



Review

## Current status of the etiology of candidiasis in Mexico

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ABSTRACT

This study presents a systematic review of the literature on the etiology of superficial and invasive candidiasis in Mexico reported from 2005 to 2015. The data have shown that *Candida albicans* is the most prevalent species with an increasing tendency of the non-*C. albicans* *Candida* species, as reported in other countries. The use of phenotypical methods in the identification of the yeasts limits the identification at the species level, particularly in species that are part of complexes, this is important because the identification only at the genus level leads to inadequate treatment due to the different susceptibility to the antifungals among species. In addition, this finding reveals the need to implement in clinical laboratories the molecular methods for the correct identification of the species involved, and the antifungal susceptibility tests to prevent the etiological changes associated with a poor therapeutic management.

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## Estado actual de la etiología de la candidiasis en México

RESUMEN

Palabras clave:

Candidiasis

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Etiología

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México

Este artículo presenta una revisión sistemática de la bibliografía sobre la etiología de las candidiasis superficiales e invasivas en México en los años 2005–2015. Los datos muestran que *Candida albicans* se sitúa en primer lugar, pero hay una tendencia al aumento de otras especies del género, como se reporta en otros países. El uso de métodos fenotípicos para la identificación de las levaduras limita la identificación de la especie, especialmente de aquellas que forman parte de complejos; esto es importante, ya que la identificación solo del género puede conducir a un tratamiento inadecuado por la diferente sensibilidad de las especies a los antifúngicos. Por ello es necesario implementar en los laboratorios clínicos los métodos moleculares necesarios para la completa identificación de las especies implicadas y las pruebas de sensibilidad a los antifúngicos, para evitar cambios etiológicos asociados con un manejo terapéutico inadecuado.

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Over the past 30 years, the incidence of opportunistic fungal infections has increased significantly and consistently worldwide. These mycoses are caused by filamentous fungi and yeasts, most notably *Aspergillus* and *Candida*, respectively, which are the most frequent causative agents. However, *Candida* is the main cause of mycosis in the world, especially in neutropenic patients, patients

with malignancies treated with immunosuppressants, patients undergoing surgery or organ transplantation, preterm infants, and HIV/AIDS patients.<sup>37,66</sup> Of the different clinical forms of candidiasis, the invasive variety is a persistent public health problem. The incidence and mortality rates associated with this infectious disease have remained unchanged for more than a decade despite major advances in the field of antifungal therapy; this persistence leads to an increase in the costs of hospital care.

The genus *Candida* includes more than 200 species, of which *Candida albicans*, *Candida glabrata*, *Candida parapsilosis*, *Candida*

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*tropicalis*, *Candida krusei*, *Candida guilliermondii* [current name *Meyerozyma guilliermondii* (Wick.)],<sup>43</sup> *Candida lusitaniae*, *Candida dubliniensis*, *Candida pelliculosa*, *Candida kefyr*, *Candida lipolytica*, *Candida famata* [current name *Debaryomyces hansenii* (Zopf)],<sup>46</sup> *Candida inconspicua*, *Candida rugosa* and *Candida norvegensis* are recognized as human pathogens more frequently.<sup>75</sup> Some of these species are found as commensals in humans but can become pathogenic as a result of alterations in their microenvironment or compromise of the host immune system, promoting proliferation of the fungus and producing endogenous candidiasis; by contrast, exogenous infections are transmitted through the hands of medical personnel or contaminated devices.<sup>36</sup>

*C. albicans* has been considered the predominant species found in bloodstream infections; however, in the last two decades, there has been a growing trend of infections caused by other species of the genus, such as *C. glabrata*, *C. parapsilosis*, *C. tropicalis*, *C. krusei*, *C. dubliniensis*, *C. kefyr*, *C. famata*, *C. lusitaniae*, *Candida lambica*, *Candida zeylanoides*, *Candida africana*, and *C. guilliermondii*.<sup>11,57,61,62</sup> In addition, within the *C. parapsilosis* complex, *Candida metapsilosis* and *Candida orthopsilosis* have been identified as infection-causing agents of superficial candidiasis; in the *C. glabrata* complex, *Candida nivariensis* and *Candida bracarensis* have been found in genital candidiasis.<sup>29,47</sup> However, it has been reported that *C. albicans* is still the most frequently isolated species among the different forms of candidiasis.<sup>61–64</sup>

This increase in non-*C. albicans* *Candida* species is attributed to the indiscriminate use of antifungals and increased use of implanted medical devices, organ transplantation and broad-spectrum antibacterial therapy.<sup>7,17,20</sup> The treatment of the infections caused by these species may be difficult due to the resistance to some antifungals. *C. krusei* (intrinsically resistant) and *C. glabrata* (dose-dependent)<sup>9</sup> are resistant to fluconazole, one of the most recommended antifungals in the treatment of invasive candidiasis, followed by echinocandins, voriconazole and amphotericin B.<sup>42,48,59</sup>

The species *C. albicans* and *C. parapsilosis* are naturally susceptible to fluconazole; however, over time, the susceptibility of *C. albicans* to this antifungal agent has decreased, as observed in clinical isolates of *C. albicans*, subjected to prolonged treatment with azoles.<sup>54,57,59</sup> In addition, the susceptibility of *C. parapsilosis* and *C. glabrata* to fluconazole has changed discreetly and varies depending on the geographic region.<sup>60,64,80</sup> It is clear that no antifungal agent is exempt from the development of resistance, so it is essential that laboratories identify not only the genus, but the species of the clinical isolates of *Candida*, and consider performing *in vitro* susceptibility tests to guide the choice of treatment. The mortality attributable to invasive candidiasis remains high, mainly due to the delay in the administration of an appropriate antifungal therapy.<sup>61</sup> Therefore, this work presents the etiology of superficial and invasive candidiasis in Mexico, as well as an overview of the antifungal susceptibility of *Candida* species.

A systematic review of the medical literature published from 2005 to 2015 was carried out to know the prevalence of the etiology of superficial and invasive candidiasis in Mexico, as well as the susceptibility of the *Candida* species to antifungals. To achieve this objective, the selection of relevant articles based on their title and abstracts was carried out. The information was searched in the following databases: PubMed, Medline and Google Scholar (January 2005 to December 2015). The searches were performed using words present in articles, key words according to the database consulted; and synonyms extracted from the databases and publications available in both English and Spanish. The articles reviewed were those which included studies carried out in Mexico on superficial or invasive candidiasis with the *Candida* isolates identified at the species level by phenotypical or genotypical methods. Studies carried out in other countries; studies that reported the identifica-

tion of *Candida* only at the genus level or that did not report the number of the isolates of *Candida* identified at the species level were not included.

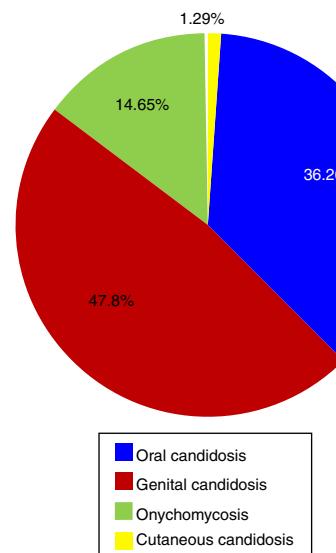
The prevalence of the *Candida* species was estimated based on the total number of isolates. The prevalence of each type of candidiasis (oral, genital, cutaneous, onychomycosis and invasive) was estimated considering the number of the patients with each type of infection and the total population.

### Etiology of superficial candidiasis

Twenty-one articles on superficial candidiasis were found published between 2005 and 2015, of which 18 were selected based on the inclusion criteria. Two thousand and ninety six patients out of 4103 patients with suspected superficial candidiasis suffered this mycosis, representing 51.1%. The data found in the literature showed that, in terms of superficial candidiasis, the most common clinical form is the genital one (47.81%),<sup>32,47,72</sup> followed by oral candidiasis (36.26%),<sup>23,30,31,38,39,70,73,74</sup> onychomycosis (14.65%)<sup>1,27,49</sup> and, finally, skin infections (1.29%)<sup>13</sup> (Table 1 and Fig. 1). These data correspond to the states of Ciudad de México, Puebla, Yucatán, San Luis Potosí, Nuevo León and Veracruz. The reports of genital candidiasis showed that *C. albicans* was the most common cause (71.36%), followed by *C. krusei* (13.77%), *C. tropicalis* (8.18%), *C. glabrata* complex (5.89%) and *C. parapsilosis* complex (0.8%).<sup>4,14,32,47,70,77</sup> (Table 1 and Fig. 2a). In 20 cases of balanitis and balanoposthitis, the most common etiologic agent was *C. albicans*, followed by *C. glabrata* complex. One case was reported as a mixed infection caused by *C. albicans* and *C. glabrata* complex.<sup>4</sup>

In oral candidiasis, the reports showed the effects on different populations,<sup>39,73</sup> including HIV and non-HIV children and adolescents,<sup>31,74</sup> patients with cancer,<sup>70</sup> diabetics,<sup>23,30</sup> malnourished children<sup>31,38</sup> and people of the Tarahumara community.<sup>31</sup> In this type of candidiasis, the most common etiologic agent was *C. albicans* (77.24%), followed by *C. glabrata* complex (13.16%), *C. tropicalis* (5.13%), *C. krusei* (3.29%), *C. parapsilosis* complex (0.79%), *C. kefyr* (0.26%), and *C. famata* (0.13%) (Table 1 and Fig. 2b).

In onychomycosis, the most common species in descending order were *C. albicans* (45.27%), *C. parapsilosis* complex (24.76%), *C. tropicalis* (7.17%), *C. guilliermondii* (6.84%), *C. glabrata* complex (5.21%), *C. famata* (3.58%), *C. lipolytica* (1.63%), *C. krusei* (1.63%), *C. zeylanoides* (0.98%), *C. dubliniensis* (0.65%), *C. lusitaniae* (0.65%),



**Fig. 1.** Clinical manifestations of superficial candidiasis in patients from Mexico during the 2005–2015 period.

**Table 1**Distribution of *Candida* species associated with superficial mycosis in Mexico during the 2005–2015 period.

Number and characteristics of the population studied	Number of patients with <i>Candida</i>	Frequency of the species identified (%)	Reference
<i>Oral candidiasis</i>			
111 patients with HIV/AIDS	72	<i>C. albicans</i> (90.27), <i>C. glabrata</i> complex (5.55), <i>C. tropicalis</i> (2.78), <i>C. parapsilosis</i> complex (1.39)	74
201 HIV-uninfected patients	112	<i>C. albicans</i> (63.39), <i>C. glabrata</i> complex (27.68), <i>C. tropicalis</i> (8.03), <i>C. parapsilosis</i> complex (0.89)	
67 children malnourished	21	<i>C. albicans</i> (100.0)	38
35 patients without symptoms of <i>Candida</i> infection	33	<i>C. albicans</i> (47.0), <i>C. tropicalis</i> (23.6), <i>C. krusei</i> (17.6), <i>C. glabrata</i> complex (11.8)	39
151 patients with different oncologic pathologies	21	<i>C. albicans</i> (100.0)	70
74 HIV-infected and HIV-uninfected patients	74 with 184 isolations	<i>C. albicans</i> (73.91), <i>C. glabrata</i> complex (19.02), <i>C. tropicalis</i> (5.98), <i>C. parapsilosis</i> complex (1.1)	73
60 children with HIV/AIDS and HAART therapy	36	<i>C. albicans</i> (83.3), <i>C. krusei</i> (5.56), <i>C. tropicalis</i> (11.1)	
55 children with malnutrition	26	<i>C. albicans</i> (61.53), <i>C. krusei</i> (30.77), <i>C. tropicalis</i> (3.85), <i>C. glabrata</i> complex (3.85)	31
57 Tarahumara patients	10	<i>C. albicans</i> (80.0), <i>C. krusei</i> (20.0)	
56 patients with DM	37	<i>C. albicans</i> (59.46), <i>C. glabrata</i> complex (18.92), <i>C. tropicalis</i> (16.22), <i>C. kefyr</i> (2.7), <i>C. famata</i> <sup>a</sup> (2.7)	23
80 individuals without DM and chronic dialysis	34	<i>C. albicans</i> (64.7), <i>C. glabrata</i> complex (17.65), <i>C. tropicalis</i> (8.82), <i>C. kefyr</i> (2.94), <i>C. parapsilosis</i> complex (5.88)	
141 patients with type 2 DM	56	<i>C. albicans</i> (85.71), <i>C. glabrata</i> complex (8.93), <i>C. krusei</i> (5.36)	30
<i>Genital candidiasis</i>			
468 gynecological patients	325	<i>C. albicans</i> (67.7), <i>C. tropicalis</i> (16.0), <i>C. glabrata</i> complex (12.6), <i>C. krusei</i> (2.5), <i>C. parapsilosis</i> complex (1.2)	32
818 gynecological patients	230	<i>C. albicans</i> (100.0)	72
300 gynecological patients	300	<i>C. albicans</i> (53.0), <i>C. krusei</i> (41.0), <i>C. tropicalis</i> (6.0)	47
110 gynecological patients	110	<i>C. albicans</i> (71.3), <i>C. glabrata</i> complex (11.7), <i>C. tropicalis</i> (8.5), <i>C. krusei</i> (5.3), <i>C. parapsilosis</i> complex (3.1)	14
150 gynecological patients	21	<i>C. albicans</i> (42.8), <i>C. krusei</i> (23.8), <i>C. glabrata</i> complex (19.0), <i>C. tropicalis</i> (14.3)	77
20 patients with balanitis or balanoposthitis	20	<i>C. albicans</i> (95.0), <i>C. glabrata</i> complex (5.0)	4
<i>Onychomycosis</i>			
936 patients with suggestive signs of onychomycosis	152	<i>C. parapsilosis</i> complex (34.86), <i>C. albicans</i> (24.34), <i>C. guilliermondii</i> <sup>b</sup> (13.81), <i>C. famata</i> <sup>a</sup> (7.23), <i>C. tropicalis</i> (4.60), <i>C. lipolytica</i> (3.28), <i>C. glabrata</i> complex (2.63), <i>C. zeylanoides</i> (1.97), <i>C. dubliniensis</i> (1.32), <i>C. lusitaniae</i> (1.32), <i>C. krusei</i> (1.32), <i>C. kefyr</i> (1.32), <i>C. lambica</i> (1.32), <i>C. intermedia</i> (0.66)	49
185 patients with candidal onychomycosis	153	<i>C. albicans</i> (66.3), <i>C. parapsilosis</i> complex (14.3), <i>C. tropicalis</i> (9.1), <i>C. glabrata</i> complex (8.0), <i>C. krusei</i> (2.0)	1
One patient with hand onychomycosis	1	<i>C. albicans</i> (33.3), <i>C. parapsilosis</i> complex (33.3), <i>C. tropicalis</i> (33.3)	27
<i>Cutaneous candidiasis</i>			
27 patients with diaper dermatitis	27	<i>C. albicans</i> (88.89), <i>C. parapsilosis</i> complex (7.41), <i>C. glabrata</i> complex (3.70)	13

AIDS, Acquired Immunodeficiency Syndrome; DM, diabetes mellitus; HAART, Highly Active Antiretroviral Therapy; HIV, Human Immunodeficiency Virus.

<sup>a</sup> Current name: *Debaryomyces hansenii* (Zopf).<sup>48</sup><sup>b</sup> Current name: *Meyerozyma guilliermondii* (Wick.).<sup>45</sup>

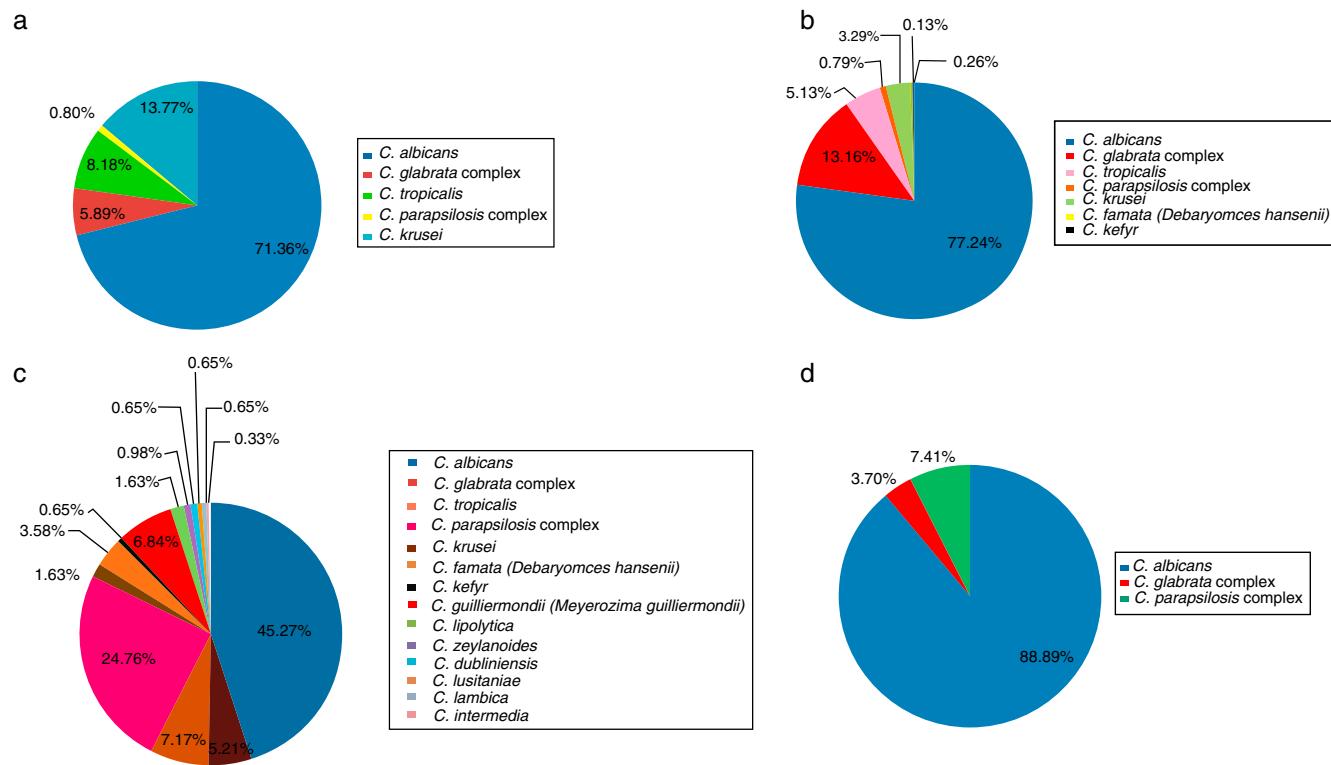
*C. kefyr* (0.65%), *C. lambica* (0.65%) and *C. intermedia* (0.33%) (Table 1 and Fig. 2c). Also, a case of a mixed infection caused by *C. albicans*, *C. parapsilosis* complex, and *C. tropicalis* was reported.<sup>27</sup> Onychomycosis was mainly reported in the fingernails of adult women and was associated with diabetes, injuries, circulatory disorders, the use of corticosteroids and the use of fake nails,<sup>1,27,49</sup> in agreement with the data reported by other countries.<sup>78</sup> *C. parapsilosis* complex predominated in toenails, while the most common fungus in fingernails was *C. albicans*.<sup>49</sup>

Regarding cutaneous candidiasis, only diaper rash reports were found during the period 2005–2015, where *C. albicans* was the most frequent species (88.89%), followed by *C. parapsilosis* complex (7.41%) and *C. glabrata* complex (3.70%)<sup>13</sup> (Table 1 and Fig. 2d).

In superficial candidiasis, the *Candida* isolates were identified by conventional methods: culture,<sup>1,4,13,14,23,32,72,74</sup> KOH (10–20%) test,<sup>1,4,13,14,27,49</sup> germ tube,<sup>1,39,70,73,74</sup> API 20C<sup>1,13,14,23,32,74</sup> and ID32<sup>49,73</sup> systems, CHROMagar<sup>4,13,27,30,31,38,39,47,49,70,73,77</sup> chlamydoconidia formation,<sup>1,14,49,73</sup> and growth at 45 °C.<sup>49</sup>

### Susceptibility to antifungal agents in superficial candidiasis

There are few studies on superficial candidiasis where antifungal susceptibility data are reported. The determination of the susceptibility to antifungals was performed using the reference procedures of broth microdilution (M27-A2 of the CLSI)<sup>32,49</sup> and disc diffusion method (M44-A),<sup>73</sup> as well as commercial systems such as Sensi-Disk<sup>32</sup> and Fungitest<sup>®</sup>,<sup>74</sup> which produce results that, in general, correlate with the reference procedures.<sup>22</sup> Based on these data, in general, oral candidiasis species showed <11% resistance to azoles.<sup>73,74</sup> In vaginal candidiasis, *C. glabrata* showed >50% resistance to fluconazole.<sup>32</sup> In onychomycosis, low resistance to azoles was reported in *C. albicans* and *C. parapsilosis* complex, high resistance to itraconazole was reported in *C. guilliermondii* and *C. famata*, and resistance was also reported to ketoconazole, itraconazole and fluconazole in four *C. glabrata* complex isolates.<sup>49</sup> In cutaneous candidiasis, susceptibility to 2% sertaconazole was reported in *C. albicans*, *C. parapsilosis* complex and *C. glabrata* complex.<sup>13</sup>



**Fig. 2.** Frequency distribution of *Candida* species causing superficial candidiasis in Mexico during the 2005–2015 period. (a) Oral candidiasis. (b) Genital candidiasis. (c) Onychomycosis. (d) Cutaneous candidiasis.

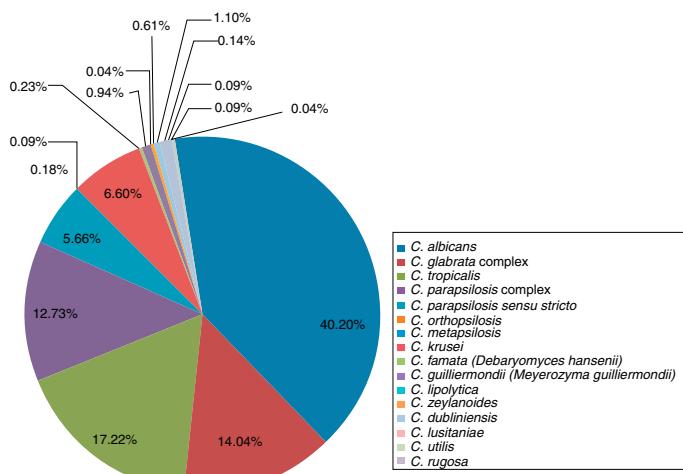
**Table 2**

Distribution of *Candida* species associated with invasive infection in Mexico during 2005–2015 period.

Number and characteristics of the population studied	Number of patients with <i>Candida</i>	Frequency of the species identified (%)	Reference
1221 neonates with candidiasis	25	<i>C. albicans</i> (56.0), <i>C. parapsilosis</i> complex (32.0), <i>C. glabrata</i> complex (8.0), <i>C. guilliermondii</i> <sup>b</sup> (4.0)	66
567 patients hospitalized (148 patients with sepsis)	18	<i>C. albicans</i> (55.55), <i>C. tropicalis</i> (11.11), <i>C. glabrata</i> complex (33.33)	18
107 pediatric patients with fever and neutropenia	31	<i>C. albicans</i> (61.29), <i>C. glabrata</i> complex (19.35), <i>C. tropicalis</i> (12.90), <i>C. krusei</i> (3.22), <i>C. parapsilosis</i> complex (3.22)	5
398 patients with nosocomial bloodstream infection	398	<i>C. parapsilosis</i> complex (37.69), <i>C. albicans</i> (31.9), <i>C. tropicalis</i> (14.82), <i>C. glabrata</i> complex (8.04), <i>C. krusei</i> (2.8), <i>C. guilliermondii</i> <sup>b</sup> (1.26), <i>C. zeylanoides</i> (0.5), <i>C. utilis</i> (0.5), <i>C. famata</i> <sup>a</sup> (1.0), <i>C. rugosa</i> (0.5), <i>C. lusitaniae</i> (0.5), <i>C. boidinii</i> (0.3)	34
Three neonates with nosocomial candidiasis	3	<i>C. parapsilosis</i> complex (100)	37
52 organ transplant patients	52	<i>C. albicans</i> (100)	67
104 patients with fever of unknown origin and immunosuppression	7	<i>C. albicans</i> (71.42), <i>C. tropicalis</i> (14.3), <i>C. guilliermondii</i> <sup>b</sup> (14.3)	52
326 pediatric patients with candidemia	358 (some patients had more than one yeast isolate)	<i>C. albicans</i> (38.95), <i>C. tropicalis</i> (22.08), <i>C. parapsilosis</i> complex (38.95) of which <i>C. parapsilosis</i> sensu stricto (95.3), <i>C. orthopsis</i> (3.1), <i>C. metapsilosis</i> (1.6)	35
15 patients with candidemia	12	<i>C. parapsilosis</i> complex (75.0), <i>C. glabrata</i> complex (16.67), <i>C. albicans</i> (8.33)	56
24 patients with blood infection	24	<i>C. albicans</i> (45.83), <i>C. tropicalis</i> (25.0), <i>C. glabrata</i> complex (12.5), <i>C. parapsilosis</i> complex (4.16), <i>C. krusei</i> (4.16), <i>C. guilliermondii</i> <sup>b</sup> (4.16), <i>C. lipolytica</i> (4.16)	21
18 patients with candidemia	11	<i>C. albicans</i> (45.45), <i>C. tropicalis</i> (27.27), <i>C. glabrata</i> complex (18.18), <i>C. famata</i> <sup>a</sup> (9.1)	24
One patient with community-acquired pneumonia	1	<i>C. lusitaniae</i> (100)	50
30 patients with candidemia	9	<i>C. albicans</i> (77.78), <i>C. parapsilosis</i> complex (22.22)	51
116 patients with candidiasis	86	<i>C. albicans</i> (71.59), <i>C. parapsilosis</i> complex (17.04), <i>C. tropicalis</i> (11.36)	33
1225 patients with candidiasis	1171	<i>C. albicans</i> (39.0), <i>C. glabrata</i> complex (21.0), <i>C. tropicalis</i> (18.0), <i>C. krusei</i> (11.0), <i>C. parapsilosis</i> complex (7.0), <i>C. dubliniensis</i> (2.0), <i>C. guilliermondii</i> <sup>b</sup> (1.0), <i>C. zeylanoides</i> (1.0)	69

<sup>a</sup>Current name: *Debaryomyces hansenii* (Zopf).<sup>48</sup>

<sup>b</sup> Current name: *Meyerozyma guilliermondii* (Wick.).<sup>45</sup>



**Fig. 3.** Frequency distribution of *Candida* species causing invasive candidiasis in Mexico during the 2005–2015 period.

### Etiology of invasive candidiasis

Regarding invasive candidiasis, 15 publications reported 2184 cases (Table 2). The affected populations included neonates<sup>56,66</sup> and pediatric patients,<sup>35,51</sup> transplant patients,<sup>52,67</sup> patients with fever of unknown origin,<sup>24,51,52</sup> patients with neutropenia,<sup>5</sup> patients with a bloodstream infection,<sup>21,24,34,35,37,51,56</sup> patients in intensive care units<sup>18,24,37,51,56,66</sup> and patients hospitalized in hematology unit,<sup>24,52</sup> infectious unit, internal medicine,<sup>24,51,52,56</sup> rheumatology,<sup>52</sup> emergency,<sup>56</sup> neurosurgery,<sup>24</sup> neurology,<sup>24</sup> pulmonology units,<sup>24</sup> and other patients hospitalized in non-specified services.<sup>21,33,34,50,69</sup> The reports found were from Nuevo León, Guadalajara, Durango, Morelia, Sonora, Sinaloa, Coahuila, Estado de México, Ciudad de México, and Tamaulipas.

The main species associated with invasive candidiasis were *C. albicans* (40.2%), *C. tropicalis* (17.22%), *C. glabrata* complex (14.04%), *C. parapsilosis* complex (12.73%), *C. krusei* (6.6%), *C. parapsilosis* sensu stricto (5.66%), *C. guilliermondii* (0.94%), *C. zeylanoides* (0.61%), *C. famata* (0.23%), *C. orthopsis* (0.18%), *C. lusitaniae* (0.14%), *Candida utilis* (0.09%), *C. metapsilosis* (0.09%), *C. rugosa* (0.09%), *C. lipolytica* (0.04%), and *Candida boidinii* (0.04%)<sup>5,18,21,24,33–35,37,50–52,56,66,67,69</sup> (Table 2 and Fig. 3).

In invasive candidiasis, *Candida* isolates were identified by conventional and molecular methods: culture,<sup>5,18,21,24,34,37,50,51,56,66,67,69</sup> KOH (10–20%) test,<sup>52,69</sup> germ tube production,<sup>5,21,35,69</sup> API 20C,<sup>5,34,35,52,69</sup> chlamydoconidia formation,<sup>33,35,52,69</sup> CHROMagar,<sup>33,50,51,56,69</sup> MicroScan<sup>33,37,51</sup> and Vitek YBC<sup>21</sup> automated systems, PCR,<sup>5,35</sup> and RFLP.<sup>50</sup>

### Susceptibility to antifungal agents in invasive candidiasis

Of the 15 studies found in the literature on invasive candidiasis, only six<sup>21,24,34,35,50,56</sup> included reports on antifungal susceptibility tests showing resistance to itraconazole in *C. tropicalis*, and to amphotericin B, fluconazole and itraconazole in *C. glabrata* complex.<sup>24,34,35,50</sup> Susceptibility tests to antifungals were performed using the broth microdilution (M27-A2<sup>34</sup> and M27-A3<sup>21,35,56</sup> of the CLSI) reference procedures and the Fungitest® commercial test.<sup>50</sup>

### Discussion

*Candida* is the main causative agent of mycosis in humans, but the epidemiology of candidiasis varies according to the

geographical region.<sup>8,16,81</sup> For this reason, it is important to conduct studies to determine the incidence of mycosis, the distribution of causative species, and their antifungal susceptibility profiles.

In Mexico, according to the morbidity yearbook published by the General Epidemiology Directorate (Dirección General de Epidemiología – [http://www.epidemiologia.salud.gob.mx/anuario/html/morbilidad\\_nacional.html](http://www.epidemiologia.salud.gob.mx/anuario/html/morbilidad_nacional.html)), urogenital candidiasis has remained among the 20 leading causes of illness nationwide, ranked between the 11th and 15th places. This is the only mycosis registered in the yearbook from 1988 to date. However, these data do not agree with the findings of the literature review, in which besides genital candidiasis other clinical forms of superficial candidiasis are also frequent. This discrepancy is due to the fact that fungal infections are not notifiable diseases. The results of this review also show that, between 2005 and 2015, the number of reports of superficial candidiasis was higher (18 publications) than that of invasive candidiasis (15 publications). The size of the population with suspected superficial candidiasis was 4103, with a positive frequency of 51.1%. In invasive candidiasis the suspected population suffering it was 4387 patients with a positivity of 49.8%. These data reveal that, in Mexico, the prevalence of superficial candidiasis is slightly greater than that of the invasive one, probably because factors predisposing to the superficial form (humidity, inadequate hygiene, pediatric age, pregnancy, hypothyroidism and type 2 diabetes mellitus) are more frequent than those predisposing to the invasive form.<sup>3</sup> It is important to mention that, in Mexico, type 2 diabetes is one of the most prevalent chronic diseases ([http://www.epidemiologia.salud.gob.mx/anuario/html/morbilidad\\_nacional.html](http://www.epidemiologia.salud.gob.mx/anuario/html/morbilidad_nacional.html)), which may be one of the most important predisposing factors to acquire superficial candidiasis compared with predisposing factors such as neutropenia, central parenteral nutrition, central venous catheter use, abdominal surgery, broad-spectrum antimicrobial use and severe immunocompromise.<sup>79</sup> This may also be due to, as in other countries, the incidence of invasive candidiasis being stable or decreased at certain institutions<sup>58</sup> or to a greater interest in invasive candidiasis because of the more frequent mortality.

Regarding the etiology of superficial candidiasis, especially the oral form, there is agreement within the international scientific literature, which reports *C. albicans* as the predominant species, followed in a different order, depending on the clinical form, by *C. krusei*, *C. tropicalis*, and *C. parapsilosis* complex. In contrast, other species such as *C. kefyr* and *C. famata* are rare.<sup>65</sup> Among the patients with oral candidiasis, there is a greater prevalence of *C. albicans* in those with HIV/AIDS than in those without HIV/AIDS.<sup>31,74</sup>

In vaginal candidiasis, the diversity of species reported in Mexico was low as only *C. albicans*, *C. tropicalis*, *C. krusei*, *C. glabrata* complex and *C. parapsilosis* complex have been reported; however, in other countries, and during the same period, *C. dubliniensis*, *C. famata*, *C. lusitaniae*, *C. africana*, *C. guilliermondii*, *C. lambica*, *C. kefyr*, *C. zeylanoides*, *C. nivariensis* and *C. bracarensis* were reported.<sup>44,76</sup> In balanitis and balanoposthitis, *C. albicans* and *C. glabrata* complex were found as causative agents, but these data cannot be compared because there are no studies on the distribution of *Candida* species in these mycoses. They are considered sexually transmitted diseases; therefore, it is thought that their distribution must be similar to that of vaginal candidiasis.<sup>2</sup>

For onychomycosis, we found that *C. parapsilosis* complex is the most frequently isolated yeast, and *C. albicans* is ranked third. The diversity of species isolated in Mexico is higher; in other countries, up to four species have been reported<sup>40</sup> and in Mexico, up to 14 species have been isolated.<sup>48</sup> This may be due to the geographical variation in the distribution of species.<sup>78</sup> In cutaneous candidiasis (diaper rash), we also noted that *C. albicans* is the most prevalent species,<sup>13</sup> as in other parts of the world.<sup>55</sup>

The etiological diversity of invasive candidiasis in Mexico coincide with that in reports from other countries, where *C. albicans*, *C. glabrata* complex, *C. krusei*, *C. tropicalis*, and *C. parapsilosis* complex cause more than 90% of cases; the isolation of *C. guilliermondii*, *C. lipolytica*, *C. famata*, *C. zeylanoides*, *C. utilis*, *C. rugosa*, and *C. boidinii* is rare.<sup>7,16</sup>

Thus, a review of the studies conducted in Mexico shows that the etiology of candidiasis in our environment is similar to that of other countries, where *C. albicans* remains the most isolated species, both in surface and invasive infections, but other species of the genus are becoming more frequent. The phenotypic methods used in the identification of *Candida* species were direct examination to observe the micromorphology, filamentation on serum to discriminate *C. albicans* from other species, culture in CHROMagar Candida to visualize the use of chromogenic substrates characteristic of *C. albicans*, *C. tropicalis* and *C. krusei*, production and clustering of chlamydoconidia to differentiate *C. albicans* from *C. dubliniensis*, and biochemical tests using the VITEK 2, API 20C, or API 32C systems. In most cases, these methods do not allow to achieve the correct identification of the species, especially in the case of infrequently isolated yeasts and those forming species complexes. For example, with filamentation on serum, other species, in addition to *C. albicans*, can also produce germ tubes.<sup>41</sup> In the case of CHROMagar Candida, which is one of the most widely used methods in clinical laboratories, there have been reports indicating that the color intensity of the colonies is not restrictive for *C. albicans* and *C. dubliniensis* species. In addition, it has been observed that the color of the colonies is gradually lost with subculturing or storage of the strains,<sup>28</sup> leading to misidentifications. However, it has been demonstrated that systems designed to identify multiple species (VITEK 2, API 20C and API 32C) are less accurate for the classification of unusual species, as reported by Desnos-Ollivier et al.<sup>26</sup> These investigators found that several isolates identified by API 32C as *C. famata* corresponded to *C. guilliermondii*, *Candida haemulonii*, *C. lusitaniae* and *Candida palmoleophila*, according to sequence analysis of gene segments. *C. parapsilosis* was also erroneously identified as *C. famata* with this method.<sup>15</sup> Another disadvantage of these identification systems is that they fail to discriminate species with similar biochemical profiles that are genetically distinct, as in the case of the species complexes *C. glabrata sensu stricto*, *C. nivariensis* and *C. bracarensis*; *C. parapsilosis sensu stricto*, *C. metapsilosis* and *C. orthopsilosis*; and *C. guilliermondii sensu stricto*, *Candida fermentati*, *Candida carpophila* and *Candida xestobii*, which can only be differentiated by molecular methods.<sup>29,44</sup> Due to the clinical importance of identifying the species of *Candida* involved, several researchers have proposed the use of molecular techniques. These include RAPD (Random Amplified Polymorphic DNA),<sup>80</sup> PCR-RFLP (Restriction Fragment Length Polymorphism)<sup>53</sup> and AFLP (Amplified Fragment Length Polymorphism).<sup>6</sup> PCR amplification of segments of specific genes, such as MP,<sup>63</sup> which encodes a mannanprotein of 65 kDa associated with morphogenesis and pathogenicity in *Candida*,<sup>12</sup> pulsed-field gel electrophoresis (PFGE),<sup>45</sup> sequencing of ITS (internal transcribed spacer) regions<sup>25</sup> and microsatellites<sup>71</sup> and multilocus sequence typing (MLST).<sup>68</sup> These techniques have been widely used in phylogenetic, taxonomic, and population structure studies and are particularly useful for the delimitation of *Candida* species.<sup>19</sup> However, the sequencing of ITS rDNA regions is the most widely used marker for the rapid, accurate identification of *Candida* species.<sup>25</sup> The importance of discriminating the species within a complex is that they differ in their susceptibility to antifungal agents, virulence and biofilm formation, so it is important for clinical laboratories to conduct antifungal susceptibility tests to administer the proper treatment. However, this review found that only 33.3% of the selected publications show antifungal susceptibility results,<sup>13,21,24,32,34,35,49,50,56,73,74</sup> which is a reflection that

these tests are not routinely performed. This situation represents an additional problem to the lack of specific identification methods because antifungal resistance may depend on the *Candida* species; thus, it is essential to perform this identification for the proper management of the patients. However, in the few reports that were found, we observed that *C. glabrata* complex showed higher resistance to azoles and amphotericin B in cases of both superficial and invasive mycosis.<sup>32,34,49,56,74</sup> This information is relevant because *C. glabrata* complex is one of the most common species in all forms of candidiasis. In addition, because discrimination between the members of *C. glabrata* complex is not reported, it is likely that a portion of the resistant isolates correspond to *C. nivariensis*.<sup>10</sup>

It is worth noting that the available epidemiological data in Mexico is limited, and the publications found corresponding to the 2005–2015 period only reflect the situation in the larger states of the country. However, these data facilitate predictions of what may be occurring in other states.

## Conclusions

The etiological behavior of candidiasis in Mexico is similar to that in other countries. However, it is important that both molecular tests for the accurate identification of yeasts and antifungal susceptibility tests are implemented and used routinely in healthcare facilities to ensure a specific diagnosis and the selection of appropriate therapeutic strategies. In addition, this strategy may help to prevent changes in the etiology of candidiasis as a result of the improper use of antifungals.

## Authors' contributions

MGFDL, MRRM, EDE, EMH, and GAA performed literature research and wrote the paper. All authors read and approved the final version.

## Conflicts of interest

The authors declare that they have no competing interests.

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## References

- Abad-González J, Bonifaz A, Ponce RM. Onicomicosis por *Candida* asociada con diabetes mellitus. Dermatol Rev Mex. 2007;51:135–41.
- Achkar JM, Fries BC. *Candida* infections of the genitourinary tract. Clin Microbiol Rev. 2010;23:253–73.
- Al-Attas SA, Amro SO. Candidal colonization, strain diversity, and antifungal susceptibility among adult diabetic patients. Ann Saudi Med. 2010;30:101–8.
- Araiza J, Montes de Oca G, Ponce Olivera RM, Bonifaz A. Balanitis y balanopostitis candidósica. Comunicación de 20 casos. Dermatol Rev Mex. 2011;55:342–6.
- Arellano-Galindo J, Moreno Galván M, Sarti E. Infecciones por hongos y neutropenia en un hospital pediátrico de tercer nivel. Salud Pública Mex. 2008;50:197–8.
- Asadzadeh M, Ahmad S, Hagen F, Meis JF, Al-Sweih N, Khan Z. Simple, low-cost detection of *Candida parapsilosis* complex isolates and molecular fingerprinting of *Candida orthopsilosis* strains in Kuwait by ITS region sequencing and amplified fragment length polymorphism analysis. PLOS ONE. 2015;10:e0142880.
- Bassetti M, Merelli M, Ansaldi F, de Florentiis D, Sartor A, Scarparo C, et al. Clinical and therapeutic aspects of candidemia: a five year single centre study. PLOS ONE. 2015;10:e0127534.
- Bassetti M, Righi E, Costa A, Fasce R, Molinari MP, Rosso R, et al. Epidemiological trends in nosocomial candidemia in intensive care. BMC Infect Dis. 2006;6:21.
- Berila N, Subik J. Opportunistic pathogen *Candida glabrata* and the mechanisms of its resistance to antifungal drugs. Epidemiol Mikrobiol Imunol. 2010;59:67–79.

10. Bertini A, de Bernardis F, Hensgens LA, Sandini S, Senesi S, Tavanti A. Comparison of *Candida parapsilosis*, *Candida orthopsilosis*, and *Candida metapsilosis* adhesive properties and pathogenicity. *Int J Med Microbiol.* 2013;303:98–103.
11. Bhai N, Tendolkar U, Baradkar V, Mathur M, Kulkarni M. Paediatric oropharyngeal and cutaneous candidiasis with special reference to *Candida dubliniensis*. *J Med Microbiol.* 2014;63:518–21.
12. Bineshian F, Yadegari MH, Sharifi Z, Akbari Eidgahi M, Nasr R. Identification of *Candida* species using MP65 gene and evaluation of the *Candida albicans* MP65 gene expression in BALB/C mice. *Jundishapur J Microbiol.* 2015;8:e18984.
13. Bonifaz A, Tirado-Sánchez A, Graniel MJ, Mena C, Valencia A, Ponce-Olivera RM. The efficacy and safety of sertaconazole cream (2%) in diaper dermatitis candidiasis. *Mycopathologia.* 2013;175:249–54.
14. Buitrón García A, Bonifaz A, Amancio Chassin O, Basurto Kuba E, Araiza J, Romero Cabello R. Correlación clínico-micológica de la candidiasis vulvovaginal. *Ginecol Obstet Mex.* 2007;75:68–72.
15. Burton MJ, Shah P, Swiatlo E. Misidentification of *Candida parapsilosis* as *C. famata* in a clinical case of vertebral osteomielitis. *Am J Med Sci.* 2011;341:71–3.
16. Caggiano G, Coretti C, Bartolomeo N, Lovero G, de Giglio O, Montagna MT. *Candida* bloodstream infections in Italy: changing epidemiology during 16 years of surveillance. *BioMed Res Int.* 2015;256580. <http://dx.doi.org/10.1155/2015/256580>.
17. Chakrabarti A, Chatterjee SS, Rao KL, Zameer MM, Shivaprakash MR, Singhji S, et al. Recent experience with fungemia: change in species distribution and azole resistance. *Scand J Infect Dis.* 2009;41:275–84.
18. Chávez García A, Cabrera Rayo A, Marín Romero MC, Villagómez Ortiz A, Méndez Reyes R, Guzmán Gómez R. Incidencia y pronóstico de candidiasis invasora en pacientes no neutropénicos de terapia intensiva. *Med Int Mex.* 2007;23:481–5.
19. Ciardo DE, Lucke K, Imhof A, Bloemberg GV, Böttger EC. Systematic internal transcribed spacer sequence analysis for identification of clinical mold isolates in diagnostic mycology: a 5-year study. *J Clin Microbiol.* 2010;48:2809–13.
20. Colombo AL, Guimarães T, Silva LR, de Almeida Monfardini LP, Cunha AK, Rady P, et al. Prospective observational study of candidemia in São Paulo Brazil: incidence rate, epidemiology, and predictors of mortality. *Infect Control Hosp Epidemiol.* 2007;28:570–6.
21. Corzo-León DE, Alvarado-Matute T, Colombo AL, Cornejo-Juárez P, Cortes J, Echevarría JJ, et al. Surveillance of *Candida* spp. bloodstream infections: epidemiological trends and risk factors of death in two mexican tertiary care hospitals. *PLOS ONE.* 2014;9:e97325.
22. Cuena-Estrella M, Alastruey-Izquierdo A, Gómez-López A, Monzón A. Estudios de sensibilidad en levaduras. Actualización y novedades. *Enferm Infect Microbiol Clin.* 2013;31 Suppl. 1:53–8.
23. De la Rosa-García E, Miramontes-Zapata M, Sánchez-Vargas LO, Mondragón Padilla A. Colonización e infección bucal por *Candida* sp. en pacientes diabéticos y no diabéticos con enfermedad renal crónica en diálisis. *Nefrología.* 2013;33:764–70.
24. De la Torre-Saldana VA, Martínez-Velázquez M, Reséndiz-Sánchez J. Factores de riesgo y epidemiología de la candidemia en el Hospital Juárez de México. *Med Int Mex.* 2014;30:121–32.
25. De Llanos Frutos R, Fernández-Espinar MT, Querol A. Identification of species of the genus *Candida* by analysis of the 5.8S rRNA gene and the two ribosomal internal transcribed spacers. *Antonie Van Leeuwenhoek.* 2004;85:175–85.
26. Desnos-Ollivier M, Ragon M, Robert V, Raoux D, Gantier JC, Dromer F. *Debaryomyces hansenii* (*Candida famata*), a rare human fungal pathogen often misidentified as *Pichia guilliermondii* (*Candida guilliermondii*). *J Clin Microbiol.* 2008;46:3237–42.
27. Díaz Molina V, Salas Espíndola Y, Sánchez de la Paz A, Sanabria Deseusa A, Ponce Olivera RM, Araiza J, et al. Onicomicosis de mano causada por tres especies de *Candida*. *Dermatol CMQ.* 2013;11:23–5.
28. Eraso E, Sahand IH, Villar-Vidal M, Marcos C, Moragues MD, Madariaga L, et al. Usefulness of *Candida* ID2 for the presumptive identification of *Candida dubliniensis*. *Med Mycol.* 2006;44:611–5.
29. Feng X, Ling B, Yang G, Yu X, Ren D, Yao Z. Prevalence and distribution profiles of *Candida parapsilosis*, *Candida orthopsilosis* and *Candida metapsilosis* responsible for superficial candidiasis in a Chinese university hospital. *Mycopathologia.* 2012;173:229–34.
30. Fernández-Martínez RF, Hernández-Pérez F, Fabián-San Miguel G, Jaimes Aveldáñez A, Arenas R. Oral *Candida* spp. carriers: its prevalence in patients with type 2 diabetes mellitus. *An Bras Dermatol.* 2013;88:222–5.
31. Gaitán-Cepeda LA, Sánchez-Vargas LO, Pavia-Ruz N, Muñoz-Hernández R, Villegas-Ham J, Caballos-Salobreña A. *Candida* bucal en niños mexicanos con VIH/sida, desnutrición o marginación social. *Rev Panam Salud Pública.* 2012;31:48–53.
32. García-Figueroa RB, Araiza-Santibáñez J, Basurto-Kuba E, Bonifaz-Trujillo A. *Candida glabrata*: un oportunitista emergente en vulvovaginitis. *Cir Cir.* 2009;77:455–60.
33. Garnica Ocegueda E, Araiza Santibáñez J, Moncada Barrón D, Arroyo Escalante S, Bonifaz A. Evaluación de sensibilidad y especificidad del agar Harina de maíz + tween 80, CHROMagar Candida® y MicroScan® para la identificación de especies aisladas de *Candida*. *Lab Acta.* 2009;21:79–84.
34. González GM, Elizondo M, Ayala J. Trends in species distribution and susceptibility of bloodstream isolates of *Candida* collected in Monterrey, Mexico, to seven antifungal agents: results of a 3-year (2004 to 2007) surveillance study. *J Clin Microbiol.* 2008;46:92902–5.
35. González GM, Treviño-Rangel RJ, Palma-Nicolás JP, Martínez C, González JG, Ayala J, et al. Species distribution and antifungal susceptibility of bloodstream fungal isolates in paediatric patients in Mexico: a nationwide surveillance study. *J Antimicrob Chemother.* 2013;68:2847–51.
36. Hani U, Shrivakumar HG, Vaghela R, Osmani RA, Shrivastava A. Candidiasis: a fungal infection – current challenges and progress in prevention and treatment. *Infect Disord Drug Targets.* 2015;15:42–52.
37. Hernández-Castro R, Arroyo-Escalante S, Carrillo-Casas EM, Moncada-Barrón D, Álvarez-Verona E, Hernández-Delgado L, et al. Outbreak of *Candida parapsilosis* in a neonatal intensive care unit: a health care workers source. *Eur J Pediatr.* 2010;169:783–7.
38. Hernández-Solís SE, Rueda-Gordillo F, Pereira-Góngora JR, Villamil-Urraiz JL. Frecuencia de portadores de *C. albicans* en un grupo de niños de una comunidad rural del estado de Yucatán. *Rev Odontol Latinoam.* 2008;1:1–4.
39. Jaimes Aveldáñez A, Hernández Pérez F, Martínez Herrera E, Rodríguez Carreón AA, Arenas-Guzmán R. Portadores de *Candida* en la mucosa oral: tipificación de 35 cepas con CHROMagar Candida. *Med Int Mex.* 2008;24:262–6.
40. Kaur R, Kashyap B, Bhalla P. Onychomycosis—epidemiology, diagnosis and management. *Indian J Med Microbiol.* 2008;26:108–16.
41. Kim D, Shin WS, Lee KH, Kim K, Young Park J, Koh CM. Rapid differentiation of *Candida albicans* from other *Candida* species using its unique germ tube formation at 39 degrees C. *Yeast.* 2002;19:957–62.
42. Kothavade RJ, Kura MM, Valand AG, Panthaki MH. *Candida tropicalis*: its prevalence, pathogenicity and increasing resistance to fluconazole. *J Med Microbiol.* 2010;59:873–80.
43. Kurtzman CP, Suzuki M. Phylogenetic analysis of ascomycete yeasts that form coenzyme Q-9 and the proposal of the new genera *Babjeviella*, *Meyeromyza*, *Milleromyza*, *Piceomyces* and *Scheffersomyces*. *Mycoscience.* 2010;51:2–14.
44. Li J, Shan Y, Fan S, Liu X. Prevalence of *Candida nivariensis* and *Candida bracarensis* in vulvovaginal candidiasis. *Mycopathologia.* 2014;178:279–83.
45. Lockhart SR, Pujol C, Dodgson AR, Soll DR. Deoxyribonucleic acid fingerprinting methods for *Candida* species. *Methods Mol Med.* 2005;118:15–25.
46. Lodder J, Kreger-van Rij JN, Kreger-van Rij, editor. *Yeasts, a taxonomic study.* 3rd ed. 1984. p. 130. Amsterdam.
47. López-García A, Ruiz-Tagle A, Pérez-Tlacomulco A, Mauleón-Montero A, Sánchez-Hernández JA, Rivera-Tapia JA. Prevalencia de diversas especies de *Candida* en mujeres con displasia cervical en un Hospital de la Ciudad de Puebla, México. *Rev Latinoam Patol Clin.* 2012;59:101–6.
48. Mann PA, McNicholas PM, Chau AS, Patel R, Mendrick C, Ullmann AJ, et al. Impact of antifungal prophylaxis on colonization and azole susceptibility of *Candida* species. *Antimicrob Agents Chemother.* 2009;53:5026–34.
49. Manzano-Gayoso P, Méndez-Tovar LJ, Arenas R, Hernández-Hernández F, Millán-Chiu B, Torres-Rodríguez JM, et al. Levaduras causantes de onicomicosis en cuatro centros dermatológicos mexicanos y su sensibilidad antifúngica a compuestos azólicos. *Rev Iberoam Micol.* 2011;28:32–5.
50. Martínez Espinosa I, González Ibarra M, Torres Guerrero HK. Identificación molecular de *Candida lusitaniae* en infección de tracto respiratorio inferior. *Rev Argent Microbiol.* 2014;46:307–10.
51. Martínez Herrera E, Esteves Jaramillo A, Tenorio Barragán I, Arroyo Escalante S, Moncada Barrón D, Arenas Guzmán R. Frecuencia de aislamientos microbílicos en hemocultivos de pacientes internados en un hospital de segundo nivel de la ciudad de México. *Med Int Mex.* 2008;24:338–41.
52. Méndez-Tovar LJ, Manzano-Gayoso P, Cumplido-Uribe C, Hernández-Hernández F, Ramos-Hernández J, López-Martínez R. Micosis invasivas en pacientes inmunodeprimidos con fiebre de origen desconocido. *Rev Med Inst Mex Seguro Soc.* 2012;50:609–14.
53. Mirhendi H, Makimura K, Khoramizadeh M, Yamaguchi H. A one-enzyme PCR RFLP assay for identification of six medically important *Candida* species. *Jpn J Med Mycol.* 2006;47:225–9.
54. Mishra NN, Prasad T, Sharma N, Payasi A, Prasad R, Gupta DK, et al. Pathogenicity and drug resistance in *Candida albicans* and other yeast species. A review. *Acta Microbiol Immunol Hung.* 2007;54:201–35.
55. Mohamadi J, Motaghi M, Panahi J, Havasian MR, Delpisheh A, Azizian M, et al. Antifungal resistance in *Candida* isolated from oral and diaper rash candidiasis in neonates. *Bioinformation.* 2014;10:667–70.
56. Morales Mendoza Y, Moncada Barrón D, Arroyo Escalante S, Sánchez MC, Manzano Gayoso P, Arenas R. Candidemias en un hospital general de la Ciudad de México: estudio de sensibilidad a antifúngicos con el método de micropelícula colorimétrica y microdilución en caldo. *Dermatol Rev Mex.* 2013;57:155–8.
57. Nakamura T, Takahashi H. Epidemiological study of *Candida* infections in blood: susceptibilities of *Candida* spp. to antifungal agents, and clinical features associated with the candidemia. *J Infect Chemother.* 2006;12:132–8.
58. Perman J, Quindós G. Aspectos actuales de las enfermedades invasoras causadas por *Candida* y otros hongos levaduriformes. *Rev Iberoam Micol.* 2016;33:133–9.
59. Pfaller MA, Andes D, Diekema DJ, Espinel-Ingroff A, Sheehan D. CLSI Subcommittee for antifungal susceptibility testing. Wild-type MIC distributions, epidemiological cutoff values and species-specific clinical breakpoints for fluconazole and *Candida*: time for harmonization of CLSI and EUCAST broth microdilution methods. *Drug Resist Updat.* 2010;13:180–95.
60. Pfaller MA, Boyken L, Hollis RJ, Kroeger J, Messer SA, Tendolkar S, et al., ARTEMIS DISK Global Antifungal Surveillance Group. Comparison of results of fluconazole and voriconazole disk diffusion testing for *Candida* spp. with results from a central reference laboratory in the ARTEMIS DISK Global Antifungal Surveillance Program. *Diagn Microbiol Infect Dis.* 2009;65:27–34.
61. Pfaller MA, Diekema DJ. Epidemiology of invasive candidiasis: a persistent public health problem. *Clin Microbiol Rev.* 2007;20:133–63.

62. Pfaller MA, Boyken L, Hollis RJ, Kroeger J, Messer SA, Tendolkar S, et al. *In vitro* susceptibility of invasive isolates of *Candida* spp. to anidulafungin, caspofungin, and micafungin: six years of global surveillance. *J Clin Microbiol.* 2008; 46:150–6.
63. Pfaller MA, Messer SA, Moet GJ, Jones RN, Castanheira M. *Candida* bloodstream infections: comparison of species distribution and resistance to echinocandin and azole antifungal agents in Intensive Care Unit (ICU) and non-ICU settings in the SENTRY Antimicrobial Surveillance Program (2008–2009). *Int J Antimicrob Agents.* 2011;38:65–9.
64. Pfaller MA, Moet GJ, Messer SA, Jones RN, Castanheira M. Geographic variations in species distribution and echinocandin and azole antifungal resistance rates among *Candida* bloodstream infection isolates: report from the SENTRY Antimicrobial Surveillance Program (2008 to 2009). *J Clin Microbiol.* 2011;49:396–9.
65. Razzaghi-Abyaneh M, Sadeghi G, Zeinali E, Alirezaee M, Shams-Ghahfarokhi M, Amani A, et al. Species distribution and antifungal susceptibility of *Candida* spp. isolated from superficial candidiasis in outpatients in Iran. *J Mycol Med.* 2014;24:e43–50.
66. Reyna Figueroa J, Fragozo Díaz A, Ortiz Ibarra FJ, Soriano Becerril D, Bermúdez G, Plazola Camacho N. Epidemiología hospitalaria de candidiasis neonatal en el Instituto Nacional de Perinatología en un período de cinco años. *Enf Inf Microbiol.* 2007;27:110–3.
67. Rivera-Sánchez R, Delgado-Ochoa D, Flores-Paz RR, García-Jiménez EE, Espinosa-Hernández R, Bazan-Borges AA, et al. Prospective study of urinary tract infection surveillance after kidney transplantation. *BMC Infect Dis.* 2010;10:245.
68. Robles JC, Koreen L, Park S, Perlin DS. Multilocus sequence typing is a reliable alternative method to DNA fingerprinting for discrimination among strains of *Candida albicans*. *J Clin Microbiol.* 2004;42:2480–8.
69. Romero-Luévano AG, Araiza-Santibáñez J, Hernández MA, Cerón-Araiza M, Hernández-Guzmán VA, Ponce RM, et al. Candidosis mixtas en aislamientos clínicos de pacientes procedentes del Hospital General de México Dr Eduardo Liceaga; identificación e importancia. *Dermatol Rev Mex.* 2014;58:239–46.
70. Rueda-Gordillo F, Hernández-Solís SE. Prevalencia de *Candida albicans* aislada de la cavidad oral de pacientes con cáncer. *Rev Odontol Latinoam.* 2008;1: 38–41.
71. Sabino R, Sampaio P, Rosado L, Stevens DA, Clemons KV, Pais C. New polymorphic microsatellite markers able to distinguish among *Candida parapsilosis sensu stricto* isolates. *J Clin Microbiol.* 2010;48:1677–82.
72. Sánchez-Hernández JA, Rivera-Tapia JA, Coyotécatl-García LL, Mendoza-López E. Incidencia de *Candida albicans* en pacientes estudiadas en la ciudad de Puebla México. *Acta Cient Estud.* 2009;7:191–5.
73. Sánchez-Vargas LO, Eraso E, Carrillo-Muñoz AJ, Aguirre JM, Gaitán-Cepeda LA, Quindós G. *In vitro* activity of voriconazole against Mexican oral yeast isolates. *Mycoses.* 2010;53:200–3.
74. Sánchez-Vargas LO, Ortiz-López NG, Villar M, Moragues MD, Aguirre JM, Cashat-Cruz M, et al. Point prevalence, microbiology and antifungal susceptibility patterns of oral *Candida* isolates colonizing or infecting Mexican HIV/AIDS patients and healthy persons. *Rev Iberoam Micol.* 2005;22:83–92.
75. Sardi JCO, Scorzoni L, Bernardi T, Fusco-Almeida AM, Mendes Giannini MJS. *Candida* species: current epidemiology, pathogenicity, biofilm formation, natural antifungal products and new therapeutic options. *J Med Microbiol.* 2013;62:10–24.
76. Shan Y, Fan S, Liu X, Li J. Prevalence of *Candida albicans*-closely related yeasts, *Candida africana* and *Candida dubliniensis*, in vulvovaginal candidiasis. *Med Mycol.* 2014;52:636–40.
77. Solís-Arias MP, Moreno Morales M, Dávalos-Tanaka M, Fernández Martínez R, Díaz Flores O, Arenas-Guzmán R. Colonización vaginal por *Candida* spp. Fre-  
cuencia y descripción de las especies aisladas en mujeres asintomáticas. *Ginecol Obstet Mex.* 2014;82:1–8.
78. Souza LK, Fernandes OF, Passos XS, Costa CR, Lemos JA, Silva MR. Epidemiological and mycological data of onychomycosis in Goiania, Brazil. *Mycoses.* 2010;53:68–71.
79. Tobar E, Silva F, Olivares R, Gaete P, Luppi M. Candidiasis invasoras en el paciente crítico adulto. *Rev Chil Infect.* 2011;28:41–9.
80. Trtkova J, Pavlicek P, Ruskova L, Hamal P, Koukalova D, Raclavsky V. Performance of optimized McRAPD in identification of 9 yeast species frequently isolated from patient samples: potential for automation. *BMC Microbiol.* 2009;9:234.
81. Xu J, Mitchell TG. Geographical differences in human oral yeast flora. *Clin Infect Dis.* 2002;36:221–4.