

Allergologia et immunopathologia

www.elsevier.es/ai



REVIEW

Diagnosing immune-mediated reactions to drugs

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KEYWORDS

Adverse drug
reactions;
Diagnosis;
Drug allergy;
Hypersensitivity drug
reactions;
Basophil activation test;
Lymphoproliferative
tests;
Skin test

Abstract

Drug allergy is a type B adverse drug reaction, which is unpredictable and difficult to prevent or manage. In patients who have a previous history of drug allergy it must be confirmed by laboratorial diagnosis. However, the diagnostic test remains a major problem in clinical practice. Skin testing is validated for some drugs, such as penicillin, but not for others. Provocation test is a confirmatory test but bears the risk of severe reactions. Lymphocyte transformation test is a reliable test but is considered as a research tool. This review addresses the most recent published literature regarding the techniques which have already been developed as well as the new tests that can be promising alternatives for diagnosis of drug allergy.

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Introduction

The percentage of the population who reports a history of penicillin allergy is estimated to be 0.7-10%; however, only 20-30% of the patients have IgE-mediated allergy¹. Adverse drug reactions (ADR) are considered to be an important public health problem and can be life-threatening. ADR is classified into two main types: type A reactions, which are dose dependent and predictable. These kinds of reactions constitute 70-80% of adverse drug reactions; and type B, which are unpredictable adverse drug reactions, include idiosyncrasy, drug intolerance, or drug allergy, and may comprise approximately 10-15% of all ADR².³. Clinical manifestations of drug allergy include anaphylaxis, bronchospasm, dermatitis, fever, granulocytopenia, haemolytic anaemia, hepatitis, lupus erythematosus-like syndrome, nephritis, thrombocytopenia, and vasculitis⁴. Symptoms and signs without

demonstration of being an immunological process are classified as non-immune hypersensitivity drug reactions, and they are generally related to nonspecific histamine release, to nonspecific complement activation, to bradykinin accumulation or to induction of leukotriene synthesis in type I hypersensitivity reactions⁵. Non-steroidal anti-inflammatory drugs and antibiotics are most often implicated in drug allergies, besides anaesthetic drugs, latex, insulin, and immunomodulators^{3,6-7} (Table I).

Drug allergy reactions are generally classified according to Coomb's classification⁶. Type I include IgE-mediated reactions, such as urticaria, anaphylaxis, and asthma; type II comprises IgM and IgG-cytotoxic mechanisms (e.g. blood cell dyscrasias); type III are related to soluble IgG and IgM immune complexes (e.g. vasculitis, nephritis). Type IV reactions include distinct manifestations (e.g. maculopapular exanthema, bullous exanthema, acute, generalized exanthemous

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Table I Clinical manifestations of drug allergy				
Agents		Clinical manifestations		
Aspirin, non-steroidal anti-inflammatory drugs		Asthma		
Penicillin		Urticaria, asthma, neutropenia, haemolytic anaemia		
Antibiotics, muscle relaxants, natural rubber latex, hynotics		Perioperative anaphylaxis		
Anticonvulsivants, antibiotics, hydralazine, procainamide		Serum sickness, drug fever, vasculitis, lymphadenopathy		
Thalidomide		Maculopapular rashes, Steven Johnson syndrome, toxic epidermal necrolysis		
Infliximab, etanercept		Anti-dsDNA antibodies, reversible systemic erythematosus lupus		
Interferon alfa		Urticaria, angioedema, anti-thyroid antibodies		
Rituximab, alemtuzumab		Dyspnoea, hypotension, urticaria		
Sulphonamides, phenytoin		Stevens Johnson syndrome		
Vancomycin		Bullous skin disease		
Data adapted from 3,6-7. Vervloet and Durham, 1998; Hepner et al., 2003; Greenberg, 2006.				

pustulosis) according to the kind of cytokines secreted by T cells. In this type of reaction, a subclassification has been suggested according to the distinct effector cells responsible for the lesions8. The type I hypersensitivity reactions are called immediate reactions when they occur within one hour after the drug intake and the common manifestations are systemic anaphylaxis, urticaria, angioedema; they are called accelerated when they occur between 1 to 72 hours after receiving the drug, and the most frequent manifestations are urticaria and maculopapular rashes. Types II to IV hypersensitivity reactions occur in general after 72 hours of the drug intake9. The most frequent clinical manifestations caused by betalactamic drugs, which are the commonest implicating agents of drug hypersensitivity, are anaphylaxis (immediate reaction); maculopapular exanthema (late reaction) and isolated urticaria (occurs at any time) 10. In a follow-up study done in two hospitals in Fortaleza, Brazil, it has been verified that from 130 paediatric patients exposed to oxacillin, a drug included in the list of essential medicines, 20.8% presented drug adverse reactions, in a mean time of 14 days after the drug exposure¹¹. Half of the patients had fever and 35.7% showed cutaneous rashes.

A pharmacovigilance evaluation can be done for monitoring a drug adverse reaction. A case report in pharmacovigilance can be defined as "a notification relating to a patient with an adverse medical event (or laboratory test abnormality) suspected to be induced by a medicine" 12. In general, case reports describe suspected adverse drug reactions 12. Various approaches have been developed for determining the likelihood of a causal relationship between drug exposure and adverse events, based on four main considerations, which are: the association in time (or place) between drug administration and event, pharmacology (including current knowledge of nature and frequency of adverse reactions), medical or pharmacological plausibility (signs and symptoms, laboratory tests, pathological findings, mechanism) (Table II).

The Uppsala Monitoring Centre (the UMC)/WHO Collaborating Centre for International Drug Monitoring ¹² has proposed a way for assessing causality categories, which have the advantages of being internationally agreed and easy to use (Table III).

Diagnosis

In the evaluation of a possible adverse drug reaction, a careful investigation of the medical history as well as the laboratorial parameters is extremely important ^{13,14}.

Clinical history investigation

A well-kept history is essential to help in the investigation of adverse drug reaction and should include: (1) timing of the onset, course and duration of symptoms; (2) description of clinical and evolutive features; (3) temporal relationship of symptoms with medication use; (4) a detailed list of all drugs taken during the onset of the reactions; (5) personal and familiar history of adverse drug reactions; and (6) co-associated diseases. Some laboratory parameters, such as full blood count; platelet count; blood sedimentation rate; serum creatinine; bilirubin; alkaline phosphatise; transaminases; biopsy, among others, and specific confirmatory tests (skin and patch tests, in vitro assays for specific IgE, also serology to check for some infectious diseases, such as cytomegalovirosis, infectious mononucleosis, parvovirosis, hepatitis B and C^{13,14}.

Diagnostic tests

Skin testing

The diagnostic value of skin tests varies depending on the drug involved in the reaction. They should be done 4 to

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Table II	Table II Reporting information for associating causal relationship between drug exposure and adverse events (UMC-WHO, 2000)		
n	Topic	Information	
1	The patient	Age, gender and brief medical history	
2	Adverse event	Description (nature, localisation, severity, characteristics), laboratorial parameters, start date, course and outcome	
3	Suspected drug(s)	Name (brand or ingredient name + manufacturer), dose, route, start/stop dates, indication for use (with particular drugs, e.g. vaccines, a batch number is important)	
4	All other drugs used (including self-medication):	Names, doses, routes, start/stop dates	
5	Risk factors	(e.g. impaired renal function, previous exposure to suspected drug, previous allergies, social drug use)	
6	Name and address of reporter	(to be considered confidential and to be used only for data verification, completion and case follow-up)	

Causality term	Assessment criteria*
Certain	 Event of laboratory test abnormality, with plausible time relationship to drug intake Cannot be explained by disease or other drugs Response to withdrawal plausible (pharmacologically, pathologically) Event definitive pharmacollogically or phenomenologically (i.e. an objective and specific medical idsorder or a recognised pharmacological phenomenon) Rechallenge satisfactory, if necessary
Probable/likely	 Event of laboratory test abnormality, with reasonable time relationship to drug intake Unlikely to be attributed to disease or ohter drugs Response to withdrawal clinically reasonable Rechallenge not required
Possible	 Event of laboratory test abnormality, with reasonable time relationship to drug intake Could also be explained by disease or other drugs Information on drug withdrawal may be lacking or unclear
Unlikey	 Event of laboratory test abnormality, with a time to drug intake that makes a relationship improbable (but not impossible) Disease or ohter drugs provide plausible explanations
Conditional/unclassified	 Event of laboratory test abnormality More data for proper assessment neeeded, or Additional data under examination
Unassessable/unclassifiable	 Report suggesting an adverse reaction Cannot be judged because information is insufficient or contradictory Data cannot be supplemented or verified

6 weeks after the reaction is resolved⁵. Their sensitivity and predictive value are considered to be very good for penicillin, myorelaxant, heterologous sera, enzymes; satisfactory for vaccines, hormones, opiates; and poor or unknown for local anaesthetics, sulphonamides, iodine radiocontrast media, non-steroidal anti-inflammatory drugs, cephalosporin⁵. Skin testing for penicillin is the only test regularly used for diagnosis of drug allergy¹⁵. The majority of patients with negative results for major and minor determinants of penicillin will tolerate the drug without risk of a severe immediate reaction¹⁶.

Penicillin is one of the most useful antimicrobial drugs⁹. A percentage of 20% of patients admitted to a hospital refer to have allergy to the drug; however, most of these reports are unreliable¹⁷ and make the physicians prescribe alternative antibiotics, which in general are more expensive, can be associated with toxic effects, and can lead to antibiotic resistance¹. Allergic reactions to penicillin are estimated to occur in 2% of the patients treated with the drug and most of them are maculopapular or urticarial rashes⁹.

Penicillin test should be performed in every patient with a background history of penicillin allergy who needs a therapy

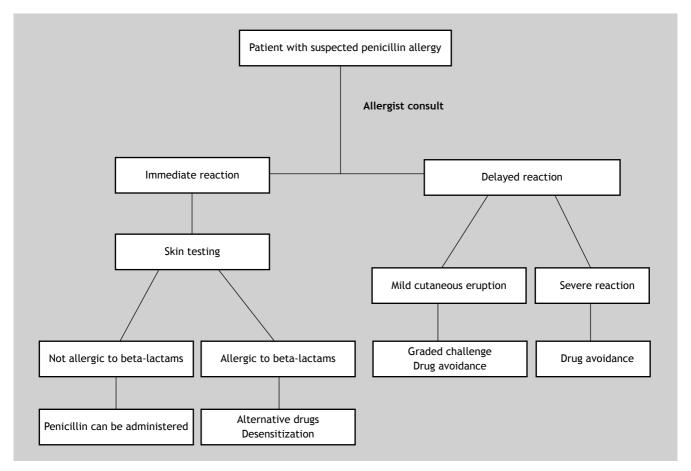


Figure 1. Representation of a clinical practice algorithm. Adapted from Forrest et al, 2001; Gruchalla and Pirmohamed, 2006.

with antibiotics¹⁸. The risk of systemic reactions to penicillin skin tests is considered to be very low and may be related to incorrect dose of the drug and the lack of performing the prick test before the intradermal test¹⁶. It is important to remember that skin testing may give false-negative results for 1 to 2 weeks or even longer after an episode of anaphylaxis⁹ (Fig. 1).

A clinical practice guideline for penicillin skin testing in patients who have a history of penicillin allergy does not increase the costs nor improve financial savings, instead, it increases the percentage of eligible patients for skin testing and, on the other hand, it reduces the necessity of alternative antibiotics1. It is necessary to take into account that the skin testing for penicillin is useful only for the diagnosis of IgE-mediated reactions to penicillins, not for delayed reactions such as Stevens-Johnson syndrome, or other reactions which are not IgE-mediated9. The penicillin skin test has a negative predictive value of 99% and a positive predictive value of 50%, when it includes major and minor determinants of the drugs. Because minor determinants are not commercially available, some authors have suggested a dilution of penicillin G, to a concentration of 10,000 units/ml with 0.9% sodium chloride¹⁷. However, the diluted solution should be freshly prepared⁹. Using benzyl penicilloyl as the major determinant, commercially named Pre-Pen, and diluted penicillin G, the negative predictive value of the test is 97%. Nonetheless, the Pre-Pen solution is nowadays unavailable commercially $^{15\cdot 16}$, which makes penicillin allergy diagnosis very difficult.

Penicillin skin testing has been proposed to be performed by using lab made reagents. Sarti¹⁹ has performed penicillin skin tests in 6,764 patients in an outpatient clinic, by employing fresh diluted penicillin G (PG) and hydrolysate penicillin as minor determinant mixtures (MDM). His data showed that only 96 patients presented positive results. None of the 6,668 patients with negative skin tests re-exposed to penicillin showed any immediate systemic reaction. According to the author, MDM correlates well with diluted penicillin G and can be employed for diagnosing patients susceptible to immediate severe reactions, such as oedema of the larynx and anaphylactic shock; although it could miss some accelerated reactions which are mainly caused by the major determinants. In our country, the Ministry of Health has adopted the mentioned schedule for penicillin drug allergy diagnosis²⁰.

The Center for Diseases Control²¹ states that as no proven alternatives to penicillin are available for treating neurosyphilis, congenital syphilis or syphilis in pregnant women, it is recommended that if the major determinants are not available, patients with a history of immediate IgE-mediated reactions should be desensitised in a hospital service.

In general, penicillin skin testing is useful for detecting any beta-lactam antibiotics¹⁸; however, it is important to note that penicillins share the beta-lactamic ring structure and the thiazolidine rings, but not necessarily the R side

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chains²². It is possible that the patient has non cross-reactive responses to various beta lactam antibiotics²³.

It is estimated that 50% of patients who experienced immediate reactions to penicillin will have a negative skin test after 5 years, and 75-80% will be negative at 10 years 9 . Nonetheless, penicillin is no longer considered the most commonly prescribed β -lactam 24 , therefore it is recommended to include other determinants in skin tests such as amoxicillin and cephalosporin $^{24\cdot25}$. According to the data from Blanca et al. 24 in patients who suffered penicillin allergy adverse reactions after treatment, the drugs most involved in the reactions were amoxicillin (87.8%), followed by benzopenicillin (8.1%), ampicillin (2.7%), and cloxacillin (1.3%).

In vitro IgE-RAST/CAP and ELISA testing

The radioallergosorbent test (RAST), cellulose fluorescent assay-IgE (CAP-IgE) or enzyme-linked immunosorbent assay for detecting anti-drug IgE has variable specificity and sensitivity. For instance, a negative result does not exclude allergy to penicillin, as the assays detect only IgE to the major determinant; however, a positive result can indicate an increased risk for allergic reactions 9,16. The specificity and sensitivity of the CAP-IgE for β -lactam are 85.7-100% and 12.5-25.0%, respectively, depending on the initial clinical manifestations²⁶. The positive predictive values are considered to be high in severe clinical conditions, such as anaphylactic shock²⁶. Therefore it could be useful in these situations if the skin test is negative, in order to avoid the necessity of drug provocation^{24,26}. However, the sensitivity and specificity of the in vitro IgE antibodies test decrease more than one year after the allergy episode²⁶.

Drug provocation test

The drug provocation test is recommended to confirm drug hypersensitivity reactions by the European Network for Drug Allergy from the European Academy of Allergology and Clinical Immunology^{10,27}. The test is cumbersome but can be considered as a reference test when skin tests are not available or not validated^{5,28}. It should be performed in cases of inconclusive or negative skin and/or CAP-IgE tests in patients with history of immediate drug allergy²⁹. In most cases, the same route of administration in the clinical history is used, unless the oral form of the culprit drug is available¹⁰.

Increasing doses of a suspected drug are given at 0.5 to 1-h intervals until the full age/weight-adjusted dose is achieved, and depending on the protocol, this procedure may take 1, 2 or more days^{23,28}. H1-antihistamines and β -blockers should be denied for 2 and 5 days, respectively, before the drug provocation test²⁷. Placebo challenges may be necessary to avoid false positive reactions²⁹. False negative results can occur and could be explained by cofactors such as exercise, sunlight, and tolerance induction²⁷. Contraindications for the test are patients with severe comorbidities such as cardiac, hepatic, renal or other diseases and patients who had experienced severe life-threatening reactions as with Stevens-Johnson syndrome, toxic epidermal necrolysis, vasculitis, drug reaction with eosinophilia and systemic symptoms (DRESS)^{27,30}.

Drug provocation tests can reproduce the hypersensitivity reactions of the clinical episode, but they are in general milder and have a shorter duration²⁷. Although they are dangerous procedures and potentially life-threatening, these tests are important to confirm drug hypersensitivity, and therefore

non-hypersensitive patients would not need to avoid the related drugs in the future if the test were negative⁵.

Intradermal and patch tests for non-immediate allergy

Intradermal and patch tests are recommended for the routine screening of patients with suspected non-immediate allergy²³. The drug skin tests should be performed 6 weeks to 6 months after resolution of the drug reaction. Barbaud et al. 13 proposed a guideline for performing skin tests in order to standardise these procedures in cutaneous adverse drug reactions. According to the authors, skin testing should be performed with the commercialised drug whenever possible, and also with the pure active products and excipients. Systemic corticosteroid or immunosuppressive therapy must be interrupted for at least one month. For patients who have experienced severe reactions such as Stevens-Johnson syndrome, the drugs must be diluted first at 0.1%; if negative then at 1.0%; and if negative up to 10% in patch tests¹³. The intradermal tests are contraindicated in these patients. Healthy volunteers with or without previous exposure to the drug can be used as negative controls. The highest frequency of positive drug patches are related to β lactam antibiotics, especially amoxicillin; cotrimoxazole; corticosteroids; heparin derivatives; carbamazepine; diazepam; and pseudoephedrine¹³. Delayed positive results in intradermal tests can be obtained in maculopapular rash, eczema, erythroderma or fixed drug reactions. The threshold of specificity has been determined for various drugs, such as β lactam antibiotics, erythromycin and spyramicin, and needs to be determined for other drugs, because of the risk of false positives in intradermal tests¹³.

Basophil activation test

The CD63 molecule was discovered in the 1990s and since then it has been used as a basophil activation marker in flow cytometry. The technique, which is named basophil activation test (BAT), relies upon anti-IgE to characterize basophiles and anti-CD63 to assess activation of these cells^{31,32}. The molecule is expressed by different types of cells, e.g. basophiles, mast cells, macrophages and platelets. In the resting cells, CD63 is anchored in the basophilic granule and weakly expressed on the surface membrane; upon activation, its expression is upregulated, besides other molecules, such as CD45, CD203c31. The BAT test allows the diagnosis of immediate drug allergy and pseudoallergy mainly for drugs that are not always detectable by CAP-IgE³³. The BAT test is more sensitive and specific than other in vitro diagnostic techniques in drug allergy³³, and is essentially complementary to skin tests and allergen-specific IgE determinations34.

This technique has been proven to be a useful tool for assessment of the diagnosis of IgE-mediated allergies, including drug allergy³³. The upregulation of CD63 and CD203c has been observed in tests for pollen, food, drugs, natural rubber latex, and recombinant pollen allergens³¹. Nevertheless, it suffers from some drawbacks. It depends on various factors, such as temperature, incubation time, a diluting buffer, type of allergen, and a positive control as anti-IgE³¹. The density of IgE and IgE receptors may vary considerably among individuals and also among basophiles from the same individual. This may play a negative role, particularly in nonatopic individuals and when the number of activated ba-

sophiles is low³⁴. The necessity of anti-IgE as a positive control in CD63 testing is considered to be an important drawback, once some patients are non-responders; also, the results might be influenced by the intake of drugs, such as glucocorticosteroids, immunossuppressives, and immunomodulators³¹. Moreover, IL-3 may be necessary for the priming of basophiles without up-regulating the activation markers, since in drug allergy it can occur that the drugs *per se* induce a low rate of basophil activation in the test³⁵.

Other alternative means of detecting basophiles have recently been identified, such as the CD203c marker, a neural cell surface differentiation antigen E-NPP3 (ectonucleotide pyrophosphatase/phosphodiesterase), which is exclusively present on basophiles among blood cells, mast cells and progenitors. The CD 203c marker is upregulated in response to allergen-specific and anti-IgE-induced IgE cross-linking similar to CD63 32,34 and can be applied in flow-assisted allergy diagnosis to improve the sensitivity of the BAT technique, but multi-centre studies are necessary in order to make the technique useful for clinical routine 35.

In a study evaluating immediate reactions to β -lactam, the sensitivity of the BAT test was 50% and its specificity 94%³⁶. Torres et al.³⁷ also assessing reactions to these antibiotics, found similar results to BAT with a sensitivity of 48.6% and specificity of 93%; moreover, the BAT for cephalosporin was positive in 77% of the patients, indicating that this test seems to be very promising for cephalosporin allergy. Nonetheless, studies in larger samples are required in order to validate the technique³⁸.

Gamboa et al. ³⁹ found a specificity of 90% and sensitivity of 60% to 70% in BAT for non-steroidal anti-inflammatory drugs (NSAIDs). The results with BAT are suggestive that when it is positive there would be no need to perform the provocation test. However, Malbran et al. ⁴⁰ showed that diclofenac activates basophiles in sensitive individuals in a way that does not induce CD63 expression, suggesting that the BAT test may not be reliable for NSAID hypersensitivity.

Lymphocyte transformation test

The lymphocyte transformation test (LTT) is a test that assesses drug-specific T cells activation after in vitro stimulation with various concentrations of the culprit drug⁴¹. It has several advantages as it is not harmful for the patient; it can be positive in drug reactions caused by different immunopathogenic mechanisms⁴². Although it is thought to be a reliable test⁴¹, it has been considered more as a research tool than for diagnostic purposes¹⁵. There are several variables which can influence its performance: time of testing; influence of therapy as corticosteroids; stimulation index; reproducibility; sensitivity; and specificity41-42. According to Kano et al.41, the right time to perform the test depends on the type of reactions, within one week from the onset of the skin rashes in patients with maculopapular exanthema or Stevens-Johnson syndrome/toxic epidermal necrolysis, and 5 to 8 weeks after the onset of skin rashes in patients with drug-induced hypersensitivity syndrome/drug rash and eosinophilia with systemic symptoms.

Detection of cytokines as IL-5

The IL-5 secretion cells can be analysed in vitro in culture supernatants from LTT after activation with specific drugs,

and improve LTT sensitivity⁴³. According to Sachs et al.⁴⁴ the determination of drug-specific IL-5 secretion was more sensitive (92%) than the LTT (75%) and patch test (55%) in patients with drug-induced maculopapular exanthemas. As Th1 and Th2 patterns occur together or not in drug allergy, and as IL-5 is a representative of the Th2 profile, the tests should include the determination of a cytokine representative of the Th1 pattern, as IFNy.

Markers of T cell activation such as CD69

Considering the several disadvantages of LTT, as mentioned above, the detection of surface markers, such as CD69 and CD25, which upregulate upon T cell activation after contact with a specific antigen, peptide or drug, can also be a promising alternative for diagnostic purposes³⁰. The test consists in culturing peripheral mononuclear cells with nontoxic concentrations of the culprit drug for 36 to 72 h and submitting them to flow cytometry after the addition of fluorochrome-labelled anti-human CD69 monoclonal antibodies³⁰.

Beeler et al. ³⁰ evaluating 15 drug-allergic, LTT positive patients, found a CD69 upregulation in both TCD4 cells and TCD8 cells after drug stimulation, except when the cells were cultivated with clavulanic acid, which induced an increase of CD4 T cells only. The test has shown a high specificity since non-allergic individuals showed no increase of CD69 detection of T cells cultivated with drugs. However, the authors demonstrated that clavulanic acid could induce CD69 upregulation without proliferation of non-allergic cells without CD69 upregulation. Therefore, the authors ³⁰ strongly suggest the necessity of an assay evaluation for every drug to be tested in non-allergic individuals.

Conclusion

The development of new tests and the validation of already developed techniques have been contributing enormously in the diagnosis of drug allergy. The diagnostic tests, together with a carefully detailed clinical history of the patient, physical examination and routine laboratorial parameters, are critical to identify the culprit drug, for the hypersensitivity drug reaction classification, and finally for taking a decision upon the treatment of patients.

Acknowledgement

This article is part of a project entitled "Incidence evaluation of allergy hypersensitivity reactions to beta-lactamics through laboratorial investigation of patients exposed to the drug", sponsored by MCT/CNPq/MS-SCTIEDECIT-DAF 54/2005, process 402509/2005-6.

Conflict of interest

The authors have no conflict of interest to declare.

References

 Forrest DM, Schellenberg RR, Thien VVS, King S, Anis AH, Dodek PM. Introduction of a practice guideline for penicillin skin testing improves the appropriateness of antibiotic therapy. CID 2001;32:1685-90. 104 Nagao-Dias AT et al

2. Gomes ER, Demoly P. Epidemiology of hypersensitivity drug reactions. Curr Opin Allergy Clin Immunol 2005;5:309-16.

- Greenberg PA. Drug allergy. J Allergy Clin Immunol 2006;117: S464-70.
- Bates DW, Leape L. Adverse drug reactions. In: Carruthers SG, Hoffman BB, Melmon KL, Nierenberg DW, editor. Clinical pharmacology: Basic principles in therapeutics. 4th ed. New York: McGraw-Hill Companies; 2000. p. 1223-54.
- 5. Demoly P. Anaphylactic reactions value of skin and provocation tests. Toxicology. 2005;209:221-3.
- Vervloet D, Durham S. ABC of allergies: adverse reactions to drugs. BMJ. 1998;316:1511-4.
- Hepner DL, Castells MC. Anaphylaxis during the perioperative period. Anesth Analg. 2003;97:1381-95.
- 8. Pichler W, Yawalkar N, Schmid S, Helbling A. Pathogenesis of drug-induced exanthems. Allergy 2002;57:884-93.
- Arroliga ME, Pien L. Penicillin allergy: considering trying penicillin again. Cleveland Clin J Med 2003;70:313-26.
- Bousquet PJ, Kvedariene V, Co-Minh HB, Martins P, Rongier M, Arnoux B, et al. Clinical presentation and time course in hypersensitivity reactions to β-lactams. Allergy 2007;62:872-876.
- Souza MOB, Araujo MCC, Santiago RA, Coelho HLL, Fonteles MMF. Adverse reactions to oxacillin in hospitalized children: a prospective study. Rev Bras Saúde Matern. Infant. 2007;7:55-61.
- 12. The Uppsala Monitoring Centre (the UMC)/WHO Collaborating Centre for International Drug Monitoring. Safety monitoring of medicinal products: Guidelines for setting up and running a Pharmacovigilance Centre, Uppsala, Sweden, 2005, 25 pp.
- Barbaud A, Goncalo M, Bruynzeel D, Bircher A. Guidelines for performing skin tests with drugs in the investigation of cutaneous adverse drug reactions. Contact Dermatitis 2001;45:321-8.
- ACAAI. Annotations of the algorithm for disease management of drug hypersensitivity. Ann Allergy Asthma Immunol 1999;83: 667-71.
- Gruchalla RS, Pirmohamed M. Antibiotic allergy. N Engl J Med 2006; 354:601-9.
- Park MA, Li JTC. Diagnosis and management of penicillin allergy. Mayo Clin Proc 2005;80:405-10.
- 17. Wall GC, Peters L, Leaders CB, Wille JA. Pharmacist-managed service providing penicillin allergy skin tests. Am J Health-Syst Pharm 2004;61:1271-5.
- 18. Shepherd GM. Hypersensitivity reactions to drugs: evaluation and management. The Mount Sinai J Med 2003;70:113-25.
- Sarti W. Routine use of skin testing for immediate penicillin allergy to 6,764 patients in an outpatient clinic. Ann Allergy 1985;55:157-61.
- Brasil. Ministério da Saúde. Secretaria de Políticas de Saúde. Coordenação nacional de DST e Aids — Manual de teste de sensibilidade à penicilina. Brasília, 1999. http://www.aids.gov.br/testes_penicilina.pdf.
- 21. Center for Diseases Control. Management of patients who have a history of penicillin allergy. Sexually transmitted diseases treatment guideline. 2006. http://cdc.gov/std/treatment/2006/penicillin-allergy.htm. access in 12/3/2007.
- 22. Gruchalla RS. Drug allergy. J Allergy Clin Immunol 2003;111: S548-59.
- 23. Padial A, Antunez C, Blanca-Lopez N, Fernandez TD, Cornejo-Garcia JA, Mayorga C, et al. Non-immediate reactions to b-lactams: diagnostic value of skin testing and drug provocation test. Clin Exp Allergy 2008;38:822-8.
- 24. Blanca M, Mayorga C, Torres MJ, Roche M, Moya MC, Rodriguez JL, et al. Clinical evaluation of Pharmacia CAP system RAST FEIA amoxicilloyl and benzylpenicilloyl in patients with penicillin allergy. Allergy 2001;56:862-70.
- 25. Torres MJ, Romano A, Mayorga C, Moya MC, Guzman AE, Reche M, et al. Diagnostic evaluation of a large group of patients with immediate allergy to penicillins: the role of skin testing. Allergy 2001;56:850-6.

26. Fontaine C, Mayorga C, Bousquet PJ, Arnoux B, Torres MJ, Blanca M, et al. Relevance of the determination of serum specific IgE antibodies in the diagnosis of immediate β -lactam allergy. Allergy 2007;62:47-52.

- Messaad D, Sahia H, Benahmed S, Godard P, Bousquet J, Demoly P. Drug provocation tests in patients with a history suggesting an immediate drug hypersensitivity reaction. Ann Intern Med 2004;140:1001-6.
- 28. Gomes ER, Fonseca J, Araujo L, Demoly P. Drug allergy claims in children: from self-reporting to confirmed diagnosis. Clin Exp Allergy 2007;38:191-8.
- 29. Wöhrl S, Vigl K, Sting G. Patients with drug reactions is it worth testing? Allergy 2006;61:928-34.
- Beeler A, Zaccaria L, Kawabata T, Gerber BO, Pichler WJ. CD69 upregulation on T cells as an in vitro marker for delayed-type drug hypersensitivity. Allergy 2008;63:181-8.
- 31. Ebo DG, Hagendorens MM, Bridts CH, Schuerwegh AJ, de Clerk LS, Stevens WJ. In vitro allergy diagnosis: should we follow the flow? Clin Exp Allergy 2004;34:332-9.
- 32. Ebo DG, Sainte-Laudy J, Bridts CH, Mertens CH, Hagendorens MM, Schuerwegh AJ, et al. Flow-assisted allergy diagnosis: current applications and future perspectives. Allergy 2006;61: 1028-39.
- Sanz ML, Gamboa PM, de Weck AL. Cellular tests in the diagnosis of drug hypersensitivity. Curr Pharm Design 2008;14: 2803-8.
- 34. De Weck AL, Sanz ML, Gamboa PM, Aberer W, Bienvenu J, Blanca M, et al. Diagnostic tests based on human basophils: more potencials and perspectives than pitfalls. II. Technical issues. J Investig Allergol Clin Immunol 2008;18:143-55.
- Erdmann SM, Sachs B, Hoffmann-Sommergruber K, Scheiner O, Merk H. Regarding Ebo DG, Hagendorens MM, Bridts CH, Schuerwegh AJ, De Clerck LS & Stevens WJ. In vitro allergy diagnosis: should we follow the flow? Clin Exp Allergy 2004;34:332-9. Clin Exp Allergy 2004;34:1498-500.
- Sanz ML, Gamboa PM, Antépara I, Uasuf C, Vila L, Garcia-Avilés C, et al. Flow cytometric basophil activation test by detection of CD63 expression in patients with immediatetype reactions to betalactam antibiotics. Clin Exp Allergy 2002; 32:277-86.
- Torres MJ, Padial A, Mayorga C, Fernández T, Sanchez-Sabate E, Cornejo-Garcia JA, et al. The diagnostic interpretation of basophil activation test in immediate allergic reactions to betalactams. Clin Exp Allergy 2004;34:1768-75.
- 38. Romano A, Demoly P. Recent advances in the diagnosis of drug allergy. Curr Opin Allergy Clin Immunol 2007;7:299-303.
- Gamboa P, Sanz ML, Caballero MR, Urrutia I, Antépara I, Esparza R, et al. The flow-cytometric determination of basophil activation induced by aspirin and other non-steroidal anti-inflammatory drugs (NSAIDs) is useful for in vitro diagnosis of the NSAID hypersensitivity syndrome. Clin Exp Allergy 2004;34: 1448-57
- Malbran A, Yeyati E, Rey GL, Galassi N. Diclofenac induces basophil degranulation without increasing CD63 expression in sensitive patients. Clin Exp Allergy 2006;147:99-105.
- 41. Kano Y, Hirahara K, Mitsuyama Y, Takahashi R, Shiohara T. Utility of the lymphocyte transformation test in the diagnosis of drug sensitivity: dependence on its timing and the type of drug eruption. Allergy 2007;62:1439-44.
- 42. Pichler WJ, Tilch J. The lymphocyte transformation test in the diagnosis of drug hypersensitivity. Allergy 2004;59:809-20.
- 43. Merck HF. Diagnosis of drug hypersensitivity: lymphocyte transformation test and cytokines. Toxicology 2005;209:217-20.
- 44. Sachs B, Erdmann S, Malte Baron J, Neis M, al Masaoudl T, Merk HF. Determination of interleukin-5 secretion from drug-specific activated ex vivo peripheral blood mononuclear cells as a test system for the in vitro detection of drug sensitization. Clin Exp Allergy 2002;32:736-44.