

only in gastrointestinal symptoms, but also in associated extra-intestinal diseases.^{4,5}

Despite suffering from CD, our patient showed no HLA-DQ2 or DQ8 alleles. This phenomenon has been described in 6% of the European celiac population.⁹ In the last few years, GWAS has been performed on large numbers of CD patients, relatives, and match controls have revealed evidence of additional non-HLA loci of CD susceptibility, most of which are related to T-cell regulation and inflammation.^{9,10} In the light of this finding, and because CD affects 2% of the European and US populations at some stage in life, it would be advisable to investigate the presence of lymphocytic enteritis in patients with SM and UP, recommending a GFD in the case of a positive result, especially given the beneficial effects observed with no side effects. Such recommendations could markedly improve patient quality of life and increase the effectiveness of currently employed treatments.

Conflict of interest

The authors have no conflict of interests to declare.

Ethical disclosures

Protection of human and animal subjects. The authors declare that the procedures followed were in accordance with the regulations of the responsible Clinical Research Ethics Committee and in accordance with those of the World Medical Association and the Helsinki Declaration.

Confidentiality of data. The authors declare that they have followed the protocols of their work centre on the publication of patient data and that all the patients included in the study have received sufficient information and have given their informed consent in writing to participate in that study.

Right to privacy and informed consent. The authors declare that no patient data appears in this article.

References

1. Hahn HP, Hornick JL. Immunoreactivity for CD25 in gastrointestinal mucosal mast cells is specific for systemic mastocytosis. *Am J Surg Pathol.* 2007;31:1669–76.

2. Valent P, Sperr WR, Schwartz LB, Horny H-P. Diagnosis and classification of mast cell proliferative disorders: delineation from immunologic diseases and non-mast cell hematopoietic neoplasms. *J Allergy Clin Immunol.* 2004;114:3–11.
3. Mansoor DK, Sharma HP. Clinical presentations of food allergy. *Pediatr Clin North Am.* 2011;58:315–26.
4. Esteve M, Rosinach M, Fernández-Bañares F, Farré C, Salas A, Alsina M, et al. Spectrum of gluten-sensitive enteropathy in first-degree relatives of patients with celiac disease: clinical relevance of lymphocytic enteritis. *Gut.* 2006;55:1739–45.
5. Kurppa K, Collin P, Viljamaa M, Haimila K, Saavalainen P, Partanen J, et al. Diagnosing mild enteropathy celiac disease: a randomized, controlled clinical study. *Gastroenterology.* 2009;136:816–23.
6. Sánchez-García S, Ibáñez MD, Martínez-Gómez MJ, Escudero C, Vereda A, Fernández-Rodríguez M, et al. Eosinophilic esophagitis, celiac disease and immunoglobulin E-mediated allergy in a 2-year-old child. *J Investig Allergol Clin Immunol.* 2011;21:73–5.
7. Pedrosa Delgado M, Martín Muñoz F, Polanco Allué I, Martín Esteban M. Cold urticaria and celiac disease. *J Investig Allergol Clin Immunol.* 2008;18:123–5.
8. Peroni DG, Paiola G, Tenero L, Fornaro M, Bodini A, Pollini F, et al. Chronic urticaria and celiac disease: a case report. *Pediatr Dermatol.* 2010;27:108–9.
9. Karell K, Louka AS, Moodie SJ, Ascher H, Clot F, Greco L, et al. European Genetics Cluster on Celiac Disease. HLA types in celiac disease patients not carrying the DQA1*05-DQB1*02 (DQ2) heterodimer: results from the European Genetics Cluster on Celiac Disease. *Hum Immunol.* 2003;64:469–77.
10. Romanos J, van Diemen CC, Nolte IM, Trynka G, Zhernakova A, Fu J, et al. Analysis of HLA and non-HLA alleles can identify individuals at high risk for celiac disease. *Gastroenterology.* 2009;137:834–40.

L. Rodrigo^a, I. Pérez-Martínez^a, A.J. Lucendo^{b,*}

^a *Department of Gastroenterology, Hospital Universitario Central de Asturias, Oviedo, Spain*

^b *Department of Gastroenterology, Hospital General de Tomelloso, Tomelloso, Ciudad Real, Spain*

* Corresponding author.

E-mail address: alucendo@vodafone.es (A.J. Lucendo).

<http://dx.doi.org/10.1016/j.aller.2012.01.005>

Occupational rhinitis caused by rice flour in a pizzeria worker[☆]

To the Editor,

Rice, the seed of monocot plant *Oryza sativa*, is nowadays a basic element in almost every culinary tradition throughout

the world, mainly in Asia, Africa and South America, where it represents as much as 20% of the total calories ingested per day. The production of this cereal grain has become comparable to that of wheat and corn, due to several factors such as price, edibility and nutritional value. Rice belongs to the *Poaceae* family; therefore cross-reactivity is suitable between this grain and other constituents of the former family.

Despite its widespread consumption, case reports in the literature regarding rice allergy, in any of its forms, remain surprisingly scarce. Hitherto, two different routes have been argued which could lead to immediate rice hypersensitivity reactions: boiled rice ingestion and

[☆] Note: This case report was partially displayed as oral communication in Reunión de Clausura XX Sesiones Interhospitalarias Sociedad Madrid-Castilla La Mancha de Alergología e Inmunología Clínica, Sigüenza (Guadalajara, Spain). June 3rd 2011.

inhalation of both boiled rice vapours and/or raw rice powder.¹

Moreover, three groups of allergens have been involved in rice allergy, a 33 kDa *Glyoxalase I* showing a unique structure consisting of two tandemly repeated homologous domains with some well-conserved sequences, similar to *Glyoxalase I* plants²; two allergens which belong to the *amylase inhibitor* family of 14 and 16 kDa, respectively; and *Lipid Transfer Proteins* (LTP), reported in an elegant work published in 2006 by Asero et al. in which they could demonstrate cross-reactivity between rice-LTP and apple/peach LTP in three cases of rice-induced anaphylaxis subsequent to ingestion of the former.

It remains elusive as to whether rice-LTP allergic potential dilutes subsequent to boiling process or by contrast this protein could form a macromolecular matrix during cooking procedures which could hamper its binding to specific IgEs, so digestion would facilitate hidden epitopes to be recognised by their specific antibodies.³ Likewise, Enrique et al. described three cases of patients with respiratory allergy to raw rice by inhalation who tolerated boiled rice ingestion.⁴

Kim et al. reported work-related rhinitis and conjunctivitis symptoms caused by occupational exposure to rice powder in the grain industry. They performed an IgE ELISA inhibition test which demonstrated cross-reactivity between rice and several grass pollen extracts.⁵ Besides, *Ory s 1*, has elicited cross-reactivity with rye and Bermuda grass pollens in patients suffering from pollen allergy and shares significant sequence identity with the major pollen allergen of rye grass (*Lol p 1*; 65.5% identity) and Bermuda grass (*Cyn d 1*; 62.9% identity).⁶

A 36-year-old woman who had been working for two years in a pizzeria kneading pizza dough reported immediate rhinoconjunctivitis while manipulating pizza dough during the past six months. She attributed typical rhinoconjunctival symptoms due to a special flour ("oak semolina") poured in the location where pizza dough was blended. Hygienic conditions at work place were appropriate and she did not have history of allergic rhinoconjunctivitis. Furthermore, she did not suffer from these symptoms on non-working days and holidays. In a successive visit, the patient indicated subsequent continuous sneezing and rhinorrhoea from April to June outside her work-place which limited her daily activities and sleep. The pizzeria where she works commonly uses rice powder, which is a flour variety known as *oak semolina* in Spain, frequently placed to enhance blending of the pizza dough and to ease its mixture. Firstly we performed skin prick testing against common aeroallergens [*D. pteronyssinus* and *Dermatophagoides pteronyssinus*, *Lepidoglyphus destructor*, animal dander (dog and cat), storage mites (*Tyrophagus putrescentiae*, *Euroglyphus maynei*, *Glycyphagus domesticus*, *Acarus siro*) *Alternaria alternata*, *Aspergillus fumigatus*, *Quercus* sp., *Populus* sp., *Willow* sp., *Elm* sp., Birch tree, Ash tree, Holm oak, *Acacia*, *Olea europaea*, *Platanus acerifolia*, *Plantago lanceolata*, grass mix (*Dactylis*, *Trisetum*, *Lolium*, *Phleum* and *Poa*), *Parietaria judaica*, *Artemisia vulgaris*, *Taraxacum officinalis*, *Cupressus arizonica*], flour battery (wheat, rye, corn, soy, oat, rice and barley), common grains (wheat, corn, oat, barley, rye) and prick-by-prick against oak semolina (rice powder flour) as well as wheat flour. Specific IgE (*CAP Phadia*, Sweden) was determined against *D. pteronyssinus*, *D.*

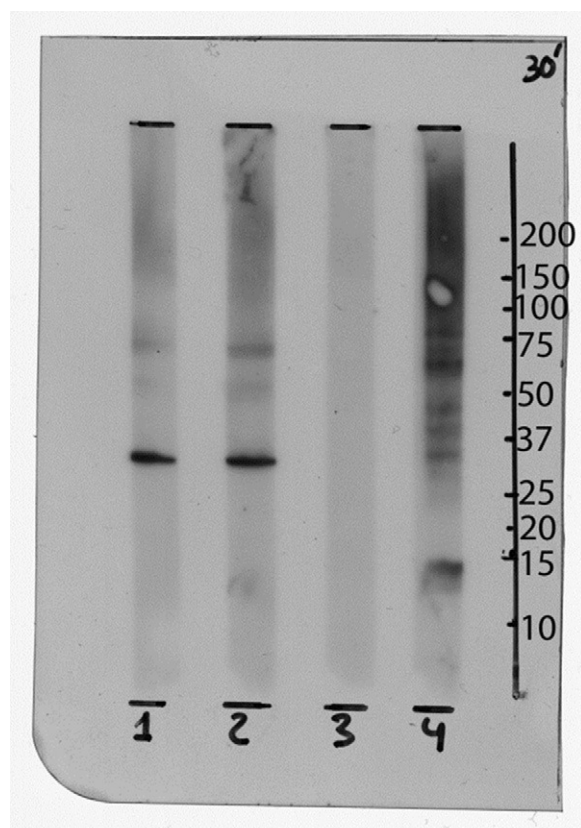
farinae, *L. destructor*, rPhl p 1-5b, rPhl p 7-12, rPrup3, and rice flour; Besides, sIgE against *Tri a 14* and wheat flour- α -amylases (Cm 3 and Cm 16) was determined by ADVIA-Centaur (Siemens). In order to discard rye flour cross-sensitisation to rice flour we performed a 2D-immunoblot essay. Microbiology department randomly selected seven flour samples and explored them by means of a high power field microscope, in order to rule out possible contaminants (i.e. storage mites and moulds). A fibre optic rhinoscopy was carried out in order to discard nasal polyps or structural abnormalities, both before and after nasal provocation test (NPT). An Anterior rhinomanometry was performed before and after NPT.

Skin prick test resulted positive against mites (*D. pteronyssinus*, *D. farinae*, *L. destructor*), storage mites (*T. putrescentiae*, *E. maynei*, *G. domesticus*, *A. siro*), pollens (*Populus* sp., *Willow* sp., *Elm* sp., Birch tree, Ash tree, Holm oak, *Acacia*, *O. europaea*, *P. acerifolia*, *Dactylis*, *Trisetum*, *Lolium*, *Phleum* and *Poa*) also rye and rice flour and negative against moulds, animal dander, alpha-amylase, enolase, papain and grains. Skin prick-by-prick resulted positive against rice powder flour and negative against wheat flour. Specific IgEs (sIgE) resulted as follows: *D. pteronyssinus*: 6.75 kU/L; *D. farinae*: 4.85 kU/L; *L. destructor*: 54.60 kU/L; rPhlp1-5b: 14.30 kU/L; rPhl7-12: 0.03 kU/L; rice flour: 0.88 kU/L; rPrup3: 0.89 kU/L. sIgE against *Tri a 14* and wheat flour- α -amylases (Cm 3 and Cm 16) resulted both negative (0.0 kU/L). Immunoblot essay did not reveal potential cross-reactive allergens among rye and rice flour (Fig. 1). Immunoblot essay performed with rye flour and rice flour did not demonstrate cross-sensitivity among the previous flours.

She did not complain about experiencing symptoms after eating boiled rice or the ingestion of prepared food.

As the recently published EAACI position paper on occupational rhinitis states,⁷ we performed a nasal provocation test in order to elucidate the culprit agent as the SEAIC Rhinoconjunctivitis Committee recently stated.^{8,9} We placed the patient in a 4 m³ room resembling her work-place where she manipulated trays which contained *oak semolina*. It is of note that the patient was completely asymptomatic and the challenge test was carried out outside the pollen season. The patient was asked to pour the content included in one tray into the other during 30 min. After 5 min of handling rice flour, our patient suffered from intense sneezing, rhinorrhoea and itchy eyes. Fifteen minutes later an anterior rhinomanometry was fulfilled which measured severe obstruction of both nostrils, differing from the one carried out before the specific nasal provocation test. Nasal smear taken right after the specific challenge with rice powder presented 30% of eosinophils. As control, a nasal provocation test with wheat flour resulted negative, following the same procedure previously described. The patient showed normal spirometry volumes and negative bronchodilator challenge. Therefore, the final diagnosis was occupational rhinitis caused by rice flour in a patient suffering from persistent moderate rhinitis.

We report an unusual case of occupational rhinitis due to rice flour in a pizzeria worker. Prick testing against rice flour, specific nasal challenge and subsequent assessment by means of endoscopy and anterior rhinomanometry as well as cessation of symptoms following strict avoidance of "oak semolina" reinforce the noteworthy diagnosis.



- 1- Rye flour-Negative Control
- 2- Rye flour-Serum
- 3- Rice flour- Negative Control
- 4- Rice flour-Serum

Figure 1 Rye and rice flour Immunoblotting.

Cross-reactivity between *Poaceae* and rice, together with LTP potential implication, could constitute the reasons of rice sensitisation in this patient, according to former research about this topic. Therefore it would be suitable to assess LTP sensitisation in grain sensitised patients. Rye sensitisation observed in our patient could be due to alpha-amylase inhibitors which are considered main rice allergens, shared among both grains. For that reason for performing immunoblotting essays which discarded a potential rye flour implication and due to the fact that sIgE against alpha-amylase inhibitors were negative, we can conclude that both alpha-amylase and rye allergens were not involved in our patient's symptoms. Outwardly *Glyoxalase I*, another rice allergen might not be implicated in this case as there is no wheat or barley sensitisation. It is of note that while wheat could be suspected as an important allergen as it is one of the ingredients of the pizza dough, rice flour is the only kind of raw flour which is poured and directly handled in this pizzeria, as the pizza dough is pre-cooked and instrumentally mashed, avoiding direct hand contact. Moreover, both skin prick testing against wheat flour and sIgE against Tri a 14, a major wheat allergen, resulted negative, therefore discarding wheat flour as the culprit allergen in this case.

As has recently been described,⁹ nasal provocation testing with standardised allergens can generally be started at an initial concentration of 1:1000 and then increased by a factor of 10. In the case of less well-known and occupational allergens, endpoint titration should be performed to identify the initial dose. Regarding occupational allergens, the irritant concentration limit for each substance must also be considered. Nonetheless, we performed a use provocation test with rice flour in a room resembling that in which the patient works to simulate real workplace conditions, handling trays containing the factory's own rice flour. It remains elusive whether rice-LTP allergic potential dilutes subsequent to boiling process or by contrast this protein could form a macromolecular matrix during cooking procedures which could hamper its binding to specific IgEs, so digestion would facilitate hidden antigenic determinants to be recognised by their specific antibodies.

Despite its widespread consumption, case reports in the literature regarding rice allergy, in any of its forms, remain sparse. Hitherto, two different routes have been argued which could lead to immediate rice hypersensitivity reactions: boiled rice ingestion and inhalation of both boiled rice vapours and/or raw rice powder. We believe this to be an important case because of the fact that rice flour was a hidden allergen in this patient and also due to the circumstance that occupational rhinitis could lead to suffering from occupational asthma resulting in socio-economic disruption. Further assessment and investigation regarding rice hypersensitivity are required in order to straighten out the former hypothesis.

Contributors

DAA, JBE, MSPB, MRR, EMV and MAM were involved in data analysis and writing manuscript. DAA was involved in patient follow-up and pharmacologic advisory. JAGR carried out anterior rhinomanometry and nasal endoscopy. LJN performed skilled technical assistance and fulfilled component resolved diagnosis on the ADVIA platform.

Conflict of interest

The authors have no conflicts of interest to declare.

Funding source

This work was partially supported by a grant from Comunidad de Madrid S2010/BMD-2502 MITIC.

Ethical disclosures

Protection of human and animal subjects. The authors declare that no experiments were performed on humans or animals for this investigation.

Confidentiality of data. The authors declare that they have followed the protocols of their work centre on the publication of patient data and that all the patients included in the study have received sufficient information and have given their informed consent in writing to participate in that study.

Right to privacy and informed consent. The authors declare that no patient data appears in this article.

References

1. Fiocchi A, Bouygue GR, Restani P, Gaiaschi A, Terracciano L, Martelli A. Anaphylaxis to rice by inhalation. *J Allergy Clin Immunol.* 2003;111:193–5.
2. Nakase M, Usui Y, Alvarez-Nakase AM, Adachi T, Urisu A, Nakamura R, et al. Cereal allergens: rice-seed allergens with structural similarity to wheat and barley allergens. *Allergy.* 1998;53 Suppl.:55–7.
3. Asero R, Amato S, Alfieri B, Folloni S, Mistrello G. Rice: another potential cause of food allergy in patients sensitized to lipid transfer protein. *Int Arch Allergy Immunol.* 2007;143:69–74 [epub 2006 Dec 28].
4. Enrique E, Ahrazem O, Bartra J, Latorre MD, Castelló JV, de Mateo JA, et al. Lipid transfer protein is involved in rhinoconjunctivitis and asthma produced by rice inhalation. *J Allergy Clin Immunol.* 2005;116:926–8 [epub 2005 Aug 1].
5. Kim JH, Kim JE, Choi GS, Hwang EK, An S, Ye YM, et al. A case of occupational rhinitis caused by rice powder in the grain industry. *Allergy Asthma Immunol Res.* 2010;2:141–3 [epub 2010 Mar 24].
6. Xu H, Theerakulpisut P, Goulding N, Suphioglu C, Singh MB, Bhalla PL. Cloning, expression and immunological characterization of Ory s 1, the major allergen of rice pollen. *Gene.* 1995;164: 255–9.
7. Moscato G, Vandenplas O, Van Wijk RG, Malo JL, Perfetti L, Quirce S, et al. EAACI position paper on occupational rhinitis. *Respir Res.* 2009;10:16.
8. Siracusa A, Desrosiers M, Marabini A. Epidemiology of occupational rhinitis: prevalence, aetiology and determinants. *Clin Exp Allergy.* 2000;30:1519–34.
9. Dordal MT, Lluch-Bernal M, Sánchez MC, Rondón C, Navarro A, Montoro J, et al. Allergen-specific nasal provocation testing: review by the Rhinoconjunctivitis Committee of the Spanish Society of Allergy and Clinical Immunology. *J Investig Allergol Clin Immunol.* 2011;21:1–12.

D. Antolin-Amerigo^{a,*}, M. Rodríguez-Rodríguez^a, J. Barbarroja Escudero^a, M.S. Pérez Bustamante^a, L. Jimeno Nogales^c, J.A. Guerrero Ríos^b, E. Mohedano-Vicente^a, M. Alvarez-Mon^a

^a Immune System Diseases-Allergy Service, Hospital Universitario Príncipe de Asturias. Department of Medicine. Universidad de Alcalá. Madrid, Spain

^b ENT Department, Hospital Universitario Alcalá de Henares, Madrid, Spain

^c ALK-Abelló. Madrid, Spain

*Corresponding author.

E-mail addresses: dario.antolin@gmail.com (D. Antolin-Amerigo), mercedesrr@gmail.com (M. Rodríguez-Rodríguez), jose.barbarroja@uah.es (J. Barbarroja Escudero), socorrobustamante@gmail.com (M.S. Pérez Bustamante), ester.mohedano@hotmail.com (E. Mohedano-Vicente), mademons@gmail.com (M. Alvarez-Mon).

<http://dx.doi.org/10.1016/j.aller.2012.01.006>

Omalizumab under 12 years old: Real-life practice

To the Editor,

Despite the major research investment in finding new efficacious and safe therapeutic options for severe uncontrolled asthmatic patients, omalizumab has been the only globally accepted recent innovative drug for allergic asthma. This recombinant DNA-derived humanised monoclonal antibody binds to IgE, ultimately decreasing allergic inflammation. Such therapeutic intervention, approved for use in Europe since 2005, had been focussed mainly on adults, notwithstanding the importance of allergy and the high prevalence of paediatric asthma. In fact, asthma is one of the most common chronic diseases in children, associated with considerable morbidity, particularly if the asthma is severe. It is the leading cause of hospital admissions in children with chronic disease, compromising both children and caregivers' quality of life.¹ These data provided a strong rationale for an investigation plan concerning omalizumab use in paediatrics, which led, in 2009, to the extension of this drug's approval to children aged 6–11 years old.² For this, clinical trials regarding children were important, showing omalizumab to be both efficacious (stressing a reduced asthma exacerbation rate) and safe in this age group.^{3,4} Further randomised, double blind, placebo-controlled clinical studies have recently been published, highlighting omalizumab's

effect on paediatric asthma control improvement and also addressing its safety.^{5–7} The next step is now to present and discuss real-life clinical data, which is the purpose of this study, concerning the recently fulfilled 52-weeks follow-up of our first omalizumab-treated patient under the age of 12. This anti-IgE therapy's effect on other allergic diseases besides asthma is also considered.

An 8-year-old boy with severe persistent uncontrolled allergic asthma since early infancy (first severe episode at the age of eight months old) was considered for omalizumab treatment. The child has concomitant moderate-severe persistent allergic rhinitis, mild-moderate atopic eczema and IgE-mediated food allergy (egg allergy until 7 years old and current peanut allergy with anaphylaxis), as well as asthma maternal history. Prior to the anti-IgE therapy, despite following an optimised asthma therapeutic plan including daily inhaled combination of long-acting β_2 -agonist (LABA) and high-dose inhaled corticosteroid (ICS), the child maintained nocturnal and diurnal wheezing, with impaired daily leisure and school activities. Compliance to therapy was closely assured by regular interview and physician-assessed correct inhalation technique as well as evaluation of medication use and prescriptions requirements. However, rescue medication had to be used often (near-daily inhaled salbutamol; oral corticosteroids at least twice a month) and the child had 12 hospital admissions due to asthma exacerbations. Systemic corticosteroids have been used for the shortest time needed, in order to avoid their unbearable side effects.