Lastly, for Embase, the search terms "('stem cell transplantation':ab,ti OR 'stem cells transplantation':ab,ti OR 'stem cells infusion':ab,ti OR 'stem cells infusion': ab,ti OR 'stem cells therapy':ab,ti OR 'stem cells therapy':ab,ti OR 'umbilical cord':ab,ti OR 'bone marrow':ab,ti OR 'mesenchymal stromal':ab,ti) AND 'liver failure':ab,ti AND 'human'/de AND ('article'/it OR 'review'/it)" were employed.

2.3. Study selection and data extraction

After eliminating redundant references, three researchers (HM, ZL and DZ) scanned the titles and abstracts in order to identify relevant studies. Should any discrepancies arise, the supervisors (IP and YZ) assumed the responsibility of making final judgments. The full texts and supplementary materials of the qualifying studies were subjected to thorough screening by two researchers (SL and HG), who worked independently of each other. YZ subsequently reviewed the final data for any inconsistencies, and facilitated the data correlation. IP reviewed, revised and finalized the manuscript. The following data were collected, when available: i) the study characteristics, i.e., the year of publication, primary author, country and study design; ii) the patient characteristics, i.e., the number of participants in each arm, etiologies, diagnostic criteria and duration of follow-up; iii) stem cells, i.e., the cell type, dosage, treatment frequency and route of administration; and iv) study outcomes, i.e., survival rates/curves, adverse events, model for end-stage liver disease (MELD) scores, the

levels of albumin (ALB), total bilirubin (TBIL), alanine aminotransferase (ALT), and the international normalized ratio (INR) at various follow-up time points. Due to the focus on a specific time point within the Kaplan-Meier (K-M) curves and the lack of a consistent follow-up duration across the studies, utilizing the survival rate alone may not adequately reflect variations in patient prognosis among the different intervention groups [16]. On the other hand, using the hazard ratio (HR), which encompasses comprehensive information relating to the entire K-M curve, has been shown to accurately reflect the treatment effects throughout the entire follow-up period [18]. For articles that did not explicitly present HR values, the graphs and extract data points were digitized using the tool IPDfromKM [19].

2.4. Quality assessment and statistical analysis

The assessment of publication bias risk for each included study was performed by two researchers (SL and HG) employing the Cochrane risk-of-bias (ROB) tool. The Cochrane ROB tool offers a structured approach to evaluate the potential for bias in studies. It encompasses five domains: i) randomization-associated bias; ii) intervention deviation-associated bias, iii) missing data-associated bias, iv) outcome measurement-associated bias; and v) reported result selection-associated bias. The bias risk within each domain was evaluated and categorized as low, high, or uncertain [20].

The continuous variables were calculated as the standard mean difference (SMD) with 95% confidence intervals (95% CI) utilizing Hedges' g methodology [21]. The estimated logarithm of the HR (logHR) and standard error (SE) obtained from IPDfromKM were employed to compute the estimated HRs and corresponding 95% CIs. To assess heterogeneity among the included studies, the Tau-squared (Tau²) value, Chi-square test and I² statistics were computed. Notable levels of heterogeneity were indicated by a large value of Tau², P < 0.05 for the Chi-square test, and I²>50%. In such cases, a random effect model was employed to pool the data; otherwise, a common effect model was utilized. Subsequently, sensitivity analysis was performed by systematically omitting one study at a time. Additionally,

publication bias was evaluated using the Funnel plot and Egger's test. In cases where an asymmetric Funnel plot existed alongside a P-value <0.05 from the Egger's test, a trim-and-fill analysis was implemented while constructing an adjusted Funnel plot. Furthermore, subgroup analyses were performed encompassing different follow-up time points, distinct cell sources (autologous or allogeneic), injection route variations (deep vessel or peripheral vessel) and differing injection frequencies (single or multiple). The entire analysis was conducted utilizing the open-source R statistical language (R-project, version 4.1.2) and relevant packages, including tidyverse [22] for reference management and selection, and meta [23] and metafor [24] for core procedures.

3. Results

3.1. Search results

The initial retrieval yielded a total of 2937 articles, encompassing 687 articles from PubMed, 606 from Embase and 1644 from WOS. Following the elimination of duplicates, unavailable and unrelated studies, 530 articles remained for further eligibility evaluation. Furthermore, 157 reviews/meta-analyses, 338 animal/experimental studies and five case reports were excluded. Thirty studies were subsequently retained for meticulous review. Eight studies were included in the final analysis, subsequent to the exclusion of 22 studies, of which nine were not specific for SCT or liver failure, three were non-English, three lacked sufficient data, and seven had uncontrolled designs. The literature retrieval and screening procedures are shown in the flowchart (Fig. 1).

3.2. Characteristics and quality of included studies

The characteristics of the selected eight studies [25-32] are shown in Table 1. These studies were published between 2017 and 2021. Out of the eight included studies, the majority were conducted by Chinese researchers (n = 6), whereas the remaining pair of articles

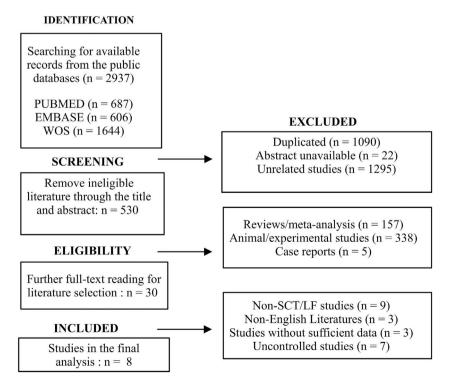


Fig. 1. The flowchart of the selection process of studies.

Table 1Characteristics of the included studies.

PMID	Author	Year	Country	Study type	Disease	Etiology	Source	Cell type	Cell dosage	Injection route	Injection times
21,608,000	Peng et al.	2011	China	Controlled clinical trial	LF	HBV	Autologous	BM-MSC	1.00E+06	Deep	Single
21,900,788	Amer et al.	2011	Egypt	RCT	CLF	HCV	Autologous	BM-MSC	1.00E+07	Deep	Single
23,197,664	Shi et al.	2012	China	RCT	ACLF	HBV	Allogeneic	UC-MSC	0.50E+06 /kg	Peripheral	Multiple
27,687,792	Li et al.	2016	China	Controlled clinical trial	ACLF	HBV	Allogeneic	UC-MSC	1.00E+08	Deep	Single
28,370,357	Lin et al.	2017	China	RCT	ACLF	HBV	Allogeneic	BM-MSC	1.00 ~10E+05 /kg	Peripheral	Multiple
30,863,450	Xu et al.	2019	China	RCT	ACLF	HBV	Allogeneic	UC-MSC	1.00E+05 /kg	Peripheral	Multiple
33,053,116	Zhu et al.	2020	China	RCT	ACLF	HBV	Autologous	PB-MSC	0.60E+08~0.9E+09	Deep	Single
34,395,335	Schacher et al.	2021	Brazil	RCT	ACLF	Various etiologies	Allogeneic	BM-MSC	1.00E+06 /kg	Peripheral	Multiple

hepatitis B virus, HBV; hepatitis C virus, HCV; bone marrow derived mesenchymal stem cells, BM-MSC; umbilical cord derived mesenchymal stem cells, UC-MSC; peripheral blood derived mesenchymal stem cells, PB-MSC; liver failure, LF; chronic liver failure, CLF; acute-on-chronic liver failure, ACLF.

originated from Egypt and Brazil. In terms of study design, 75% of the studies were RCTs, whereas the remaining 25% were controlled clinical trials. Regarding the etiology of the disease, six studies focused on HBV-associated ACLF, one study was on hepatitis C virus (HCV)-associated CLF, and one study investigated ALCF with various etiologies, of which 56% of cases were attributed to viral hepatitis. As for the source of stem cells, 62.5% (5/8 studies) were allogeneic, whereas the remaining 37.5% (3/8 studies) were autologous. In terms of stem cell type, four studies utilized BM-MSCs, three studies employed UC-MSCs and one study utilized PB-MSCs. The administration routes (deep or peripheral vessel) and the frequency of injections (single or multiple) were evenly distributed. The majority of the included studies exhibited a low overall risk of bias (Fig. 2A). The main reason for potential bias was the lack of a full description of the randomization process. Two studies were found to have a high risk of bias due to concerns regarding the randomization process, deviation from the intended intervention, and missing data (Fig. 2B).

3.3. Enhanced survival with SCT

Individual patient data from seven studies [26–32] were extracted from the K-M curves. logHR and its corresponding standard error (SE) were estimated using these data. These values were subsequently used to calculate the HR and their corresponding 95% CI for each study. Out of the seven studies, five studies [27–30,32]

compared SCT plus standard medicine treatment (SMT) with SMT alone. The SCT+SMT group consisted of 167 patients, whereas the SMT group comprised 213 patients. The follow-up time varied among these studies (follow-up times of 8, 12, 24, 48 and 72 weeks were reported), with each duration subgroup having only one available study. Heterogeneity analysis showed a low level of heterogeneity among the studies, with $Tau^2=0.033$, Chi-square test P=0.31, and $I^2=17\%$, leading to the use of a common effect model. The pooled HR was 0.695 (95% CI=0.508-0.951, P = 0.02), indicating that stem cellbased treatment had a significant effect on improving the survival of liver failure patients compared with SMT alone (Fig. 3). Three studies [26,31,32] compared the treatment strategy of plasma exchange (PE) plus SCT (SCT+PE) with the treatment of PE alone. The SCT+PE group included 51 patients, whereas the PE group comprised 84 patients. Similar to the previous comparison, only one study was available for each follow-up duration, with low heterogeneity among the studies (Tau²=0.114, Chi-square test P = 0.37, and $I^2=6\%$). The common effect model showed a pooled HR of 0.487 (95% CI=0.289-0.820, P < 0.01), indicating that the SCT+PE treatment led to an improvement in the survival rate of patients with liver failure compared with PE treatment alone (Fig. 4).

Additionally, subgroup analyses were conducted for cell sources (autologous or allogeneic), injection route variations (deep vessel or peripheral vessel), and differing injection frequencies (single or multiple). Two studies [28,31] used autologous stem cells, whereas five

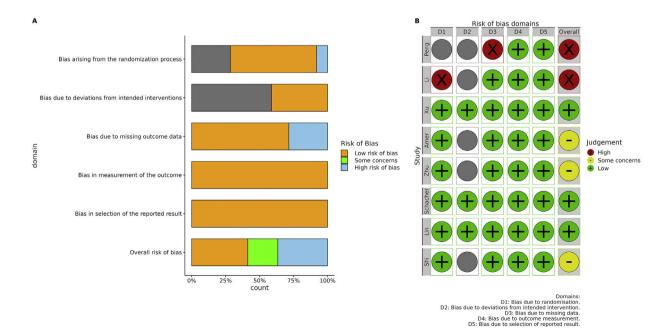


Fig. 2. The publication bias assessment. (A) Weighted bar plots display the distribution of risk-of-bias judgments within each bias domain. (B) Traffic light plots illustrate domain-level judgments for each individual study.

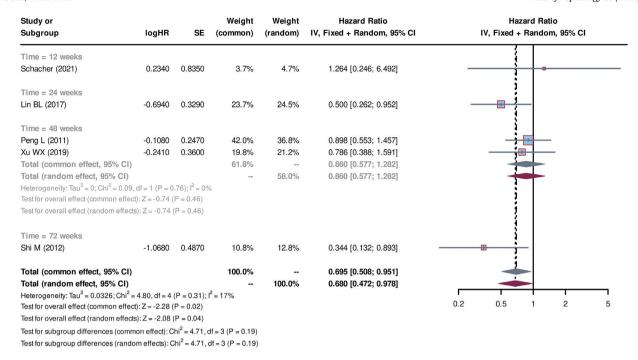


Fig. 3. Forest plots of survival hazard ratio comparing SCT + SMT with SMT.

studies [26,27,29,30,32] used allogeneic stem cells. The pooled HR for allogeneic stem cells was 0.675 (95% CI=0.507-0.900, P < 0.01, standard effect model), indicating improved survival outcomes compared with autologous stem cell treatment with a pooled HR of 0.722 (95% CI=0.472-1.104, P = 0.13, standard effect model) (Fig. 5). Regarding the injection route and injection frequencies, three studies [26,28,31] used the deep vessel route and single injection, whereas four studies [27,29,30,32] used the peripheral vessel route and multiple injections. Both administration groups were associated with a significant improvement in survival (for the deep vessel + single injection group: pooled HR=0.632, 95% CI=0.429-0.932, P = 0.02, common effect model; for the peripheral vessel + multiple injections group: pooled HR=0.727, 95% CI=0.538-0.981, P = 0.04, common effect model) (Fig. 6). The analyses of sensitivity confirmed the reliability of the

above results (Fig. S1). Additionally, the symmetry funnel plots and negative Egger's test (P > 0.05) indicated a low risk of publication bias among the studies (Fig. S2).

3.4. Effect of SCT on the MELD score

Five studies [27-30,32] compared SCT+SMT treatment with SMT treatment alone to observe the changes in MELD scores over various follow-up durations. The SCT+SMT group comprised 183 patients, whereas the control group consisted of 228 patients. The pooled SMD of MELD scores was -0.251 (95% Cl=-0.386 to -0.115, P < 0.01, the common effect model). Subgroup analyses for each follow-up duration also demonstrated a decrease in the MELD scores in the SCT

Study or Subgroup	logHR	SE	Weight (common)	Weight (random)	Hazard Ratio IV, Fixed + Random, 95% CI	Hazard Ratio IV, Fixed + Random, 95% CI
Time = <4 weeks Zhu B (2020)	0.0480	1.4150	3.5%	4.7%	1.049 [0.066; 16.799]	
Time = 12 weeks Zhu B (2020)	-1.3590	0.7910	11.3%	13.5%	0.257 [0.055; 1.211]	
Time = 24 weeks Zhu B (2020)	-1.0660	0.5930	20.1%	21.5%	0.344 [0.108; 1.101]	
Time = 48 weeks Xu WX (2019)	-0.0570	0.4430	36.0%	32.2%	0.945 [0.396; 2.251]	1
Time = 96 weeks Li YH (2016)	-1.1470	0.4930	29.1%	28.0%	0.318 [0.121; 0.835]	
Total (common effect, 95% CI) 100.0% Total (random effect, 95% CI)				 100.0%	0.487 [0.289; 0.820] 0.472 [0.254; 0.878]	0.1 0.5 1 2 10

Fig. 4. Forest plots of survival hazard ratio comparing SCT + PE with PE.

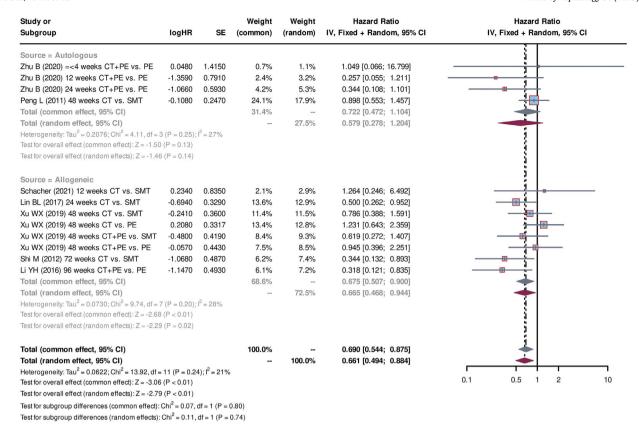


Fig. 5. Forest plots of survival hazard ratio comparing autologous stem cells with allogeneic stem cells.

Study or Subgroup	logHR	SE	Weight (common)	Weight (random)	Hazard Ratio IV, Fixed + Random, 95% CI	Hazard Ratio IV, Fixed + Random, 95% CI
Route+Injection = Deep+Single						i
Zhu B (2020) =<4 weeks CT+PE vs. PE	0.0480	1.4150	0.7%	1.1%	1.049 [0.066; 16.799]	
Zhu B (2020) 12 weeks CT+PE vs. PE	-1.3590	0.7910	2.4%	3.2%	0.257 [0.055; 1.211]	
hu B (2020) 24 weeks CT+PE vs. PE	-1.0660	0.5930	4.2%	5.3%	0.344 [0.108; 1.101]	
Peng L (2011) 48 weeks CT vs. SMT	-0.1080	0.2470	24.1%	17.9%	0.898 [0.553; 1.457]	-1 -
i YH (2016) 96 weeks CT+PE vs. PE	-1.1470	0.4930	6.1%	7.2%	0.318 [0.121; 0.835]	
otal (common effect, 95% CI)			37.5%		0.632 [0.429; 0.932]	-
Total (random effect, 95% CI)				34.7%	0.502 [0.264; 0.953]	
leterogeneity: $Tau^2 = 0.2100$; $Chi^2 = 6.44$, $df = 4$	4 (P = 0.17);	$l^2 = 38\%$				4
est for overall effect (common effect): $Z = -2.31$	(P = 0.02)					3 I
Test for overall effect (random effects): $Z = -2.11$	I (P = 0.04)					i
Route+Injection = Peripheral+Multiple						į
Schacher (2021) 12 weeks CT vs. SMT	0.2340	0.8350	2.1%	2.9%	1.264 [0.246; 6.492]	- 5 •
in BL (2017) 24 weeks CT vs. SMT	-0.6940	0.3290	13.6%	12.9%	0.500 [0.262; 0.952]	- 3 -
(u WX (2019) 48 weeks CT vs. SMT	-0.2410	0.3600	11.4%	11.5%	0.786 [0.388; 1.591]	
(u WX (2019) 48 weeks CT vs. PE	0.2080	0.3317	13.4%	12.8%	1.231 [0.643; 2.359]	j 🗉
(u WX (2019) 48 weeks CT+PE vs. SMT	-0.4800	0.4190	8.4%	9.3%	0.619 [0.272; 1.407]	- 4
(u WX (2019) 48 weeks CT+PE vs. PE	-0.0570	0.4430	7.5%	8.5%	0.945 [0.396; 2.251]	- } =
Shi M (2012) 72 weeks CT vs. SMT	-1.0680	0.4870	6.2%	7.4%	0.344 [0.132; 0.893]	
otal (common effect, 95% CI)			62.5%		0.727 [0.538; 0.981]	→
Total (random effect, 95% CI)			65.3%	0.723 [0.512; 1.022]	*	
leterogeneity: $Tau^2 = 0.0453$; $Chi^2 = 7.17$, $df = 6$		$l^2 = 16\%$!
est for overall effect (common effect): $Z = -2.08$						j
est for overall effect (random effects): Z = -1.84	1 (P = 0.07)					į
Total (common effect, 95% CI)				-	0.690 [0.544; 0.875]	
Total (random effect, 95% CI)				100.0%	0.661 [0.494; 0.884]	•
leterogeneity: $Tau^2 = 0.0622$; $Chi^2 = 13.92$, $df =$		4); $I^2 = 21\%$				0.1 0.5 1 2 1
Test for overall effect (common effect): Z = -3.06 (P < 0.01)						0.1 0.5 1 2
est for overall effect (random effects): $Z = -2.79$						
est for subgroup differences (common effect):	$Chi^2 = 0.31,$	df = 1 (P = 0)	0.58)			

Fig. 6. Forest plots of survival hazard ratio comparing deep vessel + single injection with peripheral vessel + multiple injections.

Test for subgroup differences (random effects): Chi² = 0.97, df = 1 (P = 0.33)

+SMT group compared with the control; for follow-up durations of ≤4 weeks (five studies, pooled SMD=-0.229, 95% CI=-0.428 to -0.030, P = 0.02, the common effect model), 8 weeks (one study, SMD=-0.369, 95% CI=-0.880 to -0.141), 12 weeks (two studies, pooled SMD=-0.177, 95% CI=-0.479 to 0.125, P = 0.25, the common effect model), 24 weeks (four studies, pooled SMD=-0.499, 95% CI=-1.034 to 0.035, P = 0.07, the random effect model), and 48 weeks (one study, SMD=-0.228, 95% CI=-1.178 to 0.721) (Fig. S3). Two studies [26,32] that compared SCT+PE with PE alone treatments reported on the changes of MELD scores: The pooled SMD of MELD scores was found to be -0.400 (95% CI=-0.626 to -0.173, P < 0.01, the common effect model) (Fig. S4). These results indicated that SCTs could reduce the MELD scores of patients. In terms of stem cell sources, two studies [25,28] used autologous stem cells, resulting in a pooled SMD of MELD scores of -0.523 (95% CI=-0.774 to -0.271, P < 0.01, the common effect model). On the other hand, four studies [26,27,30,32] employed allogeneic stem cells and exhibited a pooled SMD of MELD scores of -0.201 (95% CI=-0.313 to -0.088, *P* < 0.01, the common effect model) (Fig. S5). Both approaches were found to result in decreased MELD scores. With regard to injection route and frequency, three studies [25,26,28] utilized the deep vessel route with a single injection, whereas two studies [27,30] used the peripheral vessel route with multiple injections. The deep vessel + single injection approach showed a significant reduction in MELD scores (pooled SMD=-0.607, 95% CI=-803 to -0.411, P < 0.01, the common effect model), whereas the peripheral vessel + multiple injections administration route did not (pooled SMD=-0.121, 95% CI=-0.242 to 0.000, P = 0.05, the common effect model) (Fig. S6).

Sensitivity analyses were subsequently performed to confirm the reliability of the aforementioned results. However, the Egger's test (P < 0.05) revealed asymmetry in the funnel plots, suggesting the presence of publication bias among the included studies. As a result, random effect models were constructed using the trim-and-fill method to calculate the adjusted pooled SMD for each analysis, which helped to reduce the effect of publication bias (Fig. S7).

3.5. Effect of stem cell therapy on laboratory indicators

ALT, TBIL and ALB are commonly used indices of liver function in a clinical setting. Neither the levels of ALT or TBIL were found to be significantly changed following treatment in studies comparing SCT +SMT treatment with SMT or SCT+PE with PE alone (Figs. S8-11). By contrast, a significant increase in ALB was observed in the SCT+SMT treatment group compared with SMT (pooled SMD=0.475, 95% CI=0.092-0.856, P=0.01, the random-effect model). Although no significant differences in ALB levels were noted between the SCT+PE and PE groups, a trend of increased levels was observed (Figs. S12 and 13). The INR is an important indicator of coagulation function. No significant differences in INR were observed between the SCT+SMT and SMT groups; however, only two studies were included (Fig. S14). On the other hand, in studies comparing SCT+PE with PE alone, a significant decrease in INR was observed in the SCT+PE group (pooled SMD=-0.909, 95% CI=-1.375 to -0.443, P < 0.01, the random-effect model) (Fig. S15). Subsequent analyses revealed asymmetry in the funnel plot, indicating the presence of publication bias among the included studies (Egger's test, E P < 0.05). The trim-and-fill method was therefore employed to calculate the adjusted pooled SMD of INR (-0.913; 95% CI=-1.382 to -0.444) (Fig. S16).

4. Discussion

This study holds significant potential in advancing the understanding and application of SCT for liver failure, particularly highlighting the effectiveness of MSCs. Notably, no significant side effects were reported in the included clinical trials. By systematically evaluating and quantifying the benefits of SCT in improving patient

survival and clinical outcomes, the results indicated that both SCT +SMT and SCT+PE combination therapies resulted in significantly improved survival rates for the patients. This study also revealed a decrease in the MELD score following SCT, a finding that was consistent with previous studies [13,15]. Allogeneic stem cells demonstrated superior therapeutic efficacy in terms of improving patient survival compared with autologous stem cells. Both deep vessel + single and peripheral vessel + multiple injections led to significant improvements in survival. However, the deep vessel + a single injection administration route appeared to be more effective than peripheral vessel + multiple injections in terms of reducing patients' MELD scores. Consistent with previous studies [13,15], the employment of SCT may not have had a significant effect on patients' INR levels. ALB levels were found to be increased following SCT+SMT treatment, but were not significantly increased after SCT+PE treatment. The present study did not identify significant differences in TBIL and ALT levels comparing between SCTs and their respective controls, which was in contradiction with previous meta-analyses [33,15]. This discrepancy, however, may be attributed to the differences in inclusion criteria of studies, as mentioned earlier in this article.

Although researchers have made great progress in terms of SCT in liver failure, there are still some concerns that require clarification. MSCs have a unique ability to migrate to, and home in on, the injury to the liver. The homing mechanism is a crucial aspect of their therapeutic potential in liver failure. In addition, the homing process may also occur in healthy liver. In a previous study, BM-MSCs were cultured and labeled with radioactive tracer (111In-oxine) before being infused into healthy rats via deep vessel injection. The distribution of the infused MSCs was monitored using real-time imaging, and the radioactivity in various organs was measured after 48 h. The results obtained showed that MSCs were primarily detected in the lungs, followed by the liver and other organs, suggesting that MSCs have multiple homing sites [34]. Subsequently, in a study of liver injury, a thioacetamide-induced acute liver injury rat model was established. After infusion of UC-MSCs via the tail vein, the MSCs were also found to be mainly distributed in the lung and liver [35]. This phenomenon has also been observed in patients with liver cirrhosis. Following injection with ¹¹¹In-oxine-labeled BM-MSCs via the peripheral vein, the uptake of radioactivity was initially observed in the lungs, and this was gradually increased in the liver and spleen over a time period of hours to days. The spleen uptake was found to exceed that in the liver in all patients. The study also showed that the percentage of cells homing to the liver increased from 0-2.8% immediately after infusion to 13.0-17.4% on day 10. The homing process of MSCs to the liver has been demonstrated to comprise a complex interplay of various factors [36]. One key factor is the release of chemotactic signals from the damaged liver tissue. These signals include cytokines, growth factors and chemokines, which act as attractants for MSCs. The SDF-1/CXCR4 signaling pathway is one of the well-studied pathways that have been found to regulate the migration and homing of MSCs. In the rat model of acute liver injury, the expression level of SDF-1 was found to increase over time following MSC infusion, whereas the blockage of CXCR4 led to an inhibition of the migration of MSCs to the liver [37]. However, it is also important to note that the homing mechanism of MSCs to the liver is not yet fully understood, and this remains an active area of research. Both understanding and harnessing this mechanism should be able to potentially enhance the therapeutic efficacy of MSCs-based therapies for liver

Few previously published clinical studies have investigated the dose-response relationship of SCT for liver failure, and therefore the optimal dosages for treatment remain uncertain. Two RCTs evaluated bone marrow-derived mononuclear cells (BM-MNCs) in liver treatment: In one, 1×10^9 cells were administered intraportally [38], and in the other, 5×10^7 cells per kg were administered through the hepatic artery [39]. Neither trial revealed any benefit compared with

the control group. However, enriching BM-MNCs for CD133 (approximately 5×10^6 cells) or CD34/CD133 (5×10^7 cells) resulted in partial improvements, suggesting that reducing cell numbers through enrichment appeared to be beneficial [38,40]. A recent study in rat models has demonstrated dose-dependent effects of MSCs in terms of reducing mortality and liver inflammation in acetaminopheninduced acute liver failure. The researcher infused UC-MSCs into rats via the tail vein at doses ranging from 5×10^5 to 2×10^6 cells. The result showed a gradual increase in the survival rate of mice as the dose was raised from 5×10^5 to 1×10^6 cells; however, no further improvements were observed at higher doses. Furthermore, the serum transaminase levels in the surviving mice exhibited a significant dose-dependent decrease [41]. Future clinical studies on SCT should include treatment protocols with varying dose gradients to establish the optimal dosage for liver failure treatment using stem cells, thereby promoting standardization in subsequent associated studies.

There is currently no consensus on the optimal route for infusing stem cells. It is known that a significant proportion of the cells initially reside in the lungs following administration, a phenomenon referred to as the 'pulmonary-first' phenomenon [42]. Therefore, the peripheral vascular infusion of stem cells may not result in a sufficient number of cells homing to the liver. On the other hand, there are various options available for deep vascular infusion, including the hepatic artery, hepatic vein, splenic artery, femoral artery and the femoral vein. To the best of our knowledge, previous clinical studies have not compared different infusion routes. It is generally understood that using the portal vessel system can effectively bypass the pulmonary-first phenomenon, allowing an adequate number of stem cells to reach the liver. In an animal model study, researchers compared the femoral vein, femoral artery and intraperitoneal routes of infusion, and found no significant difference between the femoral vein and femoral artery administration routes in terms of the number of stem cells homing to the liver, whereas only a small fraction of cells migrated to the liver with intraperitoneal infusion [34]. In a prospective study by Amer et al., the hepatic artery and splenic artery injection routes were compared, and no significant differences in any of the clinical parameters were observed [25]. Moreover, the results of the present study indicated that both the deep vascular infusion and peripheral vascular infusion routes led to a significant improvement in patients' survival. However, combining deep vascular infusion with single injections could lead to a marked reduction in patients' MELD scores. This may be related to the deep vascular route, which can reduce the retention time of stem cells in the lung and spleen, thereby increasing the number of stem cells that reach the liver. Although deep vascular infusion carries a risk of increased bleeding due to the requirement of deep vascular puncture, which, in terms of bleeding, is of particular concern in liver failure patients with impaired coagulation function, a previous study demonstrated the safety of the usage of deep vascular injection by experienced practitioners during SCT [31].

The administration method, whether through a single or multiple infusions, is an important consideration in the clinical application of stem cells. The key consideration is to minimize the number of infusions while ensuring a sufficient therapeutic cell dose to reduce the risk of treatment-associated complications. Additionally, it is necessary to determine the optimal source of stem cells for therapy. The use of autologous stem cells is known to minimize the risk of host immune rejection. However, due to the limitations of cell harvesting, a large quantity of autologous stem cells (typically obtained from bone marrow) cannot be obtained in a single collection. A large number of bone marrow samples (typically ranging from 100-200 ml per procedure) are usually required, which inevitably places a psychological burden on the patients. Moreover, the prolonged cultivation time (typically one week) may delay the treatment of patients with liver failure [25,28]. On the other hand, allogeneic stem cells (typically

obtained from umbilical cord blood) can be prepared in sufficient quantities in a single collection, and these have good immunocompatibility [5]. Our results suggested that allogeneic stem cells had similar therapeutic effects to those of autologous stem cells, and were even superior in terms of improving the survival outcomes of patients. However, due to the limited number of included studies, this meta-analysis was unable to compare stem cells from different tissue origins.

Artificial liver support therapy is one of the treatment options for patients with end-stage liver disease, apart from LT. PE is a commonly used method in artificial liver treatment, which comprises replacement of the patient's plasma to effectively remove metabolic toxins, improve the internal environment, and promote the repair and regeneration of the liver [43,44]. Although there is currently a lack of solid evidence from evidence-based medicine to support the efficacy, for certain patients who are unable to undergo LT for various reasons, artificial liver treatment has been demonstrated to alleviate the patient's condition, even avoiding the requirement for LT eventually [45]. The present study suggested that the combination of PE with stem cell transplantation leads to an enhancement in the survival rate of patients with liver failure, as well as reducing their MELD scores and INR values.

Stem cells are highly valued in other chronic liver diseases, such as nonalcoholic fatty liver disease and autoimmune liver disease, due to their self-renewal and multipotency [8]. However, these same characteristics pose significant tumorigenic risks, restricting their use in liver cancer. ESCs can form teratomas when injected into immunodeficient mice [46,47], while iPSCs have been associated with a 20% tumor incidence in mice due to the expression of specific transcription factors [48]. In hepatocellular carcinoma (HCC) treatment, immunological approaches, particularly immune checkpoint inhibitors (ICIs), have shown promise, though clinical outcomes have yet to meet expectations [49]. MSCs, known for their "homing" effect, can migrate to sites of inflammation or tumors, making them potential vehicles for delivering pro-apoptotic drugs to multiple tumor sites. Additionally, iPSC-derived natural killer (NK) cells that inhibit the TGF- β signaling pathway have demonstrated enhanced anti-tumor activity in HCC cell line and mouse models [50]. Ongoing research is crucial to fully realize the potential of stem cell therapies in liver can-

However, this study has several limitations. The included studies had small sample sizes and limited populations, which may affect the generalizability of the findings. Additionally, restricting the analysis to English-language publications introduces potential publication bias. The heterogeneity in stem cell sources and administration routes also impacted the study's reliability, despite efforts to mitigate these issues using subgroup analysis, random effects models and trim-and-fill methods. As with any meta-analysis, controlling for study's subjects selection criteria was challenging, and the inclusion of both ACLF and CLF, along with varying diagnostic criteria, may have introduced bias. The predominance of studies in viral hepatitisrelated liver failure and conducted in China further limits the generalizability of our results. Variations in follow-up durations could have led to inaccuracies in pooled survival outcomes. Therefore, the HRs of the survival curves were estimated to refine the accuracy of the results.

While this study provides valuable insights, it also highlights significant knowledge gaps. A deeper understanding of the long-term effects and safety of SCT in liver failure is needed. Although numerous clinical trials have confirmed its safety, long-term observations are necessary to evaluate the risk of tumorigenicity [51]. The variability in treatment protocols, including differences in MSCs sources (allogeneic vs. autologous) and administration methods, underscores the importance of developing standardized guidelines. Future research should prioritize long-term follow-up studies and transplantation free survival to address these issues, as well as larger, multicenter

trials with standardized protocols to validate findings and establish best practices for SCT in liver failure. Despite the substantial resources required for stem cell research, the promising potential it has demonstrated in treating chronic and degenerative diseases has prompted some developing countries to invest in this field [52,53]. However, the significant resources needed for stem cell therapy can present challenges in low-resource settings. To make stem cell therapy more accessible in these areas, it will be essential to explore strategies such as developing cost-effective methods for cell production and storage, utilizing locally available resources, and simplifying administration techniques.

5. Conclusions

In conclusion, the present systematic meta-analysis has shown that the combinations of SCT+SMT and SCT+PE as treatment methods led to a significant improvement in patient survival, with reduced MELD scores in patients with liver failure. Allogeneic MSCs exhibited greater therapeutic effectiveness when compared with autologous MSCs. Furthermore, both the deep vessel + single injection and peripheral vessel + multiple injections administration routes were associated with improved outcomes. However, the deep vessel + single injection route appeared to be more effective.

Authors' contributions

SL, HG and YZ designed the study; HM, ZL, and DZ conducted the literature search and data collection; JP and YZ evaluated and selected the final articles. SL and HG performed the literature screening, data analysis and wrote the manuscript jointly; YZ reviewed the final data and manuscript; SL and HG contributed equally to this study and should be considered as the co-first authors.

Conflicts of interests

None.

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Supplementary materials

Supplementary material associated with this article can be found in the online version at doi:10.1016/j.aohep.2024.101586.

References

- [1] Mezzano G, Juanola A, Cardenas A, Mezey E, Hamilton JP, Pose E, et al. Global burden of disease: acute-on-chronic liver failure, a systematic review and meta-analysis. Gut 2022;71:148–55. https://doi.org/10.1136/gutjnl-2020-322161.
- [2] Sarin SK, Choudhury A, Sharma MK, Maiwall R, Al Mahtab M, Rahman S, et al. Acute-on-chronic liver failure: consensus recommendations of the Asian Pacific association for the study of the liver (APASL): an update. Hepatol Int 2019;13:353–90. https://doi.org/10.1007/s12072-019-09946-3.
- [3] Ghelichi-Ghojogh M, Rajabi A, Mohammadizadeh F, Shojaie L, Vali M, Afrashteh S, et al. Survival rate of liver transplantation in asia: a systematic review and meta-analysis. Iran J Public Health 2022;51:2207–20. https://doi.org/10.18502/ijph.v51i10.10979.

[4] Izzo A, Pellegrino RA, Locci G, Cesaretti M. Acute graft versus host disease after liver transplantation: where do we stand? Minerva Surg 2023. https://doi.org/ 10.23736/S2724-5691.23.09868-4.

- [5] Li S, Bi Y, Duan Z, Chang Y, Hong F, Chen Y. Stem cell transplantation for treating liver diseases: progress and remaining challenges. Am J Transl Res 2021;13: 3954–66
- [6] Ullah I, Subbarao RB, Rho GJ. Human mesenchymal stem cells current trends and future prospective. Biosci Rep 2015;35:e00191. https://doi.org/10.1042/BSR2015 0025
- [7] Hass R, Kasper C, Böhm S, Jacobs R. Different populations and sources of human mesenchymal stem cells (MSC): a comparison of adult and neonatal tissue-derived MSC. Cell Commun Signal 2011;9:12. https://doi.org/10.1186/1478-811X-9-12.
- [8] Hoang DM, Pham PT, Bach TQ, Ngo ATL, Nguyen QT, Phan TTK, et al. Stem cell-based therapy for human diseases. Signal Transd Targeted Therapy 2022;7:1–41. https://doi.org/10.1038/s41392-022-01134-4.
- [9] Chen S, Xu L, Lin N, Pan W, Hu K, Xu R. Activation of Notch1 signaling by marrow-derived mesenchymal stem cells through cell-cell contact inhibits proliferation of hepatic stellate cells. Life Sci 2011;89:975–81. https://doi.org/10.1016/j.lfs.2011.
- [10] Feng Y, Wang A-T, Jia H-H, Zhao M, Yu H. A brief analysis of mesenchymal stem cells as biological drugs for the treatment of acute-on-chronic liver failure (ACLF): safety and potency. CSCR 2020;15:202–10. https://doi.org/10.2174/1574888X15 666200101124317.
- [11] Lee C-W, Chen Y-F, Wu H-H, Lee OK. Historical perspectives and advances in mesenchymal stem cell research for the treatment of liver diseases. Gastroenterology 2018;154:46–56. https://doi.org/10.1053/j.gastro.2017.09.049.
- [12] Wang K, Chen X, Ren J. Autologous bone marrow stem cell transplantation in patients with liver failure: a meta-analytic review. Stem Cells Dev. 2015;24:147– 59. https://doi.org/10.1089/scd.2014.0337.
- [13] Liu Y, Dong Y, Wu X, Xu X, Niu J. The assessment of mesenchymal stem cells therapy in acute on chronic liver failure and chronic liver disease: a systematic review and meta-analysis of randomized controlled clinical trials. Stem Cell Res Ther 2022;13. https://doi.org/10.1186/S13287-022-02882-4.
- [14] Xue R, Meng Q, Dong J, Li J, Yao Q, Zhu Y, et al. Clinical performance of stem cell therapy in patients with acute-on-chronic liver failure: a systematic review and meta-analysis. J Transl Med 2018;16. https://doi.org/10.1186/S12967-018-1464-0.
- [15] Zhou GP, Jiang YZ, Sun LY, Zhu ZJ. Therapeutic effect and safety of stem cell therapy for chronic liver disease: a systematic review and meta-analysis of randomized controlled trials. Stem Cell Res Ther 2020;11. https://doi.org/10.1186/S13287-020-01935-W
- [16] Tierney JF, Stewart LA, Ghersi D, Burdett S, Sydes MR. Practical methods for incorporating summary time-to-event data into meta-analysis. Trials 2007;8:1–16. https://doi.org/10.1186/1745-6215-8-16/FIGURES/2.
- [17] Page MJ, McKenzie JE, Bossuyt PM, Boutron I, Hoffmann TC, Mulrow CD, et al. The PRISMA 2020 statement: an updated guideline for reporting systematic reviews. BMJ 2021;372:n71. https://doi.org/10.1136/bmj.n71.
- [18] Barraclough H, Simms L, Govindan R. Biostatistics primer: what a clinician ought to know: hazard ratios. Journal of Thoracic Oncology 2011;6:978–82. https://doi. org/10.1097/JTO.0b013e31821b10ab.
- [19] Liu N, Zhou Y, Lee JJ. IPDfromKM: reconstruct individual patient data from published Kaplan-Meier survival curves. BMC Med Res Methodol 2021;21:111. https://doi.org/10.1186/s12874-021-01308-8.
- [20] Sterne JAC, Savović J, Page MJ, Elbers RG, Blencowe NS, Boutron I, et al. RoB 2: a revised tool for assessing risk of bias in randomised trials. BMJ 2019;366:14898. https://doi.org/10.1136/bmj.14898.
- [21] Hedges LV, Olkin I. Statistical methods for meta-analysis 2014.
- [22] Wickham H, Averick M, Bryan J, Chang W, McGowan L, François R, et al. Welcome to the tidyverse. JOSS 2019;4:1686. https://doi.org/10.21105/joss.01686.
- [23] Balduzzi S, Rücker G, Schwarzer G. How to perform a meta-analysis with R: a practical tutorial. Evid Based Mental Health 2019;22:153–60. https://doi.org/ 10.1136/ebmental-2019-300117.
- [24] Viechtbauer W. Conducting meta-analyses in r with the metafor package. J Stat Soft 2010;36. https://doi.org/10.18637/jss.v036.i03.
- [25] Amer MEM, El-Sayed SZ, El-Kheir WA, Gabr H, Gomaa AA, El-Noomani N, et al. Clinical and laboratory evaluation of patients with end-stage liver cell failure injected with bone marrow-derived hepatocyte-like cells. Eur J Gastroenterol Hepatol 2011;23:936–41. https://doi.org/10.1097/MEG.0B013E3283488B00.
- [26] Li YH, Xu Y, Wu HM, Yang J, Yang LH, Yue-Meng W. Umbilical cord-derived mesenchymal stem cell transplantation in hepatitis b virus related acute-on-chronic liver failure treated with plasma exchange and entecavir: a 24-month prospective study. Stem Cell Rev Reports 2016;12:645–53. https://doi.org/10.1007/S12015-016-9683-3.
- [27] Lin BL, Chen JF, Qiu WH, Wang KW, Xie DY, Chen XY, et al. Allogeneic bone marrow-derived mesenchymal stromal cells for hepatitis B virus-related acute-onchronic liver failure: a randomized controlled trial. Hepatology (Baltimore, Md) 2017;66:209–19. https://doi.org/10.1002/HEP.29189.
- [28] Peng L, ying Xie D, Lin BL, Liu J, peng Zhu H, Xie C, et al. Autologous bone marrow mesenchymal stem cell transplantation in liver failure patients caused by hepatitis B: short-term and long-term outcomes. Hepatology (Baltimore, Md) 2011;54:820-8. https://doi.org/10.1002/HEP.24434.
- [29] Schacher FC, Martins Pezzi Da Silva A, Silla LMDR, Álvares-Da-Silva MR. Bone marrow mesenchymal stem cells in acute-on-chronic liver failure grades 2 and 3: a phase I-II randomized clinical trial. Can J Gastroenterol Hepatol 2021 2021. https://doi.org/10.1155/2021/3662776.
- [30] Shi M, Zhang Z, Xu R, Lin H, Fu J, Zou Z, et al. Human mesenchymal stem cell transfusion is safe and improves liver function in acute-on-chronic liver failure

- patients. Stem Cells Transl Med 2012;1:725–31. https://doi.org/10.5966/ SCTM.2012-0034.
- [31] Zhu B, You S, Rong Y, Yu Q, Lv S, Song F, et al. A novel stem cell therapy for hepatitis B virus-related acute-on-chronic liver failure. Braz J Med Biol Res 2020;53:1–9. https://doi.org/10.1590/1414-431X20209728.
- [32] xiong Xu W, liang He H, wen Pan S, li Chen Y, ling Zhang M, Zhu S, et al. Combination treatments of plasma exchange and umbilical cord-derived mesenchymal stem cell transplantation for patients with hepatitis b virus-related acute-on-chronic liver failure: a clinical trial in China. Stem Cells Int 2019:2019. https://doi.org/10.1155/2019/4130757.
- [33] Xue R, Meng Q, Li J, Wu J, Yao Q, Yu H, et al. The assessment of multipotent cell transplantation in acute-on-chronic liver failure: a systematic review and meta-analysis. Transl Res 2018;200:65–80. https://doi.org/10.1016/J.TRSL.2018.05.006.
- [34] Gao J, Dennis JE, Muzic RF, Lundberg M, Caplan Al. The dynamic in vivo distribution of bone marrow-derived mesenchymal stem cells after infusion. Cells Tissues Organs 2001;169:12–20. https://doi.org/10.1159/000047856.
- [35] Xu R, Ni B, Wang L, Shan J, Pan L, He Y, et al. CCR2-overexpressing mesenchymal stem cells targeting damaged liver enhance recovery of acute liver failure. Stem Cell Res Ther 2022;13:55. https://doi.org/10.1186/s13287-022-02729-y.
- [36] Gholamrezanezhad A, Mirpour S, Bagheri M, Mohamadnejad M, Alimoghaddam K, Abdolahzadeh L, et al. In vivo tracking of 111In-oxine labeled mesenchymal stem cells following infusion in patients with advanced cirrhosis. Nucl Med Biol 2011;38:961–7. https://doi.org/10.1016/j.nucmedbio.2011.03.008.
- [37] Xiao Ling K, Peng L, Jian Feng Z, Wei C, Wei Yan Y, Nan S, et al. Stromal derived factor-1/CXCR4 axis involved in bone marrow mesenchymal stem cells recruitment to injured liver. Stem Cells Int 2016;2016:8906945. https://doi.org/ 10.1155/2016/8906945.
- [38] Mohamadnejad M, Vosough M, Moossavi S, Nikfam S, Mardpour S, Akhlaghpoor S, et al. Intraportal infusion of bone marrow mononuclear or CD133+ cells in patients with decompensated cirrhosis: a double-blind randomized controlled trial. Stem Cells Transl Med 2016;5:87–94. https://doi.org/10.5966/sctm.2015-0004.
- [39] Lyra AC, Soares MBP, da Silva LFM, Braga EL, Oliveira SA, Fortes MF, et al. Infusion of autologous bone marrow mononuclear cells through hepatic artery results in a short-term improvement of liver function in patients with chronic liver disease: a pilot randomized controlled study. Eur J Gastroenterol Hepatol 2010;22:33–42. https://doi.org/10.1097/MEG.0b013e32832eb69a.
- [40] Salama H, Zekri A-RN, Bahnassy AA, Medhat E, Halim HA, Ahmed OS, et al. Autologous CD34+ and CD133+ stem cells transplantation in patients with end stage liver disease. World J Gastroenterol 2010;16:5297–305. https://doi.org/10.3748/wjg.v16.i42.5297.
- [41] Wang P, Cui Y, Wang J, Liu D, Tian Y, Liu K, et al. Mesenchymal stem cells protect against acetaminophen hepatotoxicity by secreting regenerative cytokine

- hepatocyte growth factor. Stem Cell Res Ther 2022;13:94. https://doi.org/ 10.1186/s13287-022-02754-x.
- [42] Ferrini E, Stellari FF, Franceschi V, Macchi F, Russo L, Murgia A, et al. Persistency of mesenchymal stromal/stem cells in lungs. Front Cell Dev Biol 2021;9:709225. https://doi.org/10.3389/fcell.2021.709225.
- [43] Li G, Zhang P, Zhu Y. Artificial liver support systems for hepatitis B virus-associated acute-on-chronic liver failure: a meta-analysis of the clinical literature. J Viral Hepat 2023;30:90–100. https://doi.org/10.1111/jvh.13767.
- [44] Maiwall R, Bajpai M, Singh A, Agarwal T, Kumar G, Bharadwaj A, et al. Standard-volume plasma exchange improves outcomes in patients with acute liver failure: a randomized controlled trial. Clin Gastroenterol Hepatol 2022;20:e831–54. https://doi.org/10.1016/j.cgh.2021.01.036.
- [45] Camus C, Lavoué S, Gacouin A, Compagnon P, Boudjéma K, Jacquelinet C, et al. Liver transplantation avoided in patients with fulminant hepatic failure who received albumin dialysis with the molecular adsorbent recirculating system while on the waiting list: impact of the duration of therapy. Ther Apher Dial 2009;13:549-55. https://doi.org/10.1111/j.1744-9987.2009.00708.x.
- [46] Bauer G, Elsallab M, Abou-El-Enein M. Concise review: a comprehensive analysis of reported adverse events in patients receiving unproven stem cell-based interventions. Stem Cell Transl Med 2018;7:676–85. https://doi.org/10.1002/sctm.17-0282.
- [47] Yang S, Lin G, Tan Y-Q, Zhou D, Deng LY, Cheng DH, et al. Tumor progression of culture-adapted human embryonic stem cells during long-term culture. Genes Chromosomes Cancer 2008;47:665–79. https://doi.org/10.1002/gcc.20574.
- [48] Knoepfler PS. Deconstructing stem cell tumorigenicity: a roadmap to safe regenerative medicine. Stem Cells 2009;27:1050–6. https://doi.org/10.1002/stem.37.
- [49] Rizzo A, Ricci AD. Challenges and future trends of hepatocellular carcinoma immunotherapy. Int J Mol Sci 2022;23:11363. https://doi.org/10.3390/ijms231911363.
- [50] Thangaraj JL, Coffey M, Lopez E, Kaufman DS. Disruption of TGF-β signaling pathway is required to mediate effective killing of hepatocellular carcinoma by human iPSC-derived NK cells. Cell Stem Cell 2024. https://doi.org/10.1016/j.stem.2024. 06.009.
- [51] Wang MF, Li YB, Gao XJ, Zhang HY, Lin S, Zhu YY. Efficacy and safety of autologous stem cell transplantation for decompensated liver cirrhosis: a retrospective cohort study. World J Stem Cells 2018;10:138–45. https://doi.org/10.4252/WJSC. V10.110.138
- [52] Bin Abdul Aziz MF, Morrison M, Kaye J. Regulating human stem cell research and therapy in low- and middle-income countries: Malaysian perspectives. New Genet Soc 2018;37:2–20. https://doi.org/10.1080/14636778.2017.1415137.
- [53] Goozner M. Innovation in biomedicine: can stem cell research lead the way to affordability? Plos Med 2006;3:e126. https://doi.org/10.1371/journal.pmed.00 30126