



Allergologia et immunopathologia

www.elsevier.es/ai



RESEARCH LETTERS

Hypersensitivity reactions following measles-mumps-rubella vaccine and dextran-specific IgG response

To the Editor,

In the context of a mass vaccination campaign, rare adverse events can be detected through passive surveillance.¹ The Brazilian Post-Vaccine Adverse Event Surveillance System received notifications of allergic reactions at a rate of 0.95 cases/100,000 doses of the Measles-Mumps-Rubella vaccine (MMR) produced in Italy (Morupar[®] Chiron), in the period from 2000 to 2003. However on August 21st 2004 (the first day of the National Measles Follow-up Campaign), an increase in allergic reactions was reported with a rate of 11.56 cases/100,000 doses administered of this vaccine. As a result, and as a preventive measure, the use of this product was discontinued in Brazil. (Ministério da Saúde. Brasília. – Nota Técnica N°97/04).

Studies have shown an association between anaphylaxis following administration of the MMR vaccine and subsequent detection of anti-gelatin IgE antibodies.^{2,3} According to the manufacturer's certification, the gelatin was not present and the Morupar[®] vaccine had residual traces of egg, hydrolysed casein and dextran 70.

Dextran, a high-molecular-weight polysaccharide used as a stabiliser in some vaccines, is rarely associated with hypersensitivity reactions. These reactions result from circulating immune complexes formed by preexisting anti-dextran IgG antibodies and dextran injected with the vaccine, causing complement activation and mast cell and basophil degranulation by anaphylatoxins.⁴

We evaluated 19 children (aged 34.9 ± 16.3 months) who reported reactions within two hours after receiving the vaccine, in Curitiba, state of Paraná. Thirty-one age/gender matched children, living in the same area, who had received on the same date and who had no adverse reactions were included as a control group. Blood samples were collected approximately five to six weeks after vaccine administration, while skin tests were performed four to seven months after vaccination.

Serum specific IgE antibodies (casein, egg) were determined by a fluoroenzyme immunoassay method (ImmunoCAP-Pharmacia[®]) and levels greater than 0.35 KU/L were considered positive.

Levels of specific IgE and IgG to dextran 70 were determined by a time-resolved fluorescent lanthanide immunoassay (DELFA, PerkinElmer, Boston, MA, US). The

dissociation-enhanced method can be used to study antibody binding to solid-phase proteins or peptides. Dextran 70 from *Leuconostoc ssp* (Molecular weight ~70 kDa, Fluka, Sigma Aldrich, Milan, Italy) was used at a concentration of 400 µg/ml to coat the Delfia plates. Serum samples were diluted 1:50 in bovine serum albumin 1% and incubated overnight at 4–8 °C. Bound antibodies were detected by a europium-labelled anti-human IgG or IgE antiserum (Perkin-Elmer). Optical density values higher than the mean plus two standard deviations of europium counts in the control group were considered positive.

For allergy skin tests we used undiluted vaccine from the same batch as that employed in the Vaccination Campaign.

The study was approved by the Federal University of Parana's Institutional Review Board of Hospital de Clinicas, and a voluntary informed-consent was signed by children's guardians.

No prior allergic reactions to vaccines were reported in either group, and there was no association with history of atopy or allergic reactions to medication and/or food.

Of the children evaluated, all developed skin manifestations (erythema, urticaria or angioedema), associated or not with other systems. All of them received oral antihistamines, two received oral corticosteroids, and subcutaneous adrenaline complemented the therapy.

Demographic and laboratory data of subjects and controls are shown in Table 1.

Casein-specific IgE was not detected in all the subjects. Two controls showed levels > 0.35 KU/L of specific IgE to egg.

Sera from the case and control groups were tested for specific IgE and IgG to dextran 70, which was the most suspected causal component in the Italian case series⁵. All the children had specific IgE to dextran 70 below the cut-off value (167,336; mean 93,092, SD 37,122). The cut-off for specific IgG was fixed at 1,433,119, which is higher than the mean plus 2 SDs of europium counts in the control group (mean, 422,899; SD, 505,109 × 2). Sixteen out of 19 cases (84%) and one out of 31 controls (3%) presented specific IgG levels above the cut-off value (Table 1).

Positive skin prick test with the Morupar vaccine was observed in five out of 18 cases and in none of the 22 controls tested. These five patients also showed high levels of specific IgG to dextran 70, with negativity of specific IgE.

The determination of specific IgE to casein and egg allowed us to exclude that these proteins were part of the cause of reactions to the Morupar vaccine. According to the results of dextran-specific antibodies, immediate allergic reactions reported in Brazil could be possibly induced by dextran 70 like those reported in Italy.⁵

Table 1 Demographic data, serologic and prick test results of cases and control subjects

| Cases | Age (months) | Gender | Specific IgE(kU/L) | | Dextran 70 IgG | Dextran 70 IgE | Prick test |
|----------|--------------|--------|--------------------|-----------|------------------|----------------|------------|
| | | | Casein | Egg white | | | |
| 1 | 40 | F | <0.35 | <0.35 | 2,046,917 | 124,299 | Positive |
| 2 | 23 | F | <0.35 | <0.35 | 1,760,366 | 94,160 | Negative |
| 3 | 43 | M | <0.35 | <0.35 | 15,943 | 33,425 | Negative |
| 4 | 34 | F | <0.35 | <0.35 | 1,858,396 | 110,011 | Negative |
| 5 | 14 | F | <0.35 | <0.35 | 1,193,181 | 98,350 | Negative |
| 6 | 55 | F | <0.35 | <0.35 | 1,969,370 | 139,781 | Negative |
| 7 | 57 | M | <0.35 | <0.35 | 2,177,109 | 113,300 | Negative |
| 8 | 54 | F | <0.35 | <0.35 | 1,564,207 | 82,105 | Negative |
| 9 | 37 | F | <0.35 | <0.35 | 1,773,746 | 144,329 | Positive |
| 10 | 41 | M | <0.35 | <0.35 | 1,798,760 | 109,966 | Positive |
| 11 | 16 | M | <0.35 | <0.35 | 2,019,567 | 119,839 | Negative |
| 12 | 32 | M | <0.35 | <0.35 | 1,501,322 | 148,892 | Negative |
| 13 | 55 | F | <0.35 | <0.35 | 1,857,291 | 129,189 | Positive |
| 14 | 13 | F | <0.35 | <0.35 | 1,923,830 | 128,222 | Negative |
| 15 | 52 | F | <0.35 | <0.35 | 2,047,325 | 106,648 | Positive |
| 16 | 15 | M | <0.35 | <0.35 | 1,785,574 | 99,988 | Negative |
| 17 | 20 | F | <0.35 | <0.35 | 1,717,535 | 99,940 | Negative |
| 18 | 14 | F | <0.35 | <0.35 | 1,894,459 | 103,162 | Negative |
| 19 | 49 | M | <0.35 | <0.35 | 592,071 | 38,380 | ND |
| Controls | | | | | | | |
| 1 | 24 | F | <0.35 | <0.35 | 818,652 | 68,916 | Negative |
| 2 | 24 | F | <0.35 | <0.35 | 1,159,819 | 104,267 | Negative |
| 3 | 58 | F | <0.35 | <0.35 | 51,912 | 67,927 | Negative |
| 4 | 25 | F | <0.35 | <0.35 | 79,455 | 76,974 | Negative |
| 5 | 27 | F | <0.35 | <0.35 | 622,043 | 128,462 | Negative |
| 6 | 57 | M | <0.35 | <0.64 | 298,816 | 17,480 | Negative |
| 7 | 50 | M | <0.35 | <0.35 | 936,896 | 68,822 | ND |
| 8 | 52 | M | <0.35 | <0.35 | 27,610 | 82,077 | Negative |
| 9 | 36 | F | <0.35 | <0.35 | 110,355 | 31,535 | Negative |
| 10 | 15 | M | <0.35 | <0.35 | 60,946 | 50,172 | ND |
| 11 | 49 | F | <0.35 | <0.35 | 16,803 | 15,281 | Negative |
| 12 | 23 | F | <0.35 | <0.35 | 777,216 | 79,284 | ND |
| 13 | 29 | F | <0.35 | <0.35 | 318,863 | 143,238 | ND |
| 14 | 24 | F | <0.35 | <0.35 | 17,682 | 65,217 | Negative |
| 15 | 15 | M | <0.35 | <0.35 | 45,897 | 90,983 | Negative |
| 16 | 13 | F | <0.35 | <0.35 | 45,887 | 99,016 | Negative |
| 17 | 44 | F | <0.35 | <0.35 | 1,991,596 | 154,322 | ND |
| 18 | 55 | F | <0.35 | <0.35 | 229,436 | 130,985 | ND |
| 19 | 32 | M | <0.35 | <0.35 | 50,786 | 70,602 | Negative |
| 20 | 49 | F | <0.35 | <0.35 | 1,068,770 | 69,044 | Negative |
| 21 | 54 | M | <0.35 | <0.35 | 1,095,377 | 112,356 | Negative |
| 22 | 55 | M | <0.35 | <0.35 | 117,545 | 120,092 | Negative |
| 23 | 32 | F | <0.35 | <0.35 | 135,736 | 139,116 | Negative |
| 24 | 58 | F | <0.35 | <0.35 | 154,005 | 140,009 | ND |
| 25 | 26 | M | <0.35 | <0.35 | 128,790 | 83,075 | Negative |
| 26 | 23 | F | <0.35 | <0.35 | 72,542 | 114,813 | Negative |
| 27 | 13 | M | <0.35 | <0.35 | 116,883 | 119,235 | Negative |
| 28 | 40 | M | <0.35 | <0.35 | 1,303,248 | 103,162 | Negative |
| 29 | 28 | M | <0.35 | <0.35 | 943,154 | 76,433 | ND |
| 30 | 24 | F | <0.35 | <0.35 | 162,615 | 124,530 | Negative |
| 31 | 47 | M | <0.35 | <0.35 | 150,347 | 144,336 | ND |

Dextran 70 IgG: fixed cutoff value, 1,433,119 Europium counts, Dextran 70 IgE: fixed cutoff value, 167,336 Europium counts (positive results are written in bold). Gender: male (M), female (F); prick test: not determined (ND).

The positive allergy skin tests to MMR vaccine observed in five cases could be due to a double pathogenetic mechanism in these children, such as in certain drug allergies⁶, involving specific IgG to dextran and IgE to a different component of the vaccine, although a non-specific reaction due to a direct degranulation of mast cells cannot be ruled out.

We conclude that residual dextran 70 in this particular brand of MMR vaccine, which induced high levels of specific IgG antibodies, could be the culprit of most of the hypersensitivity reactions reported in Brazil.

Research supported by

Programa Nacional de Imunizações, Ministério da Saúde, Brasília/Distrito Federal, Brazil

Secretaria de Estado da Saúde do Paraná, Curitiba/Paraná, Brazil

The Green Channel Reference Centre for Pre-vaccination Consultancy, University Hospital, Verona, Italy.

Conflict of interest

The authors contributed equally to this work and they have no conflict of interest.

Acknowledgments

We wish to thank the helpful discussion of Drs. Robert T. Chen, Karin R. Luhn and Fides Sbardelotto. We acknowledge Elizabeth Ferraz, Daniel R. C. de Freitas, Gisele C. B. Araújo, Kátia Terêncio and Gisele Madalosso for recruitment and serum collection.

doi:10.1016/j.aller.2010.02.006

References

1. Cunha SC, Dourado I. MMR mass vaccination campaigns, vaccine-related adverse events, and the limits of the decision making process, in Brazil. *Health Policy*. 2004;67:323–8.
2. Pool V, Braun M, Kelso JM, Mootrey G, Chen R, Yunginger JW, et al. Prevalence of anti-gelatin IgE antibodies in people with anaphylaxis after measles-mumps-rubella vaccine in the United States. *Pediatrics*. 2002;110:1–9.
3. Sakagushi M, Nakayama T, Fujita H, Toda M, Inouye S. Minimum estimated incidence in Japan of anaphylaxis to live virus vaccine including gelatin. *Vaccine*. 2001;19:431–6.
4. Ponvert C, Schenmann P. Vaccine allergy and pseudo-allergy. *Eur J Dermatol*. 2003;13:1–10.
5. Zanoni G, Puccetti A, Dolcino M, Simone R, Peretti A, Ferro A et al. Dextran-specific IgG response in hypersensitivity reactions to measles-mumps-rubella vaccine. *J Allergy Clin Immunol*. 2008; 122:1233–1235. Epub 2008 Oct 15.
6. Weiss ME, Nyhan D, Peng ZK, Horrow JC, Lowenstein E, Hirshman C, et al. Association of protamine IgE and IgG antibodies with life-threatening reactions to intravenous protamine. *N Engl J Med*. 1989;320 (14):1684–5.

I.M. Novadzki^{a,*}, N. Rosário^a, G. Zanoni^b, E. Chiesa^b, A. Puccetti^{c,d}, R. Simone^{d,e}

^a*Divisão de Alergia e Imunologia, Departamento de Pediatria, Universidade Federal do Paraná, Curitiba, PR, Brazil*

^b*Section of Immunology, Department of Pathology, University of Verona, Verona, Italy*

^c*Immunology Unit, Institute G. Gaslini, Genoa, Italy*

^d*Section of Histology, Department of Experimental Medicine, University of Genoa, Genoa, Italy*

^e*Section of Internal Medicine, Department of Clinical and Experimental Medicine, University of Verona, Verona, Italy*

*Corresponding author.

E-mail address: iolanda@ufpr.br (I.M. Novadzki).

X-linked agammaglobulinaemia. Mutation A1246G (R372G)

To the Editor:

X-linked agammaglobulinaemia (XLA) is a primary immune deficiency characterised by a reduction in circulating B lymphocytes, hypogammaglobulinaemia and recurrent infections. The disease is caused by a mutation of Bruton's tyrosine kinase (Btk). Few studies in large groups of patients have been published; a large cohort of XLA patients in Eastern and Central European countries was recently published, in which the genetic and demographic features of XLA were studied; and clinical, immunological and genetic information was collected for 122 patients from 109 families.¹¹

We present a case of XLA which we believe to be of interest in view of its classical clinical presentation, diagnostic confirmation based on molecular biological techniques, and positive familial history.

The patient was a three-year-old Caucasian male from Ecuador, who had arrived in Spain in June 2003. On occasion of the first routine primary care visit, the paediatrician noted retarded progression of body weight and height – 10.4 kg (percentile 3) and 89 cm (percentile 25) – without significant findings in the physical examination. History of disease: Starting at one year of age the patient developed two episodes of pneumonia (one requiring hospital admission), recurrent bronchitis, multiple upper airway conditions and one episode of gastroenteritis. Family history: three siblings had died as a result of respiratory infections – the first two at four and two years of age, respectively (corresponding to pneumonia and sepsis), and the third at four months of age, due to severe bronchitis. The mother explained that the first two deceased children were from