Montelukast in early childhood asthma. Predict value of IgG in clinical reply in children 2 to 5 years old?

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ABSTRACT

Background: According to current knowledge, asthma is basically an inflammatory process. Its causes and physiopathological mechanisms are various. The final result is a recurrent obstructive bronchial process, with sibilants and/or dypnea, which causes an upset in functional respiratory tests, among which the maximum respiratory peak flow meter diminished for the age, sex, and height of patient.

Aims: Our aim is to evaluate if response to treatment with Montelukast has any link with immnuloglobin values (IgG, IgA, IgM, IgE) at start of treatment.

Materials and methods: Included in the study were 32 children, of whom 2 did not begin and 1 who did not provide personal data. There were 29 patients in total, 11 girls and 18 boys. Each made three visits: first where they were instructed, together with their parents, in how to manage the meter and where they received the peak flow meter, Vitalograph, and personal data sheet, where personal and family medical history were noted. The second visit was after 4 weeks, for a clinical assessment and the third visit after 8 weeks. The value register of the PEF would be made morning and night, noting the highest value of three measurements. IgG, IgA, IgM, IgA values were quantified before treatment began. The statistic package STATA 2001 was used in the treatment of data statistics.

Results: Our between the value reached by the PEF after treatment and the IgG values at the beginning of treatment (0.712). In lesser measurement for IgA values (0.660). For each 100 mg/ml of increase in the value of IgG, an increase of 10 l/min in the PEF measurement before and following treatment with Montelukast was produced.

Conclusions: IgG values increase with age. Children with a greater IgG value at the beginning of treatment reached higher PEF values after same. It is not known if the results would be similar with another type of treatment and the way in which IgG influences the results. What appears to be confirmed by available studies is that this relation is found in a group of small children, the aim of our study.

Key words: Asthma. Childhood. IgG. Montelukast.

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RESUMEN

Antecedentes: El asma es, fundamentalmente, un proceso inflamatorio, según los actuales conocimientos. Sus causas y sus mecanismos fisiopatológicos son múltiples. El resultado final es un proceso obstructivo bronquial recurrente con sibilancias y/o disnea, que se traduce en una alteración de las pruebas funcionales respiratorias, entre ellas, el pico de flujo

espiratorio máximo, PEF, disminuido para la edad, sexo y talla del paciente.

Objetivo: El objetivo de este estudio es valorar si la respuesta al tratamiento con montelukast, tiene alguna relación con los valores de inmunoglobulinas (IgG, IgA, IgM, IgE) al inicio del tratamiento.

Material y métodos: Se incluyen en el estudio 32 niños, de los que dos no lo comienzan y uno no registra los datos. Siguen el programa previsto 29 pacientes, 11 niñas y 18 niños. Todos realizan tres visitas. Una primera en que se les instruye, junto con sus padres, en el manejo del medidor, se les entrega el *peak flow meter* (PEF), modelo Vitalograph, la hoja de registro de datos y se recogen los antecedentes familiares y personales. La segunda visita, a las 4 semanas, de valoración clínica, y la tercera visita, a las 8 semanas. El registro de los valores de PEF se hará mañana y noche, anotando el mejor valor de tres mediciones. Se cuantifican los valores de IgG, IgA, IgM e IgE antes de comenzar el tratamiento.

Resultados: Nuestra muestra tiene un 62,1 % de varones. No existen diferencias significativas, en los parámetros considerados, en razón del sexo. Existe una relación entre el valor que alcanza el PEF después del tratamiento y los valores de IgG al inicio del mismo (0,712). En menor medida, con los valores de IgA (0,660). Por cada 100 mg/ml de incremento en el valor de IgG, se produce un aumento de 10 l/min en la diferencia del PEF medido antes y después del tratamiento con montelukast.

Conclusiones: Los valores de IgG se incrementan con la edad. Los niños que tienen mayor valor de IgG al inicio del tratamiento alcanzan valores más altos en el PEF después del mismo. Desconocemos si el resultado sería similar con otro tipo de tratamiento y la manera en que influye esta IgG en el resultado. Lo que sí parece confirmado, por los estudios disponibles, es que esta relación se encuentra en el grupo de niños pequeños, objeto de nuestro estudio.

Palabras clave: Asma. Infancia. IgG. Montelukast.

INTRODUCTION

The current concept of asthma involves diverse degrees of clinical expression, as expression of complex histopathological and patho-physiological[1] base, which lead to an inflammatory process in the bronchial tree. In asthma[2], inflammation is a consequence of the embedded participation of a reticular intricate system in which cells, cytokines, chemical mediators, adhesion molecules, etc. participate in response to different types of aggression, which the bronchial tree suffers.

Due to this knowledge, the treatment of asthmatic disease has adapted toward the control of the different processes involved.

During breast feeding and early infancy, various "asthmatic subgroups" were identified: those which developed sibilants following bronchiolitis, another group which possess sibilants with a good general state ("happy wheezing"), usually overweight males and the third group where disturbances in lung function are detected[3].

Martínez[4] and colleagues describe three clearly differentiated subgroups in early infancy: one linked to viral illnesses, another associated with atopy, and the third with previous problems connected to lung function (chronic obstructive pulmonary disease). In a group of very young children, those episodes induced by virus predominate. Therefore, it is reasonable to believe that the immune situation of the child plays a large role in recovery.

In light of current knowledge, within chemical measurements, leukotriens (LT) play an outstanding role. These are fatty acids derived from arachidonic acid[5] which in turn proceed from membrane phospholipids by means of hydrolysis produced by phospholipase A2.

5 Lipoxygenase (5LO), activated by a 5LO activating protein (FLAP), acts on the arachidonic acid leading to a series of intermediate unstable compounds which result in the LT primer LTA4, which in turn is also unstable.

In neutrophils, monocytes and alveolar macrophages activated by LTA 4 hydrolase, LTB4 is obtained with known chemotactic activity and activator of neutrophils, and to a lesser degree, chemotactic of eosinophils.

In cells fundamentally implicated in inflammation, mastocytes, eosinophils and basophils, LTA4 activated by LTA4 synthase will cause the first of the cysteinil leucotrienes (cys-lt) the LTC4. This undergoes an extra cellular metabolism, which gives rise to LTD4 and LTE4[6].

The effects of LT are measured by protein G (together with BLT receptor for LTB4 and the receptors of cys-LT): cys-LT1 throughout the entire bronchial tree and cys-LT2 at the venous pulmonary system.

LT produce broncho constriction, exudance of plasma, infiltration of eosinophils, increase in mucuos production, contraction of bronchial smooth muscle, disturbance of respiratory flagellum, proliferation of smooth muscle, and specific action as stimulus of liberation of preformed IL-4 in eosinophils[7], stimulus of the synthesis of eosinopoietics cytokines[8] (GM-CSF, IL-3, IL-5), increase in expression factor of

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epidermal growth[9] IgF, stimulus in production of collagenase by fibroblasts in the lung[10], etc.

An open prospective study was designed in order to evaluate the efficiency of treatment with Montelukast, 4 mg/daily, in the control and functional recovery from bronchial asthma in children between 2 and 5 years of age, attended by consultant paediatricians with primary care, and its possible connection with initial immunoglobin values (IgG).

METHODS

Children of both sexes with a syndrome diagnoses of asthma were included in this study, who attended their paediatricians at health centers. In this age group, we define as bronchial asthma 3 or more episodes of bronchial obstruction in the last year. Exclusion criteria were established as follows: 1) no wish to participate in the study; 2) poor tolerance of medication and/or adverse effects (possible bias); 3) wish to abandon study once started; 4) appearance of another pathology causing suspension of study.

32 patients were originally included. Parents were informed that they could abandon study at any time. One week before treatment began with Montelukast 4 mg (one pill taken nightly but not with meal), the peak flow meter had to be registered morning and night, noting the best of three measurements each time. Instructions were given at the clinic on how to carry out tests using the VITALOGRAPH model (donated by PROFAS lab) and the form was provided for data.

Three programmed consultations were done: inclusion in the study for which parental permission was needed, parents were informed that they could abandon study at any time, as well as how to carry out test and collect variable data. A second consultation was held after 4 weeks and last at 8 weeks. The clinical state of the patient was checked in both, control of peak flow and correct register of data were carried out.

The general variables are: age, sex, height, daily and nightly PEF reading for the 7 days prior to treatment, as well as percentage of immunoglobins (Ig), IgG, IgA, IgM, IgE (presence and type of adverse reaction, evolutionary clinical markers, need for rescue medication, use of CI, days absent from school).

Statistical analysis

We carried out hypotheses contrasts based on non-parametric statistical tests. To contrast the difference between the proportion of men and women in the sample, an exact binomial queue was used. Hypothesis tests on gender difference for quantitative variables were obtained using a Mann-Whitney test for independent samples. For variables related to evolution, (those measures before and after treatment), a Wilcox test of designated ranges was used for matching samples. In the study for association between quantitative variables and non-parametric coefficient of correlation Spearman was used. As for peak flow, the level increase was measured, while in the rest, decrease was quantified in absolute values. For these indicators, the Spearman coefficient of correlation was constructed with the aim of measuring possible relation of improvement with other variables of interest. Finally, we carried out uni-variant regression models for variables of greater evolutionary interest. Statistic package STATA CORP. 2001[11].

RESULTS

There exists a greater number of male patients (62.1 %) although the difference is not significant. The contrasts in hypothesis concerning distribution of considered variables comparing both sexes did not reveal any significant difference (significance level of 5 %), which suggests that patients have similar characteristics regardless of sex (tables I and II).

Of the 32 patients in the study, 2 did not stard, 1 did not carry out data completion, although the later did meed some variables. 29 patients began the study. 2 patients abandoned study due to adverse reactions (diarrhoea in one, and nightmares and hives in the other). One other patients abandoned study due to a parental decision, 26 patients completed the study overall. These general results are reason for comment in another article.

We find a positive link between all Ig pairs, except between IgG and IgE. We also see an important positive link between IgG and IgA values (coefficient correlation 0.762), therefore the greater level of IgG that exists the greater level of IgA will be found and vice versa.

With regard to the PEF value reached after treatment, there exists a positive correlation with IgG values at the beginning of treatment (0.712), and similarly but to a lesser degree, with IgA values (0.660).

Those children with the highest IgG values at the start of the study, obtained the highest PEF value at the end of the 8-week treatment period with Montelukast. The same occurred in relation to the IgA value, but with a lesser correlation.

Table I

Description[a] of variables according to gender

	Male	Female	Total	p[b]
Percentage (%)	62.1 (42.3-79.3)	37.9 (20.7-57.7)	100	0.132
Age when symptoms begam (months)	11.9 (7.6-16.3)	21.0 (10.4-31.6)	15.4 (10.6-20.1)	0.141
Time of evolution from star of symptoms to treatment	28.8 (22.6-34.9)	23.9 (14.5-33.3)	26.9 (22.0-31.9)	0.233
Age when treatment began (months)	40.9 (35.5-46.3)	44.9 (37.7-52.5)	42.4 (38.2-46.6)	0.392
Size	100.9 (97.4-104.5)	103.7 (96.5-110.9)	101.9 (98.7-105.1)	0.486
Days showing symptoms in last 2 months	5.1 (3.5-6.7)	4.9 (2.1-7.8)	5.0 (3.7-6.4)	0.555
Treatment days in last 2 months	1.1 (0.2-1.9)	0.8 (0.2-1.4)	1.0 (0.4-1.5)	0.885
Peak flow prior to treatment	104.7 (73.2-136.2)	92.0 (61.7-122.3)	100.0 (78.5-121.5)	0.649
Peak flow alter treatment	142.6 (109.7-175.6)	125.5 (89.6-161.4)	136.3 (113.0-159.6)	0.256
Percentage of peak flow prior to treatment	2 (1.0-9.9)	2.5 (1.3-18.5)	2 (2-5)	0.9392
Percentage of peak flow following to treatment	50 (5.1-55)	26 (5-66.9)	42 (7.8-55)	0.5104
Daily variability prior to treatment	32.0 (21.2-42.8)	31.6 (24.5-38.7)	31.8 (24.9-38.8)	0.704
Daily variability following treatment	16.2 (10.7-21.6)	17.8 (12.7-22.8)	16.8 (13.1-20.4)	0.329
Variability in the time prior to treatment	34.2 (27.0-41.3)	36.6 (24.1-48.5)	35.0 (29.1-40.8)	0.899
Variability in the time following treatment	38.6 (28.8-48.5)	36.7 (23.6-49.7)	37.9 (30.6-45.2)	0.812
Protection index	47.2 (33.0-61.4)	38.2 (27.5-48.8)	43.8 (34.5-53.2)	0.209
Number of times under medication of asthma attacks during treatment	1.7 (0.3-3.1)	0.4 (0.0-1.0)	1.3 (0.3-2.2)	0.215
School days lost in quarter before treatment	18.6 (13.1-24.1)	13.9 (7.6-20.1)	17.1 (13.0-21.1)	0.334
School days lost in quarter following treatment	3.1 (0.3-5.8)	0.1 (0.0-0.3)	2.0 (0.2-3.8)	0.130

[a] Simple average.
[b] p-value of hypothesis contrast.

Table II

Description[a] of variables according to gender

	Male	Female	Total	p[b]
IgG	865.0 (755.4-974.7)	904.7 (753.2-1,056.2)	880.3 (797.6-963.0)	0.916
IgA	107.9 (82.0-133.9)	115.7 (59.3-172.0)	110.9 (86.4-135.3)	0.712
IgM	121.2 (87.6-154.8)	151.0 (52.0-249.9)	132.6 (93.3-172.0)	0.772
IgE	143.5 (18.8-268.2)	100.5 (34.9-166.2)	127.6 (48.9-206.3)	0.616

[a] Simple average.
[b] p-value of hypothesis contrast.

No correlation between the IgG value at the beginning of the study and the rest of the considered variables was found.

Medication during same (–0.445), as if a greater IgE value meant fewer treatment days, compared with children having lower IgE and requiring. The IgE value was curiously associated although negatively with those days when treatment was required 2 months prior to study (–0.430), and with the need for rescue less rescue medicine during the study (table III).

The adjustment of uni-variant linear regression models revealed a possible link between the IgG value and the increase in PEF, finding no significant statistical link with remaining Ig. This link is maintained even when the multi-variant model is adjusted. For each 100 mg/ml of increase in the IgG value, a rise of 10 l/min is produced in the PEF measurement, prior or following treatment, adjusted for time of evolution, protection rate, IgA and IgM (fig. 1).

As expected, the IgG values increase with age, with the proportion of 7 mg/ml for each month (fig. 2).

Table III

Non parametric correlations

	IgE	IgG	IgA	IgM
IgE	1.000			
IgG	0.228	1.000		
IgA	0.456[a]	0.762[b]	1.000	
IgM	0.458[a]	0.418[a]	0.504[a]	1.000
Difference in peak flow	−0.168	0.526[b]	0.100	0.243
Difference in peak flow percentage	−0.096	0.517[b]	0.167	0.310
Difference in daily variability	−0.027	0.104	0.069	−0.144
Difference in lost school days	−0.310	0.100	0.185	0.136
Age at onset symptoms	0.188	0.109	0.257	0.043
Evolution time from onset of symptoms to treatment	−0.147	0.164	0.133	0.080
Symptoms days in last 2 months	−0.213	−0.208	0.408[a]	0.065
Treatment days in last 2 months	−0.430[a]	−0.219	0.361	−0.148
Protection index	0.202	0.305	0.297	0.283
Number of times medication was required for asthma crises during treatment	−0.445[a]	0.162	−0.302	−0.039

[a] The correlation was significant at the level of 0.05 (bilateral).
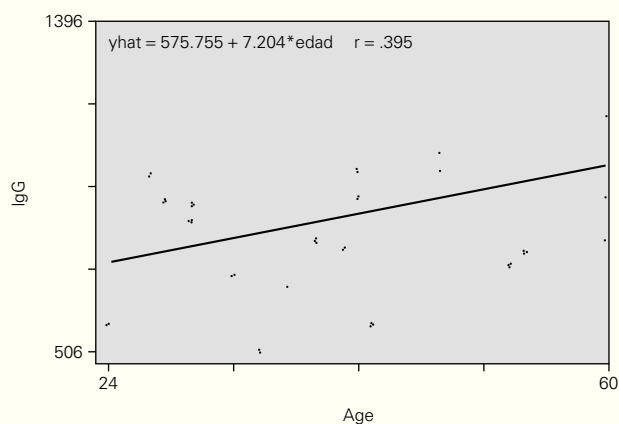[b] The correlation was significant at the level of 0.01 (bilateral).

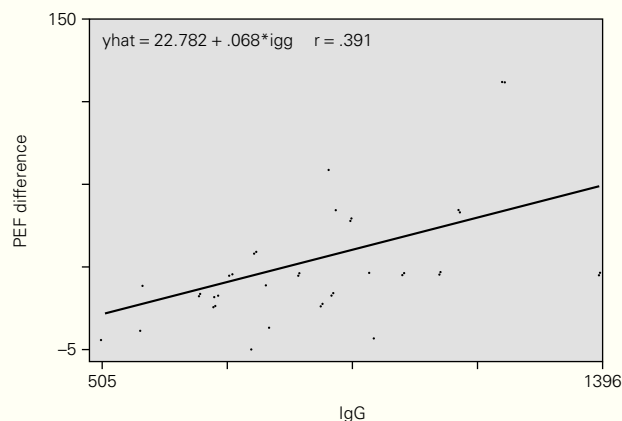Figure 1.—Time of evolution from symptom.

yhat = 575.755 + 7.204*edad   r = .395

Figure 2.—Protection index.

yhat = 22.782 + .068*igg   r = .391

DISCUSSION

That Ig values increase with age during development is well known[12,13]. This involves progressive maturation of the immune system, originating in the foetal period and accelerating from then on, in response to contact with a diversity of environmental agents. There is data which supports the existence of certain differences between boys and girls[14,15]: significantly higher levels of IgM have been found in women[14], and even in some cases IgA in a population aged between 10 and 73[15].

Children with higher IgG at the beginning of the study had elevated PEF values at the end, i.e. they showed a better functional respiratory response quantified in an increase of 10 l/min in PFM per each 100 mg/ml which increased the IgG value.

A large variety of results referring to IgG values exists as well as sub-classes of IgG, IgA, etc. and their relationship with asthma or recurrent sibilants in early childhood. It is evident that they play some role in the clinical expression of this pathology in the ages considered, however, this role is still as yet unclear.

From a sample of 37 children, Smith discovered lower levels of IgG sub-classes (IgG1, IgG2, IgG4) in intrinsic and extrinsic asthmatic children, whose proportion was higher in children who had lower initial IgG values[16], although a normal concentration of IgG

in serum does not exclude the possibility of a deficit in sub-classes. Smith suggested the possibility of an etiological role in the respiratory symptoms of some dhildren who showed deficits of the sub-classed IgG mentioned above.

A connection between the IgG sub-class deficiency and bronchitis symptoms in early childhood has recently been found[17]. This is more frequent in these age groups than later, suggesting a transitory immaturity of immunity. The etiology of recurrences is often viral in these age groups than in later age groups, meaning that in predisposed children a lower IgG "protective" activity could be involved (neutralization, complementary activation and phagocytosis).

In another recent sutdy of 42 children age between 9 months and 6 years, lower levels of IgG4 were found in children under 24 months than in the control group. From 2 to 6 years, IgE was found to be increased, while IgG3 and IgG4 ($p < 0.05$) were lower compared to the controls[18], suggesting a relationship between lower levels of IgG sub-classes and sibilants in early childhood.

In Brazilian children aged 7 to 15, lower IgA levels and sub-classes of IgG were found in the group of severe asthmatics, with an IgG3 deficiency being predominant, although this does not seem to be a reliable predictor for the development of infections in this children[19].

In Japanese children the prevalence of deficit in IgG sub-classes was 31.6 % in breast fed children of 6 to 24 months and 26.7 % in controls. There were no significant differences, so the conclusion was reached that sibilants in the breast fed child are not linked to a deficiency in IgG sub-classes, although there exists a link between recurrent sibilants and concentration of IgG sub-classes[20].

In older Asian children with asthma (average 7.5 years), no deficiency in IgG sub-classes was found. Only IgE levels were four times higher than control subjects[21], suggesting that at these ages there is greater participation of IgE responder as opposed to environmental allergens and a larger maturation of IgG responder.

IgD levels were found to be lower in children with the first signs of atopic asthma ($p < 0.001$), although these were normal in the course of the following 18 months[22]. Is this a non- specific response or an attempt to block the immune response in asthma favouring tolerance?

The transitory absence of IgA in saliva during the first years of life is linked to an increased risk of bronchial hyperactivity[23] BHR, but no link was found between this IgA transitory deficiency and the clinical diagnosis of asthma, which would favour the thesis of independence of these two clinical frameworks.

IgA and IgD levels were found to be lower both in patients with intrinsic and extrinsic asthma[24], compared with the control group. IgA was found quite high in smokers. Women had significantly higher IgA levels compared to men. Both IgG and IgA levels increased with age (14 and 24).

From a sample of 315 apparently healthy males aged between 19 and 22, D'Amelio found no relation between clinical history or IgE levels and the remaining Ig classes[25], as occurred in our study, where no link between IgE levels and the remaining IgE classes was found in such low age groups as those of our patients.

Those respiratory and non respiratory pathologies where different deficits of IgG[26] sub-classes are found are very diverse and their exact interpretation and comprehension has yet to be fully understood.

In addition, in the broncoalveolar lavage (BAL) in asthmatic patients, the IgG sub-classes were significantly higher than in healthy control subjects[27], showing a local production of 1 or more sub-classes and an increased exudance of same.

The predominant Ig in BAL is IgA which seems to be produced locally, finding significant statistics for concentration of sIgA and IgA I patients with allergic rhinitis[28]. The authors suggest the possibility of some role played by this Ig in the mediation of the atopic illness. When we study the influence of the maternal immune response in a new born baby and in the early years of life we find some data of interest. The Ac. as opposed to the IgE maternal allergens, are consistently linked to the presence of asthma in the mother, while those IgG class Ac. are linked to maternal rhinitis and eczema in the chils[29].

Moreover, treatment with intravenous Ig in patients with severe asthma often leads to clinical improvement, reduction in the need for corticoids and a decrease of cutaneous reactivity in prick-test[30]. Finally, it is important to understand the present situation regarding "hygiene hypothesis" in the development and management of bronchial asthma [31]. Good hygiene in the early years is linked to a lesser prevalence of atopic illness. In a study carried out in Denmark on an adult population and considering seropositivity for Hepatitis A, Helicobacter pylori and Toxoplasma gondii as markers of poor hygiene, a link was found between positivity to 2 of these markers and low prevalence of atopy, while colonization of the intestinal tract by potentially pathogenic bacteria is associated with high prevalence of atopy[32].

As regards infection by certain pathogens and their relation to atopic illness, new data appears again.

Bjronsson[33] et al, find a link between recent infection (IgM > 1/16) and/or previous infection (IgG < 1/512 and > 1/32) per Ch1. Pneumonic and the presence of

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sibilants. Later, Mill[34] et al, did not find any positive link between suggestive symptoms of asthma and the two previous years and Ac. IgG as opposed to C. Pneumonia.

In the population age group of our study where it is known that there exists at least 3 sub-groups of patients with recurring sibilants[4], the development of immune response must be an important factor in the clinical evolution of the illness and possibly, at the same time, participate in the genesis of same, if current evidence is taken into account. Those patients whose initial immune state is better would expect to show improved functional respiratory recovery with age considered. Perhaps the neutralization of the virus, which is responsible for reccurrences in early childhood, would be more effective in those children who finally reach greater functional recovery, In these cases, mediation of eicosanoids coming from alveolar macrophages is known, by which treatment with montelukast could have an association with this response.

In any case, it would be useful to design a study with a larger population, greater follow-up period with control in series of IgG concentrations, to evaluate if these could aid prognosis of the disease, either in the short or long term depending on what type of treatment is established.

CONCLUSIONS

The sex of the patient did not influence results. IgG values increase with age being quantified in our case in 7 mg/ml of increase for each month that age increases. The children with the highest IgG values before treatment began reached the highest PEF values after being treated with 4 mg of MONTELUKAST daily for 8 weeks (10 l/min of PEF increase for each 100 mg/ml of increase in IgG value).

The literature does not clarify the mechanisms implicated in these results. There exists various data which certify the implication of these Ig in the asthmatic process, which seems greater in breast fed and small children. Perhaps a possible explanation could be greater protection as opposed to precipitating factors of asthmatic symptoms, of viral and bacterial type, which their association would have greater impact on age groups more affected by these precipitated factors.

REFERENCES

1. Djukanovic R, Wilson JW, Britten KM et al. Quantitation of mast cell and eosinophils in the bronchial mucosa of symptomatic atopic asthmatics and healthy control subjects using immunohistochemistry. Am Rev Resp Dis 1990;142:863-71.

2. Hamid QA, Minshall EM. Molecular pathology of allergy disease lower airways. J Allerg Cin Immunol 2000;105:20-36.

3. Young S, O'Keeffe PT, Arnot J et al. Lung function, airway responsiveness and respiratory symptoms before and after bonchiolitis. Arch Dis Child 1995;72:6-24.

4. Martínez FD, Helner PJ. Types of asthma and wheezing. Eur Resp J 1998;12:53-8.

5. Holtzman MJ. Arachidonic acid metabolism. Am Rev Resp Dis 1991; 143:188-203.

6. Barnes NC, Smith LJ. Biochemistry and physiology of the leukotriens. Clin Rev Allergy Immunol 1999;17:27-42.

7. Bandeira Melo C, Hall JC, Penrose JM, Weller PF. Cysteinil leukotrienes induce IL-4 release from cord blood derived human eosinophils. J Aller Clin Immunol 2002;109:975-9.

8. Tohda Y, Nakahara H, Kubo H, Haraguchi R, Fukuoka M, Nakajima G. Effects of ONO-1078 on cytokine production in peripheral blood mononuclear cells of patients with bronchial asthma. Cli Exp Allergy 1999;29:1532-6.

9. Lexikauf GA, Claesson HE, Doupnik CA, Hybbinete S, Grafstrom RC. Cysteinil leukotrienes enhance growth of human airway epithelial cells. Am J Physiol 1990;259:1255-61.

10. Medina L, Pérez Ramos J, Ramírez R, Selman M, Pardo A. Leukotriene C4 upregulates collagenase expression and synthesis in human lung fibroblasts. Bioch Biophys Acta 1994;1224:168-74.

11. Stata corp. 20001. Stata statistical softw: Release 7.0 college station, TX: Stata corp.

12. Rauer U, Freund R. Normal levels of immunoglobulins in childhood. Monatrschr Kindreheikd 1969;117(9):559-63.

13. Berg T. The Immunoglobulin development during the first year of life. A longitudinal study. Acta Pediatr Scand 1969;58(3): 229-36.

14. Stevens WJ, de Backer W, Vermeire PA. Serum IgA, IgG, IgM and IgD in allergic (type I) and non-allergic respiratory disease. Clin Allergy 1983;13(1):11-9.

15. Stoica G, Macarie E, Michiu V, Stoica RC. Biologic variation of human immunoglobulin concentration.I.sex-age specific effects on serum levels of IgG, IgA, IgM and IgD. Med Interne 1980;18(3):223-32.

16. Smith TF, Morris EC, Bain RP. IgG subclasses in nonallergic children with chronic chest symptoms. J Pediatr 1984;105(6): 896-900.

17. De Baets F, Kint J, Pauwels R, Leroy J. IgG subclass deficiency in children with recurrent bronchitis. Eur J Pediatr 1992; 151(4):274-8.

18. Cesur Y, Uner A, Arslan S. FERUM immunoglobulins and Immunoglobulin G subclases with recurrent wheezing. Indian J Pediatr 2000;67(12):861-4.

19. Diogo CL, Kirschfink M, Grumach AS. Immunoglobulin G subclases concentrations and infections in children and adolescents with severe asthma. Pediatr Allergy Immunol 2002; 13(3):195-202.

20. Karamau O, Uguz A, Uzuner N. Immunoglobulin G subclases in wheezing infants. Acta Paediatr Jpn 1998;40(6):564-6.

21. Lee BW. Immunoglobulin (Ig) and IgG subclasses in Asian children with bronchial asthma. Ann Trop Paediatr 1995;15(4): 280-4.

22. Salpietro DC, Masarachio A, Turiaco A, di Bella MR, Toscano V, Merlino MV. Serum IgD levels in children with atopic asthma. A longitudinal study. Minerva Pediatr 2001;53(1):1-5.

23. Gleeson M, Clancy RL, Hensley MJ, Cripps AW, Henry RL, Wlodarezyk JH, Gibson PG. Development of bronchial hyperreactivity following transient absence of salivary IgA. Am J Resp Crit Care Med 2003;153(6):1785-9.

24. Jolliff CR, Cost KM, Stivrius PC, Grossman PP, Nolte CR, Franco SM et al. Reference intervals for serum IgG, IgA, IgM, C3 and C4 as determined by rate nephelometry. Clin Chem 1982;28(1):126-8.

25. D'Amelio R, Fattorossi A, Berti R, Rossi P, Paganelli R, Castaglioulo PP. Serum IgG, IgA, IgM, IgE, salivary IgA levels and lung function in subjects in a healthy male population from the Italian Air Force: A preliminary study. Ann Allergy 1984; 53(5):432-5.

26. Hanson LA, Soderstrom R, Avanzini A, Beughtsson U, Bjorkander J, Soderstrom T. Immunoglobulin subclasses deficiency. Ped Inf Dis J 1998;7(5):17-21.

27. Out TA, Van der Graaf EA, Van den Berg NJ, Janse HM. IgG subclasses in bronchoalveolar lavage fluid from patients with asthma. Scand J Immunol 1991;33(6):719-27.

28. Peebles RS Jr, Liu MC, Lichtenstein LM, Hamilton RG. IgA, IgG and IgM quantification in bronchoalveolar lavage fluids from allergic rhinitis, allergic asthmatics and normal subjects by monoclonal antibody-based immunoenzymatic assays. J Immunol Methods 1995;179(1):77-86.

29. Platts-Mills TA, Erwin EA, Allison AB, Blumenthal K, Barr M, Sredl D, Burge H, Gold D. The relevance of maternal immune responses to inhalant allergens to maternal symptoms, passive transfer to the infant and development of antibodies in the first 2 years of life. J Allergy Clin Immunol 2003;111(1):123-30.

30. Schuster A, Wahn V. Intravenous immunoglobulins in bronchial asthma: a therapeutic alternativ? Infusionsther. Transfusionsmed 1993;20(1);141-4.

31. Liu AH, Szefler SJ. Advances in childhood asthma: Hygiene hypothesis, natural history, and management. J Allergy Clin Immunol 2003;111:785-92.

32. Linneberg A, Ostergaard CH, Tvede M, Andersen LP, Nielsen NH, Madsen F et al. J Allergy Clin Immunol 2003;111:847-53.

33. Bjornsson E, Hjelm, Janson C, Fridell E, Borman G. Serology of Chlamydia in relation to asthma and bronchial hyperrresponsiveness. Scand J Infect Dis 1996;28(1):63-9.

34. Mill GD, Lindeman JA, Fawcett SP, Herbison GP, Sears MR. Chlamydia Pneumonie serological status is not associated with asthma in children or young adults. Intl J Epidemiol 2002;29(2):280-4.