

Hepatic apolipoprotein A-I gene expression in patients with cholesterol gallstones treated with ursodeoxycholic acid

Nahum Méndez-Sánchez,¹ Arturo Panduro,² Daniel Murguía,³ Ana R. Rincón,² and Misaél Uribe¹

Abstract

Objective: It has been suggested that apo A-I can inhibit cholesterol crystal nucleation *in vitro*, and ursodeoxycholic acid (UDCA) is a safe and effective treatment for selected patients with cholesterol gallstones the aim of this study was to investigate the effect of UDCA on the steady-state levels (SSL) of apo A-I mRNA in the liver, as well as serum apo A-I, in patients with cholesterol gallstones.

Design: Twenty Mexican patients with symptomatic radiolucent gallstones were randomized and assigned in a double blind fashion to groups that were administered either UDCA (4 mg/kg per day) or placebo for 10 to 15 days before cholecystectomy.

Methods: Apo A-I mRNA levels in liver and gallbladder tissues were determined by northern blot and serum levels of apo A-I by turbidimetric method.

Results: Apo A-I mRNA levels were higher in nine of the 10 patients who received UDCA and in comparison to those to the placebo group. In the gallbladder apo A-I mRNA levels were undetected. Serum levels (mg/dL) of apo A-I were similar in both UDCA and placebo groups after treatment (111.7 ± 29.8 vs 115.6 ± 25.4).

Conclusions: The results of this study shown that apo A-I mRNA gene express at the mRNA level in the liver but not in the gallbladder of patients with cholesterol gallstones treated with UDCA.

Key words: Apolipoprotein A-I, ursodeoxycholic acid, gallstones, cholesterol, bile acids.

Introduction

It has been suggested that changes to plasma HDL levels commonly reflect altered metabolism of the major HDL apolipoproteins, apo A-I and apo A-II, but the regulation of apolipoprotein metabolism is poorly understood.¹ Although a number of pharmacological and dietary factors may be mediated by these and other transcription factors to affect apo A-I levels, apo A-I gene expression has proven relatively insensitive to physiological change. Some of the states characterized by altered apo A-I gene expression in rodents include experimental hyperthyroidism (1.7–2.5-fold increased apo A-I mRNA),² and nephrotic syndrome (2-fold increased apo A-I mRNA).³ High fat diets are also associated with a 40% increase in apo A-I synthesis, reflecting increased translational efficiency of apo A-I mRNA.⁴ Interestingly, some research⁵ has reported that apolipoproteins A-I, A-II, and C-3 can inhibit cholesterol crystal formation in model biles. Furthermore, it has been observed that ursodeoxycholic acid [(UDCA) 3 α , 7 β , dihydroxy-5 β -cholanoic acid] induces an increase in serum apo A-I.⁶ UDCA also improves serum liver tests and histological features in a number of chronic cholestatic disorders, including primary biliary cirrhosis and primary sclerosing cholangitis.⁷ In addition, UDCA has been shown to prolong transplant-free survival in patients with primary biliary cirrhosis. The proposed mechanisms of action of UDCA in cholestasis involve the stimulation of impaired biliary secretion, immunomodulating effects, protection against injury of the bile ducts, and antiapoptotic effects.⁷ UDCA is also a safe and effective treatment for selected patients with radiolucent cholesterol gallstones.^{8,9} Jüngst *et al.*¹⁰ have shown that in patients with cholesterol gallstones a low dose of UDCA (250 mg/d) at bedtime for 6–10 days prior to cholecystectomy increases the time to cholesterol nucleation in gallbladder bile. In a similar group of patients, Tazuma *et al.*¹¹ reported that the administration of UDCA (300–600 mg/d) induced a significantly longer median nucleation time (16 days) compared with that in patients with cholesterol gallstones who received no preoperative treatment (4 days) ($p < 0.01$). Furthermore, three months of treatment with UDCA significantly elevated the serum concentrations of the antinu-

¹ Departments of Biomedical Research and Liver Unit, Medica Sur Clinic & Foundation, Mexico City, Mexico.

² Institute of Molecular Biology in Medicine and Hospital Civil de Belen, CUCS, University of Guadalajara, Jalisco, Mexico.

³ Gastroenterology Unit, General Hospital of Mexico City, Mexico.

Address for correspondence:

Nahum Méndez-Sánchez, M.D., PhD.

Departments of Biomedical Research and Gastroenterology, Medica Sur Clinic & Foundation, Puente de Piedra 150, Col. Toriello Guerra, Mexico City, Mexico. Phone: (+525) 606-6222, Ext. 4215 Fax: (+525) 666-4031 and 606-1651; E-mail: nmendez@medicasur.org.mx

Abbreviations: UDCA, ursodeoxycholic acid; apo A-I, apolipoprotein A-I; SSL, steady-state levels; HDL, high-density lipoprotein; LDL, low-density lipoprotein; VLDL, very low-density lipoprotein.

cleating factor apo A-I, from 133.3 ± 12.3 to 148.6 ± 13.2 mg/dL ($p < 0.05$). These findings suggest that UDCA retards cholesterol crystal nucleation, thereby inhibiting cholesterol gallstone formation. It is also possible that serum apo A-I plays a role in this process. The hypothesis that UDCA has an effect on the steady-state levels (SSL) of apo A-I mRNA in the liver, as well as serum apo A-I levels was tested in patients with cholesterol gallstones.

Patients and methods

Twenty consecutive patients who underwent elective cholecystectomy for symptomatic gallstone disease at the General Hospital of Mexico City were seen in a one-year period. Patients were selected according to the following criteria: functioning gallbladder and radiolucent gallstones, as documented by an oral cholecystogram four weeks before surgery, and a cholesterol content of $> 60\%$ in the removed stones. All patients gave their signed consent to participation in the study. Patients with clinical or laboratory evidence of diabetes mellitus, ethanol abuse, hyperlipidaemia, or liver, kidney, or thyroid function abnormalities, were excluded. No patient received oral contraceptive steroids or hormonal therapy. The experimental protocol was approved by the Ethics Committee of the General Hospital of Mexico and The Medica Sur Clinic & Foundation as conforming to the ethical guidelines of the 1975 Declaration of Helsinki, and written informed consent was obtained from all participants before entry. Eligible patients were randomized and assigned in a double blind fashion to groups that were administered either UDCA (4 mg/kg per day) or placebo capsules identical in appearance to those of UDCA (Ursofalk, Laboratorios Farmasa, S.A. de C.V. Mexico City) 10-15 days before cholecystectomy. Medication was well tolerated, and there were no side effects. Liver function tests and body weights remained unchanged during the whole treatment period. Patients were hospitalized in the Gastroenterology Surgical Unit, two to three days before surgery, and were fed, preoperatively, the standard hospital diet (carbohydrates 60%, fats 20%, proteins 20%).

All surgical procedures (open cholecystectomies) were performed between 08:00 and 10:00 am, after a 14-h fasting period. A wedge liver biopsy (100-400 mg) was routinely taken from the right lobe of the liver, frozen immediately on dry ice and acetone, transported to the laboratory, and stored at -70°C until the extraction of total RNA. Stones were removed from the gallbladder, washed with a solution of 0.9% NaCl, and the cholesterol content analysed by X-ray diffraction.¹² Immediately after removal of the gallbladder, samples of gallbladder tissue (15 x 15 mm) were taken from the neck, body, and fundus, and were frozen and stored as indicated previously.

Isolation of total cellular RNA

Total cellular RNA was isolated according to the method of Chomczynski and Sacchi, by acid guanidinium thio-

cyanate-phenol-chloroform extraction.¹³ Approximately 100 mg tissue was minced and homogenized with 2 mL denaturing solution (solution D) (4 M guanidinium thiocyanate, 25 mM sodium citrate, [pH 7.0], 0.5% sarcosyl, 0.1 M 2-mercaptoethanol). The homogenate was then transferred to a 15 mL Corex tube, and 0.2 mL 2 M sodium acetate (pH 4.0), 2 mL phenol at 65°C , and 0.4 mL chloroform were added. The final suspension was shaken, and cooled on ice for 15 min. Samples were centrifuged at 10,000 g for 20 min at 4°C , and the RNA in the aqueous phase was precipitated with 0.25 M sodium chloride and 2.5 volumes of ethanol. The pellets were resuspended in 0.4 mL solution D, and the phenol-chloroform extraction was repeated. The RNA was then dissolved in 200 μL TE buffer (10 mM Tris-HCl [pH 7.0], 1 mM EDTA).

Northern Hybridization Analysis of Total RNA

Total RNA (10 μg) was denatured for 15 min at 60°C in buffer containing 50% formamide, 6% formaldehyde, and MOPS buffer (20 mM 3-(N-morpholino) propanesulphonic acid, 5 mM sodium acetate, 1 mM $\text{Na}_2\text{-EDTA}$), and samples were loaded into separate lanes of a 1% agarose gel prepared in MOPS buffer with 6% formaldehyde. Samples were separated electrophoretically for 3-4 h at 60 mA, essentially as described previously.¹⁴ After electrophoresis, RNA was transferred to Gene Screen membrane (New England Nuclear Corp., Boston, MA, USA), according to the manufacturer's instructions, and hybridized with a radiolabelled probe. An apo A-I clone¹⁵ was radioactively labelled by primer extension, as described by Summers,¹⁶ using ^{32}P -dCTP (3000 Ci/mmol) to a specific activity of $2\text{-}6 \times 10$ cpm/ μg DNA. After hybridization, filters were washed and exposed to autoradiography at -86°C on Kodak XAR-5 film, using Dupont Lightening Plus intensifying screens. Finally, specific mRNA bands were analysed densitometrically, as described previously.¹⁷

Serum lipids and apolipoprotein A-I measurement

Serum cholesterol and triglyceride levels were determined in both groups, and total serum cholesterol concentration was measured before and after treatment, using the monotest cholesterol kit provided by Laboratories Lakeside (Mexico).¹⁸ Serum triacylglycerol concentrations were determined using an enzymatic method.¹⁹ Triacylglycerols were completely hydrolysed, and the liberated glycerol measured by colorimetry. HDL were separated from VLDL and LDL by precipitation with phosphotungstic acid.²⁰ LDL cholesterol was precipitated by adding polyvinyl sulphate to the sample, and the concentration was calculated from the difference between total cholesterol and the cholesterol in the supernatant after centrifugation. HDL and LDL cholesterol were determined as described previously.¹⁸ VLDL cholesterol was calculated as the difference between total plasma and (LDL + HDL) cholesterol.²¹ The accuracy of HDL and LDL measurements

were verified using control sera, as indicated by the manufacturer (Laboratories Lakeside). Serum apo A-I levels were determined in both groups, before and after treatment, by a turbidimetric method.²²

Statistical Analysis

All results are expressed as the mean \pm SD or median and range. Mann-Whitney U test and Student's unpaired test were used for group comparisons, with the level of statistical significance set at $p < 0.05$.²³

Results

The clinical characteristics of patients were similar in both the UDCA and placebo groups: mean age (years) 43.6 ± 14.4 vs 38.9 ± 10.5 , respectively, and body weight (kg) 62.2 ± 6.1 vs 63.3 ± 9.4 , respectively.

Steady-state levels of apo A-I mRNAs

The steady-state levels of hepatic apo A-I mRNA increased in nine of 10 patients after UDCA therapy (Figure 1), but in none of the placebo group. Studies performed on gallbladder tissue (neck, body, and fundus) in both groups of patients failed to reveal apo A-I mRNA (Figure 2).

Serum Apolipoprotein A-I

Basal serum levels (mg/dL) of apo A-I were similar in both UDCA and placebo groups (110.7 ± 13.5 vs 126.9 ± 29.1), and were not significantly different after treatment (111.7 ± 29.8 vs 115.6 ± 25.4). However, we found an increase in serum levels of apo A-I in three of nine patients (in another patient, the liver sample was insufficient to measure apo A-I) who presented with an increase in SSL of apo A-I mRNA (Table I).

Serum Lipids and Lipoproteins

Table II summarizes the serum levels of lipids (cholesterol and triglycerides) and lipoproteins (HDL, LDL, and VLDL). No significant differences were observed between UDCA and placebo groups, before or after treatment, except in serum levels of triglycerides in the placebo group, which were lower in patients after treatment (140.10 ± 47.81 vs 107.20 ± 53.48), therefore becoming significantly different from the UDCA values.

Discussion

In this study, we have observed that patients with cholesterol gallstones treated with low doses of UDCA (4 mg/kg per day) for a short period (10 to 15 days) showed higher he-

patic transcription of the apo A-I gene. However, apo A-I protein was not increased in the serum of all patients with high levels of hepatic apo A-I mRNA. Similar results have been observed under different pathophysiological conditions in experimental animal models of liver disease,²⁴ suggesting postranscriptional regulation of apo A-I expression. However, the question arises whether an increase in the expression of the apo A-I gene at the mRNA level has a beneficial effect on liver function, as well as on the biliary tract. As mentioned above, apo A-I appears to play a role as an antinucleating factor in model bile. This effect is probably important in preventing cholesterol gallstones. Swell *et al.*^{25,26} have suggested that the apo A-I present in human gallbladder bile is derived mainly from the liver. Interestingly, Poynard *et al.*^{27,28} have reported that, among drinkers, apo A-I and apo A-II serum concentrations are closely related to the degree of liver injury, reaching a maximum in patients with steatosis, decreasing in patients with fibrosis, and at a minimum in patients with severe cirrhosis. Furthermore, the prevalence of cholesterol gallstone disease in patients without fibrosis (normal liver or steatosis) is low, at 5% and 6%, respectively. In contrast, in patients with fibrosis or severe liver disease, gallstone occurrence increases to 13% and 25%, respectively.²⁷ In another prospective study on 67 male patients with cirrhosis, gallstone disease was found in 37% of the patients. The occu-



Figure 1. Steady-state levels of liver apo A-I mRNA after UDCA treatment (1 to 9) and in the placebo (©) patients. Densitometric analysis showed an increase in apo A-I mRNA levels in patients undergoing UDCA therapy compared with those in the placebo group.

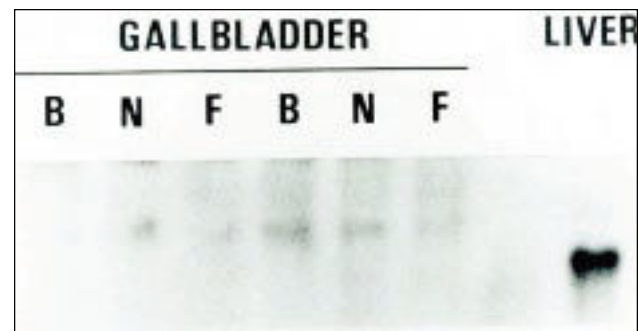


Figure 2. Steady-state levels of apo A-I mRNA expression in the liver were not detected in the gallbladder in either group of patients. B, body; N, neck; and F, fundus.

rence of gallstone disease was not related to age, weight, or the severity of liver disease.²⁹ Total serum bilirubin was higher and apo A-I levels were lower in those patients with gallstones. Apo A-I was the only factor associated independently with the occurrence of gallstones. It is important to mention that patients with chronic hepatic diseases usually develop pigmented gallstones,³⁰ and the pathogenesis of these types of stones is different from that of cholesterol gallstones. However, in both types of stones the bile was saturated with cholesterol or bilirubin, and the role of antinucleating factors such as apo A-I may be very important. In contrast, Otha *et al.*³¹ detected a decrease in the distribution of apo A-I in the liver and biliary tree in patients with intrahepatic cholesterol and brown pigmented stones. Interestingly, Pattinson *et al.*³² have identified apo A-I, apo A-II, and apo-B in normal and pathologically affected human bile ducts and in the gallbladder epithelium, using an avidin-biotin immunoperoxidase technique. They suggest a role for these apolipoproteins in the gallbladder epithelium. However, in those two publications, the researchers identified apo A-I by using an immunohistochemical technique that is less specific than the northern blot used in this study. The immunohistochemical technique is also known to produce cross-reactions.

Table I. Effects of UDCA treatment on apo A-I gene expression at the mRNA and serum levels.

Patients	Apo A-I		
	Hepatic mRNA levels ^a (Liver tissue)	Serum protein levels (mg/dL)	
		Before	After
1	4.5	132	160
2	3.8	118	120
3	6.1	116	154
4	1.8	98	82
5	1.5	92	88
6	3.2	105	69
7	2.3	118	106
8	6.0	127	119
9	5.8	104	94
10	N.D.	97	125
Mean \pm SD		110.7 \pm 13.5	111.7 \pm 29.9 ^b

^a Arbitrary units; N.D., not determined; ^b not statistically significant.

Table II. Serum lipids and lipoproteins in patients treated with a low dose of UDCA.

	UDCA	Placebo	<i>p</i> value
<i>n</i>	10	10	
Cholesterol	171.2 \pm 53.7	146.4 \pm 37.2	NS
Triglycerides	51.9 \pm 47.5	107.2 \pm 53.5	0.04
HDL	35.3 \pm 7.9	33.4 \pm 5.5	NS
LDL	108.0 \pm 48.5	92.0 \pm 28.5	NS
VLDL	25.8 \pm 8.6	21.0 \pm 10.8	NS

Values are expressed as mean \pm SD.

HDL, high-density lipoprotein; LDL, low-density lipoprotein; VLDL, very low-density lipoproteins; NS, not significant.

Secknus *et al.* have proposed a purified 15 kDa protein, derived from the whole apo A-I-bound fraction, as a novel potent biliary crystallization inhibitor protein.³³ The same group of investigators reported that apo A-I in bile has a direct effect on cholesterol crystal formation, and enhances lipid removal from gallbladder bile by gallbladder epithelial cells.³⁴ These observations support two separate roles for human biliary apo A-I, and suggest that this protein may be important in preventing the formation of cholesterol crystals (the initial step in gallstone formation) in supersaturated bile. Recently, Ginanni Corradini *et al.*³⁵ proposed that one of the main defects in cholesterol gallstone patients occurs at the **gallbladder** mucosa. They found, using an *in vitro*-isolated intra-arterially perfused **gallbladder** model, that human **gallbladder** epithelium loses its capacity to selectively and efficiently absorb cholesterol and phospholipids from bile. These observations suggest that this defect is not only relevant at the level of lipid **absorption**, but may also involve other aspects of epithelial function, such as an increased secretion of mucin or other proteins that promote cholesterol crystallization and decreased secretion of anti-crystallization proteins such as apo A-I.

On the other hand, experiments have suggested that UDCA increases bile flow in the rat,³⁶ and that apo A-I is transported to the bile at the canalicular level.³⁷ Furthermore, Van Erpecum *et al.*³⁸ studied the gallbladder bile of 13 patients with cholesterol gallstones who were treated with UDCA (10 mg/kg per day), and compared them with 13 untreated patients. They found higher concentrations of total protein in the gallbladder bile of UDCA-treated patients than in untreated patients (6.7 \pm 1.3 vs 2.8 \pm 0.6 mg/mL, respectively; *p* = 0.008) and similarly higher concentrations in the concanavilin A-binding fraction (0.42 \pm 0.07 vs 0.16 \pm 0.03 mg/mL, respectively; *p* = 0.003). They concluded that UDCA greatly reduces the levels of various proteins and has nucleation-promoting activity in bile.

Finally, in humans, low HDL cholesterol levels may be associated with defects in the synthesis or catabolism of the major HDL apolipoprotein, apoA-I, with catabolic defects being more common.^{39,40} Low HDL levels are often accompanied by hypertriglyceridaemia, obesity, and insulin resistance, and these alterations are often seen in patients with cholesterol gallstones.⁴⁰ These data suggest that apo A-I plays an important role in the pathogenesis of cholesterol gallstones.

In conclusion, the results of this study shown that apo A-I mRNA gene express at the mRNA level in the liver but not in the gallbladder of patients with cholesterol gallstones treated with UDCA.

Acknowledgements: This work was partially supported by grants from Fundación Ricardo Zevada and CONACYT (903965) to Dr. Misael Uribe and (0014-M9105) to Dr. Arturo Panduro. Dr. Nahum Méndez-Sánchez was the recipient of a Mexican Secretary of Health Scholarship as part of the PUIS Program, UNAM, and CONACYT.

References

- Silver DL, Jiang JC, Tall AR. Increased high density lipoprotein (HDL), defective hepatic catabolism of Apo A-I and Apo A-II, and decreased Apo A-I mRNA in ob/ob mice. *J Biol Chem* 1999; 274: 4140-4146.
- Mooradian AD, Wong NC, Shah GN. Age-related changes in the responsiveness of apolipoprotein A1 to thyroid hormone. *Am J Physiol* 1996; 271: R1602-7.
- Zaiou M, Azrolan N, Hayek T, Wang H, Wu L, Haghighpassand M, et al. The full induction of human apolipoprotein A-I gene expression by the experimental nephrotic syndrome in transgenic mice depends on cis-acting elements in the proximal 256 base-pair promoter region and the trans-acting factor early growth response factor 1. *J Clin Invest* 1998; 101: 1699-1707.
- Azrolan N, Odaka H, Breslow JL, Fisher EA. Dietary fat elevates hepatic apo A-I production by increasing the fraction of apolipoprotein A-I mRNA in the translating pool. *J Biol Chem* 1995; 270: 19833-19838.
- Kibe A, Holzbach RT, LaRusso NF, Mao SJT. Inhibition of cholesterol crystal formation by apolipoproteins in supersaturated mode bile. *Science* 1984; 255: 514-16.
- Hofmann AF. Pharmacology of ursodeoxycholic acid, an enterohepatic drug. *Scand J Gastroenterol Suppl* 1994; 204: 1-15.
- Poupon R, Chazouilleres O, Poupon RE. Chronic cholestatic diseases. *J Hepatol* 2000; 32(Suppl 1): 129-40.
- May GR, Sutherland LR, Shaffer EA. Efficacy of bile acid therapy for gallstone dissolution: a meta-analysis of randomized trials. *Aliment Pharmacol Ther* 1993; 7: 139-48.
- Petroni ML, Jazrawi RP, Pazzi P, Lanzini A, Zuin M, Pigozzi MG, et al. Ursodeoxycholic acid alone or with chenodeoxycholic acid for dissolution of cholesterol gallstones: a randomized multicentre trial. The British-Italian Gallstone Study Group. *Aliment Pharmacol Ther* 2001; 15: 123-8.
- Jungst D, Brenner G, Pratschke E, Paumgartner G. Low-dose ursodeoxycholic acid prolongs cholesterol nucleation time in gallbladder bile of patients with cholesterol gallstones. *J Hepatol* 1989; 8: 1-6.
- Tazuma S, Sasaki H, Mizuno, Sawaga H, Hashiba S, Horiuchi Y, et al. Effect of ursodeoxycholic acid administration on nucleation time in human gallbladder bile. *Gastroenterology* 1989; 97: 173-8.
- Sutor DJ. X-ray diffraction studies of composition of gallstones from English and Australian patients. *Gut* 1969; 10: 681-3.
- Chomczynski P, Sacchi N. Single step method of RNA isolation by acid guanidinium thiocyanate-phenol-chloroform extraction. *Annal Biochem* 1987; 162: 156-9.
- Panduro A, Shalaby F, Weiner F, Biempieca L, Zern M, Shafritz DA. Transcriptional switch from albumin to alpha-fetoprotein and changes in transcription of other genes during CC14 induced liver regeneration. *Biochemistry* 1986; 25: 1414-20.
- Panduro A, Lin-Lee Y, Chan L, Shafritz DA. Transcriptional and post-transcriptional regulation of apolipoprotein E, A-I and A-II gene expression in normal rat liver and during several pathophysiologic states. *Biochemistry* 1990; 29: 8130-35.
- Summers J. Physical map of polyoma viral DNA fragments produced by cleavage with restriction enzyme from *Haemophilus aegyptius* endonuclease R, Hae II. *J Virol* 1975; 15: 946-953.
- Maldonado V, Chan L, Meléndez J, Rincón AR, Zhu Hui-Hia, Panduro A. Regulation of apo B mRNA expression in liver and intestine during liver regeneration induced by CC14. *Biochim Biophys Acta* 1994; 1211: 1-6.
- Siedel J, Schlumberger H, Klose S, Ziegenhorn J, Wahlefeld AW. Improved reagent for the enzymatic determination of serum cholesterol. *J Clin Chem* 1981; 19: 838-9.
- Lehnus G, Smith L. Automated procedure for kinetic measurement of total triglyceride (as glycerol) in serum with the Gilford system 3500. *Clin Chem* 1978; 24: 27-31.
- Burnstein M, Scholnick HR, Morfin R. Rapid method for the isolation of lipoproteins from human serum by precipitation with polyanions. *J Lipid Res* 1970; 11: 583-595.
- Bronzert T, Brewer HB. New micromethod for measuring cholesterol in plasma lipoproteins fractions. *Clin Chem* 1977; 23: 2089-1098.
- Weiswell P, Schawardt P. Hyperlipoproteinemia: the apolipoprotein determination. *Med Lab* 1985; 13: 10-16.
- Winer BJ. *Statistical principles in experimental design*. 2nd ed. New York: McGraw-Hill; 1971: 149-160.
- Panduro A, Castrillon L, Gonzalez L, Shafritz DA. Regulation of hepatic and non-hepatic apolipoprotein A-I and E gene expression during liver regeneration. *Biochim Biophys Acta* 1993; 1167: 37-42.
- Swell RB, Mao SJ, Kawamoto T, LaRusso NF. Apolipoproteins of high, low, and very low density lipoproteins in human bile. *J Lipid Res* 1983; 24: 391-401.
- Kawamoto T, Mao SJT, LaRusso NF. Biliary excretion of apolipoprotein B by the isolated perfused rat liver. *Gastroenterology* 1987; 92: 1236-1242.
- Poynard T, Lonjon I, Mathurin P, Abella A, Musset D, Bedossa P, et al. Prevalence of cholelithiasis according to alcoholic liver disease: a possible role of apolipoproteins AI and AII. *Alcohol Clin Exp Res* 1995; 19: 75-80.
- Mathurin P, Vidaud D, Vidaud M, Bedossa P, Paradis V, Ratzu V, et al. Quantification of apolipoprotein A-I and B messenger RNA in heavy drinkers according to liver disease. *Hepatology* 1996; 23: 44-51.
- Baranda J, Ministro P, Amaro P, Rosa A, Pimenta I, Donato A, et al. Apolipoprotein A1 and biliary lithiasis in hepatic cirrhosis. *Acta Med Port* 1996; 9: 203-6.
- Cahalane MJ, Neubrand MW, Carey MC. Physical-chemical pathogenesis of pigment gallstones. *Semin Liver Dis* 1988; 8: 317-328.
- Ohta T, Nagakawa T, Takeda T, Fonseca L, Kanno M, Mori K, et al. Histological evaluation of the intrahepatic biliary tree in intrahepatic cholesterol stones, including immunohistochemical staining against apolipoprotein A-I. *Hepatology* 1993; 17: 531-7.
- Pattinson NR, Upton P, Ellingsen PJ, Chapman BA. Apolipoprotein localization in the human bile duct and gallbladder. *Pathology* 1990; 22: 55-60.
- Secknus R, Yamashita G, Ginanni Corradini S, Chernosky A, Williams C, et al. Purification and characterization of a novel human 15 kd cholesterol crystallization inhibitor protein in bile. *J Lab Clin Med* 1996; 127: 169-78.
- Secknus R, Darby GH, Chernosky A, Juvonen T, Moore EW, Holzbach RT. Apolipoprotein A-I in bile inhibits cholesterol crystallization and modifies transcellular lipid transfer through cultured human gallbladder epithelial cells. *J Gastroenterol Hepatol* 1999; 14: 446-56.
- Ginanni Corradini S, Elisei W, Giovannelli L, Ripani C, Della Guardia P, Corsi A, et al. Impaired human gallbladder lipid absorption in cholesterol gallstone disease and its effect on cholesterol solubility in bile. *Gastroenterology* 2000; 118: 912-920.
- Pérez Barriocanal F, Marin JJ, Dumont M, Erlinger S. Influence of backward perfusion on ursodeoxycholate-induced cholestasis in isolated *in situ* rat liver. *J Hepatol* 1990; 11: 165-71.
- Berr F, Jaeger H, Walli A, Bittherle TH, Wilcox H. Biliary secretion of phospholipids and apolipoprotein A-1: evidence for intracellular supply of phospholipids by a non-vesicular transport route. (Abstract). *Hepatology* 1993; 18: 179.
- Van Erpecum KJ, Portincasa P, Eckhardt E, Go PM, VanBerge-Henegouwen GP, Groen AK. Ursodeoxycholic acid reduces protein levels and nucleation-promoting activity in human gallbladder bile. *Gastroenterology* 1996; 110: 1225-37.
- Brinton EA, Eisenberg S, Breslow JL. Human HDL cholesterol levels are determined by apo A-I fractional catabolic rate, which correlates inversely with estimates of HDL particle size. Effects of gender, hepatic and lipoprotein lipases, triglyceride and insulin levels, and body fat distribution. *Arterioscler Thromb* 1994; 14: 707-20.
- Fidge N, Nestel P, Ishikawa T, Reardon M, Billington T. Turnover of apoproteins A-I and A-II of high density lipoprotein and the relationship to other lipoproteins in normal and hyperlipidemic individuals. *Metabolism* 1980; 29: 643-53.