

# New Concepts of Mechanisms of Intestinal Cholesterol Absorption

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## Abstract

The small intestine is a unique organ providing dietary and reabsorbed biliary cholesterol to the body. However, the molecular mechanisms whereby cholesterol is absorbed have not yet been fully understood. Recent research suggests that the newly identified ATP-binding cassette (ABC) transporters ABCG5 and ABCG8 are apical cholesterol export pumps that promote partial efflux of cholesterol and nearly complete efflux of plant sterols from enterocytes into the intestinal lumen after their absorption. This provides an explanation why cholesterol absorption is a selective process in that plant sterols and other non-cholesterol sterols are absorbed poorly or not at all. Furthermore, a putative cholesterol import protein has been proposed, but remains uncharacterized. The identification of such a gene should yield new insights into the mechanisms that potentially regulate the influx of cholesterol across the apical brush border membrane of the enterocyte. Combination therapy using a novel and potent cholesterol absorption inhibitor (ezetimibe) and an HMG-CoA reductase inhibitor (statins) offers an efficacious new approach to the prevention and treatment of hypercholesterolemia.

**Key words:** Bile salt, cholesterol transporter phospholipid, micelle, chylomicron, lymph, nutrition, sitosterol.

## Introduction

Cholesterol homeostasis is maintained by balancing intestinal cholesterol absorption and endogenous cholesterol synthesis with excretion of biliary cholesterol and bile acids. Since elevated plasma cholesterol is an independent risk factor for coronary heart disease<sup>1</sup> and biliary cholesterol hypersecretion is an important prerequisite for cholesterol cholelithiasis,<sup>2</sup> considerable interest has been focused on identifying biochemical, physical-chemical, and genetic determinants of intestinal cholesterol absorption. Furthermore, understanding the sequential steps in intestinal cholesterol absorption may lead to novel approaches to the treatment of these diseases that affect millions in Westernized societies.

## Physiological mechanisms of intestinal cholesterol absorption

“Absorption of cholesterol” is most accurately defined as the transfer of intraluminal cholesterol into intestinal or thoracic duct lymph. “Uptake of cholesterol” refers to entry of cholesterol into intestinal absorptive cells. According to these definitions, cholesterol absorption is a multistep process that is regulated by multiple genes.<sup>3,4</sup>

There are three sources for intestinal cholesterol: the diet, the bile, and intestinal epithelial sloughing. The average daily intake of cholesterol in the Western diet is approximately 300-500 mg. Bile provides 800-1,200 mg of cholesterol per day to the intraluminal pool. The turnover of intestinal mucosal epithelium provides a third source of intraluminal cholesterol, and estimates of this contribution are 300 mg of cholesterol per day. Although the entire length of the small intestine has the capability to absorb cholesterol from the lumen, main sites of absorption are the duodenum and proximal jejunum.

Cholesterol absorption begins in the stomach when dietary constituents are mixed with lingual and gastric enzymes. The stomach also functions to regulate the delivery of gastric chyme to the duodenum, where it is mixed with bile and pancreatic juice. This process continues within the lumen of the small intestine. Hydrolytic enzymes secreted by the pancreas and bile salts of bile solubilize the hydrolytic end products of intraluminal fat digestion,<sup>5,6</sup> and a variable proportion of dietary cholesterol is esterified to fatty acids.

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Abbreviations: ABC, ATP-binding cassette (transporter); ACAT, acyl-CoA:cholesterol acyltransferase; Mdr2, multidrug resistance gene 2; SR-BI, scavenger receptor class B type I; UDCA, ursodeoxycholic acid.

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Some of the lipolytic products, including cholesterol, are only minimally soluble in aqueous systems and are dependent on the solubilizing properties of bile salt solutions.<sup>7,8</sup> Bile salts are biological amphipathic detergents which, when present above a critical micellar concentration, spontaneously form aggregates that are able to dissolve lipids.<sup>9,10</sup> Cholesterol is only sparingly soluble in bile salt solutions, in contrast to phospholipid, monoacylglyceride and free fatty acid that are readily soluble. The addition of phospholipid or monoacylglyceride to bile salt solutions markedly increases the solubility of cholesterol.<sup>11</sup> Bile salts together with ionized and non-ionized fatty acids, monoacylglyceride, (lyso) phospholipid, and unesterified cholesterol form mixed micelles.<sup>12</sup> Excess lipids not dissolved in the micellar phase form a separate oil phase within the intestinal lumen,<sup>5,6</sup> and may be maintained as a stable emulsion by bile salt, phospholipid, monoacylglyceride, and ionized fatty acid. In addition, during lipolysis, a liquid crystalline phase composed of multilamellar products of lipid digestion forms at the surface of an emulsion droplet.<sup>13,14</sup> The liquid crystalline phase provides an accessible source of phospholipid, monoacylglyceride, fatty acid, and cholesterol for the formation of mixed micelles in the presence of bile salt.

Cholesterol is absorbed by the small intestine solely as monomers. The unstirred water layer, a series of water lamellae at the interface between the bulk water phase of the lumen and the mucosal cell membrane, and the cell membrane form two barriers through which a cholesterol molecule in the bulk phase must pass in order to be absorbed.<sup>15</sup> Diffusion through the unstirred water layer is a relatively slow process for cholesterol that is nearly insoluble in a pure aqueous system. These micelles function as a concentrated reservoir and transport vehicle for cholesterol across the unstirred water layer toward the brush border of the small intestine to facilitate uptake of monomeric cholesterol by the enterocyte.<sup>13,14</sup> Once monomers are taken up by enterocytes from the bile salt-rich reservoirs of intraluminal contents, the latter, in turn, are replete the intermicellar water with cholesterol monomers by their desorption from mixed micelles. As individual molecules of cholesterol are taken up into the cell membrane, other molecules of cholesterol move from the micelles into monomolecular solution and become available for uptake by the intestinal absorptive cells. However, it is not known how cholesterol within the intestinal lumen moves from the lumen into the intestinal absorptive cells, i.e., the entry of cholesterol molecules into cell membranes. We will further discuss it below (see Searching for intestinal cholesterol transporters).

During the absorption of cholesterol, there is little increase in the cholesterol content of the small intestine, indicating that cholesterol can be rapidly processed and exported from the mucosal cells and into the intestinal lymph.<sup>3,4</sup> Studies have shown that following an intragastric dose of cholesterol mass and radioactivity, the trans-

port of both in intestinal lymph rapidly increases and peaks after 6-8 hours.<sup>3,4</sup> Upon entering the enterocytes, approximately half the cholesterol molecules move to the endoplasmic reticulum, where they are esterified by acyl-CoA:cholesterol acyltransferase (ACAT) before incorporation into nascent chylomicron particles. Of note is that essentially all cholesterol that moves from the intestinal lumen into the intestinal mucosal cells is unesterified; however, cholesterol secreted into intestinal lymph following a cholesterol-rich meal is approximately 70-80% esterified. The cholesterol esterifying activity of the mucosa may be an important regulator of cholesterol absorption, since re-esterification of absorbed free cholesterol within the mucosal cell would enhance the diffusion gradient for free cholesterol into the cell. Further, transmucosal transport of cholesterol is reduced in rats with inhibition of mucosal ACAT.<sup>16,17</sup> It is found that ACAT1 is expressed in many tissues; however, the second enzyme, ACAT2 is localized specifically to liver and small intestine, and is most likely responsible for esterification of cholesterol absorbed from the intestine.<sup>18</sup> Also, the inhibition of intestinal 3-hydroxy-3-methylglutaryl-CoA reductase by pharmacological intervention with "statins," decreases intestinal cholesterol absorption in animals and humans.<sup>19-21</sup> Moreover, cholesterol absorption is significantly inhibited in *apolipoprotein B-48* deficient mice<sup>22</sup> because of a failure in the assembly and/or secretion of chylomicrons, suggesting that this final step in the absorption process is critically important. Although intestinal microsomal triglyceride transfer protein and APOA1/C3/A4 have been considered to possibly play some roles in the regulation of cholesterol absorption,<sup>23-25</sup> their effects will need to be further studied.

Finally, cholesterol and bile salts that escape intestinal re-absorption and are excreted as fecal neutral and acidic sterol represent the major route for sterol elimination from the body.

### Factors influencing intestinal cholesterol absorption efficiency

As cholesterol absorption is a multistep process, any factor that changes the transportation of cholesterol from the intestinal lumen to the lymph may influence intestinal cholesterol absorption efficiency. *Table 1* summarizes dietary, including administered therapeutic agents, biliary, cellular, and luminal factors that could influence intestinal cholesterol absorption. Despite these findings, it remains poorly understood which step(s) in the absorption process are rate-limiting, as well as differ inherently among individuals in any population to explain variations in intestinal cholesterol absorption efficiency.

When the dietary conditions are controlled, biliary factors such as secretion rates of biliary lipids (bile salt, cholesterol, and phospholipid), and cholesterol content of bile, as well as size, molecular composition, and hydro-

**Table I.** Possible factors influencing intestinal cholesterol absorption.

Factors	Effects on percent cholesterol absorption and type of study
a) Dietary factors	
↑ Cholesterol	↓ Human and animal feeding studies
Fat	
↑ Monounsaturated	↓ African green monkey feeding studies
↑ $\omega$ -3 polyunsaturated	↓ African green monkey feeding studies
↑ Fish oils	↓ Rat lymphatic transport studies
↑ Fiber	↓ Human and animal feeding studies
↑ Plant sterols (Phytosterols)	↓ Human and animal feeding studies
↑ Hydrophilic bile acids	↓ Human and animal feeding studies
↑ Sphingomyelin	↓ Animal feeding studies
b) Biliary factors	
↓ Biliary bile salt output	↓ Cholesterol 7 $\alpha$ -hydroxylase (–/–) mice
↓ Size of biliary bile salt pool	↓ Cholesterol 7 $\alpha$ -hydroxylase (–/–) mice
↑ Biliary phospholipid output	↓ <i>Mdr2</i> (–/–) mice <sup>b</sup>
↑ Biliary cholesterol output	↑ Diabetic mice
↑ Cholesterol content of bile	↑ Diabetic mice
↑ HI of biliary bile salt pool	↑ Diabetic mice
c) Cellular factors	
↓ ACAT2	↓ ACAT2 inhibitors
↓ HMG-CoA reductase	↓ HMG-CoA reductase inhibitors
↓ ABCA1	↓ <i>Abca1</i> (–/–) mice <sup>c</sup>
ABCG5	Sterol efflux transporter
ABCG8	Sterol efflux transporter
Cholesterol transporter	To be identified
SR-BI	(–) <i>Sr-b1</i> (–/–) mice, and ↓ <i>Sr-b1</i> transgenic mice <sup>c</sup>
Caveolin	To be identified.
MTTP	To be identified.
↓ ApoB-48	↓ ApoB-48 (–/–) mice
ApoA-I, ApoA-IV, ApoC-III	To be identified
d) Luminal factors	
↑ Small intestinal transit time	↑ <i>Cck-A</i> receptor (–/–) mice
↓ Carboxyl ester lipase	(–) Carboxyl ester lipase (–/–) mice
Sphingomyelinase	To be identified.

<sup>a</sup> ↑ represents decrease, ↓ increase, and (–) no effect.

<sup>b</sup> Abbreviations: *Mdr2*, multidrug resistance gene 2; HI, hydrophobicity index; ACAT2, acyl-CoA:cholesterol acyltransferase, isoform 2; HMG, 3-hydroxy-3-methylglutaryl; ABC, ATP-binding cassette (transporter); SR-BI, scavenger receptor class B type I; MTTP, microsomal triglyceride transfer protein; Apo, apolipoprotein; CCK, cholecystokinin.

<sup>c</sup> Contradictory results were reported by different groups (see Text for details).

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philic-hydrophobic balance of the bile salt pool,<sup>9,10</sup> could together exert major influences on the efficiency of intestinal cholesterol absorption. Any of these could explain, in part, the inter-individual and inter-strain differences in cholesterol absorption efficiency. For example, knockout of *multidrug resistance gene 2* (*Mdr2*) inhibits biliary secretion of phospholipid, and curtails intestinal cholesterol absorption efficiency.<sup>26,27</sup> Studies in homozygous and heterozygous *Mdr2* deficient mice suggest that physiological phospholipid outputs are necessary for normal intestinal cholesterol absorption.<sup>27</sup> In *cholesterol 7 $\alpha$ -hydroxylase* knockout mice,<sup>28</sup> biliary bile salt pool sizes and biliary bile salt outputs are reduced markedly and the animals absorb only trace amounts of cholesterol because of bile salt deficiency. However, cholesterol absorption is reversed readily by feeding a diet containing 0.2% cholic acid. This confirms that biliary bile salt pool size and biliary bile salt output play a crucial role in cholesterol ab-

sorption via intraluminal bile salt micellar concentrations. Changes in the hydrophilic-hydrophobic balance of the bile salt pool also influence cholesterol absorption.<sup>9</sup>

In addition, it is observed<sup>29</sup> that the higher cholesterol-absorbing C57L mice have significantly higher secretion rates of all three major biliary lipids (bile salts, cholesterol, and phospholipids) and elevated cholesterol content of bile compared with the lower cholesterol-absorbing AKR mice. These results show a positive relationship between high intestinal cholesterol absorption and augmented biliary cholesterol outputs, as well as high gallstone prevalence rates, suggesting that the effect on bile is secondary to increased intestinal cholesterol absorption.<sup>3,4</sup>

Several studies in humans and animals suggest that rapid small intestinal transit time reduces cholesterol absorption. Ponz de Leon et al<sup>30</sup> produced evidence in humans by pharmacological intervention that acceleration of small intestine transit is consistently associated with decreased

cholesterol absorption. Also, Traber and Ostwald<sup>31</sup> found that in guinea pigs resistant to the plasma effects of dietary cholesterol, intestinal transit times are more rapid than in other guinea pigs with hypercholesterolemia. Furthermore, *cholecystokinin-A receptor* deficient mice display significantly higher intestinal cholesterol absorption rates which correlate with slow small intestinal transit rates; this in turn induces biliary cholesterol hypersecretion and cholesterol gallstone formation.<sup>32</sup> However, it is surprising to find that small intestinal transit time and the length and weight of small intestine among low, middle, and high cholesterol-absorbing mouse strains are essentially identical.<sup>4</sup> Taken together, these studies reveal that under the physiological conditions, luminal factors are not responsible for the differences in intestinal cholesterol absorption efficiency in these diverse mouse strains. In addition, intestinal cholesterol absorption efficiency increases markedly with aging<sup>33,34</sup> and there are gender differences in cholesterol absorption efficiency,<sup>33-35</sup> suggesting that aging and female sex hormones could have some effects on cholesterol absorption.

### Genetic influences on variations in intestinal cholesterol absorption efficiency

It has been observed that with similar dietary cholesterol intake, inter-individual and inter-strain variations in intestinal cholesterol absorption efficiency exist in primates,<sup>36,37</sup> including humans,<sup>38-41</sup> as well as in inbred strains of rabbits,<sup>42,43</sup> rats,<sup>44</sup> and mice.<sup>4,45-47</sup> This strongly suggests that intestinal cholesterol absorption is regulated by multiple genes since diet, a key environmental factor is controlled in these studies. Recently, it is found<sup>4</sup> that there are variations in cholesterol absorption efficiency among 12 strains of inbred mice, and bile salt secretion rates and pool sizes are two key biliary factors in the regulation of intestinal cholesterol absorption. Of note is that neither molecular compositions of bile salt pool nor small intestinal transit times vary significantly among these mouse strains.<sup>4</sup> Especially, when dietary factors are controlled by feeding chow (< 0.02% cholesterol), cholesterol absorption efficiency in C57L mice with intact enterohepatic circulation of bile salt is significantly higher compared with AKR mice as measured by four independent methods, i.e., the plasma dual isotope ratio method, the fecal dual isotope ratio method, the lymphatic transport of cholesterol, and the mass balance method.<sup>3,4</sup> When these studies are repeated in mice with chronic biliary fistulae but in the setting of infusion of taurocholate and egg yolk lecithin, it is observed that the marked differences in cholesterol absorption efficiency still persist between AKR and C57L strains. Overall, these studies<sup>4</sup> suggest that the genetic factors at the enterocyte level are crucial in determining the variations of intestinal cholesterol absorption efficiency. The question arises therefore, as to which cellular step(s) in the absorption of cholesterol

might be inherently different. Furthermore, cholesterol absorption in (AKR/C57L)<sub>F<sub>1</sub></sub> mice mimics the higher-absorbing C57L parent, suggesting that high cholesterol absorption is a dominant trait in mice.

### Searching for intestinal cholesterol transporters

The molecular mechanism by which cholesterol from the intestinal lumen is transferred across the apical brush border membrane into the mucosal cell still remains poorly elucidated. The prevailing view for the molecular mechanism of cholesterol absorption has been that luminal unesterified cholesterol is shuttled across the unstirred water layer in a bile salt-containing micelle. After reaching the cell surface of the enterocyte, individual cholesterol molecules are incorporated into the brush border membrane and enter the cell interior by simple passive diffusion. This process of cholesterol transfer has no requirement for a carrier molecule to mediate cholesterol absorption. However, for water-insoluble lipids with very low monomer solubility (=10<sup>-8</sup> M) such as cholesterol, this mechanism is inefficient.

Although a simple passive diffusion process is widely assumed for intestinal cholesterol absorption, several lines of evidence support the existence of an energy-independent, protein-facilitated mechanism for cholesterol uptake by the enterocyte: (i) Transport of cholesterol from mixed micelles or small unilamellar vesicles to brush border membrane vesicles follows second order kinetics and is sensitive to protease,<sup>48,49</sup> (ii) structurally related plant sterols such as  $\beta$ -sitosterol and campesterol that differ from cholesterol only in the degree of saturation of the sterol nucleus or in the nature of the side chain at carbon-24 are less efficiently absorbed than cholesterol;<sup>50,51</sup> (iii) sitosterolemia, a rare autosomal recessive disorder is characterized by excess plant sterol absorption, suggesting that the ability to discriminate between plant sterols and cholesterol appears to have been lost in this disease;<sup>52,53</sup> (iv) intestinal cholesterol absorption can be specifically inhibited by cholesterol absorption inhibitors of different chemical structure: 2-azetidinones and sterol glycosides, each class showing profound structure-activity relationships.<sup>54,55</sup>

The *in vitro* study from Hauser and co-workers<sup>56</sup> suggests the scavenger receptor class B type I (SR-BI) mediates intestinal cholesterol absorption. Their immunoblotting data demonstrated that SR-BI protein is expressed in brush border membrane preparations and Caco-2 cells and preincubation with anti-SR-BI antibody partially inhibits cholesterol and cholesteryl ester uptake by brush border membrane vesicles and Caco-2 cells compared with no antibody control incubations. Their results suggest that SR-BI may be involved in the absorption of dietary lipids, and might be a putative sterol transporter. However, knockout of the *Sr-b1* gene in mice appears to have little effect on intestinal cholesterol absorption.<sup>57-59</sup>



Repa and colleagues<sup>60</sup> showed that intestinal ATP-binding cassette (ABC) transporter ABCA1 serves to efflux cholesterol from the enterocyte back into the intestinal lumen, thereby modulating net cholesterol absorption efficiency,<sup>61,62</sup> as well as cholesterol absorption and the transcriptional regulation of *Abca1* gene expression are mediated by the RXR/LXR heterodimer. More recently, it is found that ABCA1 protein expression is not located at the apical brush border membrane of the enterocyte, but on the basolateral membrane.<sup>63,64</sup> It is hypothesized therefore that transfer of cholesterol into lymph by ABCA1 may involve in HDL particles. Clearly, additional studies are required to pursue this tantalizing observation.

Mutations in the genes for these two proteins, ABCG5 and ABCG8,<sup>65,66</sup> result in sitosterolemia, a rare autosomal recessive disorder characterized by hyperabsorption of sitosterol and other plant sterols. Sitosterolemic individuals also absorb cholesterol more efficiently and are often hypercholesterolemic, implying a role of ABCG5 and ABCG8 in modulating the efficiency of cholesterol absorption. This suggests that cholesterol absorption is a selective process in which plant sterols and other non-cholesterol sterols are absorbed poorly or not at all. Recent research on the sterol efflux pumps ABCG5 and ABCG8 provides an explanation for this selectivity.<sup>67-72</sup> These studies yield new insights into the mechanisms that ABCG5 and ABCG8 promote partial efflux of cholesterol and nearly complete efflux of plant sterols from the enterocyte into the intestinal lumen, and may play a critical role in modulating the amount of cholesterol that reaches the lymph from the intestinal lumen. One question that remains is whether ABCG5/G8 alone is sufficient and required or whether other proteins are integral to this process.

Over the past decade, several groups have been searching for a cholesterol transporter that is located at the apical brush border membrane of the enterocyte.<sup>48,49,73-76</sup> More recently, Kramer and co-workers<sup>77</sup> found that by photoaffinity labeling using photoreactive derivatives of cholesterol and 2-azetidinone cholesterol absorption inhibitor, an 80-kDa and a 145-kDa integral membrane protein are identified as specific binding proteins for cholesterol and cholesterol absorption inhibitor, respectively, in the brush border membrane of the enterocyte. The 80-kDa cholesterol-binding protein does not interact with cholesterol absorption inhibitor and vice versa, as well as cholesterol or plant sterols do not interfere with the 145-kDa molecular target for cholesterol absorption inhibitor. Both proteins show an identical tissue distribution and are exclusively found at the anatomical sites of cholesterol absorption: duodenum, jejunum, and ileum. Furthermore, both proteins are different from the hitherto described candidate proteins for the intestinal cholesterol transporters such as SR-BI, ABCA1, ABCG5, or ABCG8. Their results strongly suggest that intestinal cholesterol absorption is not facilitated by a single transporter protein but occurs by a complex machinery. Furthermore, a novel se-

lective cholesterol absorption inhibitor ezetimibe decreases intestinal cholesterol absorption, but does not influence gene expression of intestinal *Sr-b1*, *Abca1*, *Abcg5*, and *Abcg8*, thus suggesting that this drug might curtail the activity of a putative sterol transporter at the brush border membrane of the enterocyte that actively facilitates the uptake of cholesterol.<sup>78</sup>

## Pharmacological control of intestinal cholesterol absorption

The use of cholesterol absorption inhibitors for treating hypercholesterolemia has a long history, with several classes of compounds having been developed. In this review, we will discuss bile salts, plant sterols, and ezetimibe, as well as their actions on intestinal cholesterol absorption because they have been shown to have marked effects on lowering plasma or biliary cholesterol levels in the human.

Bile salts. Ursodeoxycholic acid (UDCA) has been used to treat cholesterol gallstones for more than twenty years,<sup>79</sup> and decreasing intestinal cholesterol absorption is one of its major therapeutic actions.<sup>80-82</sup> Recently, Wang and colleagues<sup>9</sup> explored the influence of the hydrophilic-hydrophobic balance of a series of natural bile acids on cholesterol absorption in the mouse. Because bacterial and especially hepatic biotransformations of specific bile acids occur, hydrophobicity indices of the resultant bile salt pools differ from the fed bile acids. They<sup>9</sup> observed a significant and positive correlation between hydrophobicity indices of the bile salt pool and percent cholesterol absorption. The principal mechanism whereby hydrophilic bile acids inhibit cholesterol absorption appears to be via the uptake step by curtailing micellar cholesterol solubilization intraluminally.<sup>9,83-85</sup> Hence, decreasing the hydrophobicity index of the biliary bile salt pool reduces cholesterol's bioavailability for absorption by enterocytes. Furthermore, gene expression of the intestinal sterol efflux transporters *Abcg5* and *Abcg8* is up-regulated by feeding cholic acid, but not by the hydrophilic  $\beta$ -muri-cholic acid nor by the hydrophobic deoxycholic acid.<sup>9</sup> Their study<sup>9</sup> suggests that natural hydrophilic bile acids efficiently suppress cholesterol absorption, and may act as potent plasma and biliary cholesterol-lowering agents, even more so than UDCA, for prevention of cholesterol deposition diseases in humans. An example of the latter is a recent study<sup>86</sup> showing that  $\beta$ -muricholic acid efficiently prevents cholesterol gallstone formation in gallstone-susceptible C57L mice by chronically inhibiting intestinal cholesterol absorption.

**Plant sterols (Phytosterols):** Plant sterols have the same basic function in plants as cholesterol in animals; that is, they play a key role in cell membrane function. Over the past decade, the possibility of using plant sterols as ingredients in functional foods has led to numerous research studies<sup>87-89</sup> in relation to their ability to reduce

plasma cholesterol. The main conclusion is that the effective doses are between 1.5 and 3g/day, leading to a reduction between 8% and 15% in plasma LDL-cholesterol. Unlike cholesterol, plant sterols have a very low capacity for intestinal absorption,<sup>90</sup> which, together with their high rate of biliary secretion by the liver, leads to an extremely low level of plant sterols in the plasma. Cholesterol absorption from the dietary and the biliary sources is strongly reduced in the presence of plant sterols, and the unabsorbed cholesterol is excreted in the feces. The commonly accepted, basic mechanism of action of these compounds is that, in appropriate conditions, they can become efficiently incorporated into the micelles in the intestinal lumen, displace the cholesterol, and lead to its precipitation with other, non-solubilized plant sterols.<sup>91-94</sup> Furthermore, competition between cholesterol and plant sterols for incorporation into micelles, for transfer into the brush border membrane, as well as competition within the cell for ACAT, could explain the effect of large amounts of plant sterols to inhibit cholesterol absorption. This process reduces both the cholesterol and the triglycerides in the liver, which is compensated for by two different mechanisms: an increase in cholesterol synthesis, detected as an increase in its precursors lathosterol and desmosterol, and an increase in the LDL receptors. In contrast, cholesterol synthesis is strongly inhibited by the  $\Delta^{22}$ -sterols (stigmaterol), by the competitive inhibition of sterol  $\Delta^{24}$ -reductase, which is an interesting secondary mechanism for future research.

**Ezetimibe:** Ezetimibe (SCH 58235) and its analogs SCH 48461 and SCH 58053 are novel potent and selective inhibitors of cholesterol absorption,<sup>95,96</sup> and markedly lowering plasma LDL-cholesterol levels at a very low dose range in the human is similar to that at which statins are routinely given. A daily dose of 10 mg of ezetimibe alone effects an average reduction of about 18% in plasma LDL-cholesterol levels.<sup>97</sup>

Ezetimibe is glucuronidated in the enterocyte during its first pass. Both ezetimibe and its glucuronide are circulated enterohepatically,<sup>98</sup> repeatedly delivering the agent back to the site of action on the luminal surface of the enterocyte. This agent does not change the physicochemical nature of the intraluminal environment, nor does it increase the expression of proteins ABCG5/G8 that drive sterol efflux from the enterocyte. Furthermore, ezetimibe and its analogs are relatively small molecular structures that do not affect the enterohepatic flux of bile acids, but rather appear to act by disrupting the uptake of sterol across the microvillous membrane, i.e., inhibiting a putative sterol transporter that facilitates the movement of cholesterol into the intestinal cell. This mechanism remains to be the subject of intensive investigation. In addition, there is a marked compensatory increase in cholesterol synthesis in the liver, but not in the peripheral organs, and an accelerated loss of cholesterol in the feces with little or no change in the rate of conversion of cho-

lesterol to bile salts. Recent studies<sup>99-101</sup> show that the combination of ezetimibe with either atorvastatin or simvastatin offers a powerful new therapeutic approach to the control of LDL-cholesterol levels in the general population, as well as provides a complementary treatment strategy for patients with homozygous familial hypercholesterolemia, a high-risk population.

## Conclusions

The significant inter-individual differences found in the human and inter-strain variations observed in the inbred mouse strains strongly suggest that multiple genes are involved; however, the question as to which cellular step(s) in the absorption of cholesterol is inherently different remains unresolved. Cholesterol absorption is a selective process in that plant sterols are absorbed poorly or not at all, and recent research on the sterol efflux transporters ABCG5 and ABCG8 provides an explanation for this selectivity. Also, growing genetic and biochemical evidence suggests a sterol transporter that facilitates the movement of cholesterol into the intestinal cell, and the identification of such a gene is predicted to be elucidated in the near future, which should yield new insights into the mechanisms that potentially regulate the influx of cholesterol across the enterocyte. Understanding the molecular mechanisms whereby cholesterol is absorbed will provide a powerful novel strategy for the prevention and treatment of coronary heart disease and cholesterol gallstones.

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## References

1. The National Cholesterol Education Program Expert Panel. Executive Summary of the Third Report of the National Cholesterol Education Program (NCEP) Expert Panel on Detection, Evaluation, and Treatment of High Blood Cholesterol in Adults (Adult Treatment Panel III). *J Amer Med Assoc* 2001; 285: 2486-2497.
2. Apstein MD, Carey MC. Pathogenesis of cholesterol gallstones: a parsimonious hypothesis. *Eur J Clin Invest* 1996; 26: 343-352.
3. Wang DQ-H, Carey MC. Measurement of intestinal cholesterol absorption by plasma and fecal dual-isotope ratio, mass balance, and lymph fistula methods in the mouse: an analysis of direct versus indirect methodologies. *J Lipid Res* 2003; 44: 1042-1059.
4. Wang DQ-H, Paigen B, Carey MC. Genetic factors at the enterocyte level account for variations in intestinal cholesterol absorption efficiency among inbred strains of mice. *J Lipid Res* 2001; 42: 1820-1830.
5. Hofmann AF, Borgström B. Physico-chemical state of lipids in intestinal content during their digestion and absorption. *Gastroenterology* 1963; 21: 43-50.
6. Hofmann AF, Borgström B. The intraluminal phase of fat digestion in man: the lipid content of the micellar and oil phases of intestinal

- content obtained during fat digestion and absorption. *J Clin Invest* 1964; 43: 247-257.
7. Hofmann AF, Small DM. Detergent properties of bile salts: correlation with physiological function. *Annu Rev Med* 1967; 18: 333-376.
  8. Siperstein MD, Chaikoff IL, Reinhardt WO. C<sup>14</sup>-Cholesterol. V. Obligatory function of bile in intestinal absorption of cholesterol. *J Biol Chem* 1952; 198: 111-114.
  9. Wang DQ-H, Tazuma S, Cohen DE, Carey MC. Feeding natural hydrophilic bile acids inhibits intestinal cholesterol absorption: studies in the gallstone-susceptible mouse. *Am J Physiol* 2003; 285: G494-G502.
  10. Wang DQ-H, Lammert F, Cohen DE, Paigen B, Carey MC. Cholic acid aids absorption, biliary secretion, and phase transitions of cholesterol in murine cholelithogenesis. *Am J Physiol* 1999; 276: G751-G760.
  11. Wang DQ-H, Carey MC. Complete mapping of crystallization pathways during cholesterol precipitation from model bile: Influence of physical-chemical variables of pathophysiologic relevance and identification of a stable liquid crystalline state in cold, dilute and hydrophilic bile salt-containing systems. *J Lipid Res* 1996; 37: 606-630.
  12. Eckhardt ER, Wang DQ-H, Donovan JM, Carey MC. Dietary sphingomyelin suppresses intestinal cholesterol absorption by decreasing thermodynamic activity of cholesterol monomers. *Gastroenterology* 2002; 122: 948-956.
  13. Staggars JE, Hernell O, Stafford RJ, Carey MC. Physical-chemical behavior of dietary and biliary lipids during intestinal digestion and absorption. 1. Phase behavior and aggregation states of model lipid systems patterned after aqueous duodenal contents of healthy adult human beings. *Biochemistry* 1990; 29: 2028-2040.
  14. Hernell O, Staggars JE, Carey MC. Physical-chemical behavior of dietary and biliary lipids during intestinal digestion and absorption. 2. Phase analysis and aggregation states of luminal lipids during duodenal fat digestion in healthy adult human beings. *Biochemistry* 1990; 29: 2041-2056.
  15. Westergaard H, Dietschy JM. The mechanism whereby bile acid micelles increase the rate of fatty acid and cholesterol uptake into the intestinal mucosal cell. *J Clin Invest* 1976; 58: 97-108.
  16. Bennett Clark S, Tercyak AM. Reduced cholesterol transmembrane transport in rats with inhibited mucosal acyl CoA: cholesterol acyltransferase and normal pancreatic function. *J Lipid Res* 1984; 25: 148-159.
  17. Heider JG, Pickens CE, Kelly LA. Role of acyl CoA: cholesterol acyltransferase in cholesterol absorption and its inhibition by 57-118 in the rabbit. *J Lipid Res* 1983; 24: 1127-1134.
  18. Buhman KK, Accad M, Novak S, Choi RS, Wong JS, Hamilton RL, Turley S, Farese RV Jr. Resistance to diet-induced hypercholesterolemia and gallstone formation in ACAT2-deficient mice. *Nat Med* 2000; 6: 1341-1347.
  19. Nielsen LB, Stender S, Kjeldsen K. Effect of lovastatin on cholesterol absorption in cholesterol-fed rabbits. *Pharmacol Toxicol* 1993; 72: 148-151.
  20. Hajri T, Ferezou J, Laruelle C, Lutton C. Crivastatin, a new 3-hydroxy-3-methylglutaryl-coenzyme A reductase inhibitor, inhibits cholesterol absorption in genetically hypercholesterolemic rats. *Eur J Pharmacol* 1995; 286: 131-136.
  21. Vanhanen H, Kesäniemi YA, Miettinen TA. Pravastatin lowers serum cholesterol, cholesterol-precursor sterols, fecal sterols, and cholesterol absorption in man. *Metabolism* 1992; 41: 588-595.
  22. Young SG, Cham CM, Pitas RE, Burri BJ, Connolly A, Flynn L, Pappu AS, Wong JS, Hamilton RL, Farese RV Jr. 1995. A genetic model for absent chylomicron formation: mice producing apolipoprotein B in the liver, but not in the intestine. *J Clin Invest* 1995; 96: 2932-2946.
  23. van Greevenbroek MM, Robertus-Tuinissen MG, Erkelens DW, de Bruin TW. Participation of the microsomal triglyceride transfer protein in lipoprotein assembly in Caco-2 cells: interaction with saturated and unsaturated dietary fatty acids. *J Lipid Res* 1998; 39: 173-185.
  24. Gordon DA, Jamil H, Gregg RE, Olofsson SO, Boren J. Inhibition of the microsomal triglyceride transfer protein blocks the first step of apolipoprotein B lipoprotein assembly but not the addition of bulk core lipids in the second step. *J Biol Chem* 1996; 271: 33047-33053.
  25. Ordoas JM, Schaefer EJ. Genetic determinants of plasma lipid response to dietary intervention: the role of the APOA1/C3/A4 gene cluster and the APOE gene. *Br J Nutr* 2000; 83: S127-S136.
  26. Voshol PJ, Havinga R, Wolters H, Ottenhoff R, Princen HM, Oude Elferink RP, Groen AK, Kuipers F. Reduced plasma cholesterol and increased fecal sterol loss in multidrug resistance gene 2 P-glycoprotein-deficient mice. *Gastroenterology* 1998; 114: 1024-1034.
  27. Wang DQ-H, Lammert F, Cohen DE, Paigen B, Carey MC. Hyposecretion of biliary phospholipids significantly decreases the intestinal absorption of cholesterol in *Mdr2*(-/-) and (+/-) mice. *Gastroenterology* 1998; 114: A913.
  28. Schwarz M, Russell DW, Dietschy JM, Turley SD. Alternate pathways of bile acid synthesis in the cholesterol 7  $\alpha$ -hydroxylase knockout mouse are not upregulated by either cholesterol or cholestyramine feeding. *J Lipid Res* 2001; 42: 1594-1603.
  29. Wang DQ-H, Lammert F, Paigen B, Carey MC. Phenotypic characterization of *Lith* genes that determine susceptibility to cholesterol cholelithiasis in inbred mice. Pathophysiology of biliary lipid secretion. *J Lipid Res* 1999; 40: 2066-2079.
  30. Ponz de Leon M, Iori R, Barbolini G, Pompei G, Zaniol P, Carulli N. Influence of small-bowel transit time on dietary cholesterol absorption in human beings. *N Engl J Med* 1982; 307: 102-103.
  31. Traber MG, Ostwald R. Cholesterol absorption and steroid excretion in cholesterol-fed guinea pigs. *J Lipid Res* 1978; 19: 448-456.
  32. Wang DQ-H, Schmitz F, Kopin AS, Carey MC. Targeted disruption of the murine cholecystokinin-1 receptor promotes intestinal cholesterol absorption and susceptibility to cholesterol cholelithiasis. *J Clin Invest* 2003; (submitted).
  33. Wang DQ-H. Aging per se is an independent risk factor for cholesterol gallstone formation in gallstone susceptible mice. *J Lipid Res* 2002; 43: 1950-1959.
  34. Duan L-P, Wang DQ-H. Role of the jejunal and ileal ATP-binding cassette (ABC) transporters A1, G5 and G8 (ABCA1/G5/G8) in intestinal cholesterol absorption: age and gender effects. *Hepatology* 2002; 36: 306A.
  35. Jolley CD, Dietschy JM, Turley SD. Genetic differences in cholesterol absorption in 129/Sv and C57BL/6 mice: effect on cholesterol responsiveness. *Am J Physiol* 1999; 276: G1117-G1124.
  36. Bhattacharyya AK, Eggen DA. Cholesterol absorption and turnover in rhesus monkey as measured by two methods. *J Lipid Res* 1980; 21: 518-524.
  37. Lofland HB Jr, Clarkson TB, St. Clair RW, Lehner NDM. Studies on the regulation of plasma cholesterol levels in squirrel monkeys of two genotypes. *J Lipid Res* 1972; 13: 39-47.
  38. Kesäniemi YA, Miettinen TA. Cholesterol absorption efficiency regulates plasma cholesterol level in the Finnish population. *Eur J Clin Invest* 1987; 17: 391-395.
  39. Sehayek E, Nath C, Heinemann T, McGee M, Seidman CE, Samuel P, Breslow JL. U-shape relationship between change in dietary cholesterol absorption and plasma lipoprotein responsiveness and evidence for extreme interindividual variation in dietary cholesterol absorption in humans. *J Lipid Res* 1998; 39: 2415-2422.
  40. McNamara DJ, Kolb R, Parker TS, Batwin H, Samuel P, Brown CD, Ahrens EH Jr. Heterogeneity of cholesterol homeostasis in man. Response to changes in dietary fat quality and cholesterol quantity. *J Clin Invest* 1987; 79: 1729-1739.
  41. Bosner MS, Lange LG, Stenson WF, Ostlund RE Jr. Percent cholesterol absorption in normal women and men quantified with dual stable isotopic tracers and negative ion mass spectrometry. *J Lipid Res* 1999; 40: 302-308.
  42. Beynen AC, Meijer GW, Lemmens AG, Glatz JFC, Versluis A, Katan MB, van Zutphen LFM. Sterol balance and cholesterol absorption in inbred strains of rabbits hypo- or hyperresponsive to dietary cholesterol. *Atherosclerosis* 1989; 77: 151-157.
  43. Van Zutphen LFM, Fox RR. Strain differences in response to dietary cholesterol by JAX rabbits: correlation with esterase patterns. *Atherosclerosis* 1977; 28: 435-446.
  44. Van Zutphen LFM, Den Bieman MGCW. Cholesterol response in inbred strains of rats, *Rattus norvegicus*. *J Nutr* 1981; 111: 1833-1838.
  45. Kirk EA, Moe GL, Caldwell MT, Lernmark JA, Wilson DL, LeBoeuf RC. Hyper- and hypo-responsiveness to dietary fat and cholesterol



- among inbred mice: searching for level and variability genes. *J Lipid Res* 1995; 36: 1522-1532.
46. Carter CP, Howles PN, Hui DY. Genetic variation in cholesterol absorption efficiency among inbred strains of mice. *J Nutr* 1997; 127: 1344-1348.
  47. Schwarz M, Davis DL, Vick BR, Russell DW. Genetic analysis of intestinal cholesterol absorption in inbred mice. *J Lipid Res* 2001; 42: 1801-1811.
  48. Thurnhofer H, Hauser H. Uptake of cholesterol by small intestinal brush border membrane is protein-mediated. *Biochemistry* 1990; 29: 2142-2148.
  49. Compassi S, Werder M, Boffelli D, Weber FE, Hauser H, Schulthess G. Cholesteryl ester absorption by small intestinal brush border membrane is protein-mediated. *Biochemistry* 1995; 34: 16473-16482.
  50. Moreau RA, Whitaker BD, Hicks KB. Phytosterols, phytostanols, and their conjugates in foods: structural diversity, quantitative analysis, and health-promoting uses. *Prog Lipid Res* 2002; 41: 457-500.
  51. Piironen V, Lindsay DG, Miettinen TA, Toivo J, Lampi AM. Plant sterols: biosynthesis, biological function and their importance to human nutrition. *J Sci Food Agric* 2000; 80: 939-966.
  52. Salen G, Shore V, Tint GS, Forte T, Shefer S, Horak I, Horak E, Dayal B, Nguyen L, Batta AK, et al. Increased sitosterol absorption, decreased removal, and expanded body pools compensate for reduced cholesterol synthesis in sitosterolemia with xanthomatosis. *J Lipid Res* 1989; 30: 1319-1330.
  53. Salen G, Tint GS, Shefer S, Shore V, Nguyen L. Increased sitosterol absorption is offset by rapid elimination to prevent accumulation in heterozygotes with sitosterolemia. *Arterioscler Thromb* 1992; 12: 563-568.
  54. Harwood HJ Jr, Chandler CE, Pellarin LD, Bangerter FW, Wilkins RW, Long CA, Cosgrove PG, Malinow MR, Marzetta CA, Pettini JL, et al. Pharmacologic consequences of cholesterol absorption inhibition: alteration in cholesterol metabolism and reduction in plasma cholesterol concentration induced by the synthetic saponin  $\beta$ -tigogenin cellobioside (CP-88818; tiqueside). *J Lipid Res* 1993; 34: 377-395.
  55. Burnett DA, Caplen MA, Davis HR Jr, Burrier RE, Clader JW. 2-Azetidinones as inhibitors of cholesterol absorption. *J Med Chem* 1994; 37: 1733-1736.
  56. Hauser H, Dyer JH, Nandy A, Vega MA, Werder M, Bieliauskaite E, Weber FE, Compassi S, Gemperli A, Boffelli D, Wehrli E, Schulthess G, Phillips MC. Identification of a receptor mediating absorption of dietary cholesterol in the intestine. *Biochemistry* 1998; 37: 17843-17850.
  57. Mardones P, Quinones V, Amigo L, Moreno M, Miquel JF, Schwarz M, Miettinen HE, Trigatti B, Krieger M, VanPatten S, Cohen DE, Rigotti A. 2001. Hepatic cholesterol and bile acid metabolism and intestinal cholesterol absorption in scavenger receptor class B type I-deficient mice. *J Lipid Res* 2001; 42: 170-180.
  58. Altmann SW, Davis HR Jr, Yao X, Laverty M, Compton DS, Zhu LJ, Crona JH, Caplen MA, Hoos LM, Tetzloff G, Priestley T, Burnett DA, Strader CD, Graziano MP. The identification of intestinal scavenger receptor class B, type I (SR-BI) by expression cloning and its role in cholesterol absorption. *Biochim Biophys Acta* 2002; 1580: 77-93.
  59. Wang DQ-H, Carey MC. Susceptibility to murine cholesterol gallstone formation is not affected by partial disruption of the HDL receptor SR-BI. *Biochim Biophys Acta* 2002; 1583: 141-150.
  60. Repa JJ, Turley SD, Lobaccaro JA, Medina J, Li L, Lustig K, Shan B, Heyman RA, Dietschy JM, Mangelsdorf DJ. 2000. Regulation of absorption and ABC1-mediated efflux of cholesterol by RXR heterodimers. *Science* 2000; 289: 1524-1529.
  61. McNeish J, Aiello RJ, Guyot D, Turi T, Gabel C, Aldinger C, Hoppe KL, Roach ML, Royer LJ, De Wet J, Brocardo C, Chimini G, Francone OL. High density lipoprotein deficiency and foam cell accumulation in mice with targeted disruption of ATP-binding cassette transporter-1. *Proc Natl Acad Sci USA*. 2000; 97: 4245-4250.
  62. Drobnik W, Lindenthal B, Lieser B, Ritter M, Christiansen Weber T, Liebisch G, Giesa U, Igel M, Borsukova H, Buchler C, Fung-Leung WP, Von Bergmann K, Schmitz G. ATP-binding cassette transporter A1 (ABCA1) affects total body sterol metabolism. *Gastroenterology* 2001; 120: 1203-1211.
  63. Mulligan JD, Flowers MT, Tebon A, Bitgood JJ, Wellington C, Hayden MR, Attie AD. ABCA1 is essential for efficient basolateral cholesterol efflux during the absorption of dietary cholesterol in chickens. *J Biol Chem* 2003; 278: 13356-13366.
  64. Attie AD, Hamon Y, Brooks-Wilson AR, Gray-Keller MP, MacDonald ML, Rigot V, Tebon A, Zhang LH, Mulligan JD, Singaraja RR, Bitgood JJ, Cook ME, Kastelein JJ, Chimini G, Hayden MR. Identification and functional analysis of a naturally occurring E89K mutation in the ABCA1 gene of the WHAM chicken. *J Lipid Res* 2002; 43: 1610-1617.
  65. Berge KE, Tian H, Graf GA, Yu L, Grishin NV, Schultz J, Kwiterovich P, Shan B, Barnes R, Hobbs HH. Accumulation of dietary cholesterol in sitosterolemia caused by mutations in adjacent ABC transporters. *Science* 2000; 290: 1771-1775.
  66. Lee MH, Lu K, Hazard S, Yu H, Shulenin S, Hidaka H, Kojima H, Allikmets R, Sakuma N, Pegoraro R, Srivastava AK, Salen G, Dean M, Patel SB. Identification of a gene, ABCG5, important in the regulation of dietary cholesterol absorption. *Nat Genet* 2001; 27: 79-83.
  67. Yu L, Hammer RE, Li-Hawkins J, Von Bergmann K, Lutjohann D, Cohen JC, Hobbs HH. Disruption of Abcg5 and Abcg8 in mice reveals their crucial role in biliary cholesterol secretion. *Proc Natl Acad Sci USA*. 2002; 99: 16237-16242.
  68. Yu L, Li-Hawkins J, Hammer RE, Berge KE, Horton JD, Cohen JC, Hobbs HH. Overexpression of ABCG5 and ABCG8 promotes biliary cholesterol secretion and reduces fractional absorption of dietary cholesterol. *J Clin Invest* 2002; 110: 671-680.
  69. Yu L, York J, von Bergmann K, Lutjohann D, Cohen JC, Hobbs HH. Stimulation of cholesterol excretion by the liver X receptor agonist requires ATP-binding cassette transporters G5 and G8. *J Biol Chem* 2003; 278: 15565-15570.
  70. Graf GA, Li WP, Gerard RD, Gelissen I, White A, Cohen JC, Hobbs HH. Coexpression of ATP-binding cassette proteins ABCG5 and ABCG8 permits their transport to the apical surface. *J Clin Invest* 2002; 110: 659-669.
  71. Duan L-P, Wang DQ-H. Sterols influence intestinal cholesterol absorption through mediating expression of the ileal ATP-binding cassette transporters G5 and G8 (ABCG5/G8). *Gastroenterology* 2002; 122: A403.
  72. Morales VM, Wang DQ-H. Expression of intestinal ATP-binding cassette transporters G5 and G8 (ABCG5/G8) plays a major role in determining variations in cholesterol absorption efficiency in inbred mice. *Gastroenterology* 2002; 122: A58.
  73. Sparrow CP, Patel S, Baffic J, Chao YS, Hernandez M, Lam MH, Montenegro J, Wright SD, Detmers PA. A fluorescent cholesterol analog traces cholesterol absorption in hamsters and is esterified *in vivo* and *in vitro*. *J Lipid Res* 1999; 40: 1747-1757.
  74. Kramer W, H. Glombik, S. Petry, H. Heuer, H. Schafer, W. Wendler, D. Corsiero, F. Girbig, C. Weyland. 2000. Identification of binding proteins for cholesterol absorption inhibitors as components of the intestinal cholesterol transporter. *FEBS Lett* 2000; 487: 293-297.
  75. Hernandez M, Montenegro J, Steiner M, Kim D, Sparrow C, Detmers PA, Wright SD, Chao YS. 2000. Intestinal absorption of cholesterol is mediated by a saturable, inhibitable transporter. *Biochim Biophys Acta* 2000; 1486: 232-242.
  76. Detmers PA, Patel S, Hernandez M, Montenegro J, Lisnock JM, Pikounis B, Steiner M, Kim D, Sparrow C, Chao YS, Wright SD. A target for cholesterol absorption inhibitors in the enterocyte brush border membrane. *Biochim Biophys Acta* 2000; 1486: 243-252.
  77. Kramer W, Girbig F, Corsiero D, Burger K, Fahrenholz F, Jung C, Muller G. Intestinal cholesterol absorption: identification of different binding proteins for cholesterol and cholesterol absorption inhibitors in the enterocyte brush border membrane. *Biochim Biophys Acta* 2003; 1633: 13-26.
  78. Repa JJ, Dietschy JM, Turley SD. Inhibition of cholesterol absorption by SCH 58,053 in the mouse is not mediated via changes in the expression of mRNA for ABCA1, ABCG5, or ABCG8 in the enterocyte. *J Lipid Res* 2002; 43: 1864-1874.
  79. Tokyo Cooperative Gallstone Study Group. Efficacy and indications of ursodeoxycholic acid treatment for dissolving gallstones. A multicenter double-blind trial. *Gastroenterology* 1980; 78: 542-548.
  80. Ponz de Leon M, Carulli N, Loria P, Iori R, Zironi F. Cholesterol absorption during bile acid feeding. Effect of ursodeoxycholic acid (UDCA) administration. *Gastroenterology* 1980; 78: 214-219.



81. Lanzini A, Northfield TC. Effect of ursodeoxycholic acid on biliary lipid coupling and on cholesterol absorption during fasting and eating in subjects with cholesterol gallstones. *Gastroenterology* 1988; 95: 408-416.
82. Hardison WGM, Grundy SM. Effect of ursodeoxycholate and its taurine conjugate on bile acid synthesis and cholesterol absorption. *Gastroenterology* 1984; 87: 130-135.
83. Watt SM, Simmonds WJ. Effects of four taurine-conjugated bile acids on mucosal uptake and lymphatic absorption of cholesterol in the rat. *J Lipid Res* 1984; 25: 448-55.
84. Leiss O, von Bergmann K, Streicher U, Strotkoetter H. Effect of three different dihydroxy bile acids on intestinal cholesterol absorption in normal volunteers. *Gastroenterology* 1984; 87: 144-149.
85. Uchida K, Akiyoshi T, Igimi H, Takase H, Nomura Y, Ishihara S. Differential effects of ursodeoxycholic acid and ursocholic acid on the formation of biliary cholesterol crystals in mice. *Lipids* 1991; 26: 526-530.
86. Wang DQ-H, Tazuma S. Effect of  $\beta$ -muricholic acid on the prevention and dissolution of cholesterol gallstones in C57L/J mice. *J Lipid Res* 2002; 43: 1960-1968.
87. Miettinen TA, Puska P, Gylling H, Vanhanen H, Vartiainen E. Reduction of serum cholesterol with sitostanol-ester margarine in a mildly hypercholesterolemic population. *N Engl J Med* 1995; 333: 1308-1312.
88. Ostlund RE Jr, Racette SB, Okeke A, Stenson WF. Phytosterols that are naturally present in commercial corn oil significantly reduce cholesterol absorption in humans. *Am J Clin Nutr* 2002; 75: 1000-1004.
89. Maki KC, Davidson MH, Umporowicz DM, Schaefer EJ, Dicklin MR, Ingram KA, Chen S, McNamara JR, Gebhart BW, Ribaya-Mercado JD, Perrone G, Robins SJ, Franke WC. Lipid responses to plant-sterol-enriched reduced-fat spreads incorporated into a National Cholesterol Education Program Step I diet. *Am J Clin Nutr* 2001; 74: 33-43.
90. Heinemann T, Axtmann G, von Bergmann K. Comparison of intestinal absorption of cholesterol with different plant sterols in man. *Eur J Clin Invest* 1993; 23: 827-831.
91. Nissinen M, Gylling H, Vuoristo M, Miettinen TA. Micellar distribution of cholesterol and phytosterols after duodenal plant stanol ester infusion. *Am J Physiol* 2002; 282: G1009-G1015.
92. Ikeda I, Tanabe Y, Sugano M. Effects of sitosterol and sitostanol on micellar solubility of cholesterol. *J Nutr Sci Vitaminol* 1989; 35: 361-369.
93. Ikeda I, Tanaka K, Sugano M, Vahouny GV, Gallo LL. Discrimination between cholesterol and sitosterol for absorption in rats. *J Lipid Res* 1988; 29: 1583-1591.
94. Ikeda I, Tanaka K, Sugano M, Vahouny GV, Gallo LL. Inhibition of cholesterol absorption in rats by plant sterols. *J Lipid Res* 1988; 29: 1573-1582.
95. Rosenblum SB, Huynh T, Afonso A, Davis HR Jr, Yumibe N, Clader JW, Burnett DA. Discovery of 1-(4-fluorophenyl)-(3R)-[3-(4-fluorophenyl)-(3S)-hydroxypropyl]-(4S)-(4-hydroxyphenyl)-2-azetidinone (SCH 58235): a designed, potent, orally active inhibitor of cholesterol absorption. *J Med Chem* 1998; 41: 973-980.
96. Van Heek M, France CF, Compton DS, McLeod RL, Yumibe NP, Alton KB, Sybertz EJ, Davis HR Jr. *In vivo* metabolism-based discovery of a potent cholesterol absorption inhibitor, SCH58235, in the rat and rhesus monkey through the identification of the active metabolites of SCH48461. *J Pharmacol Exp Ther* 1997; 283: 157-163.
97. Bays HE, Moore PB, Dreihobl MA, Rosenblatt S, Toth PD, Dujovne CA, Knopp RH, Lipka LJ, Lebeaut AP, Yang B, Mellars LE, Cuffie-Jackson C, Veltri EP; Ezetimibe Study Group. Effectiveness and tolerability of ezetimibe in patients with primary hypercholesterolemia: pooled analysis of two phase II studies. *Clin Ther* 2001; 23: 1209-1230.
98. van Heek M, Farley C, Compton DS, Hoos L, Alton KB, Sybertz EJ, Davis HR Jr. Comparison of the activity and disposition of the novel cholesterol absorption inhibitor, SCH58235, and its glucuronide, SCH60663. *Br J Pharmacol* 2000; 129: 1748-1754.
99. Dujovne CA, Ettinger MP, McNeer JF, Lipka LJ, LeBeaut AP, Suresh R, Yang B, Veltri EP. Ezetimibe Study Group. Efficacy and safety of a potent new selective cholesterol absorption inhibitor, ezetimibe, in patients with primary hypercholesterolemia. *Am J Cardiol* 2002; 90: 1092-1097.
100. Sudhop T, Lutjohann D, Kodal A, Igel M, Tribble DL, Shah S, Perevozskaya I, von Bergmann K. Inhibition of intestinal cholesterol absorption by ezetimibe in humans. *Circulation* 2002; 106: 1943-1948.
101. Gagne C, Gaudet D, Bruckert E; Ezetimibe Study Group. Efficacy and safety of ezetimibe coadministered with atorvastatin or simvastatin in patients with homozygous familial hypercholesterolemia. *Circulation* 2002; 105: 2469-2475.