



Boletín Médico del Hospital Infantil de México

www.elsevier.es/bmhim



RESEARCH ARTICLE

Clinical characterization and antimicrobial resistance of *Escherichia coli* in pediatric patients with urinary tract infection at a third level hospital of Quito, Ecuador



CrossMark

David Garrido^{a,*}, Santiago Garrido^a, Miguel Gutiérrez^b, Leonel Calvopiña^c, Amani Sunday Harrison^a, Michelle Fuseau^d, Ramiro Salazar Irigoyen^e

^a Facultad de Ciencias Médicas, Universidad Central del Ecuador, Quito, Ecuador

^b Servicio de Pediatría, Hospital de Especialidades de las Fuerzas Armadas N.^o 1, Quito, Ecuador

^c Laboratorio de Microbiología, Hospital de Especialidades de las Fuerzas Armadas N.^o 1, Quito, Ecuador

^d Hospital de Especialidades de las Fuerzas Armadas N.^o 1, Quito, Ecuador

^e Laboratorio de Microbiología, Facultad de Ciencias Médicas, Universidad Central del Ecuador, Quito, Ecuador

Received 29 September 2016; accepted 7 February 2017

Available online 8 July 2017

KEYWORDS

Urinary tract infections;
Pediatrics;
Escherichia coli

Abstract

Background: Urinary tract infections (UTI) are among the most common infections in pediatric patients. The main etiopathogenic agent is *Escherichia coli*. The purpose of this study was to determine the antimicrobial resistance pattern of *E. coli* in pediatric patients and to understand their main clinical and laboratory manifestations.

Methods: Fifty-nine patients were included in the study and classified into two groups: hospitalization (H) and external consultation (EC). Every patient presented urine cultures with the isolation of *E. coli* that included an antibiogram. Clinical signs and symptoms, urinalysis, complete blood count (CBC) and serum inflammatory markers were analyzed.

Results: The most common clinical manifestations were fever (H: 76.5%; EC: 88%), vomiting (H: 32.4%; EC: 32%), hyporexia (H: 20.6%; EC: 16%), abdominal pain (H: 20.6%; EC: 28%), and dysuria (H: 14.7%; EC: 32%). Ten patients (16.95%) presented UTI for extended spectrum beta-lactamase (ESBL) *E. coli*. Ampicillin, nalidixic acid, and trimethoprim-sulfamethoxazole showed a higher resistance rate, being ampicillin the most significant (H: 88.2%; EC: 92%). Leukocyturia, bacteriuria and urine nitrites were frequent alterations in urinalysis (H: 52.9%; EC: 92%). In ESBL *E. coli* patients, a positive correlation was found between leukocytes in CBC and C-reactive protein ($r = 0.9$, $p < 0.01$). Diarrhea and foul-smelling urine were associated with *E. coli* resistance.

* Corresponding author.

E-mail address: david_labinmuno@hotmail.com (D. Garrido).

Conclusions: The presence of leukocytes, bacteria, nitrites and the Gram stain are the most common indicators. Nitrofurantoin and phosphomycin are good therapeutic options. However, an antibiogram must be conducted to determine the best therapeutic agent.

© 2017 Hospital Infantil de México Federico Gómez. Published by Masson Doyma México S.A. This is an open access article under the CC BY-NC-ND license (<http://creativecommons.org/licenses/by-nc-nd/4.0/>).

PALABRAS CLAVE

Infección del tracto urinario;
Pediatría;
Escherichia coli

Caracterización clínica y resistencia antimicrobiana de *Escherichia coli* en pacientes pediátricos con infección del tracto urinario en un hospital de tercer nivel en Quito, Ecuador

Resumen

Introducción: Las infecciones de tracto urinario (ITU) se encuentran entre las más frecuentes en pediatría, siendo *Escherichia coli* el principal agente etiopatogénico. El objetivo de este estudio fue determinar el patrón de resistencia antimicrobiana de *E. coli* en pacientes pediátricos y conocer sus principales manifestaciones clínicas y de laboratorio.

Métodos: Se incluyeron en el estudio 59 pacientes en dos grupos: hospitalización (H) y consulta externa (CE). En cada uno se realizó un urocultivo y un antibiograma con aislamiento de *E. coli*. Se analizaron signos y síntomas, uroanálisis, hemograma y marcadores séricos de inflamación.

Resultados: Las manifestaciones clínicas más frecuentes fueron fiebre (H: 76.5%; CE: 88%), vómito (H: 32.4%; CE: 32%), hiporexia (H: 20.6%; CE: 16%), dolor abdominal (H: 20.6%; CE: 28%) y disuria (H: 14.7%; CE: 32%). Diez pacientes (16.95%) presentaron ITU por *E. coli* beta-lactamasa de espectro extendido (BLEE). La ampicilina, ácido nalidíxico y trimetroprim con sulfametoxazol mostraron alta resistencia, siendo ampicilina la más significativa (H: 88.2%, CE: 92%). Leucocituria, bacteriuria y nitritos en orina fueron frecuentes en el uroanálisis. En pacientes con *E. coli* BLEE se encontró una correlación positiva entre los leucocitos y la proteína C reactiva ($r = 0.9$, $p < 0.01$). La diarrea y el mal olor en la orina se asociaron con resistencia de *E. coli*.

Conclusiones: La leucocituria, la bacteriuria, los nitritos y la tinción Gram son los indicadores más frecuentes de ITU. La nitrofurantoina y fosfomicina son buenas opciones terapéuticas. Sin embargo, debe realizarse un antibiograma para determinar el mejor tratamiento.

© 2017 Hospital Infantil de México Federico Gómez. Publicado por Masson Doyma México S.A. Este es un artículo Open Access bajo la licencia CC BY-NC-ND (<http://creativecommons.org/licenses/by-nc-nd/4.0/>).

1. Introduction

Urinary tract infections (UTIs) are diseases caused primarily by bacteria, which migrate from the perineal region and proliferate in the urethra, bladder, ureters or kidneys.¹ However, migration may take place by hematogenous dissemination, direct contamination in medical procedures or traumas, or even through the bacterial translocation from the rectum to the urethra.² The risk of urinary tract infection in the first ten years of life is 1% for males and 3% for females.³ Principal risk factors include constipation, family and personal history of urinary tract infections, improper hygiene, failure to thrive, and anorexia.⁴ The recurrence of UTIs is common (15-20%), especially in the first year after an initial occurrence and the risk increases with the number of prior occurrences (up to 60-75% of the cases with three or more occurrences).⁵ The absence of a diagnosis upon the occurrence of a urinary tract infection or its poor management may favor irreversible kidney damage.⁶

The diagnosis of UTIs with safety and reliability is essential to any treatment plan and its subsequent follow-up, taking into consideration that a poor diagnosis may have repercussions on the future life of the child and cause a burden from unnecessary examinations in the child, his family, and the health system.^{7,8} The estimated direct cost of the treatment that UTIs may require is approximately 6 billion dollars annually on health services worldwide.⁹

Appropriate clinical assessment of pediatric patients would be of great assistance in controlling infant morbidities and avoiding the possible complications of recurrent infections.¹⁰

The purpose of this study was to determine the pattern of antimicrobial resistance presented by *Escherichia coli* in pediatric patients admitted to the Department of Pediatrics, Hospital de Especialidades de las Fuerzas Armadas No. 1 (HE-1), and to identify the main clinical and laboratory manifestations of the UTIs. These results could provide general practitioners and pediatricians with statistical data for an accurate diagnostic of UTIs.

2. Methods

We carried out an epidemiological, observational, descriptive case series study at the HE-1 in the city of Quito, Ecuador, from January to September 2015. We collected urine samples from 132 patients between 3 months and 14 years of age with clinical features suggestive of UTI. Seventy-three subjects were excluded since their respective cultures did not show *E. coli* growth, or because they were on antibiotic treatment before the sample collection. Neoplastic disease was also an exclusion criterion. The diagnosis of UTI was proposed by the medical history and physical examination of the patients. These data, together with a urine test (which was confirmed with a positive urine culture), suggested wherein a pathogen was isolated. Fifty-nine patients from the Department of Pediatrics participated in the study; these subjects were classified into two groups: hospitalization (H) and external consultation (EC). Based on the recommendations of the Instituto Nacional de Investigación en Salud Pública [National Health Research Institute] (INSP) and the regulations of HE-1, different antibiotic disks in the antibiogram were tested on each group. However, in antibiograms positive for extended spectrum beta-lactamase (ESBL) *E. coli*, all required disks were tested. The elemental and microscopic urine and Gram stain tests were performed on all samples. All blood samples were taken for a complete blood count (CBC), C-reactive protein (CRP) and erythrocyte sedimentation rate (ESR) tests.

The Ethics Committee of the Universidad Central del Ecuador [Central University of Ecuador] supervised the study. All participants signed informed consents and were informed of the research applications that the results from their samples would have.

2.1. Urine sample collection

After washing the periurethral and perineal regions, urine samples were collected from each patient (under nursing supervision). Preferably, the sample was collected in the morning or with a 4-hour interval since the last micturition. In children younger than two years of age, the urine sample was taken through catheterization; in children older than two years of age, it was collected from a clean mid-stream urinary sample. The samples were kept and processed by the clinical and microbiological laboratories of the HE-1.

2.2. Blood sample collection

Peripheral blood samples were taken by venopuncture before the initiation of antimicrobial therapy. Two samples per patient were taken in red and lilac cap tubes and were conserved and processed in the HE-1 Clinical Laboratory.

2.3. Urinalysis

The rapid urinary screening tests are urinary sediment, dipstick, and Gram stain. The dipstick test includes pH, proteins, glucose, ketones, blood, bilirubin, urobilinogen, nitrites, leukocyte esterase and urine density. With the urine sediment test, the following data were obtained:

bacteria, nitrites, erythrocytes, hemoglobin, epithelial cells, and leukocytes.

2.4. Urine cultures

Urine cultures and antibiogram tests of the urine samples of all patients with a suspected UTI diagnosis were performed; these samples were processed according to the international standards from the Microbiology Laboratory of the HEP-1. Various antibiotic disks were tested in both groups of patients. All patients whose cultures contained the minimum quantity of bacteria were included in the study. The results obtained were recorded in the hospital software.

Methods used for the antibiogram were both automated (Vitek compact 2) and disk diffusion (Kirby-Bauer) by the NCCL 2015, both for the minimum inhibitory concentration (MIC) and for the diameter of the inhibition halo.

HE-1 Microbiology Laboratory determines the presence of ESBL strains using various methods: identifying the species, testing broad panels of antibiotics and investigating rare phenotypes.

The double disk diffusion technique is based on double disk synergy. Briefly, a Müller-Hinton agar plate is inoculated with a bacterial suspension; then, cephalosporin and β-lactamase disks are placed at a certain distance (30 mm or 20 mm for more sensitivity) from the clavulanic acid. If there is an increase in the space between the inhibition halos of some of the antibiotics and the disk with the β-lactamase inhibitor, an ESBL is deemed to exist. In our case, the presence of ESBL was confirmed when the inhibition halo was equal to 5 mm on cephalosporin alone.

For the E-test technique, paper strips impregnated with antibiotics were used. One-half contained cephalosporin in decreasing concentrations, and the other half contained cephalosporin in decreasing concentrations with clavulanic acid in a fixed concentration of 2 µg. The synergy of these antibiotics was considered positive when the MIC decreased in two or more dilutions.

2.5. Analysis of clinical histories

The clinical history number and UTI diagnosis were registered using the logbooks from the Pediatrics Department. Each clinical history was reviewed, and the software from the HE-1 was used to record the data from the anamnesis of each patient. The clinical signs or symptoms were registered to detect the most common clinical manifestations.

The collection of the microbiological and antibiogram data of each patient was performed with the assistance of the Microbiology Laboratory personnel.

2.6. Statistical analysis

Data were recorded and analyzed. Analyses were performed using SPSS software and Excel, both in their latest versions for Windows 10. The qualitative variables were compared to each other using a Pearson correlation. The comparison between the quantitative and qualitative variables was carried out with an odds ratio (OR). We calculated the risk

Table 1 Clinical manifestations of patients with urinary tract infection.

| | H | EC |
|--|-------------|-------------|
| <i>Group size (n)</i> | 34 | 25 |
| <i>Male [n (%)]</i> | 7 (20.59) | 2 (8) |
| <i>Age in years [mean (SD)]</i> | 4.23 (3.90) | 3.35 (3.57) |
| <i>Urinary clinical manifestations</i> | | |
| Hematuria [n (%)] | 1 (2.9) | 0 (0) |
| Pollakiuria [n (%)] | 3 (8.8) | 3 (12) |
| Foul-smelling urine [n (%)] | 5 (14.7) | 3 (12) |
| Dysuria [n (%)] | 5 (14.7) | 8 (32) |
| Urinary urgency [n (%)] | 2 (5.9) | 3 (12) |
| <i>Non-urinary clinical manifestations</i> | | |
| Fever [n (%)] | 26 (76.5) | 22 (88) |
| Headache [n (%)] | 2 (5.9) | 0 (0) |
| Lower back pain [n (%)] | 1 (2.9) | 2 (8) |
| Congestive/erythematous oropharynx [n (%)] | 13 (38.2) | 10 (40) |
| Cough [n (%)] | 2 (5.9) | 3 (12) |
| Abdominal pain [n (%)] | 7 (20.6) | 7 (28) |
| Vomiting [n (%)] | 11 (32.4) | 8 (32) |
| Irritability [n (%)] | 6 (17.6) | 8 (32) |
| Diarrhea [n (%)] | 6 (17.6) | 5 (20) |
| Petechiae [n (%)] | 1 (2.9) | 0 (0) |
| Palpebral edema [n (%)] | 1 (2.9) | 0 (0) |
| Hyporexia [n (%)] | 7 (20.6) | 4 (16) |
| Genital erythema [n (%)] | 2 (5.9) | 3 (12) |
| Genital secretion [n (%)] | 0 (0) | 2 (8) |
| Convulsions [n (%)] | 2 (5.9) | 1 (4) |
| <i>Associated factors</i> | | |
| Prior UTI [n (%)] | 12 (35.3) | 10 (40) |
| Low weight < -2SD [n (%)] | 10 (29.4) | 2 (8) |
| Stunted growth < -2SD [n (%)] | 8 (23.5) | 4 (16) |
| Constipation [n (%)] | 5 (14.71) | 3 (12) |

H, hospitalization; EC, external consultation; SD, standard deviation; UTI, urinary tract infection.

association between clinical findings and the resistance pattern of *E. coli*.

3. Results

3.1. Clinical and laboratory findings

Clinical and laboratory data from patients were registered (**Tables 1 and 2**). Females presented UTIs with a higher frequency (H: 79.41%, EC: 92%), under five years of age (H: 4.23, EC: 3.35) in both groups hospitalization (H) and external consultation (EC). The most common urinary and non-urinary clinical manifestations observed were fever (H: 76.5%, EC: 88%), congestive oropharynx (H: 38.2%, EC: 40%), vomiting (H: 32.4%, EC: 32%), hyporexia (H: 20.6%, EC: 16%), abdominal pain (H: 20.6%, EC: 28%), dysuria (H: 14.7%, EC: 32%), and foul-smelling urine (H: 14.7%, EC: 12%). It is important to take into consideration that in the EC group, a big percentage of patients showed irritability (32%). **Table 3** shows the frequency of the

Table 2 Laboratory results from patients with urinary tract infections.

| | H | EC |
|--|-------------|----------|
| <i>Group size (n)</i> | 34 | 25 |
| <i>Urinalysis</i> | | |
| Cloudy urine [n (%)] | 32 (94.12) | 25 (100) |
| Positive nitrites [n (%)] | 18 (52.9%) | 14 (56%) |
| Infectious Gram [n (%)] | 24 (70.6%) | 24 (96%) |
| White blood cells > 5/field [n (%)] | 28 (82.35%) | 23 (92%) |
| Red blood cells > 4/field [n (%)] | 13 (38.24%) | 9 (36%) |
| Low epithelial cells > 5/field [n (%)] | 4 (11.76%) | 6 (24%) |
| Bacteria > 358/ μ l [n (%)] | 30 (88.24%) | 21 (84%) |
| <i>Complete blood count</i> | | |
| Hemoglobin < 11.8 [n (%)] | 11 (32.4%) | 10 (40%) |
| Leukocytes > 12000 [n (%)] | 20 (58.8%) | 12 (48%) |
| Neutrophils > 7000 [n (%)] | 17 (50%) | 15 (60%) |
| Lymphocytes > 6250 [n (%)] | 4 (11.8%) | 7 (28%) |
| Monocytes > 850 [n (%)] | 19 (55.9%) | 14 (56%) |
| <i>Inflammatory markers</i> | | |
| ESR > 15 [n (%)] | 10 (29.4%) | 10 (40%) |
| CRP > 5 [n (%)] | 10 (29.4%) | 12 (48%) |

H, hospitalization; EC, external consultation; SD, standard deviation; ESR, erythrocyte sedimentation rate; CRP, C reactive protein.

Table 3 Altered parameters on urinalysis, complete blood count and inflammatory markers.

| | H | EC |
|-----------------------------|------------|-----------|
| <i>Urinalysis</i> | n (%) | n (%) |
| 0 | 2 (5.9%) | 0 (0%) |
| ≥ 1 | 32 (93.1%) | 25 (100%) |
| <i>CBC</i> | | |
| 0 | 3 (8.8%) | 0 (0%) |
| ≥ 1 | 31 (91.2%) | 25 (100%) |
| <i>Inflammatory markers</i> | | |
| 0 | 18 (52.9%) | 8 (32%) |
| ≥ 1 | 12 (47.1%) | 12 (48%) |

CBC, complete blood count; H, hospitalization; EC, external consultation.

number of clinical signs and alterations in the laboratory parameters that patients presented in the study.

3.2. Antimicrobial resistance of *E. coli*

Table 4 indicates the percentage of resistance of *E. coli* found in the antibiogram. Ten patients (16.95%) were found with ESBL *E. coli* UTIs. The antibiogram for third generation cephalosporins for patients from the EC group was only performed in cultures with a high probability of β lactamase-producing strains. Five patients (EC: 20%) were corroborated with ESBL *E. coli*.

Table 4 Antimicrobial resistance of *E. coli*.

| | H (%) | 95% CI | EC (%) | 95% CI |
|---------|-------|-------------|--------|-------------|
| AMP | 88.2 | 73.4-95.33 | 92 | 75.03-97.78 |
| SAM | 29.4 | 16.83-46.17 | — | — |
| TZP | 14.7 | 6.45-30.13 | — | — |
| FEP | 14.7 | 6.45-30.14 | — | — |
| CRO | 14.7 | 6.45-30.15 | — | — |
| CXM | 14.7 | 6.45-30.16 | 20 | 8.86-39.13 |
| CZO | 20.6 | 10.35-36.80 | 48 | 30.03-66.50 |
| LVX | 35.3 | 21.49-52.09 | — | — |
| NAL | — | — | 68 | 48.41-82.79 |
| TMP-SMX | 61.8 | 45.04-76.10 | 84 | 66.35-93.60 |
| NIT | 2.9 | 0.52-14.92 | 4 | 0.71-19.54 |
| GEN | 0 | 0 | 12 | 4.17-42.77 |
| IPM | 0 | 0 | — | — |
| FOS | — | — | 4 | 0.71-19.54 |

AMP, ampicillin; SAM, ampicillin/sulbactam; TZP, piperacillin/tazobactam; FEP, cefepime; CRO, ceftriaxone; CXM, cefuroxime; CZO, cefazolin; LVX, levofloxacin; NAL, nalidixic acid; TMP-SMX, trimethoprim/sulfamethoxazole; NIT, nitrofurantoin; GEN, gentamicin; IPM, imipenem; FOS, fosfomycin; H, hospitalization; EC, external consultation; CI, confidence interval.

3.3. Comparison of qualitative variables

The comparisons between data from the urinalysis and CBC and serological studies, and between weight and height by weight were non-statistically significant. However, a direct correlation between the number of leukocytes in the CBC and the quantity of CRP ($r=0.39$, $r^2=0.15$, $p<0.01$), and between the percentage of lymphocytes from CBC and the number of lymphocytes in urine ($r=0.29$, $r^2=0.08$, $p<0.05$) was found. Also, there was an inverse relationship between the number of bacteria in urine and the percentage of monocytes in CBC ($r=-0.26$, $r^2=0.01$, $p<0.05$); however, these correlations were weak.¹⁰

The same correlation was found between leukocytes in CBC vs. CRP ($r=0.9$, $r^2=0.81$, $p<0.01$) in patients who presented ESBL *E. coli*. In these patients, the quantity of CRP ($r=0.67$, $r^2=0.44$, $p<0.05$) was greater at higher percentiles.

3.4. Temporal association between the risk of antimicrobial resistance and clinical findings

We compared clinical variables obtained from the patients with the antimicrobial resistance of *E. coli* in the H group, and we found 30 times more risk of carrying a strain resistant to aminopenicillin as a cause of UTI in patients who presented a previous cough (OR 31; CI 95% 1.02-94.0). On second generation cephalosporins, it was shown that patients with foul-smelling urine had 20 times more risk of having a UTI caused by a strain resistant to this family of antibiotics (OR 20.25; CI 95% 2.04-20.8). Similarly, it was found that patients who presented foul-smelling urine had a nine-time higher risk of being carriers of strains resistant to first generation cephalosporins isolated in urine cultures (OR 9.38; CI 95% 1.17-74.84). Another symptom that indicated an association with the resistance to antibiotics was diarrhea since it was observed that patients with this symptom had

13-times higher risk of being carriers of strains resistant to ureidopenicillins + β -lactamase inhibitor (BLI) (OR 13; CI 95% 1.51-111.8). Regarding fever, it was shown that in patients with this symptom the risk of presenting strains resistant to nitrofurans decreased by 24% (OR 0.76; CI 95% 0.63-0.92).

In the EC group, no significant association was found between resistance to antimicrobials and clinical symptoms.

4. Discussion

UTIs and respiratory tract infections are the most common pediatric bacterial diseases.¹¹ It is estimated that between 8-10% of female and 2-3% of male patients will have a symptomatic UTI.⁵ The worldwide prevalence of pediatric UTIs is estimated at 5%, with an annual incidence of 3.1/1,000 girls (0-14 years) and 1.7/1,000 boys (0-14 years).¹²

After the neonatal period, the most common uropathogens are *E. coli*, *Klebsiella*, *Proteus*, *Enterobacter*, *Citrobacter*, *Staphylococcus saprophyticus* and *Enterococcus*,¹³ while group B streptococcal infections occur with greater frequency in newborns.²

In this study, we found that UTIs are clinically non-specific, which is consistent with the findings presented in other publications, even stating that making a UTI diagnosis based upon clinical manifestations has a 33% margin of error.^{14,15} Our data agree with other reports, where fever, abdominal pain, vomiting, dysuria, foul-smelling urine, and irritability are reported as frequent signs and symptoms of UTIs.^{4,16} In the urine sample study, we found that the presence of leukocytes, bacteria, nitrites and the Gram stain are the most common indicators of UTI and should be analyzed together. However, the absence of any of these does not exclude the diagnosis; therefore, urine culture is still the gold standard test.^{17,18}

Ampicillin is the antibiotic for which we identified the most resistance, which is similar to other studies wherein resistance between 50-90% is reported. We have also found that there is resistance exceeding 20% for fluoroquinolones, first generation cephalosporins, trimethoprim/sulfamethoxazole and nalidixic acid, which is also consistent with other studies.^{19,20} Bacterial resistance to aminoglycosides, nitrofurantoin, and phosphomycin was low, which is consistent with the Ecuadorian National Bacterial Resistance Network (REDNARBEC).^{19,21} The percentage of ESBL *E. coli* is similar to other reports, between 5.5-33.3%.^{9,19,22} However, these values are high in comparison to a study reporting ESBL *E. coli* in 7.5% of the Pediatrics service.²³ According to the antimicrobial resistance pattern found, the pharmacological groups that should be avoided as empirical therapy in this population are aminopenicillins, aminopenicillins+BLI, first generation cephalosporins, quinolones, fluoroquinolones, and folate pathway inhibitors.²⁴

This study has important limitations that should be considered; one of them is the reduced number of patients. Based on this, it is necessary to increase the number of patients. Another limitation is the fact that we only considered data from our hospital for the analysis. It is important to use data from other health institutions for further investigations and to make a consensus for a better understanding of UTIs in our population. Also, it would be interesting to

analyze the association between clinical or laboratory data and the time of hospitalization of the patients, or the association of ESBL *E. coli* with hospitalization length of stay.

In conclusion, and according to our findings and the increasing incidence of infections by broad spectrum beta-lactamase producing strains, we confirm the importance of not starting antibiotic therapy without a prior microbiological culture, especially in patients at risk of ESBL *E. coli* infection.²⁵ However, initiation of empiric nitrofurantoin or phosphomycin therapy is a good option in the event of suspicion of cystitis, both in hospitalization and in outpatient services.^{26,27} When clinical manifestations suggesting gravity or upper urinary tract infection are present, therapy with aminoglycosides may be used, as they are also effective against ESBL *E. coli*.^{28,29}

Ethical responsibilities

Protection of human and animal subjects. The authors declare that no experiments were performed on humans or animals for this study.

Confidentiality of data. The authors declare that they have followed the protocols of their work center on the publication of patient data.

Right to privacy and informed consent. The authors have obtained the written informed consent of the patients or subjects mentioned in the article. The corresponding author is in possession of this document.

Funding

This study was financed using the funds of the investigators.

Conflict of interests

The authors declare no conflicts of interest of any nature.

Acknowledgements

We would like to thank the residents of the HE-1 Department of Pediatrics: Mónica Tonato, Carolina Paucarima, Óscar Yanez, Veronica Torres, and Sofía Jiménez for their assistance in collecting clinical data, and Dr. Guillermo Fuenmayor for his valuable comments. We also thank the HE-1 Microbiology Laboratory for conducting the urine cultures, which were the basis for this research.

References

1. Foxman B. Urinary tract infection syndromes: occurrence, recurrence, bacteriology, risk factors, and disease burden. *Infect Dis Clin North Am.* 2014;28:1-13.
2. Ma JF, Shortliffe LD. Urinary tract infection in children: etiology and epidemiology. *Urol Clin North Am.* 2004;31:517-26.
3. Calderón-Jaimes E, Casanova-Román G, Galindo-Fraga A, Gutiérrez-Escoto P, Landa-Juárez S, Moreno-Espinosa S, et al. Diagnóstico y tratamiento de las infecciones en vías urinarias: un enfoque multidisciplinario para casos no complicados. *Bol Med Hosp Infant Mex.* 2013;70:3-10.
4. Hadjipanayis A, Grossman Z, Del Torso S, van Esso D, Dornbusch HJ, Mazur A, et al. Current primary care management of children aged 1-36 months with urinary tract infections in Europe: large scale survey of paediatric practice. *Arch Dis Child.* 2015;100:341-7.
5. González Rodríguez JD, Rodríguez Fernández LM. Infección de vías urinarias en la infancia. *Protoc Diagn Ter Pediatr.* 2014;1:91-108.
6. Montini G, Tullus K, Hewitt I. Febrile urinary tract infections in children. *N Engl J Med.* 2011;365:239-50.
7. Martínez-Suárez V, Santos-Rodríguez F. Infección de las vías urinarias (ITU) en el niño: plan diagnóstico-terapéutico. *Bol Pediatr.* 2006;46:222-9.
8. Roberts KB. Revised AAP guideline on UTI in febrile infants and young children. *Am Fam Physician.* 2012;86:940-6.
9. Hanna-Wakim RH, Ghanem ST, El Helou MW, Khafaja SA, Shaker RA, Hassan SA, et al. Epidemiology and characteristics of urinary tract infections in children and adolescents. *Front Cell Infect Microbiol.* 2015;5:45.
10. Mukaka MM. A guide to appropriate use of correlation coefficient in medical research. *Malawi Med J.* 2012;24:69-71.
11. De Merech LF, García Bellenzier V, Franco Valdez M. Infección urinaria: aspectos relevantes y puesta al día. *Pediatría (Asunción).* 2014;41:57-64.
12. Wald E. Urinary tract infections in infants and children: a comprehensive overview. *Curr Opin Pediatr.* 2004;16:85-8.
13. Nagaraj S, Sukhlal Kalal BS, Kamath N, Muralidharan S. Microbiological and antimicrobial profile of pathogens associated with pediatric urinary tract infection: a one-year retrospective study from a tertiary care teaching hospital. *NJLM.* 2014;3:4-7.
14. Shaikh N, Morone NE, Bost JE, Farrell MH. Prevalence of urinary tract infection in childhood: a meta-analysis. *Pediatr Infect Dis J.* 2008;27:302-8.
15. Working Group of the Clinical Practice Guidelines for Urinary Tract Infection in Children. Clinical Practice Guidelines for Urinary Tract Infection in Children; Ministry of Health National Health Service Quality Plan, Social and Equality Policy. Aragon Institute of Health Sciences; 2011. pp. 71-5.
16. McMullen JA, Mahant S, DeGroot JM, Stephens D, Parkin PC. Predictors of long length of stay in infants hospitalized with urinary tract infection. *Hosp Pediatr.* 2014;4:291-7.
17. Mandal J. Pathogenesis and laboratory diagnosis of childhood urinary tract infection. *EMJ Urol.* 2016;4:101-7.
18. Subcommittee on Urinary Tract Infection, Steering Committee on Quality Improvement, Management. Urinary tract infection: clinical practice guideline for the diagnosis and management of the initial UTI in febrile infants and children 2 to 24 months. *Pediatrics.* 2011;128:595-607.
19. Salles MJ, Zurita J, Mejía C, Villegas MV, Latin America Working Group on Bacterial Resistance. Resistant gram-negative infections in the outpatient setting in Latin America. *Epidemiol Infect.* 2013;141:2459-72.
20. Bryce A, Hay AD, Lane IF, Thornton HV, Wootton M, Costelloe C. Global prevalence of antibiotic resistance in paediatric urinary tract infections caused by *Escherichia coli* and association with routine use of antibiotics in primary care: systematic review and meta-analysis. *BMJ.* 2016;352:i939.
21. Zurita J. Urinary *E. coli*: resistance trend in Ecuador from 1999 to 2007. *Rev Ecuat Ginecol Obstet.* 2009;15:29-35.
22. Fan NC, Chen HH, Chen CL, Ou LS, Lin TY, Tsai MH, et al. Rise of community-onset urinary tract infection caused by extended-spectrum β-lactamase-producing *Escherichia coli* in children. *J Microbiol Immunol Infect.* 2014;47:399-405.
23. Hernández JR, Pascual A, Cantón R, Martínez-Martínez L, Grupo de Estudio de Infección Hospitalaria (GEIH). *Escherichia*

- coli* y *Klebsiella pneumoniae* productores de betalactamasas de espectro extendido en hospitales españoles (Proyecto GEIH-BLEE 2000). Enferm Infect Microbiol Clin. 2003;21: 77–82.
- 24. Russell G. Antibiotic resistance in children with *E. coli* urinary tract infection. BMJ. 2016;352:i1399.
 - 25. Osthoff M, McGuinness SL, Wagen AZ, Eisen DP. Urinary tract infections due to extended-spectrum beta-lactamase-producing Gram-negative bacteria: identification of risk factors and outcome predictors in an Australian tertiary referral hospital. Int J Infect Dis. 2015;34:79–83.
 - 26. Moore CE, Sona S, Poda S, Putchhat H, Kumar V, Sophey S, et al. Antimicrobial susceptibility of uropathogens isolated from Cambodian children. Paediatr Int Child Health. 2016;36:113–7.
 - 27. Butler C, O'Brien K, Wootton M, Pickles T, Hood K, Howe R, et al. Empiric antibiotic treatment for urinary tract infection in preschool children: susceptibilities of urine sample isolates. Fam Pract. 2016;33:127–32.
 - 28. Walton JM. Antibiotics for acute pyelonephritis in children. Pediatr Child Health. 2015;20:349–50.
 - 29. Lee SJ. Clinical guideline for childhood urinary tract infection (second revision). Child Kidney Dis. 2015;19:56–64.