



Review

Advances and challenges in the management of feline sporotrichosis

Isabella Dib Ferreira Gremião^{a,*}, Luisa Helena Monteiro de Miranda^b,
Gabriela Reis Pereira-Oliveira^a, Rodrigo Caldas Menezes^a, Ana Caroline de Sá Machado^a,
Anderson Messias Rodrigues^c, Sandro Antonio Pereira^a

^a Laboratory of Clinical Research on Dermatозoonoses in Domestic Animals, Oswaldo Cruz Foundation (Fiocruz), Rio de Janeiro, Brazil

^b Sydney School of Veterinary Science, University of Sydney, Sydney, Australia

^c Laboratory of Emerging Fungal Pathogens, Federal University of São Paulo, São Paulo, Brazil



ARTICLE INFO

Article history:

Received 21 March 2022

Accepted 23 May 2022

Available online 13 July 2022

Keywords:

Cats

Sporotrichosis

Sporothrix

Therapy

ABSTRACT

The domestic cat is the most susceptible host to *Sporothrix* infection, developing severe clinical forms. Few effective antifungal agents are available for treating feline sporotrichosis, and cases of treatment failure are common. Treatment success depends on cat health status, therapy-related factors, as well as social/economic issues, but it is mainly contingent upon the host–fungus interaction. The owner's adherence is critical and should be reinforced throughout the treatment to increase the chances of a successful outcome. The antifungal agents described for feline sporotrichosis are most often used in monotherapy regimens. Due to cases in which the treatment with itraconazole failed, the use of antifungal agents in combination should be considered to achieve synergy. The combination of itraconazole and potassium iodide represents an important option for the treatment of naïve cats presenting multiple cutaneous lesions, nasal mucosal lesions and/or respiratory signs, as well as for refractory cases. However, the therapeutic options for unsuccessfully treated cases are scarce. Therefore new options are needed, even more taking into account that there are many *in vitro* potential molecules not available for use in cats yet. More studies are necessary to correlate *in vitro* antifungal susceptibility tests results and the outcome of cats treated due to sporotrichosis. This review will briefly discuss both the antifungal drugs and treatment protocols used in cats with sporotrichosis, as well as the determinants of treatment failure.

© 2022 Asociación Española de Micología. Published by Elsevier España, S.L.U. All rights reserved.

Avances y desafíos en el manejo de la esporotricosis felina

RESUMEN

El gato doméstico es el huésped más susceptible a la infección por *Sporothrix*, llegando a desarrollar formas clínicas graves. Hay pocos agentes antimicóticos efectivos disponibles para tratar la esporotricosis felina, y los casos de fracaso terapéutico son habituales. El éxito del tratamiento depende del estado de salud del gato, los factores relacionados con la terapia y los problemas sociales/económicos, pero se asocia principalmente con la interacción huésped-hongo. El cumplimiento del tratamiento por parte del propietario es fundamental y debe reforzarse durante todo el proceso para aumentar las posibilidades de éxito. Los agentes antimicóticos descritos para la esporotricosis felina se usan con mayor frecuencia en monoterapia. Debido a los casos de fallo terapéutico en el tratamiento con itraconazol se debe considerar el uso combinado de agentes antifúngicos en sinergia. La combinación de itraconazol y yoduro de potasio es una buena opción en el caso de gatos no tratados previamente y con múltiples lesiones cutáneas, en la mucosa nasal y/o con signos respiratorios, así como para casos refractarios. Sin embargo, las opciones terapéuticas para la mayoría de los casos que fracasan son escasas. Por tanto, son necesarias nuevas opciones terapéuticas, más aún cuando existen muchas moléculas potenciales *in vitro*, no disponibles de momento para su uso en gatos. Son necesarios más estudios que correlacionen los resultados de las

Palabras clave:

Gatos

Esporotricosis

Sporothrix

Terapia

* Corresponding author.

E-mail address: isabella.dib@ini.fiocruz.br (I.D.F. Gremião).

pruebas de sensibilidad in vitro a los antifúngicos con aquellos del tratamiento de gatos con esporotricosis. Esta revisión discutirá brevemente los fármacos antimicóticos y los protocolos terapéuticos utilizados para tratar gatos con esporotricosis, así como los factores determinantes del fracaso del tratamiento.

© 2022 Asociación Española de Micología. Publicado por Elsevier España, S.L.U. Todos los derechos reservados.

Sporothrix brasiliensis and *Sporothrix schenckii* are the main causative agents of feline sporotrichosis.^{23,49,79} *Sporothrix globosa*,⁵⁴ *Sporothrix humicola*⁵⁹ and *Sporothrix pallida*⁹² are scarcely reported. Cases of feline sporotrichosis have been reported in the United States, Mexico, Argentina, Paraguay, Malaysia, Spain, Germany, Australia, Japan, Thailand, United Kingdom and Brazil.^{26,27,29,43,59} In this last country, there is a predominance of *S. brasiliensis*, and epizootics have been reported since 1998.⁴⁵ On the other hand, *S. schenckii* has been described as the main causal agent in some Asian countries.⁴⁹

Feline sporotrichosis ranges from a single lesion to multiple skin lesions and fatal disseminated systemic forms.⁸⁴ *Sporothrix brasiliensis* is the most virulent species in terms of mortality in a murine model,²⁴ and infections are frequently associated with atypical and severe clinical manifestations in humans,² with increasing virulence over time.³⁵ Remarkably, the fatal disseminated form of the disease in cats has not been reported outside Brazil, where non-*S. brasiliensis* species are the major agents.^{48,49} The cat is considered the most susceptible animal species to *Sporothrix* infection, leading to severe clinical forms and more difficult treatment in many cases.^{64,88} The treatment of cats with sporotrichosis is an important measure of disease control as it induces a quick reduction of the fungal burden.⁶⁴ However, it requires a long period of daily care, and cats do not always respond well to the treatment, leading to therapeutic failure.^{42,66,78,86}

Few effective antifungal agents are available, and scarce clinical studies on cats with sporotrichosis have been conducted. Itraconazole (ITZ) as monotherapy or associated with potassium iodide (KI) is the most common therapeutic regimen. Ketoconazole (KTZ), sodium iodide (NaI), fluconazole (FLZ), amphoterin B deoxycholate (AMB), terbinafine (TRB), posaconazole (POS), local heat therapy, cryosurgery and surgical removal of the lesions have also been described for treating this disease in cats.⁴⁴ Some progress has recently been made in searching for new antifungal therapies for sporotrichosis. Potential candidates that showed *in vitro* activity against *Sporothrix* species include terpinen-4-ol and farnesol, TCAN26 (a structural analogue of miltefosine), H3 (a 24-sterol methyltransferase inhibitor), clotrimazole, β -dihydrofuran naphthoquinone isomers (naphthoquinone), coumarin derivatives, extracts from members of the Piperaceae family (*Piper abutiloides*), marjoram (*Origanum majorana* L.), rosemary (*Rosmarinus officinalis* L.),⁸⁰ pentathiepin compounds,⁵ buparvaquone,¹⁰ curcumin⁵⁰ and derivatives from the acylhydrazones.⁸ Many of these compounds show synergism with ITZ and significant antifungal activity against ITZ-resistant isolates,⁸⁰ making them candidates for further studies in cats. However, the path taken from the *in vitro* findings, then the *in vivo* drug tests to check effectiveness and safety, and the approval of the final product at the end is a long one. Although miltefosine was long considered a candidate in the drug repositioning strategy,^{9,25} it was not effective in a study that included cats with sporotrichosis.⁸⁵

This review will briefly discuss about the current antifungal drugs, the treatment protocols used in cats with sporotrichosis, and the determinant factors in treatment failure.

Therapeutic options

Potassium iodide (20% or as a saturated solution) was the first treatment described for feline sporotrichosis.³³ However, few successful treatment outcomes were reported in isolated cases^{15,36} or series of cases.^{28,34} The occurrence of dose-related adverse reactions (anorexia, vomiting, lethargy and fever) was common (10–20 mg/kg every 12–24 h).^{3,20,21,28,33,68} When KI concentration was lower (2.5–20 mg/kg every 24 h), the clinical cure rate was 47.9%, and clinical adverse reactions were observed in 52% of the cases.⁷⁵ Despite the adverse reactions when used in monotherapy, KI remains being an option for feline sporotrichosis when talking about oral capsules with a lower dose than that established in the literature and combining with ITZ, a combination therapy that was proposed.^{76,78}

Sodium iodide was also used for the treatment of cats with sporotrichosis, with adverse reactions similar to those of KI.^{14,16,20,65,69,71,84} Cases of clinical cure were reported in a single case¹⁴ and a series of cases.^{16,84} However, in these case series, the percentage of clinical cure ranged from 21.4 to 23%, and the occurrence of gastrointestinal adverse reactions ranged from 35.7 to 40%.^{16,84}

Licensed in 1959, AMB (polyene antifungal antibiotic) became the mainstay for the treatment of invasive fungal diseases,⁵⁸ including the disseminated forms of human sporotrichosis.⁵⁵ However, its use in cats with sporotrichosis was associated with a lack of clinical response and adverse effects, especially nephrotoxicity.^{28,56,69} The imidazole KTZ became available in 1979, and the results of recovery and failure in feline sporotrichosis were published thereafter, with doses of KTZ varying from 5 to 27.7 mg/day.^{14,21,28,48,60,65,68,73,84} KTZ is less effective than ITZ (28.6%, 38.3%, respectively), and it is linked to higher occurrence of adverse reactions (42.1%, 30.9%, respectively), which include gastrointestinal alterations (nausea, anorexia, vomiting)⁷³ and hepatotoxicity.³⁹ The first reported case of feline sporotrichosis treated with ITZ occurred one year after its introduction in 1992.⁷¹ This drug became the first-line treatment for sporotrichosis in cats due to its effectiveness and improved safety profile compared with AMB and KTZ.^{20,48,57,64,73,81,84,88} The dosage established for treating cats with sporotrichosis (50–100 mg/day) resulted in variable clinical cure rates (38–77%).^{64,73,88} In comparison to KTZ, the use of ITZ implies lower occurrence of gastrointestinal and liver adverse reactions (30.9%).^{39,73} Dosage adjustments are not necessary in cases with renal impairment, but the dose must be lowered if administered with hepatic disease or dysfunction.³⁸

The association of ITZ (50–100 mg/day) and KI (2.5–20 mg/kg/day) is the most studied combination so far in the treatment of sporotrichosis in cats.^{64,76,78} This option increases the likelihood of recovery in cats with sporotrichosis that did not respond to ITZ monotherapy,⁷⁸ as well as for naïve cats, particularly those presenting multiple cutaneous lesions, nasal mucosal lesions and/or respiratory signs^{64,76} (Fig. 1). In cats with sporotrichosis refractory to ITZ or ITZ + KI, subcutaneous or intralesional AMB combined with ITZ is another therapeutic option.^{40–42}

It is worth noting that in the 1990s, the lipid-based formulations of AMB reached the market. These drugs have lower toxicity, can be offered at higher doses, and may effectively treat

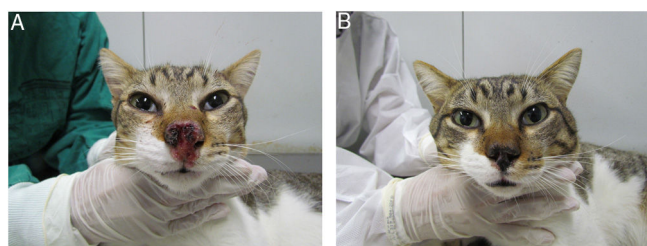


Fig. 1. Cat with sporotrichosis successfully treated with itraconazole and potassium iodide at the Laboratory of Clinical Research on Dermatозoonoses in Domestic Animals at INI/Fiocruz, Rio de Janeiro, Brazil (2021). (A) Cat with sporotrichosis presenting epiphora, ulcerated skin lesion and crusts on the nasal planum, philtrum and above the upper lip before receiving the antifungal treatment. (B) The clinical cure was achieved after 28 weeks of itraconazole (100 mg/24 h) and potassium iodide (5 mg/kg/24 h) therapy.

resistant or AMB-unresponsive mycoses. However, the cost may be prohibitive for its use in animals.³⁷ Intravenous administration of liposomal amphotericin B (L-AMB) (1 mg/kg) and oral ITZ (100 mg/day) were successfully used in a cat presenting skin and mucosal lesions on the nasal region refractory to ITZ 100 mg/day, AMB administered both subcutaneously (0.5 mg/kg) and intralesional (1 mg/kg).⁴² However, when intravenous L-AMB associated with oral ITZ (100 mg/day) was used in three cats with poor general health, multiple lesions and respiratory signs, one cat died due to the progression of the disease, and the others did not respond to the treatment.⁷²

Fluconazole, a broad-spectrum triazole whose use was approved in early 1990, is recommended for systemic antifungal infections, like cryptococcosis, involving difficult-to-reach tissues (central nervous system).³⁹ In a single study, FLZ (10 mg/day) was used in monotherapy to treat one cat with sporotrichosis that had cutaneous lesions and respiratory signs. Due to the recurrence of the respiratory signs, this drug was prescribed again shortly after the first treatment had end, and the clinical cure was further achieved.²⁰ Thus, there is no therapeutic or cost advantage that justifies the use of FLZ over ITZ for sporotrichosis infections.

The use of terbinafine, an allylamine antifungal agent, primarily recommended for dermatophytosis, was approved in 1990.⁴⁶ Terbinafine (30 mg/day) as monotherapy and in combination with itraconazole (5–10 mg/kg/day) was used in the treatment of feline sporotrichosis in one study, with no information on the clinical cure rate.⁸⁴ The effectiveness of this drug has been confirmed in humans^{31,32} and in two dogs⁹⁴ with sporotrichosis, but not in cats so far.

Posaconazole is a high-cost drug and an analog of ITZ, and was approved in 2006.⁸³ The use of POS was reported in a cat from Australia that presented with a non-ulcerated nodule on its nasal bridge due to an infection of a member within the *S. pallida* complex. After the ITZ-induced hepatic injury, therapy was changed to POS (40 mg/day), which eventually resulted in regression of residual infected tissues and clinical resolution.⁹²

When considering cryosurgery or administration of intralesional AMB, some factors should be considered, such as the location of the residual skin lesions and the requirement of anesthetic procedures.^{40,41,87}

Different treatment protocols used in feline sporotrichosis are described in Table 1.

Factors affecting the therapeutic response in feline sporotrichosis

Cat-related factors

A factor that may hinder reaching a complete cure is the presence of lesions on the bridge of the nose, nasal mucosa lesions

Table 1

Feline sporotrichosis treatment protocols described since 1956.

Year	Authors	Origin	Treatment
1956	Freitas et al. ³³	Brazil	KI
1965	Freitas et al. ³⁴	Brazil	KI
1973	Anderson et al. ³	USA	KI
1983	Burke et al. ¹⁴	USA	KTZ + NaI
1983	Nusbaum et al. ⁶⁹	USA	NaI
1986	Dunstan et al. ²⁸	USA	20% KI, SSKI, KTZ
1986	Mackay et al. ⁵⁶	AUS	20% KI
1989	Gonzalez Cabo et al. ³⁶	Spain	20% KI
1991	Carvalho et al. ¹⁵	USA	KI
1993	Peaston ⁷¹	USA	NaI, ITZ
1993	Marques et al. ⁶⁰	Brazil	KTZ
1996	Davies & Troy ²¹	USA	KI, KTZ
1996	Nakamura et al. ⁶⁵	Japan	NaI, KTZ
2001	Nobre et al. ⁶⁸	Brazil	KTZ, KI, ITZ
2004	Schubach et al. ⁸⁴	Brazil	NaI, KTZ, TRB, ITZ, ITZ + FLZ, ITZ + TRB
2009	Crothers et al. ²⁰	USA	NaI, KI, ITZ, FLZ
2009	Gremião et al. ⁴⁰	Brazil	ITZ + AMB IL
2010	Pereira et al. ⁷³	Brazil	ITZ, KTZ
2010	Madrid et al. ⁵⁷	Brazil	ITZ
2011	Gremião et al. ⁴¹	Brazil	ITZ + AMB IL
2012	Reis et al. ⁷⁵	Brazil	KI
2013	Rossi et al. ⁸¹	Brazil	ITZ
2015	Gremião et al. ⁴²	Brazil	AMB SC + ITZ, L-AMB IV + ITZ
2015	Pereira et al. ⁷²	Brazil	L-AMB IV + ITZ
2016	Reis et al. ⁷⁶	Brazil	ITZ + KI
2016	Souza et al. ⁸⁷	Brazil	Cryosurgery + ITZ
2017	Han et al. ⁴⁸	Malaysia	KTZ, ITZ
2018	Rocha et al. ⁷⁸	Brazil	ITZ + KI
2018	Carvalho et al. ¹⁶	Brazil	NaI
2018	Miranda et al. ⁶⁴	Brazil	ITZ, ITZ + KI
2018	Souza et al. ⁸⁸	Brazil	ITZ
2019	Thomson et al. ⁹²	Australia	ITZ, POS

KI: potassium iodide; NaI: sodium iodide; SSKI: super saturated potassium iodide solution; ITZ: itraconazole; KTZ: ketoconazole; TRB: terbinafine; FLZ: fluconazole; AMB IL: intralesional amphotericin B; L-AMB IV: intravenous liposomal amphotericin B; POS: posaconazole.

and respiratory signs, which are a risk of failure in cats.^{73,88} The host immune response may also negatively influence the prognosis. Cats with poor general condition and disseminated sporotrichosis show lesions with a high fungal load, which is associated with treatment failure and a longer healing time of the lesion. In these cases, the histopathological findings reveal the predominance of poorly formed granulomas, a reduced number of macrophages, neutrophils and lymphocytes, suggesting an ineffective immune response against *S. brasiliensis* infection. An optimal lymphocyte CD4/CD8 ratio seems to be determinant in the outcome of the patient, and high levels of CD4 lymphocytes in the peripheral blood are linked to better clinical outcomes, more organized granulomatous inflammation and reduced fungal load.^{63,88}

Interestingly, treatment failure does not seem to be associated with feline immunodeficiency virus (FIV) or feline leukemia virus (FeLV) infections. However, the histological changes observed in skin lesion samples of coinfecting cats suggest a less efficient immune response when compared to non-coinfecting cats, with the subsequent lower capacity to eliminate the fungus.⁸⁸ Moreover, cats with sporotrichosis with high levels of IL-10 during FIV and/or FeLV co-infections, and low levels of IL-4 (FeLV-positive) and IL-12 (FIV-positive) seem to develop severe clinical presentations and poor general condition. Nevertheless, severe clinical presentations and treatment failure are frequently described in the absence of feline retrovirus coinfections.⁶³

Considering the role of host factors in host–*Sporothrix* interaction, immunomodulation has been explored as a potential adjuvant therapy for sporotrichosis. In fact, it seems that those better outcomes when administering KI are related to an influence of iodides

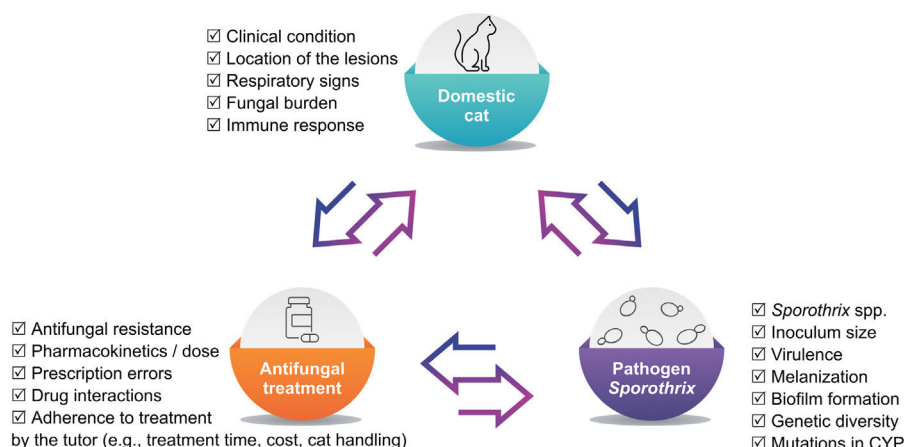


Fig. 2. Major factors influencing the therapeutic response in feline sporotrichosis.

on the immune response. The use of KI seems to induce considerable anti-inflammatory effects, notably in neutrophils.^{52,70,93} In this scenario, it has been shown that KI inhibits *Sporothrix* species biofilm metabolic activity in both filamentous and yeast forms.¹³ Other studies also described enhanced antibacterial and antiviral properties in the respiratory mucosa with the use of iodides.³⁰ Although it is tempting to correlate these findings with good results of KI use in the treatment of cats with nasal mucosa involvement, there is no evidence of this association in cats.

New vaccine candidates and immunomodulatory alternatives are currently being investigated in animal models with promising results for prophylactic and therapeutic purposes. A strong protective Th1-mediated response is induced with the use of the enolase-based vaccine and the Montanide PetGel A as an adjuvant.⁹⁰ Experimental passive immunization of mice with monoclonal antibodies against a *Sporothrix* 70-kDa glycoprotein has also shown a fungal burden reduction in tissue and an increased interferon- γ production.^{1,22,67} The treatment with complex carbohydrates as beneficial adjuvants to antifungal therapy, such as β -glucans, that are innate immunity agonists, has also been described.⁸⁹ These alternatives have been little explored in cats so far, and the current immunomodulators described do not show sufficient evidence to support their use in feline sporotrichosis.

Social and economic factors

Time commitment and the cost of the antifungal drugs are major causes of abandonment. Other economic factors like transportation costs, or costs associated with supplementary medication, negatively influence adherence.^{7,66,74} In addition, cases of illness in family members are another important reason that leads owners to request euthanasia for their cats⁸⁴ or abandon them. In Brazil the provision of drugs is free of charge, but it is not enough to guarantee compliance: treatment dropout is frequently described (34–38.5%) and mainly occurs once the clinical signs have disappeared.^{17,84} Thus, it is essential to emphasize the importance of both compliance since the treatment is started and the risk of relapse if the medication is discontinued before achieving the clinical cure.

Most treatment protocols are based on daily oral administration of the antifungal drugs, but some cats can be difficult to handle⁴⁴; giving medication to cats is usually more difficult when compared with dogs due to their discriminating sense of taste.⁹¹ Keeping cats indoors or confined within a limited space until reaching the clinical cure is recommended; however, this situation entails a substantial challenge to owners, especially when owning non-neutered cats.⁶

Therapy-related factors

The median treatment duration in cats with sporotrichosis is four months, and the treatment should be continued at least four weeks after reaching the clinical cure,⁴⁴ which might compromise the owner's adherence.^{53,74} Importantly, pet owners should be aware that an inadequate length of the therapy or an irregular administration may result in prolonged treatment schedule, failure or relapse, which usually occurs 3–18 months after the completion of the therapy.^{17,84} The occurrence of adverse drug reactions seems to be unrelated with treatment abandonment,¹⁷ but may interfere with the therapy schedule and prolong treatment duration. An inadequate drug prescription can also lead to treatment failure. The use of ITZ with H₂ receptor antagonists or proton pump blockers should be avoided because alkalinity decreases ITZ absorption.⁵¹ The bioavailability of ITZ in oral capsules is also erratic, but it can be maximized through the intake of food or fat at the same time.³⁹

As ITZ may be cost-prohibitive in brand name form, the generic antifungal medication is a satisfactory alternative. In the case of compounded ITZ, this should be avoided whenever possible. Studies in dogs and cats demonstrated that the generic and compounded ITZ are not bioequivalent to the reference drug. Pharmacokinetic data obtained with the generic formulation showed that therapeutic concentrations could be achieved, but compounded ITZ led to low plasma concentrations, being unlikely to be effective.^{62,77} Furthermore, a study described that the ITZ content in oral capsules acquired from two brand-name products, and three ITZ capsules from compounding pharmacies (for human and veterinary use in all cases) did not match the concentration reported by the manufacturers.⁹⁶

Antifungal resistance

The widespread use of antifungal agents is presumed to be a factor that promotes drug resistance.^{4,19} The emergence of *Sporothrix* species with *in vitro* antifungal resistance has been evidenced, and resistance development may be due to the melanin production capacity, genetic diversity and mutations in cytochrome P450.⁹⁵ The ability of the filamentous saprophytic phase of *Sporothrix* species to form biofilms has been previously described and is also associated with increased resistance to antifungal agents.^{12,25}

Antifungal-resistant strains of *S. brasiliensis* recovered from cats have been reported in Brazil.^{66,82,95} In cats with sporotrichosis there is a trend in relating *in vitro* resistance to treatment failure (clinical resistance); however, the latter can come of many reasons, not only of drug resistance (Fig. 2).

The *in vitro* activity of ITZ, KTZ, TRB and AMB was evaluated against 47 isolates of *S. brasiliensis* from cats followed-up at the Laboratory of Clinical Research on Dermatозoonoses in Domestic Animals at INI/Fiocruz, Rio de Janeiro, Brazil (unpublished data). These isolates were collected before the treatment with ITZ or KTZ. The filamentous phase was subjected to *in vitro* susceptibility testing using the liquid microdilution method according to the Clinical and Laboratory Standards Institute protocol M38-A2.¹⁸ TRB was the most effective antifungal, with *in vitro* activity showing the lowest MIC values (0.015–0.12 µg/mL). AMB showed a MIC range of 0.25–2 µg/mL. Among the azoles, KTZ (0.12–2 µg/mL) showed better *in vitro* activity than ITZ (0.50–2 µg/mL). These results were similar to those in other studies with *S. brasiliensis* strains recovered from cats.^{11,61,82} Curiously, *S. schenckii* isolates from Malaysia showed low susceptibility towards terbinafine.⁴⁸ In the same study, when assessing the outcome of the antifungal treatment on 33 cats (17 achieving clinical cure and 16 therapeutic failure), no significant difference was observed concerning the MICs obtained for KTZ and ITZ, suggesting that there was no correlation between the *in vitro* and *in vivo* findings in the studied sample. In another study from Southern Brazil that selected 12 feline cases of infection by *S. brasiliensis* with high MIC values (≥ 4 µg/mL) to ITZ, a correlation between the *in vivo* therapeutic response and the *in vitro* antifungal susceptibility assay was observed due to the occurrence of refractory cases with ITZ-resistant *S. brasiliensis*.⁶⁶

Additionally, no significant differences were observed between the general condition of one infected cat (presence of lymphadenomegaly, distribution of skin lesions and presence of respiratory signs) and the MIC values obtained from other cats in Rio de Janeiro. This lack of significant association between clinical characteristics and MIC values was already described in a study with human patients from the same region.⁴⁷ Therefore, the evaluation of the results of the antifungal susceptibility tests must be managed with great caution.

Conclusions

Some of the major advances and significant challenges that remain in the management of feline sporotrichosis have been reviewed. The veterinarian's role is essential to enable owners to increase adherence to therapeutic protocols and to develop easy-to-administer medications for cats (e.g., injectable depot formulations for long-term controlled drug release). Due to the few therapeutic options, studies on drug repositioning, vaccines and immunotherapy candidates should be encouraged. As prompt treatment in cats can rapidly reduce the fungal burden and risk of transmission of *Sporothrix* from cats, the availability of a first-line treatment for these animals at health facilities in affected areas is essential.

Conflict of interest

None declared.

Acknowledgements

Laboratório de Pesquisa Clínica em Dermatозoonoses em Animais Domésticos, Instituto Nacional de Infectologia Evandro Chagas, Fundação Oswaldo Cruz, Rio de Janeiro, RJ, Brazil for the technical support.

References

- Almeida SR. Therapeutic monoclonal antibody for sporotrichosis. *Front Microbiol.* 2012;3, <http://dx.doi.org/10.3389/fmicb.2012.00409>.
- Almeida-Paes R, de Oliveira MM, Freitas DF, do Valle AC, Zancopé-Oliveira RM, Gutierrez-Galhardo MC. Sporotrichosis in Rio de Janeiro, Brazil: *Sporothrix brasiliensis* is associated with atypical clinical presentations. *PLoS Negl Trop Dis.* 2014;8:e3094, <http://dx.doi.org/10.1371/journal.pntd.0003094>.
- Anderson NV, Ivoghli D, Moore WE, Leipold HW. Cutaneous sporotrichosis in a cat: a case report. *J Am Anim Hosp Assoc.* 1973;9:526–9.
- Antonovics J, Abbate JL, Baker CH, Daley D, Hood ME, Jenkins CE, et al. Evolution by any other name: antibiotic resistance and avoidance of the E-word. *PLoS Biol.* 2007;5:e30, <http://dx.doi.org/10.1371/journal.pbio.0050030>.
- Asquith CRM, Machado ACS, Miranda LHM, Konstantinova LS, Almeida-Paes R, Rakitin OA, et al. Synthesis and identification of pentathiepin-based inhibitors of *Sporothrix brasiliensis*. *Antibiotics (Basel).* 2019;8, <http://dx.doi.org/10.3390/antibiotics8040249>.
- Barros MBL, Schubach AO, Valle ACF, Gutierrez-Galhardo MC, Conceição-Silva F, Schubach TMP, et al. Cat-transmitted sporotrichosis epidemic in Rio de Janeiro, Brazil: description of a series of cases. *Clin Infect Dis.* 2004;38:529–35, <http://dx.doi.org/10.1086/381200>.
- Barros MBL, Schubach TMP, Coll JO, Gremião IDF, Wanke B, Schubach A. Esporotricose: a evolução e os desafios de uma epidemia. *Rev Panam Salud Publica.* 2010;27:455–60.
- Bonilla JJA, Honorato L, Haranahalli K, Gremião IDF, Pereira SA, Guimarães A, et al. Antifungal activity of acylhydrazones derivatives against *Sporothrix* spp. *Antimicrob Agents Chemother.* 2021;65:e02593–2620, <http://dx.doi.org/10.1128/AAC.02593-20>.
- Borba-Santos LP, Ishida K, Calogeropoulou T, De Souza W, Rozental S. Adamantylidene-substituted alkylphosphocholine TCAN26 is more active against *Sporothrix schenckii* than miltefosine. *Mem Inst Oswaldo Cruz.* 2016;111:523–7, <http://dx.doi.org/10.1590/0074-02760160088>.
- Borba-Santos LP, Barreto TL, Vila T, Chi KD, Monti FS, de Farias MR, et al. *In vitro* and *in vivo* antifungal activity of buparvaquone against *Sporothrix brasiliensis*. *Antimicrob Agents Chemother.* 2021;65:e00699–721, <http://dx.doi.org/10.1128/AAC.00699-21>.
- Brilhante RSN, Silva NF, Marques FJ, Castelo-Branco DS, Lima RA, Malaquias AD, et al. *In vitro* inhibitory activity of terpenic derivatives against clinical and environmental strains of the *Sporothrix schenckii* complex. *Med Mycol.* 2015;53:93–8, <http://dx.doi.org/10.1093/mmy/myu085>.
- Brilhante RSN, de Aguiar FRM, Silva MLQ, de Oliveira JS, de Camargo ZP, Rodrigues AM, et al. Antifungal susceptibility of *Sporothrix schenckii* complex biofilms. *Med Mycol.* 2018;56:297–306, <http://dx.doi.org/10.1093/mmy/myx043>.
- Brilhante RSN, Silva MLQ, Pereira VS, de Oliveira JS, Maciel JM, Silva INGD, et al. Potassium iodide and miltefosine inhibit biofilms of *Sporothrix schenckii* species complex in yeast and filamentous forms. *Med Mycol.* 2019;57:764–72, <http://dx.doi.org/10.1093/mmy/myy119>.
- Burke MJ, Grauer GF, Macy DW. Successful treatment of cutaneous lymphatic sporotrichosis in cat with ketoconazole and sodium iodine. *J Am Anim Hosp Assoc.* 1983;19:542–7.
- Carvalho JJ, Caldwell JB, Radford BL, Feldman AR. Feline-transmitted sporotrichosis in the southwestern United States. *West J Med.* 1991;154:462–5.
- Carvalho BW, Pereira SA, Figueiredo ABF, Miranda LHM, Pereira-Oliveira GR, Schubach TMP, et al. Iodeto de sódio: uma alternativa de tratamento para a esporotricose felina? *Acta Sci Vet.* 2018;46, <http://dx.doi.org/10.22456/1679-9216.88706>.
- Chaves AR, Campos MP, Barros MBL, Carmo CN, Gremião IDF, Pereira SA, et al. Treatment abandonment in feline sporotrichosis – study of 147 cases. *Zoonoses Public Health.* 2013;60:149–53, <http://dx.doi.org/10.1111/j.1863-2378.2012.01506.x>.
- CLSI. Reference method for broth dilution antifungal susceptibility testing of filamentous fungi. Wayne: CLSI document M-28 A; 2008.
- Cowen LE, Steinbach WJ. Stress, drugs, and evolution: the role of cellular signaling in fungal drug resistance. *Eukaryot Cell.* 2008;7:747–64, <http://dx.doi.org/10.1128/EC.00041-08>.
- Crothers SL, White SD, Ihrke PJ, Affolter VK. Sporotrichosis: a retrospective evaluation of 23 cases seen in northern California (1987–2007). *Vet Dermatol.* 2009;4:249–59, <http://dx.doi.org/10.1111/j.1365-3164.2009.00763.x>.
- Davies C, Troy GC. Deep mycotic infections in cats. *J Am Anim Hosp Assoc.* 1996;32:380–91, <http://dx.doi.org/10.5326/15473317-32-5-380>.
- De Almeida JRF, Kaihama GH, Jannuzzi GP, De Almeida SR. Therapeutic vaccine using a monoclonal antibody against a 70-kDa glycoprotein in mice infected with highly virulent *Sporothrix schenckii* and *Sporothrix brasiliensis*. *Med Mycol.* 2014;53:42–50, <http://dx.doi.org/10.1093/mmy/myu049>.
- De Carvalho JA, Beale MA, Hagen F, Fisher MC, Kano R, Bonifaz A, et al. Trends in the molecular epidemiology and population genetics of emerging *Sporothrix* species. *Stud Mycol.* 2021;100:100129, <http://dx.doi.org/10.1016/j.simyco.2021.100129>.
- Della Terra PP, Rodrigues AM, Fernandes GF, Nishikaku AS, Burger E, de Camargo ZP. Exploring virulence and immunogenicity in the emerging pathogen *Sporothrix brasiliensis*. *PLoS Negl Trop Dis.* 2017;11:e0005903, <http://dx.doi.org/10.1371/journal.pntd.0005903>.
- Dos Santos GMP, Borba-Santos LP, Vila T, Gremião IDF, Pereira SA, De Souza W, et al. *Sporothrix* spp. biofilms impact in the zoonotic transmission route: feline claws associated biofilms, itraconazole tolerance, and potential repurposing for miltefosine. *Pathogens.* 2022;11, <http://dx.doi.org/10.3390/pathogens11020206>.

26. Duangkhaew L, Yurayart C, Limsivilai O, Chen C, Kasornrondkua C. Cutaneous sporotrichosis in a stray cat from Thailand. *Med Mycol Case Rep.* 2019;23:46–9, <http://dx.doi.org/10.1016/j.mmcr.2018.12.003>.
27. Duarte JMG, Acosta VRW, Viera PMLF, Caballero AA, Matiauda GAG, De Oddone VBR, et al. Sporotrichosis transmitted by domestic cat. A family case report. *Rev Del Nacional (Itauguá).* 2017;9:67–76, <http://dx.doi.org/10.18004/rdn2017.0009.02.086-095>.
28. Dunstan RW, Langham RF, Reimann KA, Wakenell PS. Feline sporotrichosis: a report of five cases with transmission to humans. *J Am Acad Dermatol.* 1986;15:37–45, [http://dx.doi.org/10.1016/S0190-9622\(86\)70139-4](http://dx.doi.org/10.1016/S0190-9622(86)70139-4).
29. Etcheopaz A, Toscanini MA, Gisbert A, Mas J, Scarpa M, Iovannitti CA, et al. *Sporothrix brasiliensis*: a review of an emerging South American fungal pathogen, its related disease, presentation and spread in Argentina. *J Fungi.* 2021;7:170, <http://dx.doi.org/10.3390/jof703017>.
30. Fischer AJ, Lennemann NJ, Krishnamurthy S, Póca P, Durairaj L, Launspach JL, et al. Enhancement of respiratory mucosal antiviral defenses by the oxidation of iodide. *Am J Respir Cell Mol Biol.* 2011;45:874–81, <http://dx.doi.org/10.1165/rcmb.2010-0329OC>.
31. Francesconi G, Valle ACF, Passos SL, Reis R, Galhardo MC. Terbinafine (250 mg/day): an effective and safe treatment of cutaneous sporotrichosis. *J Eur Acad Dermatol Venerol.* 2009;23:1273–6, <http://dx.doi.org/10.1111/j.1468-3083.2009.03306.x>.
32. Francesconi G, Valle ACF, Passos SL, Barros MBL, Almeida-Paes R, Curi ALL, et al. Comparative study of 250 mg/day terbinafine and 100 mg/day itraconazole for the treatment of cutaneous sporotrichosis. *Mycopathologia.* 2011;171:349–54, <http://dx.doi.org/10.1007/s11046-010-9380-8>.
33. Freitas DC, Miglino MF, Zani Neto L. Esporotricose – observação de caso espontâneo em gato doméstico (*F. catus*). *Rev Fac Med Vet Univ S P.* 1956;5:601–4, <http://dx.doi.org/10.11606/issn.2318-5066.v7i2p381-388>.
34. Freitas DC, Moreno G, Saliba AM, Bottino JA, Mós EN. Esporotricose em cães e gatos. *Rev Fac Med Vet Univ S P.* 1965;7:381–8, <http://dx.doi.org/10.11606/issn.2318-5066.v7i2p381-388>.
35. Freitas DF, Santos SS, Almeida-Paes R, de Oliveira MM, do Valle AC, Gutierrez-Galhardo MC, et al. Increase in virulence of *Sporothrix brasiliensis* over five years in a patient with chronic disseminated sporotrichosis. *Virulence.* 2015;6:112–20, <http://dx.doi.org/10.1080/21505594.2015.1014274>.
36. Gonzalez Cabo JF, De las Heras Guillon MV, Latre Cequiel MV, Garcia de Jalón Ciercoles JA. Feline sporotrichosis: a case report. *Mycopathologia.* 1989;108:149–54, <http://dx.doi.org/10.1007/BF00436219>.
37. Greene CE. Antimicrobial drug formulary. In: Greene EC, editor. *Infectious diseases of the dog and cats.* Missouri: Elsevier; 2012. p. 581.
38. Greene CE. Antimicrobial drug formulary. In: Greene EC, editor. *Infectious diseases of the dog and cats.* Missouri: Elsevier; 2012. p. 1269.
39. Greene CE, Calpin J. Antimicrobial drug formulary. In: Greene EC, editor. *Infectious diseases of the dog and cats.* Missouri: Elsevier; 2012. p. 583.
40. Gremião IDF, Schubach TMP, Pereira SA, Rodrigues AM, Chaves AR, Barros MBL. Intralesional amphotericin B in a cat with refractory localised sporotrichosis. *J Feline Med Surg.* 2009;11:720–3, <http://dx.doi.org/10.1016/j.jfms.2009.01.012>.
41. Gremião IDF, Schubach TMP, Pereira SA, Rodrigues AM, Honse CO, Barros MBL. Treatment of refractory feline sporotrichosis with a combination of intralesional amphotericin B and oral itraconazole. *Aust Vet J.* 2011;89:346–51, <http://dx.doi.org/10.1111/j.1751-0813.2011.00804.x>.
42. Gremião IDF, Menezes RC, Schubach TMP, Figueiredo ABF, Cavalcanti MCH, Pereira SA. Feline sporotrichosis: epidemiological and clinical aspects. *Med Mycol.* 2015;53:15–21, <http://dx.doi.org/10.1093/mmy/myu061>.
43. Gremião IDF, Miranda LHM, Reis EG, Rodrigues AM, Pereira SA. Zoonotic epidemic of sporotrichosis: cat to human transmission. *PLoS Pathog.* 2017;13:1006077, <http://dx.doi.org/10.1371/journal.ppat.1006077>.
44. Gremião IDF, Da Rocha EMS, Montenegro H, Carneiro AJB, Xavier MO, De Farias MR, et al. Guideline for the management of feline sporotrichosis caused by *Sporothrix brasiliensis* and literature revision. *Braz J Microbiol.* 2020;52:107–24, <http://dx.doi.org/10.1007/s42770-020-00365-3>.
45. Gremião IDF, Oliveira MME, Miranda LHM, Freitas DFS, Pereira SA. Geographic expansion of sporotrichosis, Brazil. *Emerg Infect Dis.* 2020;26:621–4, <http://dx.doi.org/10.3201/eid2603.190803>.
46. Gupta M, Narang T, Kaur RJ, Manhas A, Saikia UM, Dogra S. A prospective case series evaluating efficacy and safety of combination of itraconazole and potassium iodide in rhinofacial conidiobolomycosis. *Int J Dermatol.* 2015;55:208–14, <http://dx.doi.org/10.1111/ijd.12966>.
47. Gutierrez-Galhardo MC, De Oliveira RMZ, Valle ACF, Almeida-Paes R, Silvatavares PM, Monzon A, et al. Molecular epidemiology and antifungal susceptibility patterns of *Sporothrix schenckii* isolates from a cat-transmitted epidemic of sporotrichosis in Rio de Janeiro, Brazil. *Med Mycol.* 2008;46:141–51, <http://dx.doi.org/10.1080/13693780701742399>.
48. Han HS, Kano R, Chen C, Noli C. Comparison of two in vitro antifungal sensitivity tests and monitoring during therapy of *Sporothrix schenckii* sensu stricto in Malaysian cats. *Vet Dermatol.* 2017;28, <http://dx.doi.org/10.1111/vde.12417>, 156–e32.
49. Han HS, Kano R. Feline sporotrichosis in Asia. *Braz J Microbiol.* 2021;52:125–34, <http://dx.doi.org/10.1007/s42770-020-00274-5>.
50. Huang L, Zhang J, Song T, Yuan L, Zhou J, Yin H, et al. Antifungal curcumin promotes chitin accumulation associated with decreased virulence of *Sporothrix schenckii*. *Int Immunopharmacol.* 2016;34:263–70, <http://dx.doi.org/10.1016/j.intimp.2016.03.010>.
51. Itraconazole. Drugs.com on 2021. Available from: <https://www.drugs.com/mtm/itraconazole.html> [accessed 04.03.22].
52. Jeong KU, Lee HS, Hwang JS. Effects of short-term potassium iodide treatment for thyrotoxicosis due to Graves disease in children and adolescents. *Ann Pediatr Endocrinol Metab.* 2014;19:197–201, <http://dx.doi.org/10.6065/apem.2014.19.4.197>.
53. Jin J, Sklar GE, Sen Oh VM, Chuen Li S. Factors affecting therapeutic compliance: a review from the patient's perspective. *Ther Clin Risk Manag.* 2008;4:269–86, <http://dx.doi.org/10.2147/TCRM.S1458>.
54. Kano R, Okubo M, Siew HH, Kamata H, Hasegawa A. Molecular typing of *Sporothrix schenckii* isolates from cats in Malaysia. *Mycoses.* 2015;58:220–4, <http://dx.doi.org/10.1111/myc.12302>.
55. Kauffman CA, Bustamante B, Chapman SW, Pappas PG. Clinical practice guidelines for the management of sporotrichosis: 2007 update by the Infectious Diseases Society of America. *Clin Infect Dis.* 2007;45:1255–65, <http://dx.doi.org/10.1086/522765>.
56. Mackay BM, Menrath VH, Ridley MF, Kellym WR. Sporotrichosis in a cat. *Aust Vet Pract.* 1986;16:3–5.
57. Madrid IM, Mattei A, Martins A, Nobre M, Meireles M. Feline sporotrichosis in the southern region of rio grande do sul, Brazil: clinical, zoonotic and therapeutic aspects. *Zoonoses Public Health.* 2010;57:151–4, <http://dx.doi.org/10.1111/j.1863-2378.2008.01227.x>.
58. Maertens JA. History of the development of azole derivatives. *Clin Microbiol Infect.* 2004;10:1–10, <http://dx.doi.org/10.1111/j.1470-9465.2004.00841.x>.
59. Makri N, Paterson GK, Gregge F, Urquhart C, Nuttall T. First case report of cutaneous sporotrichosis (*Sporothrix* species) in a cat in the UK. *J Fel Med Surg Open Rep.* 2020;6:1–5, <http://dx.doi.org/10.1177/2055116920906001>.
60. Marques SA, Franco SRVS, Camargo RMP, Dias LDF, Haddad Júnior V, Fabris VE. Sporotrichosis of the domestic cat (*Felis catus*): human transmission. *Rev Inst Med. Trop S P.* 1993;35:4, <http://dx.doi.org/10.1590/S0036-46651993000400004>.
61. Maschio-Lima T, Marques MDR, Lemes TH, Brizzotti-Mazuchi NS, Caetano MH, de Almeida BG, et al. Clinical and epidemiological aspects of feline sporotrichosis caused by *Sporothrix brasiliensis* and in vitro antifungal susceptibility. *Vet Res Commun.* 2021;45:171–9, <http://dx.doi.org/10.1007/s11259-021-09795-2>.
62. Mawby DI, Whitemore JC, Genger S, Papich MG. Bioequivalence of orally administered generic, compounded, and innovator-formulated itraconazole in healthy dogs. *J Vet Intern Med.* 2014;28:72–7, <http://dx.doi.org/10.1111/jvim.12219>.
63. Miranda LHM, Santiago MA, Schubach TMP, Morgado FN, Pereira SA, de Oliveira RVC, et al. Severe feline sporotrichosis associated with an increased population of CD8 low cells and a decrease in CD4+ cells. *Med Mycol.* 2016;54:29–39, <http://dx.doi.org/10.1093/mmy/myv079>.
64. Miranda LHM, Silva JN, Gremião IDF, Menezes RC, Almeida-Paes R, Reis EG, et al. Monitoring fungal burden and viability of *Sporothrix* spp. in skin lesions of cats for predicting antifungal treatment response. *J Fungi.* 2018;4:92, <http://dx.doi.org/10.3390/jof4030092>.
65. Nakamura Y, Sato H, Watanabe S, Takahashi H, Koide K, Hasegawa A. *Sporothrix schenckii* isolated from a cat in Japan. *Mycoses.* 1996;39:125–8, <http://dx.doi.org/10.1111/j.1439-0507.1996.tb00114.x>.
66. Nakasu CCT, Waller SB, Ripoll MK, Ferreira MRA, Conceição FR, Gomes AR, et al. Feline sporotrichosis: a case series of itraconazole-resistant *Sporothrix brasiliensis* infection. *Braz J Microbiol.* 2020;52:163–71, <http://dx.doi.org/10.1007/s42770-020-00290-5>.
67. Nascimento RC, Espíndola NM, Castro RA, Teixeira PAC, Loureiro, Penha CV, et al. Passive immunization with monoclonal antibody against a 70-kDa putative adhesin of *Sporothrix schenckii* induces protection in murine sporotrichosis. *Eur J Immunol.* 2008;38:3080–9, <http://dx.doi.org/10.1002/eji.200838513>.
68. Nobre MO, Castro AP, Caetano D, De Souza LL, Meireles MCA, Ferreiro L. Recurrence of sporotrichosis in cats with zoonotic involvement. *Rev Iberoam Micol.* 2001;18:137–40.
69. Nusbaum BP, Gulbas N, Horwitz SN. Sporotrichosis acquired from a cat. *J Am Acad Dermatol.* 1983;8:386–91, [http://dx.doi.org/10.1016/S0190-9622\(83\)80325-9](http://dx.doi.org/10.1016/S0190-9622(83)80325-9).
70. Okamura K, Sato K, Fujikawa M, Bandai S, Ikenoue H, Kitazono T. Remission after potassium iodide therapy in patients with Graves' hyperthyroidism exhibiting thionamide-associated side effects. *J Clin Endocrinol Metab.* 2014;99:3995–4002, <http://dx.doi.org/10.1210/jc.2013-4466>.
71. Peaston A. Sporotrichosis. *J Vet Intern Med.* 1993;7:44–5, <http://dx.doi.org/10.1111/j.1939-1676.1993.tb03168.x>.
72. Pereira COA, Gremião IDF, Pereira SA, Antonio IMS, Carvalho BW, Schubach TMP. Liposomal amphotericin B in the treatment of cats with refractory sporotrichosis. In: ISHAM 2015, Melbourne. 19th Congress of the International Society for human and Animal Mycology. 2015.
73. Pereira SA, Passos SRL, Silva JN, Gremião IDF, Figueiredo FB, Teixeira JL, et al. Response to azolic antifungal agents for treating feline sporotrichosis. *Vet Rec.* 2010;166:290–4, <http://dx.doi.org/10.1136/vr.166.10.290>.
74. Read SI, Sperling LC. Feline sporotrichosis. Transmission to man. *Arch Dermatol.* 1982;118:429–31, <http://dx.doi.org/10.1001/archderm.118.5.429>.
75. Reis EG, Gremião IDF, Kitada AAB, Rocha RFDB, Castro VSP, Barros MBL, et al. Potassium iodide capsule treatment of feline sporotrichosis. *J Feline Med Surg.* 2012;14:399–404, <http://dx.doi.org/10.1177/1098612X12441317>.
76. Reis EG, Schubach TMP, Pereira SA, Silva JN, Carvalho BW, Quintana MSB, et al. Association of itraconazole and potassium iodide in the treatment of feline sporotrichosis: a prospective study. *Med Mycol.* 2016;54:684–90, <http://dx.doi.org/10.1093/mmy/myw027>.
77. Renschler J, Albers A, Sinclair-Mackling N, Wheat LJ. Comparison of compounded, generic, and innovator-formulated itraconazole

- in dogs and cats. J Am Anim Hosp Assoc. 2018;54:195–200, <http://dx.doi.org/10.5326/JAAHA-MS-6591>.
78. Rocha RFDB, Schubach TMP, Pereira SA, Reis EG, Carvalho BW, Gremião IDF. Refractory feline sporotrichosis treated with itraconazole combined with potassium iodide. J Sm Anim Pract. 2018;59:720–1, <http://dx.doi.org/10.1111/jsap.12852>.
 79. Rodrigues AM, Teixeira MM, de Hoog GS, Schubach TMP, Pereira SA, Fernandes GF, et al. Phylogenetic analysis reveals a high prevalence of *Sporothrix brasiliensis* in feline sporotrichosis outbreaks. PLoS Negl Trop Dis. 2013;7:e2281, <http://dx.doi.org/10.1371/journal.pntd.0002281>.
 80. Rodrigues AM, Della Terra PP, Gremião ID, Pereira SA, Orofino-Costa R, De Camargo ZP, et al. The threat of emerging and re-emerging pathogenic *Sporothrix* species. Mycopathologia. 2020;185:813–42, <http://dx.doi.org/10.1007/s11046-020-00425-0>.
 81. Rossi CN, Odagui J, Larsson CE. Retrospective assessment of the treatment of sporotrichosis in cats and dogs using itraconazole. Acta Sci Vet. 2013;41:1112.
 82. Sanchotene KO, Brandolt TM, Klafke GB, Poester VR, Xavier MO. In vitro susceptibility of *Sporothrix brasiliensis*: comparison of yeast and mycelial phases. Med Mycol. 2017;55:869–76, <http://dx.doi.org/10.1093/mmy/myw143>.
 83. Schiller DS, Fung HB. Posaconazole: an extended-spectrum triazole antifungal agent. Clin Ther. 2007;29:1862–86, <http://dx.doi.org/10.1016/j.clinthera.2007.09.015>.
 84. Schubach TM, Schubach A, Okamoto T, Barros MB, Figueiredo FB, Cuzzi T, et al. Evaluation of an epidemic of sporotrichosis in cats: 347 cases (1998–2001). J Am Vet Med Assoc. 2004;224:1623–9, <http://dx.doi.org/10.2460/javma.2004.224.1623>.
 85. Silva FS, Cunha SCS, Baptista ARS, Baptista VS, Silva KVGC, et al. Miltefosine administration in cats with refractory sporotrichosis. Acta Sci Vet. 2018;46:1571, <http://dx.doi.org/10.22456/1679-9216.83639>.
 86. Silva FS, Cunha SCS, Moraes VA, Leite JS, Ferreira AMR. Refractory feline sporotrichosis: a comparative analysis on the clinical, histopathological, and cytopathological aspects. Pesq Vet Bras. 2022;42:e06923, <http://dx.doi.org/10.1590/1678-5150-PVB-6923>.
 87. Souza CP, Lucas R, Ramadinho RHR, Pires TBCP. Cryosurgery in association with itraconazole for the treatment of feline sporotrichosis. J Feline Med Surg. 2016;18:137–43, <http://dx.doi.org/10.1177/1098612X15575777>.
 88. Souza EW, Borba CM, Pereira SA, Gremião IDF, Langohr IM, de Oliveira MM, et al. Clinical features, fungal load, coinfections, histological skin changes, and itraconazole treatment response of cats with sporotrichosis caused by *Sporothrix brasiliensis*. Sci Rep. 2018;8:9074, <http://dx.doi.org/10.1038/s41598-018-27447-5>.
 89. Téllez-Martínez D, Batista-Duharte A, Portuondo DL, Zeppone Carlos I. Prophylactic and therapeutic vaccines against sporotrichosis. Feasibility and prospects. Microbes Infect. 2019;21:432–40, <http://dx.doi.org/10.1016/j.micinf.2019.05.003>.
 90. Téllez-Martínez D, Portuondo DL, Loesch ML, Batista-Duharte A, Zeppone Carlos I. A recombinant Enolase-Montanide™ PetGel A vaccine promotes a protective Th1 immune response against a highly virulent *Sporothrix schenckii* by toluene exposure. Pharmaceutics. 2019;11:144, <http://dx.doi.org/10.3390/pharmaceutics11030144>.
 91. Thombre AG. Oral delivery of medications to companion animals: palatability considerations. Adv Drug Deliv Rev. 2004;56:1399–413, <http://dx.doi.org/10.1016/j.addr.2004.02.012>.
 92. Thomson J, Trott DJ, Malik R, Galgut B, McAllister MM, Nimmo J, et al. An atypical cause of sporotrichosis in a cat. Med Mycol Case Rep. 2019;23:72–6, <http://dx.doi.org/10.1016/j.mmcr.2019.01.004>.
 93. Van der Spek AH, Fliers E, Boelen A. Thyroid hormone metabolism in innate immune cells. J Endocrinol. 2017;232:67–81, <http://dx.doi.org/10.1530/JOE-16-0462>.
 94. Viana PG, Figueiredo ABF, Gremião IDF, Miranda LHM, Antonio IMS, Boechat JS, et al. Successful treatment of canine sporotrichosis with terbinafine: case reports and literature review. Mycopathologia. 2018;183:471–8, <http://dx.doi.org/10.1007/s11046-017-0225-6>.
 95. Waller SB, Dalla Lana DF, Quattrin PM, Ferreira MRA, Fuentefria AM, Mezzari A. Antifungal resistance on *Sporothrix* species: an overview. Braz J Microbiol. 2021;52:73–80, <http://dx.doi.org/10.1007/s42770-020-00307-z>.
 96. Waller SB, Ripoll MK, Madrid IM, Acunha T, Cleff MB, Chaves FC, et al. Susceptibility and resistance of *Sporothrix brasiliensis* to branded and compounded itraconazole formulations. Braz J Microbiol. 2021;52:155–62, <http://dx.doi.org/10.1007/s42770-020-00280-7>.